# Inhibitory Effect of Fermented Papaya Preparation on Hydroxyl Radical Generation from Methylguanidine

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Summary We have previously shown that extremely high level of guanidino compounds such as methylguanidine (MG), known as a neurotoxin and also a nephrotoxin, generate reactive oxygen species (ROS) using an electron spin resonance (ESR) technique with spin trap 5,5dimethyl-1-pyrroline-N-oxide (DMPO). In this *in vitro* study, the inhibitory effect of fermented papaya preparation (SAIDO-PS501:PS-501) on hydroxyl radical (·OH) generation from MG was examined using an ESR spectrometry, and it was found that PS-501 suppressed ·OH generation from MG in a dose-dependent manner. The ID<sub>50</sub> value of PS-501 was 8 mg/ml. On the contrary, glucose itself did not suppress ·OH generation from MG up to100 mg/ml, whereas PS-501 almost completely suppressed ·OH generation from MG at a dose of 100 mg/ ml. These results imply that PS-501 itself may have a beneficial effect of preventing ROS- and MG-related diseases.

Key Words: methylguanidine, papaya, hydroxyl radical

### Introduction

Methylguanidine (MG) is known as not only a nephrotoxin but also as a neurotoxin. Greatly increased level of MG was demonstrated in the plasma from chronic uremic patient by Giovannetti *et al.* [1] in 1968. An importance of MG as a uremic toxin has been well established in animals with chronic renal failure [2–5].

The increased levels of guanidine-like substances in the blood of epileptic patients was first reported by Murry *et al.* [6] in 1940. In 1966, Mori *et al.* [7] have first shown that  $\gamma$ -guanidinobutyric acid induced convulsions in rabbits, and then demonstrated that many guanidino compounds, including MG [8], could induce convulsion in experimental

animals, e.g., rats and rabbits, by intracisternally injection with them [9-11]. In addition, sustained increased level of MG was found in the brain of experimental models of epilepsy: amygdala- or hippocampus-kindled rats [12, 13].

Meanwhile, a natural antioxidant, papaya (*Carica papaya L*.), is know to have beneficial properties such as the prevention not only against the lipid peroxidation, but also other protection, for example, the enhancement of the phase II enzyme activity such as glutathione S-transferase in hepatocytes [14], hepatoprotective activity against CCl<sub>4</sub> [15], the protection against the iron-mediated oxidative DNA damage in the plasmid [16] and also  $\beta$ -amyloid-mediated copper neurotoxicity in  $\beta$ -amyloid precursor protein in the APPsw in cell culture system [17].

Recently, Mori *et al.* [18, 19] demonstrated that extremely high level of guanidino compounds generated reactive oxygen species (ROS) using an electron spin resonance (ESR) spectrometry with spin trap 5,5-dimethyl-1pyrroline-N-oxide (DMPO).

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In this *in vitro* study, the inhibitory effect of the fermented papaya preparation (SAIDO-PS501: PS-501) on hydroxyl radical (·OH) generation from MG was examined using an ESR technique.

### **Materials and Methods**

### Sample

Fermented Papaya (*Carica papaya L.*) preparation, fermented the unripe fruits including the seeds, (SAIDO-PS501: PS-501) (SAIDO Co., Ltd., Fukuoka, Japan) was used. The sample solution was dissolved in the medium just before analysis. Glucose was used as a control of PS-501, as PS-501 contains 95% of glucose in the preparation.

### Reagents

MG hydrochloride was obtained from Tokyo Kasei Kogyo Co. (Tokyo, Japan). Xanthine oxidase (1 U/ml, from caw milk) was obtained from Roche Diagnostics (Indianapolis, IN) EPC-K<sub>1</sub> ( $\alpha$ -tocopheryl-L-ascorbate-2-O-phosphate diester), is a gift from Dr. Kazumi Ogata (Oga Research, Osaka, Japan) [20]. Superoxide dismutase (SOD) and catalase were from Sigma Chemical Co. (St. Louis, MO). The spin trap, DMPO was from Labotec Co (Tokyo, Japan). All other chemicals were the highest grade available.

