



## Antiviral activities of extracts from Hong Kong seaweeds\*

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**Abstract:** We extracted six Hong Kong brown seaweed species with hot water for their antiviral properties. The cytotoxicity and antiviral activity of these extracts were tested by MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] method, cytopathic effect reduction assay, and plaque reduction assay. The antiviral effect was further determined by flow cytometric analysis. The results showed that most of these extracts inhibited the propagation of herpes simplex virus types 1 and 2 (HSV-1 and HSV-2) standard strains with very low cytotoxicity to the host cells. The extracts of *Hydroclathrus clathratus* and *Lobophora variegata* showed more potential anti-HSV activities than the extracts of the other four seaweeds. They also had moderate anti-respiratory syncytial virus (RSV) activities but could not inhibit influenza A virus. *Hydroclathrus clathratus* was further extracted by diluted acid and alkali and the antiviral effects of the extracts were also detected. The result showed that the hot water extract contained the main carbohydrate components that exhibited the antiviral activities against various strains of HSV, including the acyclovir-resistant strain. HI-3, a compound fractionated from this hot water extract, showed a dose-dependent anti-HSV activity in flow cytometric analysis and plaque reduction assay.

**Key words:** Hong Kong seaweed, *Hydroclathrus clathratus*, Antiviral activity, Herpes simplex virus (HSV), Flow cytometry  
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### INTRODUCTION

Viruses are responsible for many serious illnesses. Respiratory syncytial virus (RSV), for example, is a major cause of lower respiratory tract disease especially in infants, children and the elderly (Scott *et al.*, 2007). Despite much effort, it has not yet been possible to develop an effective vaccine against RSV and therapy for respiratory tract disease caused by RSV is still not available (Kneyber *et al.*, 2000; Becker, 2007). Even if such vaccines exist, problems may continue to arise since virus changes rapidly, so vaccine developed for it must be reformulated frequently. The nucleoside analog ribavirin is used currently in antiviral therapy against RSV and influenza virus, but the efficacy, value and safety of ribavirin remain to be clarified (de Clercq, 1996). Herpes virus

is a kind of ancient virus that causes different types of infections. Approximately 80% of the adult population worldwide are infected with herpes simplex virus (HSV) type 1 (HSV-1) and approximately 20% of them are also infected with HSV type 2 (HSV-2) (Chutkowski *et al.*, 2002). Until now, a number of nucleoside analogues, especially the guanosine analogue acyclovir, have been developed as antiherpetic agents (Alché *et al.*, 2002). The therapeutic limitation of these nucleoside analogues is that drug resistant strains develop readily through mutations in viral genes for thymidine kinase and/or polymerase. Therefore, the continuous search for new compounds as antiviral agents is urgently needed (Ammendolia *et al.*, 2007).

New types of antiviral agents from natural sources, especially those that have high efficacy on resistant mutant viral strains and low toxicity to host, are considered to be most promising. Products from marine organisms show many interesting activities. Their constituents are more novel than those of many

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land plants. Seaweeds have long been recognized as rich and valuable natural resources of bioactive compounds because of their various biological properties (Mayer and Lehmann, 2000). The water-soluble extracts of seaweeds have been shown to exhibit antiviral activity against a wide spectrum of viruses (Witvrouw and de Clercq, 1997). There are more than 200 species of seaweeds in Hong Kong coastal waters (Ang, 2005), but research on their antiviral activity is very limited (Zhu *et al.*, 2003). In this study, crude water extracts of six species of seaweeds from Hong Kong coastal waters were examined for their cellular toxicity and antiherpetic activity. Their inhibitory effects on RSV and influenza A virus were also tested. Aqueous extracts by ethanol, acid and alkali extracts of one of these species, *Hydroclathrus clathratus* (C. Agardh) M.A. Howe, were further evaluated for their antiviral activities as well.

