



Published in final edited form as:

Neuroscience. 2008 March 27; 152(3): 753–760. doi:10.1016/j.neuroscience.2008.01.022.

Modulation of The Balance Between Cannabinoid CB1 and CB2 Receptor Activation During Cerebral Ischemic/Reperfusion Injury

Ming Zhang, MD, PhD,

1 Center for Substance Abuse Research, Temple University School of Medicine, 208 MRB, 3420 N Broad Street, Philadelphia, PA 19140

2 Department of Physiology, Temple University School of Medicine, 208 MRB, 3420 N Broad Street, Philadelphia, PA 19140; Fax 2157074003; Phone 2157073398; Email mzhang@temple.edu

Billy R Martin, PhD,

Department of Pharmacology and Toxicology, Virginia Commonwealth University School of Medicine, 410 North 12th Street, Richmond, VA 23298; Fax 8048282117; Phone: 8048088407; Email martinb@vcu.edu

Martin W. Adler, PhD,

Center for Substance Abuse Research, Temple University School of Medicine, 305 OMS, 3400 N Broad Street, Philadelphia, PA 19140; Fax 2157071904; Phone 2157073243; Email baldeagl@temple.edu

Raj K. Razdan, PhD,

Organix Inc, 240 Salem Street, Woburn, MA 01801; Fax 7819336695; Phone 7819324142; Email: razdanrk@aol.com

Doina Ganea, PhD, and

1 Center for Substance Abuse Research, Temple University School of Medicine, 709 MRB, 3420 N Broad Street, Philadelphia, PA 19140

2 Department of Microbiology and Immunology, Temple University School of Medicine, 709 MRB, 3420 N Broad Street, Philadelphia, PA 19140; Fax 2157074003; Phone 2157079921; Email: dganea@temple.edu

Ronald F Tuma, PhD

1 Center for Substance Abuse Research, Temple University School of Medicine, 231 OMS, 3400 N Broad Street, Philadelphia, PA 19140

2 Department of Physiology, Temple University School of Medicine, 231 OMS, 3400 N Broad Street, Philadelphia, PA 19140; Fax 2157074003; Phone 2157075485; Email tumarf@temple.edu (corresponding author)

Abstract

Cannabinoid receptor activation has been shown to modulate both neurotransmission (CB₁) and neuroinflammatory (CB₂) responses. There are conflicting reports in the literature describing the influence of cannabinoid receptor activation on ischemic/reperfusion injury. The goal of this study

Section Editor: Dr. Yoland Smith (Neuropharmacology) Yerkes National Primate Research Center, Emory University, 954 Gatewood Road NE, Atlanta, GA 30329, USA

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

was to evaluate how changing the balance between CB₁ and CB₂ activation following cerebral ischemia influences outcome. CB₁ and CB₂ expression were tested at different times after transient middle cerebral artery occlusion (MCAO) in mice by real-time RT-PCR. Animals subjected to 1 hour MCAO were randomly assigned to receive different treatments: a CB₁ antagonist, a CB₂ antagonist, a CB₂ agonist, a CB₁ antagonist plus CB₂ agonist, a CB₂ antagonist plus CB₂ agonist or an equal volume of vehicle as control. Cerebral blood flow was continuously monitored during ischemia; cerebral infarction and neurological deficit were tested 24 hours after MCAO. Cerebral CB₁ and CB₂ mRNA expression undertook dynamic changes during cerebral ischemia. The selective CB₁ antagonist significantly decreased cerebral infarction by 47%; the selective CB₂ antagonist increased infarction by 26% after 1 hour MCAO followed by 23 hours reperfusion in mice. The most striking changes were obtained by combining a CB₁ antagonist with a CB₂ agonist. This combination elevated the cerebral blood flow during ischemia and reduced infarction by 75%. In conclusion, during cerebral ischemia/reperfusion injury, inhibition of CB₁ receptor activation is protective while inhibition of CB₂ receptor activation is detrimental. The greatest degree of neuroprotection was obtained by combining an inhibitor of CB₁ activation with an exogenous CB₂ agonist.

Keywords

endogenous cannabinoids; cerebral ischemia/reperfusion injury; cerebral blood flow; inflammatory responses

INTRODUCTION

The endocannabinoid system refers to two major types of cannabinoid receptors (termed CB₁ and CB₂), the endogenous ligands for those receptors and specific enzymes responsible for their degradation and inactivation (Rodriguez de Fonseca et al., 2005). The CB₁ receptor is primarily expressed in the central nervous system (CNS), exhibiting a presynaptic location and playing a prominent role in synaptic neurotransmission (Pazos et al., 2005, Rodriguez de Fonseca et al., 2005). The CB₂ receptor is expressed predominantly by cells of the immune system, such as lymphocytes and neutrophils, but is also expressed on resident inflammatory cells within the CNS. CB₂ stimulation has been shown to have immunomodulatory properties, such as decreasing the activity of antigen presenting cells (APC) and down-regulating cytokine (IFN- γ and TNF- α) production during inflammatory responses (Berdyshev, 2000, Walter and Stella, 2004, Klein and Cabral, 2006, Lombard et al., 2007).

A number of investigations have shown that CB₂ receptor activation has anti-inflammatory therapeutic potential in various CNS diseases, such as multiple sclerosis, traumatic brain injury and Alzheimer's disease (Grundy et al., 2001, Molina-Holgado et al., 2002, Croxford, 2003, Ni et al., 2004, Ramirez et al., 2005). Because inflammatory responses have been shown to be important contributors to secondary injury following cerebral ischemia; the CB₂ receptor has been investigated as a potential therapeutic target in stroke. It was demonstrated that selective activation of CB₂ receptor attenuated cerebral ischemia/reperfusion injury in mice which was associated with decreased leukocyte/endothelial cell interactions (Zhang et al., 2007). The CB₁ receptor has also been studied in cerebral ischemia/reperfusion injury (Nagayama et al., 1999, Jin et al., 2000, Parmentier-Batteur et al., 2002, Hayakawa et al., 2004). However, to date there are few studies focusing on the roles of CB₁ and CB₂ activation by endogenous cannabinoids in cerebral ischemic injury. In this investigation, we evaluated how ischemia/reperfusion injury influences cannabinoid receptor expression and how modification of the balance between CB₁R and CB₂R activation by endogenous and exogenous cannabinoids influences outcome after stroke.