#### ESR measurements

For the measurements of  $\cdot$ OH and O<sub>2</sub><sup>--</sup> scavenging activity, ESR spectrometer (JES-REIX HR, JEOL, Tokyo, Japan) equipped with manganese oxide (MnO) as an internal standard was used. ROS,  $\cdot$ OH and O<sub>2</sub><sup>--</sup>, were analyzed as the spin adducts of DMPO, DMPO-OH and DMPO-OOH, respectively. ESR settings are the following: magnetic field: 335.6 ± 5 or 331.5 ± 10 mT; microwave power: 8 mT; microwave frequency: 9.4 GHz; modulation frequency 100 kHz; modulation width: 1 × 0.16000 mT; response time: 0.1 s; gain; 1 × 200–1 × 790; sweep width: 5.000 or 10.000 mT; sweep time: 2 min; accumulation: ×4 (for the hydroxyl radical detection from methylguanidine); temperature: 25°C.

Methods for the spin trapping of  $\cdot$ OH and O2<sup>--</sup> were based on earlier reports [21–23].

### Hydroxyl radical scavenging activity

All solutions except FeSO<sub>4</sub> were dissolved in 0.1 M phosphate buffer (pH 7.4); FeSO<sub>4</sub> was dissolved in distilled water. The sample solution (50 µl), 18 or 1.8 mM DMPO (50 µl) (final concentration of DMPO: 4.5 or 0.45 mM), 2 mM H<sub>2</sub>O<sub>2</sub> (50 µl), and 0.2 mM FeSO<sub>4</sub> (50 µl) were put into the ESR quartz flat cell (capacity: 200 µl). Exactly 30 s after the addition of FeSO<sub>4</sub>, the ESR spectra of the DMPO-OH spin adducts were recorded [24, 25].

# Superoxide anion radical scavenging activity (SOD-like activity)

All solution except dimethylsulfoxide (DMSO) was in 0.1 M phosphate buffer (pH 7.4). Fifty microliters of 4 mM hypoxanthine, DMSO (30  $\mu$ l), sample solution (50  $\mu$ l), 4.5 M DMPO (20  $\mu$ l), and xanthine oxidase (0.4 units/ml) (50  $\mu$ l) were mixed, and transferred into the ESR quartz flat cell. Exactly 30 s after the addition of xanthine oxidase, the ESR spectra of the DMPO-OOH spin adducts were recorded [24, 25].

# Inhibitory effect of PS-501 on hydroxyl radical generation from MG

The ESR measurements were basically performed as follows according to the procedure by Noda *et al.*: The exact amount of MG was added 2.2 M DMPO in pure water (10 µl) followed by 0.2 M Tris buffer (pH 6.0) (100 µl), the sample in pure water (40 µl) (control: pure water without sample) and 0.2 mM ferrous sulfate in pure water (50 µl). The mixture was mixed quickly by a vortex mixer, and the mixture was immediately put into an ESR quartz flat cell (capacity: 200 µl). ESR recordings were started exactly 30 seconds after the addition of ferrous ion (Fe<sup>2+</sup>). The intensity of the DMPO-spin adducts of hydroxyl radicals (DMPO-OH) was normalized by an internal standard manganese oxide (MnO) equipped to the ESR spectrometer.

### Results

#### Hydroxyl radical scavenging activity of PS-501

The PS-501 scavenged ·OH generated by Fenton reaction in a dose-dependent manner (Fig. 1a). The ID<sub>50</sub> values were 1 and 0.1 mg/ml when the DMPO concentrations were 4.5 and 0.45 mM (each final concentration), respectively (Fig. 1b). The ID<sub>50</sub> value shifted 10-fold lower when a 10fold lower concentration of DMPO was used.

# Superoxide anion radical scavenging activity of PS-501 (SOD-like activity)

Fig. 2 shows representative ESR spectra of DMPO-spin adducts of  $O_2^{-}$  (DMPO-OOH) generated from hypoxanthinexanthine oxidase system with or without PS-501 (control: without PS-501). The PS-501 hardly scavenged  $O_2^{-}$  in this assay system.