## MATERIALS AND METHODS

### Seaweed samples

Six species of brown seaweeds, including *Codium sinuosa* (Mertens ex Roth), *Dictyota dichotoma* (Hudson) J.V. Lamouroux, *Hydroclathrus clathratus*, *Lobophora variegata* (Lamouroux) Womersley ex Oliveira, *Padina australis* Holmes and *Sargassum hemiphyllum* (Turner) C. Agardh, were collected from Hong Kong coastal waters in March, April and May, 2002. Voucher specimens of these materials were deposited for reference in the Marine Science Laboratory, the Chinese University of Hong Kong. These seaweeds were stored at  $-20^{\circ}\text{C}$  until use.

### Viruses, cells and positive control compounds

HSV-1, HSV-2 clinical strains and influenza virus A H1N1 strain were obtained from the Department of Microbiology, Prince of Wales Hospital, the Chinese University of Hong Kong. HSV-1 15577 strain (standard strain), HSV-2 8702 strain (standard strain), HSV-1 DM2.1 strain (acyclovir-resistant strain with thymidine kinase deficiency) and RSV Long strain were kindly provided by Dr. Spence H.S. Lee (Department of Microbiology and Immunology, Dalhousie University, Halifax, N.S., Canada). The viral titer was tested by the cytopathic end-point assay

(Burleson *et al.*, 1992) and expressed as 50% tissue culture infective dose (TCID<sub>50</sub>).

Vero cell (ATCC CCL-81, African green monkey kidney cell line), HEp-2 cell (ATCC CCL-23, human larynx epidermoid carcinoma cell line) and MDCK cell (ATCC CCL-34, Madin Darby canine kidney cell line) were used as the hosts for HSV, RSV and influenza A virus, respectively. All of the cells were cultured in growth medium (GM), the Eagle's minimum Essential medium (EMEM, GIBCO™, USA) supplemented with 10% (v/v) heat-inactivated fetal bovine serum (FBS, GIBCO™, USA) and incubated at  $37^{\circ}\text{C}$  in 95% humidified atmosphere and 5% (v/v) CO<sub>2</sub> incubator. Maintenance medium (MM) was the same EMEM but containing 1% (v/v) fetal bovine serum (FBS).

In antiviral assay, acyclovir (ACV, the Wellcome Foundation Ltd., England) and ribavirin, two clinically used medicines for treating viral infections, and dextran sulfate (DS, MW 10000, Sigma, USA), a sulfated polysaccharide with well-known antiherpetic properties isolated from seaweed, were used as positive control compounds.

### Extraction

The seaweeds were thoroughly washed with running water to remove salt and all visible epiphytes. They were put out to air-dry for 24 h and then minced with a blender separately. The extraction was carried out in boiling water for 2 h. After centrifugation and filtration, the supernatant was condensed and then lyophilized to obtain brown or dark brown crude water extracts.

The seaweed (*Hydroclathrus clathratus*) that showed the highest antiviral activity was chosen for further treatment. The crude water extract was precipitated by ethanol at the final concentration of 30% and 70% (v/v). The residue of water extract was divided into two parts and extracted by 0.1 mol/L of hydrochloric acid and 1 mol/L of sodium hydroxide, respectively, at  $4^{\circ}\text{C}$  and kept for overnight. After centrifugation, the supernatants were adjusted to about pH 7.0 and precipitated by ethanol with the final concentration of 80% (v/v). All precipitates were dissolved in distilled water, dialyzed and lyophilized, and four components were obtained. The antiviral activities of these compounds against HSV were then tested separately.

### Cellular toxicity test

The morphological changes of cells induced by test samples were observed under inverted microscope (de Clercq, 1985). Confluent cell monolayers were treated with the extracts in MM at a serial twofold dilution or only with MM (as cell control) at 37 °C for 3 d. A disruption of the cell monolayer was observed using an inverted microscope (Nikon CMM 214, Japan). The maximal non-cytotoxic concentration (MNCC) of the extract was determined by comparing the shape of the treated cells with that of the untreated control.

The cytotoxicity of seaweed extracts was tested by MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide] assay as previously described (Wang *et al.*, 2007a). After the cells were treated with extract solutions at various concentrations for 3 d, MTT (Sigma) solution [5 mg/ml in phosphate buffered saline (PBS)] was added (12 µl/well) and incubated for additional 4 h. The old medium and redundant MTT were replaced by acid-isopropanol (0.04 mol/L HCl in isopropanol, 150 µl per well) to dissolve the dark blue crystals. The absorbance of the solution was then read at 570 nm with a reference wavelength of 630 nm. The 50% cytotoxicity concentration (CC<sub>50</sub>) was estimated from the plots graphically.