EXPERIMENTAL PROCEDURES

Animals and surgical procedures

The cerebral ischemia/reperfusion studies were carried out in 8 week old male C57BL/6 mice (weighing 23 to 27g; Taconic, Hudson NY) and conducted in accordance with the guidelines approved by the Institutional Animal Care and Use Committee at Temple University.

Middle Cerebral Artery Occlusion and Reperfusion (MCAO/R)

The animals were anesthetized by intraperitoneal injection of a mixture of Ketamine (100mg/ml) - Xylazine (20mg/kg) (1:1) at a dose of 1ml/kg. Body and cerebral temperature were maintained at $37\pm 5^{\circ}\text{C}$ by a heating lamp and heating pad. Middle cerebral artery occlusion was achieved by the intraluminal filament method (Hata et al., 1998). Briefly, a midline neck incision was made under an operating microscope. The right common carotid artery (CCA), external carotid artery (ECA) and internal carotid artery (ICA) were isolated. The ECA was ligated with 6-0 silk suture distal from the ICA-ECA branch and then cut distal from the ligated point. Another 6-0 silk suture was tied loosely around ECA close to the origin at the CCA. A blunted 5-0 monofilament nylon suture coated with poly-L-lysine (0.1% in deionized water, Sigma Inc, St Louis MO, USA) (Belayev et al., 1999) was introduced through a small incision in ECA and advanced into the circle of Willis, finally occluding the origin of the middle cerebral artery. The silk suture around the ECA stump was tied tightly to prevent bleeding and secured with a silk suture. The nylon suture was removed after 60 minutes occlusion and the ECA was permanently tied. Reperfusion was confirmed when pulsations were again observed in ICA.

Regional Cerebral Blood Flow (rCBF)

A laserPro Blood Perfusion Monitor (TSI Inc, Shoreview, MN, USA) was used to monitor and record regional cerebral blood flow (rCBF) prior to ischemia, during MCAO and reperfusion. A 1mm diameter microfiber laser-Doppler probe was attached to the skull 4mm lateral and 2mm posterior of Bregma. The MCAO was considered adequate if rCBF showed a sharp drop to 25% of baseline (pre-ischemia) level, otherwise, animals were excluded (Tsuchiya et al., 2003).

Injection of cannabinoid receptor agonist and antagonists in MCAO/R

The CB₁ antagonist (SR141716) and CB₂ antagonist (SR144528) were dissolved in a DMSO: cremophor: saline mixed solution (1:1:18). The antagonists (5mg/kg or 20mg/kg) or an equal volume of vehicle were administered 1 hour before MCAO *i.p.* The CB₂ agonist (O-1966) was dissolved in a pure ethanol: emulphor: saline mixed solution at 1:1:18. The CB₂ agonists (1mg/kg) or an equal volume of vehicle were administered as an intravenous injection into the jugular vein or intraperitoneally in a separate experimental group 1 hour before MCAO.

Real time PT-PCR

CB1 and CB2 expression were detected by the SYBR green-based real time RT-PCR technique. Animals were euthanized and transcardially perfused with cold PBS to remove the blood from vessels. Total RNA was isolated from brain specimens at 1, 3, 6 or 24 hours after MCAO by using Ultraspec reagent (Biotecx Laboratories, Huston TX, USA). Normal brain was used as sham. cDNA was prepared by reverse transcription. The 20 μl (total volume) of the PCR mixture consists of 4 μl diluted cDNA, 10 μl SYBR green-containing PCR master mixture (2X) and 150 μM of each primer. The CB1 and CB2 primers for real-time RT-PCR were designed by using the Primer Express software from Applied Biosystems (Fostercity, CA), and are as follows: CB₁ sense: 5'-TGA AGT CGA TCT TAG ACG GCC-3' and antisense: 5'GTG GTG ATG GTA CGG AAG GTA-3'; CB₂ sense: 5'-TGA ATG AGC AGA CCG ACA GG-3' and antisense: 5'-AGA GAT GTT TGC TGG GTG GC-3'; β -actin sense: 5'-TCC ACC ACC ACA

GCT GAG AGG-3', and antisense: 5'-CAG CTT CTC TTT GAT GTC ACG-3' Real Time RT-PCR was performed using the Stratagene Mx3005P, and the cycling conditions used were 95°C for 15 sec, 60°C for 1 min, for 40 cycles, followed by a melting point determination or dissociation curves. The expression level of each gene is indicated by the cycle numbers needed for the cDNA to be amplified to reach a threshold. The amount of DNA is calculated from the cycle numbers by using standard curves and the results are normalized to the housekeeping gene β -actin from the same sample.

Infarct Volume Assessment

The animals were euthanized with an overdose of pentobarbital (200mg/kg *i.p.*) 24 hours after MCAO and the brains were removed, and chilled on ice for 10 minutes to slightly harden the tissue. Five 2mm coronal sections were cut using a mouse brain matrix (Zivic lab, Pittsburg, PA, USA). The brain sections were placed in 2% triphenyltetrazolium chloride (TTC) (Sigma Inc, St Louis, MO, USA) dissolved in saline and stained for 20 minutes at 37°C in the dark. The brain sections were then fixed in 4% paraformaldehyde at 4°C for 24 hours and the anterior and caudal face of each section was scanned by a flatbed color scanner (Microtek Inc, Carson, CA, USA). The resulting images were captured as JPEG files and analyzed with NIH image software. The hemispheric infarct volumes were corrected for brain edema/swelling: the hemispheric infarct volume in each section was calculated by subtracting the area of normal, TTC stained tissue in the hemisphere ipsilateral to the ligation from the contralateral nonischemic area to generate the infarct fraction (%), as described by Swanson *et al.* and Lin *et al.* (Swanson *et al.*, 1990, Lin *et al.*, 1993).