### Hydroxyl radical generation from MG

The relationship between the concentrations of MG and the intensity of the DMPO-spin adducts of ·OH (DMPO-OH) linearly increased at the range of 5 to 20 mg/ml/assay under the experimental condition (Fig. 3).

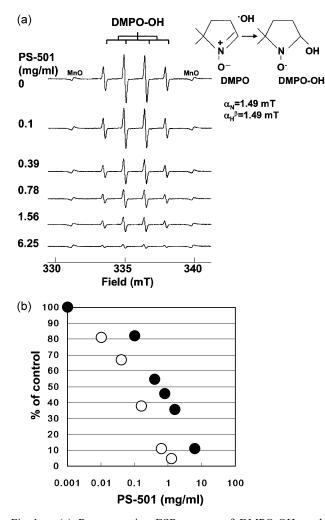


Fig. 1. (a) Representative ESR spectra of DMPO-OH at the various concentrations of PS-501 (DMPO: 4.5 mM). Hydroxyl radicals were generated by Fenton reaction. (b) A dose-response curve of hydroxyl radical scavenging activity for PS-501 (Fenton reaction). The relative peak height of the second peak of DMPO-OH spin adduct was plotted against the concentration of MG. The intensity of the DMPO-OH spin adducts was normalized by an internal standard, MnO. The symbol of the closed circles and opened circles represent the data points obtained by using 4.5 and 0.45 mM DMPO (each final concentration), respectively.

# Inhibitory effect of PS-501 or antioxidants on hydroxyl radical generation from MG

In our preliminary experiments, the intensity of the DMPO spin adducts of ·OH generated from MG was almost constant between the pH ranges of 5.5 to 6.5 under the experimental condition. Therefore, the following experiments were performed at pH 6.0. The PS-501 suppressed ·OH generation from MG (Fig. 4a). At a dose of 100 mg/ml, the PS-501 almost completely inhibited ·OH generation

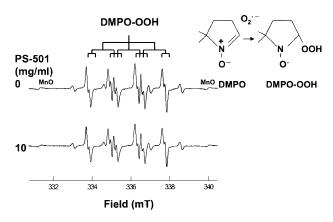


Fig. 2. Representative ESR spectra of DMPO-OOH with or without PS-501 (DMPO: 450 mM). Superoxide anion radicals were generated by hypoxanthine-xanthine oxidase system. Control: without PS-501.

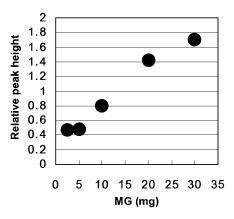


Fig. 3. The relationship between MG concentrations and hydroxyl radical generation. The relative peak height of the second peak of DMPO-OH spin adduct was plotted against the amount of MG. The intensity of the DMPO-OH spin adducts was normalized by an internal standard, MnO. Sweep time: 2 min; accumulation: ×4 times.

from MG. On the contrary, glucose itself showed no effect on  $\cdot$ OH generation from MG up to the concentration of 100 mg/ml. In contrast, a known hydroxyl radical scavenger ascorbate or EPC-K<sub>1</sub> effectively suppressed  $\cdot$ OH generation from MG in a dose-dependent manner. Ascorbate and EPC-K<sub>1</sub> completely inhibited at a dose of 1 µg/ml and 100 µg/ml, respectively.

The PS-501 suppressed ·OH generation from MG in a dose-dependent manner (Fig. 5a). The value of the ID<sub>50</sub> for PS-501 was 8.0 mg/ml (Fig. 5b).