### Cytopathic effect reduction assay

The antiviral activities of the six samples were first tested by cytopathic effect (CPE) reduction assay (Serkedjieva and Ivancheva, 1998). In brief, a serial twofold dilution of extracts in MM (50 µl per well) and equal volume of virus suspension (1 000 TCID<sub>50</sub>) were added to the confluent cell monolayers in a 96-well plate (IWAKI, Japan). The virus induced CPE was scored on Day 3 post infection (p.i.) for HSV and influenza A and Day 5 p.i. for RSV. The reduction of virus multiplication was calculated as the percentage of virus control: virus control (%) =  $CPE_{exp} / CPE_{virus\ control} \times 100$ . The 50% effective concentration (EC<sub>50</sub>) was estimated from the plots graphically.

### Plaque reduction assay

The antiviral activity of the aqueous extract against HSV was also determined by plaque reduction assay (PRA) and performed as described previously (Li *et al.*, 2005) but with some modification. Briefly, confluent Vero cells in a 12-well plate (IWAKI, Japan)

were infected with 80 plaque forming units per well (PFU per well) of HSV with or without (as control) the extract solutions. After adsorption at 37 °C for 1 h, residual inoculum was replaced with 1 ml of MM containing 0.6% (w/v) agarose and corresponding extract solution. These were incubated for 3 d to allow plaque formation. The EC<sub>50</sub> was then estimated from the plots graphically.

### Flow cytometric analysis

In this assay, Vero cells were infected by HSV in the presence of different concentrations of the carbohydrate fraction of algal extracts. All of the cells were then gathered, fixed and stained firstly with mouse anti-human HSV-1 (clone H62) or HSV-2 (clone HH-2) monoclonal antibody (Argène-Biosoft, Varihles, France), and then with FITC-conjugated goat anti-mouse IgG polyclonal antibodies (PharMingen, USA). The stained cells were then subjected to analysis with BECKMAN EPICS XL Flow Cytometry System (USA). Fluorescein-5-isothiocyanate (FITC)-fluorescence from the anti-HSV antibody was measured at photomultiplier 1 (PMT1, 525 nm) (Chi-Ming Chiu *et al.*, 2004).

## RESULTS

### Cellular toxic effect of extracts

When different cells were exposed to the extracts at the final concentration ranging from 31.25 µg/ml to 1 000 µg/ml for 72 h separately, the hot water extract of *S. hemiphyllum* showed the highest toxicity on Vero cells. By observing the morphological changes of cells, extracts from the other five algae had moderate to low cytotoxicity. The MNCC of these extracts were all higher than 250 µg/ml. The extracts of *D. dichotoma* and *P. australis* were only slightly toxic to Vero cells with high CC<sub>50</sub> values of 925 and 1 000 µg/ml, respectively (Table 1). The extracts of *H. clathratus* and *L. variegata* also showed low toxic effect on HEp-2 and MDCK cells, with the CC<sub>50</sub> values being higher than 500 µg/ml for both these cell lines.

### Antiviral activity assay

The six crude extracts were studied for their inhibitory effect on HSV replication in Vero cells firstly by measuring the CPE reduction. The monolayers of

Vero cells in 96-well plates were infected with HSV-1 and HSV-2 at the multiplicity of infection (MOI) of 1000 TCID<sub>50</sub>/ml in the absence or presence of serially twofold diluted crude extracts. The CPE induced by the virus was scored on Day 3 p.i.