Neurological Function Evaluation

The severity of neurological deficits was evaluated 24 hours after ischemic insult using a five-point deficit score (0=normal motor function; 1=flexion of torso and of contralateral forelimb upon lifting of the animal by tail; 2=circling to the contralateral side but normal posture at rest; 3=leaning to contralateral side at rest; and 4=no spontaneous motor activity) (Hata *et al.*, 1998).

Statistical analysis

Bonferroni's test after one way ANOVA was used for analyzing differences in infarct volume, neurological score and average of rCBF. The mRNA expression of CB1 and CB2 were analyzed by two way ANOVA (times, hemispheres) followed by Bonferroni's test. Data were presented as means \pm SEM. A statistically significant difference was assumed at $P<0.05$.

RESULTS

CB₁ and CB₂ mRNA expression in brain during MCAO

There were no differences in CB₁ mRNA expression in the non-ischemic hemisphere compared to normal control at 1, 3, 6 and 24 hours after MCAO. CB₁ expression in the ischemic hemisphere increased at 1 hour after ischemia and was maximal at 6 hours. Similarly there were no significant differences in CB₂ mRNA expression in the non-ischemic hemispheres compared to control. However, CB₂ expression in the ischemic hemispheres decreased during first 3 hours following ischemia, followed by a gradual increase until the 24 hour measurement time (Figure 1).

Effects of CB antagonists and agonist on cerebral blood flow during Occlusion

During MCAO, rCBF decreased to approximately 25% of baseline value. Combined administration of the CB₁ antagonist (SR141716, 20mg/kg, *i.p.*) and CB₂ agonist (O-1966, 1mg/kg, *i.v.*) 1 hour prior to occlusion increased rCBF during the occlusion period when

compared with the vehicle-treated group, whereas other treatments failed to alter rCBF during MCAO (Figure 2A). Lowering the dose of the CB₁ and CB₂ antagonist did not alter their influence on blood flow changes during occlusion; combination of CB₂ agonist (*i.p.* injection) and low dose CB₁ antagonist (5mg/kg) also increased rCBF (Figure 2B)

Effects of CB antagonists and agonist on cerebral infarction

Administration of the CB₁ antagonist (SR141716, 20mg/kg, *i.p.*) 1 hour before MCAO significantly decreased cerebral infarct fraction (12.3±0.86%) while administration of the CB₂ antagonist (SR144528, 20mg/kg *i.p.*) alone increased cerebral infarct volume (31.1 ±2.45%) compared with vehicle-treated group (24.7±1.16%) (Figure 3A). Administration of a CB₂ agonist (O-1966, 1mg/kg, *i.v.*) alone decreased cerebral infarction (15.6±1.67%) compared with vehicle treated group (22.4±1.08%), and the protection was totally reversed by co-administration of a CB₂ antagonist (23.4±3.2%). Administration of the CB₁ antagonist plus CB₂ agonist 1 hour before MCAO further dramatically reduced infarct volume (6.1 ±2.33%) (Figure 4B). Reducing the dose of the antagonists to 5mg/kg and administration of the CB₂ agonist at same dose by *i.p.* did not change the results obtained (Figure 3C).

Effects on neurological function

In parallel with the changes in infarct size, administration of a CB₁ antagonist (20mg/kg *i.p.*) improved motor function significantly, while administrations of a CB₂ antagonist (20mg/kg *i.p.*) tended to worsen motor function, although this value did not reach statistical significance (Figure 4A). Animals treated with either CB₂ agonist (1mg/kg, *i.v.*) alone or the combined CB₁ antagonist and CB₂ agonist had significantly improved motor function compared to the vehicle treated animals (Figure 4B).

DISCUSSION

The primary goal of this study was to investigate whether modification of the endocannabinoid system could influence outcome following cerebral ischemia/reperfusion injury. The hypothesis that modification of the endocannabinoid system could influence outcome following ischemia was based upon prior reports that this system can have direct effects on neuronal function and can also modify inflammatory responses (Baker et al., 2001, van der Stelt et al., 2001, Muthian et al., 2004, McCollum et al., 2007). Both of these actions could have significant impact on cerebral ischemia/reperfusion injury.

Endogenous cannabinoids, are derivatives of arachidonic acid, and serve as natural ligands for the cannabinoid receptors with similar pharmacological properties to those of plant derived cannabinoids and synthetic analogs. 2-arachidonoylglycerol (2-AG) and anandamide are two major endogenous cannabinoids and are produced in relatively high concentrations in the CNS (Pazos et al., 2005, Rodriguez de Fonseca et al., 2005). Neuromodulatory properties of endogenous cannabinoids have been investigated in animal models of multiple sclerosis and brain trauma as well as in vitro studies (Baker et al., 2001, Panikashvili et al., 2001). It has previously been reported that the production of certain endogenous cannabinoids such as anandamide was increased in brain following ischemia (Muthian et al., 2004). There are also reports showing that the expression of the cannabinoid receptors were up-regulated in rat brain after cerebral ischemia (Jin et al., 2000, Ashton et al., 2007), but the changes of cerebral cannabinoid receptors expression in a mouse model of ischemic injury have not been previously examined. We found that CB₁ receptor mRNA expression increased following ischemia and reached at peak 6 hours post ischemia in the ischemic hemisphere, while the non ischemic hemisphere CB₁ mRNA content remained unchanged. Interestingly, cerebral CB₂ mRNA content decreased over the first 3 hours after MCAO in the ischemic hemisphere, but increased after that. The difference in the time course of expression of these two receptors could, at least

in part be due to changes in the cell types that express them. The delayed increase in CB₂ receptor expression could reflect the time required for immune cell invasion of the CNS following ischemia (Heinel et al., 1994, Tomita and Fukuuchi, 1996). Microglia expression of CB₂ receptors may also contribute (Nunez et al., 2004, Maresz et al., 2005).