#### Discussion

The PS-501 scavenged OH generated by Fenton reaction in a dose-dependent manner. When the concentration of

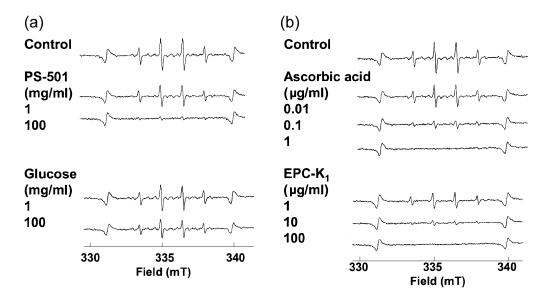


Fig. 4. Representative ESR spectra of DMPO-OH generated from MG with PS-501, glucose, ascorbate or EPC-K1. Hydroxyl radicals were generated from MG (10 mg/assay). Sweep time: 2 min; accumulation: ×4 times.

DMPO was 10-fold lowered, the value of ID<sub>50</sub> lowered 10fold, indicating that the PS-501 directly scavenged ·OH when encountered to ·OH without inhibiting the ·OH generation system such as chelating with ferrous iron (Fe<sup>2+</sup>) [26–28]. The ID<sub>50</sub> value of PS-501 was 1 or 0.1 mg/ml when the concentration of DMPO was 4.5 or 0.45 M (each final concentration). Ttrolox, a water-soluble vitamin analogue, is known to be a ·OH scavenger, for example, showed ID<sub>50</sub> value of 0.50 mg/ml when the concentration of DMPO was 4.5 mM (final concentration) (data not shown).

The PS-501 hardly scavenged O<sub>2</sub><sup>--</sup> generated by xanthinexanthine oxidase system.

In the previous study by Mori *et al.* [19], the mechanism for the ·OH generation from MG has been shown that the oxidation of guanidine compounds such as MG, guanidinosuccinic acid, guanidinoacetic acid or arginine, MG itself generates ·OH *via* the conversion of surrounding oxygen to  $O_2^-$ , then quickly producing hydrogen peroxide followed by ·OH [19]. In this study, highly purified DMPO was used. The relationship between the concentrations of MG and the intensity of the DMPO-spin adducts of ·OH (DMPO-OH) were dose-dependent on the concentrations of MG under this experimental condition. By the addition of superoxide dismutase (SOD) (200 U/assay) or catalase (650 U/assay) in MG, the ·OH generation was suppressed, and ethanol (20%, v/v) or DMSO (20%, v/v) inhibited ·OH generation from MG (data not shown).

Meanwhile, PS-501 suppressed ·OH generation from MG in a dose-dependence manner. The value of ID<sub>50</sub> for PS-501 was 8.0 mg/ml. Interestingly, glucose itself did not show the inhibition against ·OH generation from MG, when the concentration of glucose was examined up to 100 mg/ml, while PS-501 inhibited almost completely at a does of 100 mg/ml under the same experimental condition (Fig. 4). In PS-501, glucose is contained because of the requirement for the fermentation process. When glucose alone was examined for the scavenging activity on  $\cdot$ OH generation by Fenton reaction, the level of ID<sub>50</sub> for glucose was similar to that of PS-501 at the same concentration of DMPO. Therefore, the brief mechanism for the inhibitory effect of PS-501 on hydroxyl radical generation from MG is not yet clear, however, it could be most likely due to its  $\cdot$ OH scavenging and probably other factors could be involved in.

Not only the DMPO spin adducts, DMPO-OOH, can be converted into DMPO-OH, or spin trapping, nitroxides, can be reduced to its hydroxylamines by reductants such as ascorbate depending upon the structures of nitroxides [29-31], but also DMPO-OH and DMPO-OOH could be reduced by reductants. Therefore, although ascorbate is known to act as potent reducing agent, which can reduce more reactive species such as ·OH and O2<sup>--</sup>, for example, by donating electron to the ·OH system (standard one-electron reduction potential for the following couples are; ascorbate, H<sup>+</sup>/ascorbate monoanion: 282 mV, ·OH, H<sup>+</sup>/H<sub>2</sub>O: 2310 mV, O2<sup>--</sup>, 2H<sup>+</sup>/H2O2: 940 mV) [32], and thus apparently scavenging these radicals, further study will elucidate the brief mechanism, i.e, the direct scavenging and/or reducing the nitroxide or reducing of DMPO-OOH and DMPO-OH themselves, by using more stable spin probes.