**Table 1 Cellular toxicity of aqueous extracts of Hong Kong seaweeds on Vero, HEp-2 and MDCK cells**

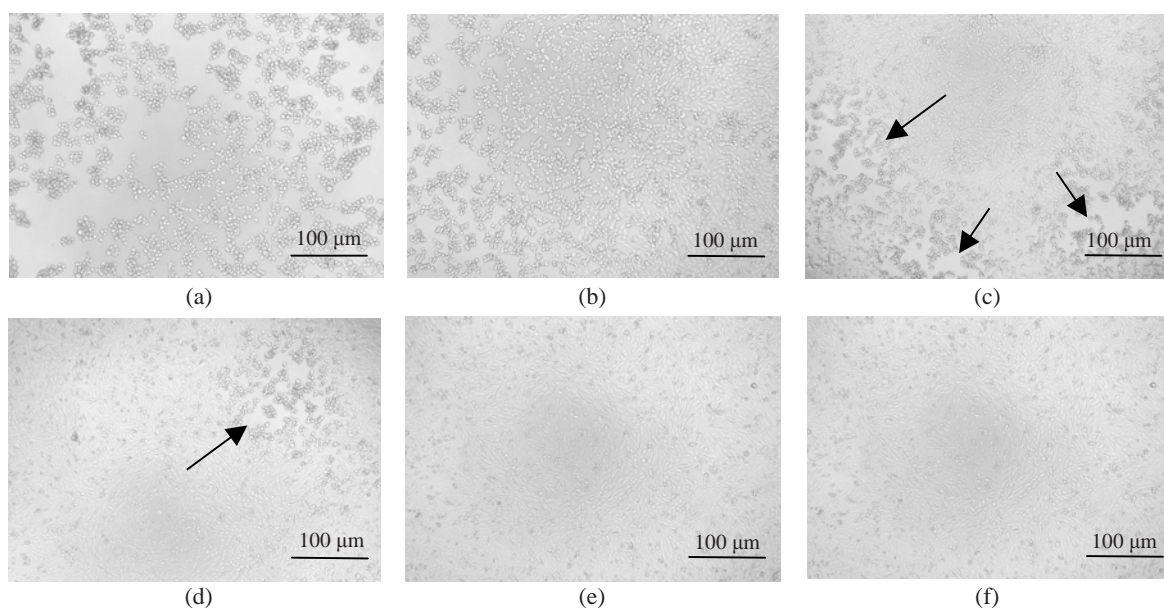
Seaweed species	MNCC (μg/ml)			CC <sub>50</sub> (μg/ml)		
	Vero cell	HEp-2 cell	MDCK cell	Vero cell	HEp-2 cell	MDCK cell
A	250	ND	ND	450	ND	ND
B	500	ND	ND	925	ND	ND
C	500	250	500	780	560	780
D	400	500	500	700	620	>1000
E	750	ND	ND	1000	ND	ND
F	62.5	ND	ND	125	ND	ND

A: *Colpomenia sinuosa*; B: *Dictyota dichotoma*; C: *Hydroclathrus clathratus*; D: *Lobophora variegata*; E: *Padina australis*; F: *Sargassum hemiphyllum*. MNCC: maximum non-cytotoxic concentration on cells; CC<sub>50</sub>: the concentration that reduced 50% of viable cells tested by MTT method; ND: not detected. All data were the means of two separate tests

All of the seaweed extracts could, in different degrees, protect the Vero cells from the infection of HSV-1 and HSV-2. The extract of *H. clathratus* had the highest antiviral effect on HSV-1 and HSV-2. This is followed by the extract of *L. variegata*. The others were moderate anti-HSV agents. When

observed under the inverted microscope, the CPE induced by HSV decreased after being treated by these extracts. Fig.1 shows the reduction of CPE induced by HSV-1 in Vero cells after being treated with various concentrations of *H. clathratus* extract for 3 d. The effect was concentration-dependent, with less and less CPE developed when compared with the viral control that showed 100% CPE (Fig.1a) as the extract concentration increased. At the dose of 25 μg/ml (Fig.1e), no CPE appeared and growth of the host cells was the same as that of the control cells (Fig.1f). Similar CPE reduction was also observed in Vero cells infected by HSV-2 and treated with *H. clathratus* extract (data not shown).

These results were confirmed and quantified by plaque reduction assay. Among the six seaweeds, the extract of *H. clathratus* had the highest anti-HSV activity with the lowest EC<sub>50</sub> values of 6.25 μg/ml against the standard strains of HSV-1 and HSV-2 (Table 2). Based on its low cytotoxicity and high antiviral effect, the selective index (SI) of *H. clathratus* extract was the highest. This is followed by the extract of *L. variegata* that showed higher antiviral effect especially against HSV-2 (Table 2). For its high cellular toxicity, the extract of *S. hemiphyllum* gave a low SI value indicating that its antiviral activity was related to its cytotoxicity to the host cells.