Previous investigations performed in our laboratory have demonstrated the neuroprotective effects of administration of an exogenous CB₂ ligand in a mouse model of cerebral ischemia/reperfusion injury (Zhang et al., 2007). Prior to this investigation, most studies have focused on the activation of the CB₁ receptor rather than the activation of the CB₂ receptor. WIN55212-2, which stimulates both the CB₁ and CB₂ receptor, with greater affinity for the CB₂ receptor, has been shown to be neuroprotective in both global and focal models of ischemia. Based upon the use of WIN55212-2 in combination with a CB₁ antagonist, these effects were interpreted to be the result of CB₁ receptor activation (Nagayama et al., 1999). A later study indicated that the protective effect derived from stimulation of the CB₁ receptor was the result of induced hypothermia, and was eliminated when temperature was maintained at the normal level (Hayakawa et al., 2004). Another study implicated a protective effect for the CB₁ receptor by utilizing CB₁ knockout mice which shown an increase in infarct size compared to wild type animals (Parmentier-Batteur et al., 2002). When interpreting these results it must be recognized that embryonic deletion of CB₁ receptor may lead to abnormal CNS development, making these animals more susceptible to ischemia and may not reflect the acute contributions of the CB₁ receptor in attenuating ischemic damage. In a separate investigation, and in agreement with the results presented in our study, other investigators have reported that the CB₁ receptor antagonist SR141716 was found to reduce infarct volume in a rat MCAO model (Muthian et al., 2004).

Importantly, all of the investigations previously described have involved manipulation of either the CB₁ or CB₂ receptor alone. The current study examined the effects of changing the activity of both receptors simultaneously through either endogenous or exogenous cannabinoids. The results of this study showed that that the inhibition of CB₁ receptor and activation of the CB₂ receptor are both neuroprotective. Animals treated with either a CB₁ antagonist alone or CB₁ and CB₂ antagonist together had smaller infarct volume compared to control animals, while CB₂ antagonist treatment resulted in a larger infarct volume. This would indicate that the two major cannabinoid receptors, CB₁ and CB₂, play different roles in lesion formation during cerebral ischemia/reperfusion injury. These findings are consistent with results from our previous study showing that exogenous selective CB₂ agonists reduced infarct volume and improved motor function in a mouse cerebral ischemia/reperfusion model (Zhang et al., 2007). Since the CB₂ receptor is important in signaling immune cells as well as in the modulation of inflammatory responses, it is likely that CB₂ activation exerts its protective mechanism at least in part through attenuation of inflammatory responses after stroke. Inflammation has been shown to be an important contributor to damage to the brain following ischemia/reperfusion injury (White et al., 2000, Danton and Dietrich, 2003). Within minutes of ischemia, cerebral vascular endothelium is activated and leukocytes begin to roll and adhere on inflamed endothelial cells, followed by transmigration into brain tissue (Kishimoto and Rothlein, 1994). Neutrophils are the first leukocytes to infiltrate at the site of inflammation and monocytes are subsequently recruited. Leukocytes activation and migration have been implicated as primary contributors to ischemia/reperfusion injury (Vasthare et al., 1990). In addition to their role in physical obstruction of capillaries, they participate in inflammatory responses and cause brain tissue damage by various mechanisms. For example, pro-inflammatory cytokines (TNF- α and IL-1 β) secreted by leukocytes not only activate vascular endothelial cells and amplify inflammatory response but also directly induce neuronal injury (Wood, 2003). These studies highlight the involvement of immune cells and inflammatory cytokines in exacerbating ischemic injury, and numerous studies have shown that protection

could be offered by interfering the inflammatory responses following ischemic/reperfusion injury (Kanemoto et al., 2002, Weaver et al., 2002, Sughrue et al., 2004).

CB₂ is a G_i protein coupled-receptor and its activation triggers a series of signal transduction pathways which eventually leads to either up-or down-regulation of gene transcription. In most cases, the genes involved are coded for pro-inflammatory cytokines (Klein et al., 2001) and CB₂ activation has been shown to inhibit some pro-inflammatory cytokines such as TNF- α and IL-6 in *in vivo* and *in vitro* studies (Klein and Cabral, 2006). We have shown in our previous experiments that CB₂ activation is protective in cerebral ischemic injury as well as CNS demyelinating diseases, and this protection is associated with attenuated leukocyte/endothelial interactions in brain (Ni et al., 2004, Zhang et al., 2007). Whether this attenuation is mediated through leukocytes, endothelial cells, or both is still under investigation.

