

Am Chem Soc. Author manuscript; available in PMC 2009 October 22.

Published in final edited form as:

J Am Chem Soc. 2008 October 22; 130(42): 13820–13821. doi:10.1021/ja802683u.

Bcl-X_L-templated assembly of its own protein-protein interaction modulator from fragments decorated with thio acids and sulfonyl azides

Xiangdong Hu[†], Jiazhi Sun[‡], Hong-Gang Wang[‡], and Roman Manetsch^{*,†}

†Department of Chemistry, University of South Florida, CHE205, 4202 East Fowler Avenue, Tampa, Florida 33620

‡H. Lee Moffitt Cancer Center and Research Institute, 12902 Magnolia Drive, Tampa, Florida 33612

Abstract

Protein-protein interactions have key importance in various biological processes and modulation of particular protein-protein interactions has been shown to have therapeutic effects. However, disrupting or modulating protein-protein interactions with low-molecular-weight compounds is extremely difficult due to the lack of deep binding pockets on protein surfaces. Herein we describe the development of an unprecedented lead synthesis and discovery method that generates only biologically active compounds from a library of reactive fragments. Using the protein $Bcl-X_L$, a central regulator of programmed cell death, we demonstrated that an amidation reaction between thio acids and sulfonyl azides is applicable for $Bcl-X_L$ -templated assembly of inhibitory compounds. We have demonstrated for the first time that kinetic target-guided synthesis can be applied not only on enzymatic targets but also for the discovery of small molecules modulating protein-protein interactions.

Protein-protein interactions are central to many biological processes and hence represent a large and important class of potential targets for human therapeutics. Recent discovery of a variety of low-molecular-weight compounds interfering with protein-protein complexes launches and validates viable routes for a large number of new therapies. However, disrupting or modulating protein-protein interactions with low-molecular-weight compounds remains challenging. Most protein-protein interfaces lack deep pockets that might provide binding sites for small molecules and they are composed of two relatively large protein surfaces that are complementary with respect to shape and electrostatics. Moreover, the adaptive and flexible nature of amino acid residues on protein surfaces creates additional challenges for lead compound design and discovery.

The Bcl-2 family consists of both anti- and pro-apoptotic members, which are central regulators of programmed cell death. The anti-apoptotic proteins like Bcl-2, Bcl- X_L , and Mcl-1 heterodimerize with the pro-apoptotic constituents, which include the multidomain molecules Bax and Bak, and BH3-only proteins such as Bim, Bad, Bid, Noxa or Puma, through the conserved BH3 domain. The relative ratios of pro- and anti-apoptotic Bcl-2 family proteins determine the ultimate sensitivity or resistance of cells to a wide variety of apoptotic signals. Meanwhile, small molecules have been reported to modulate the extent of heterodimerization between anti- and pro-apoptotic Bcl-2 family members inducing apoptosis in cancer cells. Using a combination of SAR by NMR screening, parallel synthesis and structure-guided lead

E-mail: manetsch@cas.usf.edu.

design, the Abbott laboratories developed ABT-737 and a large series of analogues displaying inhibition constants in the nanomolar or subnanomolar range.

Herein, we report our progress toward the development of lead synthesis and discovery methods that generate only biologically active compounds targeting protein-protein interactions. In the last decade, various fragment-based lead discovery approaches have been reported in which the biological target is actively engaged in the assembly of its own multidentate inhibitor from a pool of smaller reactive fragments. These approaches can roughly be divided into three different categories: (1) dynamic combinatorial chemistry $(DCC)^8$, (2) catalyst/reagent-accelerated target-supported assembly⁹, and (3) kinetic target-guided synthesis (TGS)¹⁰. While DCC utilizes reversible reactions to create a library of all possible heterodimeric compounds, kinetic TGS approaches employ irreversible reactions generating only the biologically active bidentate members of the entire virtual library. In situ click chemistry ^{10c}, in particular, has shown to produce potent triazole inhibitors of the enzymes acetylcholine esterase 11, carbonic anhydrase 12, and HIV protease 13. So far, kinetic TGS approaches have been applied only for the discovery of enzyme inhibitors but not for proteinprotein interaction modulators (PPIMs). Based on studies by Abbott laboratories, we investigated whether a kinetic TGS approach is a viable route for the discovery of PPIMs targeting the anti-apoptotic protein Bcl-X_L.