**Fig.1 CPE reduction assay showing the inhibitory effect of *H. clathratus* extract on HSV-1**

Vero cells were infected with HSV-1 15577 strain and treated with the test sample at the final concentrations of (a) 0 μg/ml (acted as viral control), (b) 3.1 μg/ml, (c) 6.25 μg/ml, (d) 12.5 μg/ml, and (e) 25 μg/ml. The cell control is exhibited in (f). The arrows point out distinct CPE induced by the virus

**Table 2 Anti-HSV activities of aqueous extracts of seaweeds tested by plaque reduction assay (PRA)**

Extracts of Seaweeds	$EC_{50}$ ( $\mu\text{g/ml}$ )		$SI$	
	HSV-1	HSV-2	HSV-1	HSV-2
A	22.1	12.5	20.4	36.0
B	24.3	25.0	38.1	24.3
C	6.25	<6.25	124.8	>124.8
D	18.5	9.0	37.8	77.8
E	58.9	40.0	17.0	25.0
F	19.1	12.5	6.5	10.0

A: *Colpomenia sinuosa*; B: *Dictyota dichotoma*; C: *Hydroclathrus clathratus*; D: *Lobophora variegata*; E: *Padina australis*; F: *Sargassum hemiphyllum*.  $SI$ : selective index, the value of  $CC_{50}/EC_{50}$ ; HSV-1, HSV-2: antiviral effect on standard strain of HSV-1 and HSV-2. All data were the means of three repeated tests

The extracts of *H. clathratus* and *L. variegata* also showed some inhibitory effect against RSV with the  $EC_{50}$  of 25 and 100  $\mu\text{g/ml}$ , respectively. But both of them had no anti-influenza A virus effect even at the concentration of 200  $\mu\text{g/ml}$  (Table 3).

**Table 3 Antiviral activities of extracts from *H. clathratus* and *L. variegata* on RSV and influenza A virus**

Viruses	$EC_{50}$ ( $\mu\text{g/ml}$ )			
	<i>H. clathratus</i>	<i>L. variegata</i>	DS	Ribavirin
RSV*	25	100	3.1	3.0
Influenza A*	NA	NA	ND	5.0

\*MOI=100 TCID<sub>50</sub>/ml; NA: no activity at concentrations up to 200  $\mu\text{g/ml}$ ; ND: not detected

The extracts of *H. clathratus* and *L. variegata* showed excellent antiherpetic activities, including the inhibitory ability on acyclovir (ACV)-resistant HSV-1 (DM2.1 strain) and the clinical strains. Moreover, these two crude extracts exhibited a slightly higher effect on HSV-2 than on HSV-1 (Table 4).

**Table 4 Antiviral activities of extracts from *H. clathratus* and *L. variegata* on various strains of HSV as tested by PRA method on Vero cells**

Viruses	$EC_{50}$ ( $\mu\text{g/ml}$ )			
	<i>H. clathratus</i>	<i>L. variegata</i>	DS	ACV
A	6.25	18.5	1.6	0.125
B	6.25	18.2	ND	ND
C	9.0	12.5	ND	0.125
D	<6.25	9.0	0.8	ND
E	<6.25	6.25	ND	ND

A: HSV-1 standard strain; B: HSV-1 DM2.1 strain; C: HSV-1 clinical strain; D: HSV-2 standard strain; E: HSV-2 clinical strain. The data were the means of two repeated tests; ND: not detected

*Hydroclathrus clathratus* was extracted by hot water, diluted acid and alkali solution, and four components were obtained. Two fractions precipitated from aqueous extract by 30% and 70% (v/v) ethanol were named as HI-2 and HI-3, respectively, and another two fractions HII-A and HIII-A were obtained from acid and alkali solution extracts, respectively.