Our most striking result was the finding that simultaneous administration of a CB₁ antagonist and a CB₂ agonist exerted the strongest effect in reducing the cerebral infarction following ischemic injury, which was associated with improved regional cerebral blood flow during ischemia. This might indicated a separate mechanism of action for the agonist and antagonist, which appear to have a synergistic neuroprotective effect. Although neither a CB₁ antagonist alone nor a CB₂ agonist alone had an effect on rCBF during occlusion, the combination significantly improved cerebral blood flow during ischemia. The underlying mechanisms contributing to this phenomenon are still under investigation. There are however, a number of potential targets that may be responsible. It has been reported that endogenous cannabinoids such as 2-AG and anandamide may induce vasodilation through a non-cannabinoid receptor (such as vanilloid receptor) present on endothelial cells (Golech et al., 2004, McCollum et al., 2007). It has been shown that cannabidiol, the nonpsychoactive constituent of cannabis, also reduced cerebral infarction due to increases in rCBF during ischemia, through activation of the serotonergic 5-hydroxytryptamine_{1A} receptor (Mishima et al., 2005). An additional mechanism could be through changes in the rheological contribution of leukocytes during the ischemic period. During periods of reduced cerebral perfusion pressure in stroke, activated leukocytes exaggerate microcirculatory dysfunction by direct occlusion of microvessels or releasing vasoactive factors such as thromboxane A₂ causing platelets aggregation and increasing microvascular procoagulant activity (Ritter et al., 2000, Ishikawa et al., 2004), which further decrease perfusion in the ischemic brain. The increased blood flow in the ischemic region during occlusion in animals treated with the combination of a CB₁ antagonist and a CB₂ agonist could be the result of a reduction in vascular resistance due to an enhanced effect on the inhibition of leukocytes activation through greater CB₂ receptor activation in the presence of the CB₁ antagonist. It is also possible that the elevation in flow to the ischemic region during occlusion is the result of improved collateral blood flow. It has been reported that CB₁ and CB₂ receptors are found on different types of endothelial cells and their activation may participate in the control of vascular resistance (Zoratti et al., 2003, Golech et al., 2004).

In conclusion, the results of this investigation demonstrate dynamic changes in the expression of CB₁ and CB₂ receptors during cerebral ischemic/reperfusion injury in mice. The effects of stimulation of these receptors on damage ischemia/reperfusion injury differed dramatically. Stimulation of the CB₂ receptor was found to be neuroprotective, while inhibition of the CB₁ receptor was also protective, too. The combination of a CB₂ agonist and a CB₁ antagonist provided the greatest degree of protection and indicated a synergistic effect derived from combining these agents. Therefore, changing the balance of stimulation of these receptors by endogenous cannabinoids may provide an important therapeutic strategy during stroke.

Acknowledgements

This project is funded, in part, under a grant with the Pennsylvania Department of Health, a contract from BTG (London) and grants from DA P30 13429, DA 03672 and DA 05488 from the National Institute on Drug Abuse.

List of Abbreviations

CB₁R	cannabinoid CB ₁ receptor
CB₂	cannabinoid CB ₂ receptor
MCAO	middle cerebral artery occlusion
CNS	central nervous system
APC	antigen presenting cell
CCA	common carotid artery
ECA	external carotid artery
ICA	internal carotid artery
rCBF	regional cerebral blood flow
TTC	triphenyltetrazolium chloride
2-AG	2-arachidonoyglycerol

References

- Ashton JC, Rahman RM, Nair SM, Sutherland BA, Glass M, Appleton I. Cerebral hypoxia-ischemia and middle cerebral artery occlusion induce expression of the cannabinoid CB₂ receptor in the brain. *Neurosci Lett* 2007;412:114–117. [PubMed: 17123706]
- Baker D, Pryce G, Croxford JL, Brown P, Pertwee RG, Makriyannis A, Khanolkar A, Layward L, Fezza F, Bisogno T, Di Marzo V. Endocannabinoids control spasticity in a multiple sclerosis model. *FASEB J* 2001;15:300–302. [PubMed: 11156943]
- Belayev L, Busto R, Zhao W, Fernandez G, Ginsberg MD. Middle cerebral artery occlusion in the mouse by intraluminal suture coated with poly-L-lysine: neurological and histological validation. *Brain Res* 1999;833:181–190. [PubMed: 10375693]
- Berdyshev EV. Cannabinoid receptors and the regulation of immune response. *Chem Phys Lipids* 2000;108:169–190. [PubMed: 11106790]
- Croxford JL. Therapeutic potential of cannabinoids in CNS disease. *CNS Drugs* 2003;17:179–202. [PubMed: 12617697]
- Danton GH, Dietrich WD. Inflammatory mechanisms after ischemia and stroke. *J Neuropathol Exp Neurol* 2003;62:127–136. [PubMed: 12578222]