The reaction combining two fragments in the presence of a biological template into a larger molecule plays a crucial role for kinetic TGS. For example, the slow nature and the bioorthogonality of the Huisgen cycloaddition, the compatibility of the reactants with water, and the stability of the triazole products are distinct characteristics of in situ click chemistry. ^{10c} Recently, Williams and co-workers developed an amidation reaction displaying a reactivity profile suitable for kinetic TGS applications. ¹⁴ This particular amidation reaction between thio acids 1 and sulfonyl azides 2 to give corresponding acylsulfonamides 3 is effective at room temperature in both organic and aqueous solvents (Scheme 1-A).

To probe if this amidation reaction is suited for kinetic TGS applications targeting protein-protein interactions, building blocks, of which some are structurally related to fragments of ABT-737, were decorated with sulfonyl azide or thio acid groups (Scheme 1-B). Thio acids **TA1–TA3** and sulfonyl azides **SZ1–SZ6** were incubated as binary mixtures in the presence of buffer solution containing Bcl- X_L for 6 hours at 38°C. As a control, 18 identical binary building block combinations were kept under the same incubation conditions in buffer without Bcl- X_L . All incubations were then analyzed by HPLC with product detection by electrospray ionization in the positive selected ion mode (LC/MS-SIM). ^{11b} From all the screened samples, only one incubation sample led to an increased amount of acylsulfonamide **SZ4TA2** ^{7e} in the Bcl- X_L -containing sample compared to the incubation without the protein. We synthesized this compound for hit validation. ¹⁵ Comparison of the LC/MS-SIM traces of the Bcl- X_L incubation mixture with the trace of the corresponding synthesized **SZ4TA2** clearly confirmed that Bcl- X_L templates the formation of the characterized hit compound (Figure 1. A – C).

To assess whether the Bcl- X_L -templated reactions occur at the BH3 binding pocket or randomly elsewhere on the protein surface, control experiments have been performed, in which the reactive fragments SZ4 and TA2 were incubated with Bcl- X_L and various pro-apoptotic BH3-containing peptides. ¹⁵ Bak BH3 and Bim BH3 peptides bind to Bcl- X_L through their BH3 domain and theoretically compete with the reactive building blocks for binding during these incubations. In comparison, their mutants exhibit lower affinity towards Bcl- X_L and hence do not suppress the Bcl- X_L -templated acylsulfonamide formation to the same extent. Comparison of the LC/MS-SIM traces (Figure 1. D–G) between the Bcl- X_L incubations with and without these peptides suggests that the generation of SZ4TA2 occurs at the BH3 binding site on Bcl- X_L .

An advantage of kinetic TGS approaches is the increased throughput screening capability with incubations containing more than two reactive building blocks. Experiments were undertaken to test whether our amidation kinetic TGS screening can be performed as incubations containing more than two complimentary reacting building blocks. Best results were obtained with reactions containing one thio acid and six sulfonyl azides. ¹⁵ Samples of Bcl- X_L with all 9 reactive building blocks at the same time failed at giving clear results. Although the multicomponent screening of the entire library failed, these experiments prove that the amidation kinetic TGS approach can be tested in an enhanced screening throughput.

To investigate the ability of the hit compound to disrupt the interaction between Bcl- X_L and Bak, we performed the well-established fluorescence polarization competition assay using Bcl- X_L and fluorescein-labeled Bak BH3 peptide. ¹⁶ Abbott laboratories reported that **SZ4TA2** is a good Bcl- X_L PPIM with a Ki constant of 19 nM, as determined by a competitive fluorescence polarization assay using a fluorescein-labeled Bad-BH3 peptide. ^{7e} Consistently, compound **SZ4TA2** is validated again as a Bcl- X_L inhibitor with an IC₅₀ constant of 78.8 nM by our assay. ¹⁵ To compare the activity of acylsulfonamides, not assembled by Bcl- X_L , with that of the hit compound **SZ4TA2**, compounds **SZ2TA1**, **SZ2TA2**, **SZ2TA3**, **SZ4TA1**, **SZ5TA1** and **SZ5TA2** have been synthesized and tested for their binding to Bcl- X_L . The IC₅₀ values of these compounds have been determined to be 5 μ M or higher. ¹⁵ Finally, we also determined the IC₅₀ constants for the corresponding reactive building blocks **SZ4** and **TA2** to be higher than 100 μ M. Taken together, these results indicate that the hit compound **SZ4TA2** identified through the kinetic TGS screening is indeed a respectable ligand of the biological target, which underscores the utility of kinetic TGS as a valuable approach to PPIM discovery and optimization.