The results listed in Table 5 show that most of the carbohydrates in *H. clathratus* could be extracted by hot water. This part also showed the highest antiviral effect against HSV-1 and HSV-2 when compared with the acid and alkali extracts. HI-3 precipitated by 70% (v/v) of ethanol from water crude extract had excellent inhibitory effects against both HSV-1 and HSV-2 with very low  $EC_{50}$  values at 1.6 and 0.8  $\mu\text{g/ml}$ , respectively (Table 5). This is comparable to that of the positive control compound DS (Table 4).

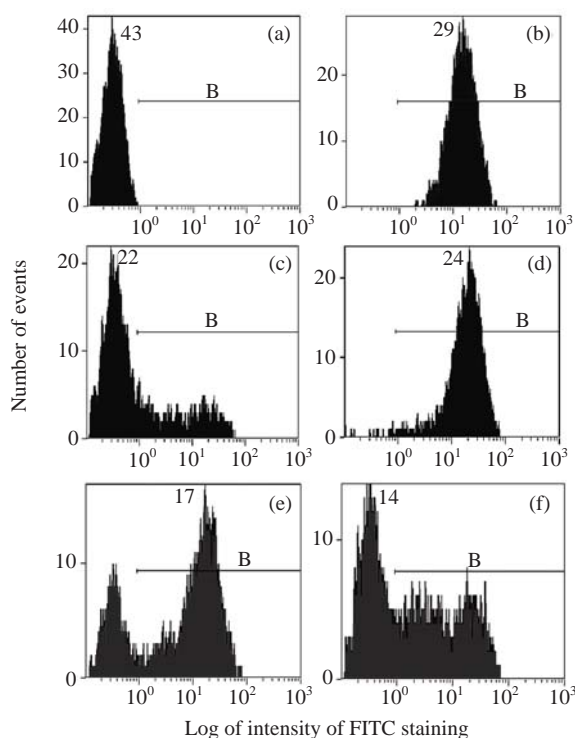
**Table 5 Inhibitory effects of crude water extract and other four fractions isolated from *H. clathratus* on plaque formation of HSV-1 and HSV-2 standard strain in Vero cells**

Fractions	Yield rate (%) <sup>*</sup>	$EC_{50}$ ( $\mu\text{g/ml}$ )	
		HSV-1 15577 strain	HSV-2 8702 strain
Water crude extract	2.030	6.0	3.1
HI-2	0.360	6.5	3.1
HI-3	0.780	1.6	0.8
HII-A	0.034	6.3	1.6
HIII-A	0.187	6.3	3.1

\*Yield rate was calculated based on wet weight of the seaweed samples

The efficacy value for HI-3 against HSV was also obtained from flow cytometric assay, and the FITC-fluorescence level from the anti-HSV antibody in the cells was measured. The  $EC_{50}$  values of HI-3 determined by flow cytometry were statistically similar to those obtained by plaque reduction assay. Fig.2 illustrates the typical histogram of fluorescence level in Vero cells exposed to HSV-1 15577 strain under the treatment with different concentrations of HI-3. Compared with the results of cell control (Fig.2a), virus control (Fig.2b) and the dextran sulfate positive control (Fig.2c), the replication of HSV-1 15577 strain was repressed by a dose-dependent response to HI-3 (Figs.2d~2f). HI-3 could therefore

protect the Vero cells from the infection of HSV. The higher the concentration of this extract presented, the less the amount of viral antigen could be detected in the infected host cells. HI-3 also showed an obvious dose-related antiviral effect against HSV-2 8702 strain when similarly tested by flow cytometry. The resulting histograms depicting FITC-fluorescence level from the anti-HSV-2 antibody are similar to those shown in Fig.2 and are therefore no longer shown here.



**Fig.2 Antiviral effect of HI-3 against standard strain of HSV-1 as analyzed by flow cytometric system**

These histograms depict FITC-fluorescence level from the anti-HSV-1 antibody in (a) cell control, (b) HSV-1 control, and HSV-1-infected Vero cells that had been treated with (c) 3.13  $\mu\text{g/ml}$  of dextran sulfate (positive control), or (d) 1.56  $\mu\text{g/ml}$  HI-3, (e) 3.13  $\mu\text{g/ml}$  HI-3, (f) 6.25  $\mu\text{g/ml}$  HI-3. In each histogram, B bar indicates the proportion of cells that were positive for the HSV-1 antigen

## DISCUSSION

Until now, chemically synthesized or modified compounds have been the major source of selective antiviral agents, particularly in the case of antiherpetic compounds (Alché *et al.*, 2002). For combating drug resistant viral strain, it has been suggested that the new antiherpes drugs should be non-nucleosides.