- Golech SA, McCarron RM, Chen Y, Bemby J, Lenz F, Mechoulam R, Shohami E, Spatz M. Human brain endothelium: coexpression and function of vanilloid and endocannabinoid receptors. *Brain Res Mol Brain Res* 2004;132:87–92. [PubMed: 15548432]
- Grundy RI, Rabuffetti M, Beltramo M. Cannabinoids and neuroprotection. *Mol Neurobiol* 2001;24:29–51. [PubMed: 11831553]
- Hata R, Mies G, Wiessner C, Fritze K, Hesselbarth D, Brinker G, Hossmann KA. A reproducible model of middle cerebral artery occlusion in mice: hemodynamic, biochemical, and magnetic resonance imaging. *J Cereb Blood Flow Metab* 1998;18:367–375. [PubMed: 9538901]
- Hayakawa K, Mishima K, Abe K, Hasebe N, Takamatsu F, Yasuda H, Ikeda T, Inui K, Egashira N, Iwasaki K, Fujiwara M. Cannabidiol prevents infarction via the non-CB1 cannabinoid receptor mechanism. *Neuroreport* 2004;15:2381–2385. [PubMed: 15640760]
- Heinel LA, Rubin S, Rosenwasser RH, Vasthare US, Tuma RF. Leukocyte involvement in cerebral infarct generation after ischemia and reperfusion. *Brain Res Bull* 1994;34:137–141. [PubMed: 8044688]
- Ishikawa M, Zhang JH, Nanda A, Granger DN. Inflammatory responses to ischemia and reperfusion in the cerebral microcirculation. *Front Biosci* 2004;9:1339–1347. [PubMed: 14977549]
- Jin KL, Mao XO, Goldsmith PC, Greenberg DA. CB1 cannabinoid receptor induction in experimental stroke. *Ann Neurol* 2000;48:257–261. [PubMed: 10939579]
- Kanemoto Y, Nakase H, Akita N, Sakaki T. Effects of anti-intercellular adhesion molecule-1 antibody on reperfusion injury induced by late reperfusion in the rat middle cerebral artery occlusion model. *Neurosurgery* 2002;51:1034–1041; discussion 1041–1032. [PubMed: 12234414]
- Kishimoto TK, Rothlein R. Integrins, ICAMs, and selectins: role and regulation of adhesion molecules in neutrophil recruitment to inflammatory sites. *Adv Pharmacol* 1994;25:117–169. [PubMed: 7515640]
- Klein TW, Cabral G. Cannabinoid-Induced Immune Suppression and Modulation of Antigen-Presenting Cells. *J Neuroimmune Pharmacol* 2006;1:50–64. [PubMed: 18040791]
- Klein TW, Newton CA, Friedman H. Cannabinoids and the immune system. *Pain Res Manag* 2001;6:95–101. [PubMed: 11854771]
- Lin TN, He YY, Wu G, Khan M, Hsu CY. Effect of brain edema on infarct volume in a focal cerebral ischemia model in rats. *Stroke* 1993;24:117–121. [PubMed: 8418534]
- Lombard C, Nagarkatti M, Nagarkatti P. CB2 cannabinoid receptor agonist, JWH-015, triggers apoptosis in immune cells: potential role for CB2-selective ligands as immunosuppressive agents. *Clin Immunol* 2007;122:259–270. [PubMed: 17185040]
- Maresz K, Carrier EJ, Ponomarev ED, Hillard CJ, Dittel BN. Modulation of the cannabinoid CB2 receptor in microglial cells in response to inflammatory stimuli. *J Neurochem* 2005;95:437–445. [PubMed: 16086683]
- McCollum L, Howlett AC, Mukhopadhyay S. Anandamide-mediated CB1/CB2 receptor-independent NO production in rabbit aortic endothelial cells. *J Pharmacol Exp Ther*. 2007
- Mishima K, Hayakawa K, Abe K, Ikeda T, Egashira N, Iwasaki K, Fujiwara M. Cannabidiol prevents cerebral infarction via a serotonergic 5-hydroxytryptamine1A receptor-dependent mechanism. *Stroke* 2005;36:1077–1082. [PubMed: 15845890]
- Molina-Holgado E, Vela JM, Arevalo-Martin A, Almazan G, Molina-Holgado F, Borrell J, Guaza C. Cannabinoids promote oligodendrocyte progenitor survival: involvement of cannabinoid receptors and phosphatidylinositol-3 kinase/Akt signaling. *J Neurosci* 2002;22:9742–9753. [PubMed: 12427829]
- Muthian S, Rademacher DJ, Roelke CT, Gross GJ, Hillard CJ. Anandamide content is increased and CB1 cannabinoid receptor blockade is protective during transient, focal cerebral ischemia. *Neuroscience* 2004;129:743–750. [PubMed: 15541895]
- Nagayama T, Sinor AD, Simon RP, Chen J, Graham SH, Jin K, Greenberg DA. Cannabinoids and neuroprotection in global and focal cerebral ischemia and in neuronal cultures. *J Neurosci* 1999;19:2987–2995. [PubMed: 10191316]
- Ni X, Geller EB, Eppihimer MJ, Eisenstein TK, Adler MW, Tuma RF. Win 55212-2, a cannabinoid receptor agonist, attenuates leukocyte/endothelial interactions in an experimental autoimmune encephalomyelitis model. *Mult Scler* 2004;10:158–164. [PubMed: 15124761]