The biologically active components in PS-501 have not yet been identified, although it is kwon that it contains carbohydrate, proteins, and some amino acids, but not ascorbate (in our personal communication by SAIDO Co., Ltd). Interestingly, it is reported that papaya fruits and seeds

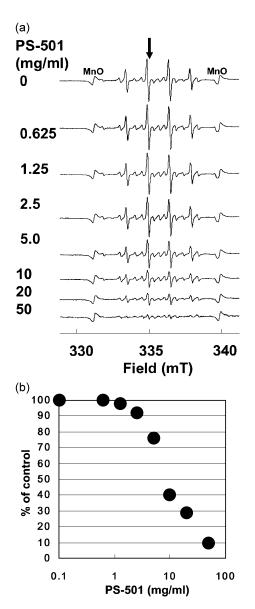


Fig. 5. (a) A dose-response of PS-501 on hydroxyl radical generation from MG. Control: without PS-501. Hydroxyl radicals were generated from MG (10 mg/assay). Sweep time: 2 min; accumulation: ×4 times. (b) Inhibitory effect of PS-501 on hydroxyl radical generation from MG. The relative peak height of the second peak of DMPO-OH spin adducts was plotted against the concentration of MG. The intensity of the DMPO-OH spin adducts were normalized by an internal standard, MnO.

contain benzyl isothiocyanate [33-37]. Due to the fermentation, the components may differ from those in the fresh fruits and seeds. Therefore, the identification of the active components in the fermented papaya preparation, which suppress the  $\cdot$ OH generation from MG, would be of interest.

Guanidino compounds including MG are widely present in mammals. However, the level of MG significantly increases under the pathophysiological condition such as convulsion and chronic renal failure. For example, in the case of left-hippocampus kindling rats, the level of MG in the left amygdala increased up to 124% or 110% for 7-days or 28-days after the completion of the kindling when compared to the control (control: 0.5 nmol/g tissue wet weight), indicating that long-lasting increase of MG may be important in producing neuronal hyperexcitability in kindled epileptogenesis [13]. Meanwhile, the level of MG both in the serum and urine in the patients with renal failure is known to increase markedly comparing with normal control [38].

In conclusion, PS-501 itself effectively inhibited ·OH generation from MG, although the precise mechanism for the inhibitory effect of PS-501 against the ·OH generation from MG is not yet clear at this stage. However, these results imply that PS-501 may have a beneficial effect on preventing ROS- and MG-related diseases. Further studies will elucidate the possible effective components in the PS-501 and its possible beneficial effects on the diseases such as convulsion and/or chronic renal failure in the future *in vivo* studies.

## References

- Giovannetti, S., Biagini, M., and Cioni, L.: Evidence that methylguanidine is related in chronic renal failure. *Experientia*, 24, 341–343, 1968.
- [2] Giovannetti, S., Biagini, M., Balestri, P.L., Navalesi, R., Giagnoni, P., De Matteis, S., Ferro-Milone, P., and Perffetti, C.: Uremia-like syndrome in dogs chronically intoxicated with methylguanidine and creatinine. *Clin. Sci.*, 36, 445–452, 1969.
- [3] Giovannetti, S., Balestri, P.L., and Barsotti, G.: Methylguanidine in uremia. Arch. Intern. Med., 131, 709–713, 1973.
- [4] Yokozawa, T., Mo, Z.L., and Oura, H.: Comparison of toxic effects of methylguanidine, guanidinosuccinic acid and creatinine in rats with adenine-induced chronic renal failure. *Nephron*, **51**, 388–392, 1989.
- [5] Yokozawa, T., Fujitsuka, N., and Oura, H.: Variations in the distribution of methylguanidine with the progression of renal failure after methylguanidine loading. *Nephron*, **52**, 347–351, 1989.
- [6] Murry, M. and Hoffman, A.B.: The occurrence of guanidinelike substances in the blood in essential epilepsy. J. Lab. Clin. Med., 25, 1072–1073, 1940.
- [7] Jinnnai, D., Sawai, A., and Mori, A.: γ-Guanidinobutyric acid as a convulsive substance. *Nature*, **212**, 617, 1966.
- [8] Matsumoto, M., Kobayashi, K., Kishikawa, H., and Mori, A.: Convulsive activity of methylguanidine in cats and rabbits. *IRCS Med. Sci.*, 4, 65, 1976.
- [9] Mori, A., Watanabe, Y., and Akagi, M.: Guanidino compound anomalies in epilepsy, *in Advances in Epileptology: XIIIth Epilepsy International Symposium*, eds. By Akimoto, H., Kazamatsuri, H., Seino, M., and Ward, A., Raven Press, New York, pp. 347–351, 1982.

