

Video Article

Preparation of Gene Gun Bullets and Biolistic Transfection of Neurons in Slice Culture

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Abstract

Biolistic transfection is a physical means of transfecting cells by bombarding tissue with high velocity DNA coated particles. We provide a detailed protocol for biolistic transfection of rat hippocampal slices, from the initial preparation of DNA coated bullets to the final shooting of the organotypic slice cultures using a gene gun. Gene gun transfection is an efficient and easy means of transfecting neurons and is especially useful for fluorescently labeling a small subset of cells in tissue slice. In this video, we first outline the steps required to coat gold particles with DNA. We next demonstrate how to line the inside of plastic tubing with the gold/DNA bullets, and how to cut this tubing to obtain the plastic cartridges for loading into the gene gun. Finally, we perform biolistic transfection of rat hippocampal slice cultures, demonstrating handling of the Bio-Rad Helios gene gun, and offering trouble shooting advice to obtain healthy and optimally transfected tissue slices.

Protocol

PROTOCOL FOR BIOLISTIC TRANSFECTION

Making bullets

Bullets are good for up to 6 months, but may begin to decrease in transfection efficiency after 2-3 months. Bullets should be stored at 4°C in the presence of desiccant pellets. Always let the scintillation vials, in which bullets are stored, warm to room temperature before opening the vial.

Before getting started have ready:

- Spermidine (0.05M)
- CaCl₂ (1M)
- EtOH (100%-high grade-unopened bottle)
- Autoclaved H₂O
- DNA to be transfected
- Polyvinylpyrrolidone (PVP-20 mg/ml stock)
- 2 X 15 ml conical (2/bullet set)
- Tubing (cut into 1 X ~ 30 inches/ bullet set)
- Scintillation vials (1/bullet set)
- Desiccation pellets
- 10 ml syringe w/tubing on the end

Prepare tubing:

1. Dry ~ 30 inches of tubing (about 2 inches extending beyond right O-ring) in the tubing station w/ N₂ gas (0.4 pressure) for a minimum of 20 min.

Precipitating DNA on gold beads:

1. Prepare DNA to be transfected in 50 mL total volume in microcentrifuge tube (maximum 50 mg total DNA)
2. Weigh out 6-8 mg of gold and transfer to an microcentrifuge tube
3. Add 100 mL of 0.05M spermidine to gold and sonicate for 20 sec
4. Add the 50 mL of DNA to the gold/spermidine
5. Vortex (around setting 5) w/cap open
6. Add drop wise 100 mL of 1M CaCl₂
7. Sonicate briefly (< 5 sec)
8. Allow to precipitate for 10 min at room temp
9. Sonicate briefly

Prepare PVP solution during sonication:

1. Make dilute PVP solution in 15 ml conical, 3.5mL dilute PVP for each set of bullets (add 7.5 mL of 20mg/ml PVP stock to 3.5 ml 100 % EtOH)

Rinsing gold beads with EtOH:

1. Spin down gold beads at full speed for 10 sec in microcentrifuge
2. Discard supernatant, but keep a small amount of liquid on top of gold beads
3. Wash 3X with 1ml EtOH, sonicate briefly each time, if necessary

Resuspend gold/DNA in PVP solution:

1. Resuspend the gold pellet in 200 mL of the 3.5 ml dilute PVP/EtOH solution
2. Vortex the gold pellet to resuspend (briefly sonicate if necessary)
3. Transfer the 200 mL of gold/PVP solution to a new 15 mL conical
4. Continue in this way, adding 200 mL of PVP at a time until all the gold has been transferred to the 15 mL conical, and the final volume is 3.2 mL

Coating tubing:

1. Turn off N₂ at the tubing station and remove dried tubing
2. Attach dried tubing to the 10 ml syringe
3. Vigorously shake the 15 mL conical filled with gold/PVP
4. Place the end of the tubing in gold/PVP solution and suck up slowly being careful to avoid bubbles
5. Leave ~ 2 inches dry from both ends of the tubing
6. With the tubing still attached to syringe and filled with the gold/PVP solution, carefully place the tubing back into station
7. Mark the ends of the tubing where the solution sits
8. Let the gold settle for 5 min with the syringe attached
9. Suck-up the solution very slowly by pulling the syringe so that the settled gold is left behind (make sure not to back-wash)
10. Disconnect syringe
11. Rotate 90° and let sit 2 seconds
12. Rotate 180° and let sit another 2 seconds
13. Rotate tube for 5 sec
14. Turn on the N₂ so that the valve reads between 0.4 - 0.5 and spin the gold-coated tubing while drying for 5 min.

Cutting tubing:

1. Remove the tubing from the tubing prep-station and cut off the ends at the marks.
2. Put a labeled scintillation vial under the tubing chopper to catch cut cartridges
3. Place tubing in the tubing chopper and cut the tubing with clean razor
4. Place a desiccation pellets in scintillation vial
5. Store cartridges at 4°C

Shooting Tissue Slices

When shooting cultured slices, it is important to employ a relatively sterile technique in order to avoid contamination. Remember, that when shooting two different constructs, both sets of bullets can be loaded into the same cartridge holder, but the barrel-liner and screen should be changed between constructs.