In a proof-of-concept study, we have shown for the first time that kinetic TGS can be applied not only for enzymatic targets but also for protein-protein interaction disruption by using the recently-reported amidation reaction between sulfonyl azides and thio acids. In the future, we will study in great detail the kinetics and mechanism of the kinetic TGS approach. Additionally, we will investigate the scope and limitations of the herein reported lead discovery and optimization method targeting other protein-protein interactions related to various diseases.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgment

We are grateful to the James and Esther King Biomedical Research Program (NIR Grant 07KN-08 to R.M.) and the National Cancer Institute, National Institutes of Health (Grant P01CA118210 to H-G.W.) for financial support.

References

- 1. (a) Arkin MR, Wells JA. Nat. Rev. Drug Discov 2004;3:301–317. [PubMed: 15060526] (b) Berg T. Angew. Chem. Int. Ed. Engl 2003;42:2462–2481. [PubMed: 12800163]
- (a) Clackson T, Wells JA. Science 1995;267:383–386. [PubMed: 7529940] (b) Arkin M. Curr. Opin. Chem. Biol 2005;9:317–324. [PubMed: 15939335] (c) Bogan AA, Thorn KS. J. Mol. Biol 1998;280:1–9. [PubMed: 9653027] (d) DeLano WL. Curr. Opin. Struct. Biol 2002;12:14–20. [PubMed: 11839484]
- (a) Preissner R, Goede A, Frommel C. J. Mol. Biol 1998;280:535–550. [PubMed: 9665855] (b) Mccoy AJ, Epa VC, Colman PM. J. Mol. Biol 1997;268:570–584. [PubMed: 9159491] (c) Lo Conte L, Chothia C, Janin J. J. Mol. Biol 1999;285:2177–2198. [PubMed: 9925793] (d) Nooren IMA, Thornton JM. J. Mol. Biol 2003;325:991–1018. [PubMed: 12527304]
- 4. (a) DeLano WL, Ultsch MH, de Vos AM, Wells JA. Science 2000;287:1279–1283. [PubMed: 10678837] (b) Wodak SJ, Janin J. Protein Modules and Protein-Protein Interactions 2003;61:9–73.