- Y. Noda et al.
- [10] Mori, A.: Biochemistry and neurotoxicology of guanidino compounds: History and recent advances. *Pavlov. J. Biol. Sci.*, 22, 85–94, 1987.
- [11] Mori, A.: Reactive oxygen species and mechanism of induction of seizure by guanidino compounds, *in Free Radicals in Brain Physiology and Disorders*, Academic Press, San Diego, pp. 3–14, 1966.
- [12] Hirayasu, Y., Morimoto, K., and Otsuki, S.: Increase of methylguanidine and guanidinoacetic acid in the brain of amygdala-kindled rats. *Epilepsia*, **32**, 761–766, 1991.
- [13] Shimizu, Y., Morimoto, K., Kuroda, S., and Mori, A.: Sustained increase of methylguanidine in the rats after amygdala and hippocampal kindling. *Epilepsy Res.*, 21, 11– 17, 1995.
- [14] Nakamura, Y., Morimitsu, Y., Uzu, T., Ohigashi, H., Murakami, A., Naito, Y., Nagasawa, Y., Osawa, T., and Uchida, K.A.: Glutathione S-transferase inducer from papaya: rapid screening, identification and structure-activity relationship of isothiocyanates. *Cancer Lett.*, **157**, 193–200, 2000.
- [15] Rajkapoor, B., Jayakar, B., Kavimani, S., and Murugesh, N.: Effect of dried fruits of *Carica papaya* Linn on hepatotoxicity. *Biol. Pharm. Bull.*, 25, 1645–1646, 2002.
- [16] Rimbach, G., Guo, Q., Akiyama, T., Matsugo, S., Moini, H., Virgili, F., and Packer, L.: Ferric nitrilotriacetate induced DNA and protein damage: inhibitory effect of a fermented papaya preparation. *Anticancer Res.*, **20**, 2907–2914, 2000.
- [17] Zhang, J., Mori, A., Chen, Q., and Zhao, B.L.: Fermented papaya preparation attenuates β–amyloid precursor protein: β-amyloid-mediated copper neurotoxicity in β–amyloid precursor protein and in β–amyloid precursor Swedish mutation overexpressing SH-SY5Y cells. *Neuroscience*, **143**, 63–72, 2006.
- [18] Mori, A., Kohno, M., Masumizu, T., and Packer, L.: α-Guanidinoglutaric acid as a free radical generator. *Biochem. Mol. Biol. Int.*, **37**, 371–374, 1995.
- [19] Mori, A., Kohno, M., Masumizu, T., Noda, Y., and Packer, L.: Guanidino compounds generate reactive oxygen species. *Biochem. Mol. Biol. Int.*, 40, 135–143, 1996.
- [20] Mori, A., Edamatsu, R., Kohno, M., and Ohmori, S.: A new hydroxyl radical scavenger: EPC-K<sub>1</sub>. *Neurosciences*, 15, 371–376, 1989.
- [21] Rosen, G.M. and Rauchman, E.J.: Spin trapping of superoxide and hydroxyl radicals, *in Methods in Enzymology*. *Oxygen Radicals in Biological Systems*, vol. 105, ed. By Packer, L., Academic Press, San Diego, pp. 198–209, 1984.
- [22] Lagercrantz, C.: Spin trapping of some short-lived radicals by the nitroxide method. J. Phys. Chem., 75, 3466–3476, 1971.
- [23] Janzen, E.G.: Spin trapping. Acc. Chem. Res., 4, 31-40, 1971.
- [24] Noda, Y., Anzai, K., Mori, A., Kohno, M., Shimmei, M., and Packer, L.: Hydroxyl and superoxide anion radical scaveng-

ing activities of natural antioxidants using the computerized JES-FR 30 ESR spectrometer system. *Biochem. Mol. Biol. Int.*, **42**, 35–44, 1997.