- Nunez E, Benito C, Pazos MR, Barbachano A, Fajardo O, Gonzalez S, Tolon RM, Romero J. Cannabinoid CB2 receptors are expressed by perivascular microglial cells in the human brain: an immunohistochemical study. *Synapse* 2004;53:208–213. [PubMed: 15266552]
- Panikashvili D, Simeonidou C, Ben-Shabat S, Hanus L, Breuer A, Mechoulam R, Shohami E. An endogenous cannabinoid (2-AG) is neuroprotective after brain injury. *Nature* 2001;413:527–531. [PubMed: 11586361]
- Parmentier-Batteur S, Jin K, Mao XO, Xie L, Greenberg DA. Increased severity of stroke in CB1 cannabinoid receptor knock-out mice. *J Neurosci* 2002;22:9771–9775. [PubMed: 12427832]
- Pazos MR, Nunez E, Benito C, Tolon RM, Romero J. Functional neuroanatomy of the endocannabinoid system. *Pharmacol Biochem Behav* 2005;81:239–247. [PubMed: 15936805]
- Ramirez BG, Blazquez C, Gomez del Pulgar T, Guzman M, de Ceballos ML. Prevention of Alzheimer's disease pathology by cannabinoids: neuroprotection mediated by blockade of microglial activation. *J Neurosci* 2005;25:1904–1913. [PubMed: 15728830]
- Ritter LS, Orozco JA, Coull BM, McDonagh PF, Rosenblum WI. Leukocyte accumulation and hemodynamic changes in the cerebral microcirculation during early reperfusion after stroke. *Stroke* 2000;31:1153–1161. [PubMed: 10797180]
- Rodriguez de Fonseca F, Del Arco I, Bermudez-Silva FJ, Bilbao A, Cippitelli A, Navarro M. The endocannabinoid system: physiology and pharmacology. *Alcohol Alcohol* 2005;40:2–14. [PubMed: 15550444]
- Sughrue ME, Mehra A, Connolly ES Jr, D'Ambrosio AL. Anti-adhesion molecule strategies as potential neuroprotective agents in cerebral ischemia: a critical review of the literature. *Inflamm Res* 2004;53:497–508. [PubMed: 15597143]
- Swanson RA, Morton MT, Tsao-Wu G, Savalos RA, Davidson C, Sharp FR. A semiautomated method for measuring brain infarct volume. *J Cereb Blood Flow Metab* 1990;10:290–293. [PubMed: 1689322]
- Tomita M, Fukuuchi Y. Leukocytes, macrophages and secondary brain damage following cerebral ischemia. *Acta Neurochir Suppl* 1996;66:32–39. [PubMed: 8780794]
- Tsuchiya D, Hong S, Kayama T, Panter SS, Weinstein PR. Effect of suture size and carotid clip application upon blood flow and infarct volume after permanent and temporary middle cerebral artery occlusion in mice. *Brain Res* 2003;970:131–139. [PubMed: 12706254]
- van der Stelt M, Veldhuis WB, van Haaften GW, Fezza F, Bisogno T, Bar PR, Veldink GA, Vliegenthart JF, Di Marzo V, Nicolay K. Exogenous anandamide protects rat brain against acute neuronal injury in vivo. *J Neurosci* 2001;21:8765–8771. [PubMed: 11698588]
- Vasthare US, Heinel LA, Rosenwasser RH, Tuma RF. Leukocyte involvement in cerebral ischemia and reperfusion injury. *Surg Neurol* 1990;33:261–265. [PubMed: 2326731]
- Walter L, Stella N. Cannabinoids and neuroinflammation. *Br J Pharmacol* 2004;141:775–785. [PubMed: 14757702]
- Weaver M, Leshley K, Sands H, Gritman KR, Legos JJ, Tuma RF. LEX032, a novel recombinant serpin, protects the brain after transient focal ischemia. *Microvasc Res* 2002;63:327–334. [PubMed: 11969309]
- White BC, Sullivan JM, DeGracia DJ, O'Neil BJ, Neumar RW, Grossman LI, Rafols JA, Krause GS. Brain ischemia and reperfusion: molecular mechanisms of neuronal injury. *J Neurol Sci* 2000;179:1–33. [PubMed: 11054482]
- Wiley JL, LaVecchia KL, Karp NE, Kulasegram S, Mahadevan A, Razdan RK, Martin BR. A comparison of the discriminative stimulus effects of delta(9)-tetrahydrocannabinol and O-1812, a potent and metabolically stable anandamide analog, in rats. *Exp Clin Psychopharmacol* 2004;12:173–179. [PubMed: 15301634]
- Wood, PL. *Neuroinflammation: mechanisms and management*. Humana Press; Totowa, N.J.: 2003.
- Zhang M, Martin BR, Adler MW, Razdan RK, Jallo JI, Tuma RF. Cannabinoid CB(2) receptor activation decreases cerebral infarction in a mouse focal ischemia/reperfusion model. *J Cereb Blood Flow Metab* 2007;27:1387–1396. [PubMed: 17245417]
- Zoratti C, Kipmen-Korgun D, Osibow K, Malli R, Graier WF. Anandamide initiates Ca(2+) signaling via CB2 receptor linked to phospholipase C in calf pulmonary endothelial cells. *Br J Pharmacol* 2003;140:1351–1362. [PubMed: 14645143]

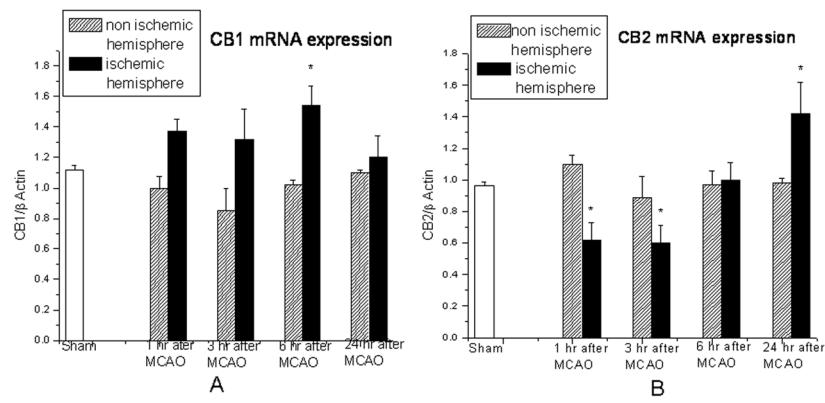


Figure 1. CB₁ mRNA expression in ischemic hemisphere increased from 1 hour after ischemia and was highest at 6 hours after ischemia. However, CB₂ mRNA expression in ischemic hemisphere decreased during first 3 hours following ischemia but then continued to increase until the 24 hour measurement time. There were no significant differences in both CB₁ and CB₂ mRNA expression in non-ischemic hemisphere compared to normal animals. (Samples were tested as triplicates, n=3 in each group, data were expressed as Mean±SEM, * $p < 0.05$ vs sham)

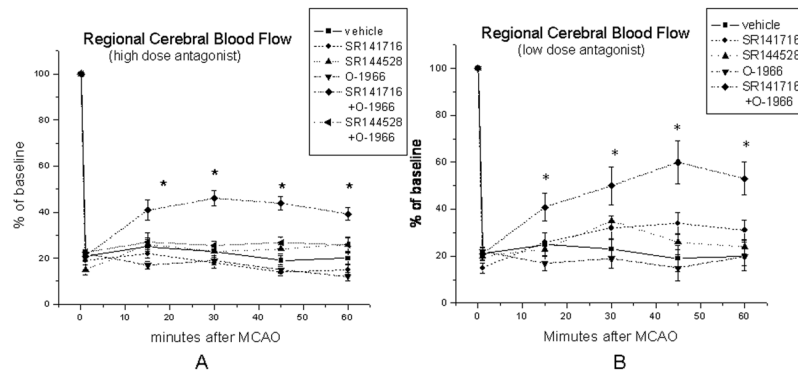


Figure 2.