- 5. Walensky LD. Cell Death Differ 2006;13:1339–1350. [PubMed: 16763614]
- (a) Danial NN, Korsmeyer SJ. Cell 2004;116:205–219. [PubMed: 14744432] (b) Huang DCS, Strasser
 A. Cell 2000;103:839–842. [PubMed: 11136969] (c) Kelekar A, Thompson CB. Trends Cell Biol 1998;8:324–330. [PubMed: 9704409]
- (a) Oltersdorf T, et al. Nature 2005;435:677–681. [PubMed: 15902208] (b) Wendt MD, et al. J. Med. Chem 2006;49:1165–1181. [PubMed: 16451081] (c) Park CM, Oie T, Petros AM, Zhang H, Nimmer PM, Henry RF, Elmore SW. J. Am. Chem. Soc 2006;128:16206–16212. [PubMed: 17165773] (d) Petros AM, et al. J. Med. Chem 2006;49:656–663. [PubMed: 16420051] (e) Wendt MD, et al. J. Med. Chem 2006;49:1165–1181. [PubMed: 16451081] (f) Bruncko M, et al. J. Med. Chem 2007;50:641–662. [PubMed: 17256834] (g) Tse C, et al. Cancer Res 2008;68:3421–3428. [PubMed: 18451170]
- 8. (a) Huc I, Lehn J-M. Proc. Natl. Acad. Sci. U.S.A 1997;94:2106–2110. [PubMed: 9122156] (b) Corbett PT, Leclaire J, Vial L, West KR, Wietor J-L, Sanders JKM, Otto S. Chem. Rev 2006;106:3652–3711. [PubMed: 16967917]
- (a) Nicolaou KC, Hughes R, Cho SY, Winssinger N, Smethurst C, Labischinski H, Endermann R. Angew. Chem. Int. Ed. Engl 2000;39:3823–3828.
 (b) Nicolaou KC, Hughes R, Cho SY, Winssinger N, Labischinski H, Endermann R. Chem.-Eur. J 2001;7:3824–3843.
- (a) Inglese J, Benkovic SJ. Tetrahedron 19991;47:2351–2364.
 (b) Nguyen R. Huc. Angew. Chem. Int. Ed. Engl 2001;40:1774–1776.
 (c) Sharpless KB, Manetsch R. Expert Opinion on Drug Discovery 2006;1:525–538.
- 11. (a) Lewis WG, Green LG, Grynszpan F, Radic Z, Carlier PR, Taylor P, Finn MG, Sharpless KB. Angew. Chem. Int. Ed. Engl 2002;41:1053–1057. [PubMed: 12491310] (b) Manetsch R, Krasinski A, Radic Z, Raushel J, Taylor P, Sharpless KB, Kolb HC. J. Am. Chem. Soc 2004;126:12809–12818. [PubMed: 15469276] (c) Krasinski A, Radic Z, Manetsch R, Raushel J, Taylor P, Sharpless KB, Kolb HC. J. Am. Chem. Soc 2005;127:6686–6692. [PubMed: 15869290]
- (a) Mocharla VP, Colasson B, Lee LV, Roeper S, Sharpless KB, Wong C-H, Kolb HC. Angew. Chem. Int. Ed. Engl 2005;44:116–120. [PubMed: 15599912] (b) Wang J, Sui G, Mocharla VP, Lin RJ, Phelps ME, Kolb HC, Tseng H. Angew. Chem. Int. Ed. Engl 2006;45:5276–5281. [PubMed: 16927322]
- Whiting M, Muldoon J, Lin Y-C, Silverman SM, Lindstrom W, Olson AJ, Kolb HC, Finn MG, Sharpless KB, Elder JH, Fokin VV. Angew. Chem. Int. Ed. Engl 2006;45:1435–1439. [PubMed: 16425339]
- 14. (a) Shuangguan N, Katukojvala S, Greenberg R, William LJ. J. Am. Chem. Soc 2003;125:7754–7755. [PubMed: 12822965] (b) Kolakowski RV, Shuangguan N, Sauers RR, Greenberg R, William LJ. J. Am. Chem. Soc 2006;128:5695–5702. [PubMed: 16637636]
- 15. Please see supporting information for details.
- 16. Yin H, Lee G, Sedey KA, Rodriguez JM, Wang H-G, Sebti SM, Hamilton AD. J. Am. Chem. Soc 2005;127:5463–5468. [PubMed: 15826183]

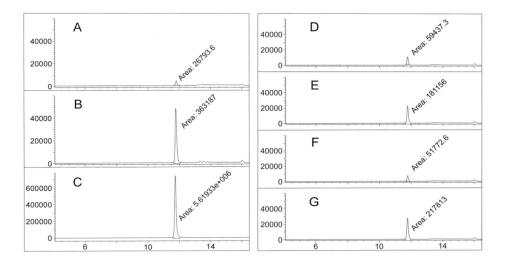


Figure 1. Hit identification of acylsulfonamide SZ4TA2 by LC/MS-SIM. (A) Incubation of SZ4 and TA2 in buffer without Bcl-X_L. (B) Bcl-X_L-templated reaction after 6 hours of incubation. (C) Synthesized SZ4TA2 as reference. (D) Suppression of Bcl-X_L-templated reaction by Bak BH3 peptide. (E) Bcl-X_L-templated incubation in presence of mutant Bak BH3 peptide. (F) Suppression of Bcl-X_L-templated reaction by Bim BH3 peptide. (G) Bcl-X_L-templated incubation in presence of mutant Bim BH3 peptide.

A) Amidation reaction

B) Bcl-X_L-templated screening

Scheme 1.

 $Bcl-X_L$ -templated assembly of acylsulfonamide ${\bf SZ4TA2}$ from fragments decorated with thio acid or sulfonyl azide functionalities.

SZ4TA2