- [25] Noda, Y., Kohno, M., Mori, A., and Packer, L.: Automated electron spin resonance free radical detector assyas for antioxidant activity in natural extracts, *in Methods in Enzymology. Oxidants and antioxidants part A, vol. 299*, ed. By Packer, L., Academic Press, San Diego, pp. 28–34, 1999.
- [26] Kimura, K.: Mechanism of active oxygen species reduction by non-steroid anti-inflammatory drugs. *Int. J. Biochem. Cell Biol.*, 29, 437–446, 1997.
- [27] Noda, Y., Kaneyuki, T., Igarashi, K., Mori, A., and Packer, L.: Antioxidant activity of nasunin, an anthocyanin in eggplant. *Res. Commun. Mol. Pathol. Pharmacol.*, **102**, 175– 187, 1998.
- [28] Noda, Y., Kaneyuki, T., Mori, A., and Packer, L.: Antioxidant activities of pomegranate fruit extract and its anthocyanidins: delphinidin, cyanidin, and pelargonidin. *J. Agric. Food Chem.*, **50**, 166–171, 2002.
- [29] Smith, I.C.P.: in Biological Applications of Electron Spin Resonance, eds. By Swartz, H.M., Bolton, J.R., and Borg, D.C., Wiley-Interscience, New York, pp. 483–539, 1972.
- [30] Quintanilha, A.T. and Packer, L.: Surface localization of sites of reduction of nitroxide spin-labeled molecules inmitochondria. *Proc. Natl. Acad. Sci.*, 74, 570–574, 1977.
- [31] Finkelstein, E., Rosen, G.M., and Rauckman, E.J.: Spin trapping of superoxide and hydroxyl radical: practical aspects. Arch. Biochem. Biopys., 200, 1–16, 1980.
- [32] Buettner, G.R.: The pecking order of free radicals and antioxidants: lipid peroxidation, α-tocopherol, and ascorbate. *Arch. Biochem. Biopys.*, **300**, 535–543, 1993.
- [33] Tang, C.S.: Benzyl isothiocyanate in papaya fruit. *Phyto-chemistry*, **12**, 117–121, 1971.
- [34] Flath, R.A. and Forrey, R.R.: Volatile components of papaya (*Carica papaya* L. solo variety). J. Agric. Food Chem., 25, 103–109, 1977.
- [35] Shen, F. and Shyn, Y.T.: Determination of benzyl isothiocyanate in papaya fruit by solid phase extraction and gas chromatography. J. Food Drug Anal., 4, 327–334, 1996.
- [36] Kermanshai, R., McCarry, B.E., Rosenfeld, J., Summers, P.S., Weretilnyk, E.A., and Sorger, G.J.: Benzyl isothiocyanate is the chief or sole anthelmintic in papaya seed extracts. *Phytochemistry*, 57, 427–435, 2001.
- [37] Nakamura, Y., Yoshimoto, M., Murata, Y., Shimoishi, Y., Asai, Y., Park, E.Y., Sato, K., and Nakamura, Y.: Papaya seed represents a rich source of biologically active isothiocyanate. *J. Agric. Food Chem.*, 55, 4407–4413, 2007.
- [38] Marescau, B., Nagels, G., Possemiers, I., De Broe, M.E., Because, I., Billiouw, J.-M., Lornoy, W., and De Deyn, P.P.: Guanidino compounds in serum and urine of nondialyzed patients with chronic renal insufficiency. *Metabolism*, 46, 1024–1031, 1997.