Before getting started have ready:

- DNA bullets (let scintillation vial warm to room temperature before opening)
- Tissue slices
- Helios Gene Gun
- Helium tank with gene gun hose attached
- Cartridge holder
- Barrel liner with plastic O-ring in place
- Diffuser (modified)
- Diffuser Screen
- Forceps

Prepping the gene gun:

1. Using forceps place the gold coated plastic cartridges into the cartridge holder.
2. Place the cartridge holder into the gene gun by first pushing back lock
3. Secure the cartridge holder with lock
4. Obtain a barrel liner with an O-ring and the diffuser screen securely in place
5. Screw the barrel-liner into the gene gun

Shooting cultured slices with the gene gun:

1. Put on ear muffs
2. Plug the gene gun into hose connected Helium gas tank
3. Turn up pressure on tank to 180 PSI
4. Shoot a blank into the air in order to remove any dust or debris from the diffuser screen. To shoot the gun, first depress the safety interlock button until the gun beeps, then pull the trigger.
5. After shooting the blank, remove the slices from the incubator
6. Advance gun to first construct
7. Shoot with the diffuser screen about 0.5 - 1 inch from the slice
8. Advance the cartridge between wells.
9. After shooting the last well, place the slices back into the incubator
10. Turn off gas and release pressure before detaching the gun from the Helium hose
11. Unscrew the barrel liner, and remove the cartridge and the used shells

Cleaning cartridge holders and barrel liners:

1. Place cartridges, barrel liners, and diffuser screens in a beaker with soapy water

2. Place beaker in bath sonicator and sonicate for 20 min
3. Thoroughly rinse with water until all soap residues are removed
4. Soak in 70% EtOH for an hour
5. Place on dry paper towel overnight to dry
6. Clean cartridges, barrel liners, and screens can then be reused in subsequent biolistic transfections

Trouble shooting tips for Biolistic Transfection

Biolistic transfection can sometimes result in suboptimal transfection conditions. It can lead to too few or too many cells transfected, too high or too low expression levels, or may cause tissue slices to become generally unhealthy. Often unhealthy slices result from the pressure blast. Here are some tips to obtain optimally transfected tissue slices.

If slices appear to be unhealthy (or if *too many* cells are transfected/ slice), try:

- *increasing* shooting distance
- *decreasing* pressure on gas tank
- *decreasing* the amount of gold
- cutting out the center of the Bio-Rad diffuser and replacing the center with a diffuser screen.

If *too few* cells are transfected, try:

- *decreasing* shooting distance
- *increasing* pressure on gas tank
- *increasing* the amount of gold
- *increasing* the number of tissue slices/ well
- make sure the O-ring in the barrel-liner is intact.

If you obtain slices with both *too many* AND *too few* cells transfected during the same shooting *make sure*:

- that you are holding the gun symmetrically above well
- that gold is evenly coating the inside of tubing. (If you are getting a *line of gold*, draw out the supernatant from the tubing at a faster rate once the gold has settled and rotate the gold longer before turning on the nitrogen. If, on the other hand, you are getting *streaking of gold*, this is likely due to too much moisture exposure during bullet making: make sure you are using an unopened bottle of EtOH, that the tubing is thoroughly dry before coating, and that you capping lids and preparing bullets in a timely manner).

If transfection efficiency seems optimal (# cells transfected/ slice), but the expression level seems too high or too low:

- adjust the ratio DNA to gold. (e.g., if it is *too low* maintain the amount of gold but *increase* the amount of DNA.)
- adjust the amount of time between shooting and data collection

In the case of dual transfection, if you are getting too little expression of one of your constructs, adjust the ratio of the two constructs in the appropriate manner and make sure that both constructs are under the same promoter, so as to make sure that one promoter will not out-compete the other.

Discussion

Biolistic transfection is a physical means of transfecting cells via bombardment of living tissue with high velocity DNA coated gold particles. In particle mediated gene transfer, in general transfected cells result when the bullet comes to rest in the nucleus. While many different transfection methods exist, such as microinjection, lipofection, calcium-phosphate transfection, electroporation and viral transfection, biolistic transfection can offer a less labor intensive, and efficient alternative for transfecting cells that are not easily transfected using these other methods. In fact, it was first developed as a technique for gene transfer in plants since the presence of the cell wall made transfection difficult using preexisting methods¹. Similarly, biolistics has gained popularity in the field of neurobiology since post-mitotic neurons are notoriously difficult to transfect^{2,3}. In particular, it is widely recognized as the principle technique to be used in experiments aimed at assessing fine morphology of single neurons in intact brain slices. The methods described here provide a detailed and comprehensive explanation of how to perform biolistic transfection of cultured brain slices using a hand held gene-gun (the Helios Gene Gun system; Bio-Rad).

Biolistic transfection has become the preferred method for transfecting neurons in slice culture because it allows transfection of a sparse number of cells throughout the brain slice. In this way, individual neurons can be examined in isolation. Since this technique is particularly well suited for transfecting neurons in brain slice, it is often used in conjunction with 2-photon microscopy, since 2-photon excitation enables visualization of fluorescently labeled cells deep within light scattering tissue⁴. Indeed the high magnification images of single pyramidal neurons presented throughout this video were captured using a custom built 2-photon laser scanning microscope. In conclusion biolistic transfection is an efficient means of transfecting individual cells within the context of a high density of surrounding cells, and as such has proven extremely useful for fluorescently labeling individual neurons in neuronal slice culture.

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