rCBF dropped to 25% of baseline level within the first 1 minute in all pretreatment groups. Administration of the high dose CB₁ antagonist (SR141716, 20mg/kg, *i.p.*) and CB₂ agonist (O-1966, 1mg/kg, *i.v.*) together 1 hour prior to occlusion increased rCBF during the 1 hour occlusion period when compared with the vehicle-treated group (A); similarly, administration of low dose CB₁ antagonist (SR141716, 5mg/kg, *i.p.*) and CB₂ agonist (O-1966, 1mg/kg) by *i.p.* injection also increased rCBF, whereas other treatments failed to alter rCBF during MCAO (B). (* $p < 0.05$ vs vehicle treated control group, $n = 8$ in each group)

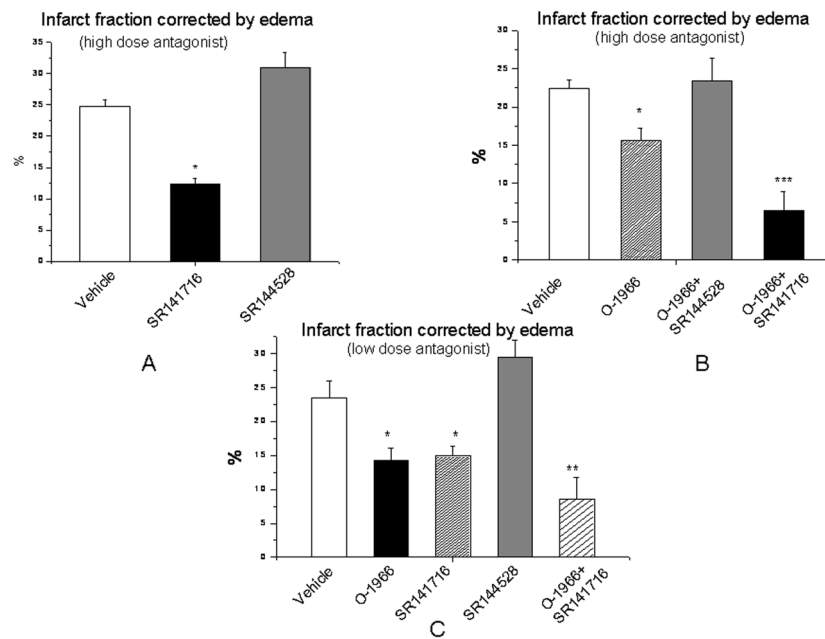


Figure 3.

Effects of modulation of CB₁ and CB₂ receptor activation on cerebral infarction after MCAO. Administration of the CB₁ antagonist (SR141716, 20mg/kg, *i.p.*) 1 hour before MCAO significantly decreased cerebral infarction while administration of CB₂ antagonist (SR144528, 20mg/kg, *i.p.*) alone increased cerebral infarction compared with vehicle-treated group (A). CB₂ agonist (O-1966, 1mg/kg, *i.v.*) alone decreased infarct volume which was completely reversed by CB₂ antagonist; while administration of CB₂ agonist with CB₁ antagonist together dramatically further reduced cerebral infarction following MCAO (B). When the CB₂ agonist (O-1966) was administered *i.p.* and antagonists were given at a lower dosage at 5mg/kg *i.p.* 1 hour before MCAO, both the CB₂ agonist and CB₁ antagonist still significantly reduced cerebral infarction, while CB₂ antagonist tended to increase the cerebral infarction although not achieving statistical significance. The combination of both CB₂ agonist (1mg/kg, *i.p.*) and CB₁ antagonist (5mg/kg, *i.p.*) dramatically reduced cerebral infarction compared to vehicle treated group (C). (n=5–8 in each group, data were expressed as Mean±SEM, **p*<0.05 vs vehicle treated control group, ***p*<0.01 vs vehicle treated control group, ****p*<0.001 vs vehicle treated control group)

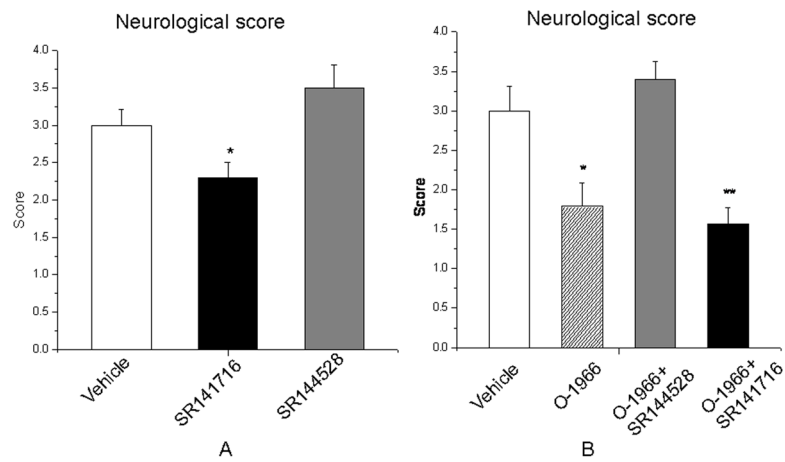


Figure 4.

Effects of modulation of CB₁ and CB₂ receptor activation on neurological function after MCAO. In parallel with the changes in cerebral infarction, administration of a CB₁ antagonist (20mg/kg, *i.p.*) improved motor function, while administrations of a CB₂ antagonist tended to worsen motor function (A). Administration of CB₂ agonist (1mg/kg, *i.v.*) alone improved motor function and this protection which was totally reversed by CB₂ antagonist (20mg/kg, *i.p.*); while administration of CB₂ agonist with CB₁ antagonist together significantly attenuated neurological deficits after MCAO (B). (n=7–8 in each group, data were expressed as Mean ± SEM, **p*<0.05 vs vehicle treated control group, ***p*<0.01 vs vehicle treated control group)