Thus, many efforts have been carried out to screen for antiviral agents from natural sources. Marine algae have shown their potential as important sources of antiviral as well as other bioactive compounds (Ahn *et al.*, 2002).

In this study, we examined hot water extracts of six species of brown seaweeds and found that they showed antiviral activities against herpes simplex virus and had low cytotoxicity to Vero cells, except *S. hemiphylum* extract. These extracts could inhibit the replication of various strains of HSV, including the ACV-resistant strain. Earlier works also showed that the aqueous extracts from five out of eight species of Hong Kong seaweeds exhibited some potential antiviral activities and very low cytotoxicity (Zhu, 2002). These studies indicate that the antiviral activities of these seaweed extracts were not due to their cytotoxic effect on host cells. Therefore, Hong Kong seaweeds could be a promising source of natural products with antiherpetic effect at non-toxic concentrations, just like seaweeds from other geographical regions.

It has been reported that seaweed extracts can be active against various enveloped viruses, including DNA and RNA viruses such as human immunodeficiency virus (HIV), HSV, cytomegalovirus (CMV), RSV and influenza virus (Baba *et al.*, 1988). In the present investigation, the best inhibitory effect on HSV was shown by the hot water extract of *H. clathratus*, followed by that from *L. variegata*. They also showed some anti-RSV activities in HEp-2 cells but could not inhibit influenza A virus in MDCK cells even at a high concentration (200  $\mu\text{g/ml}$ ). HSV is an enveloped DNA virus while RSV and influenza A are both enveloped RNA viruses. Influenza A contains segmented genome. Therefore, it seems that the extracts from these brown seaweeds could exhibit higher antiviral effect on enveloped DNA virus than on RNA virus, and had no effect on RNA virus with segmented genome, e.g., the influenza virus. However, Damonte *et al.*(1994) reported that the water extract of a red seaweed showed high inhibitory effect on RSV in HeLa cells and influenza A virus in MDCK cells with very low  $\text{EC}_{50}$  values. These findings suggest that there are possible differences in the virus-inhibitory effects of the seaweed extract. These differences may not only depend on the choice of cell lines used in the assay, but also on the seaweed species collected from different habitats, geographical

regions or even at different collecting time (Zhu, 2002).

All the six crude extracts in the current screening showed positive results in phenol-sulfuric acid reaction (data not shown), suggesting that the main effective components in these extracts were polysaccharides. As can be found in the literature, a number of polyanionic substances or water-soluble sulfated polysaccharides are potential antiviral agents (Béress *et al.*, 1993; Ponce *et al.*, 2003). It is commonly suggested that these negatively charged molecules disturb the proliferation cycle of enveloped viruses at the early stage, therefore inhibiting the viral infection (Witvrouw and de Clercq, 1997). However, Nakashima *et al.* (1987) found that a sulfated seaweed polysaccharide selectively inhibited reverse transcriptase (RT) enzyme of human immunodeficiency virus (HIV) and its replication *in vitro*. Other studies showed that sulfated polysaccharides could prevent viral protein synthesis (González *et al.*, 1987) or were capable of blocking various steps during the life cycle of HSV (Huleihel *et al.*, 2001). The pleiotropic modes of antiviral actions of sulfated polysaccharides make it less likely for the virus to develop resistant mutants.

The carbohydrates contained in seaweed could be extracted by water, diluted acid and alkali (Witvrouw and de Clercq, 1997). In the present experiment, these three parts of carbohydrates from *H. clathratus* were tested. They all showed antiviral activities against HSV-1 and HSV-2 and the largest portion, the hot water extract, has the most potential for developing into antiviral agent. HI-3, which was fractionated from the hot water extract, exhibited an excellent effect in antiviral test using both the plaque reduction and flow cytometric assays. Further fractionation of HI-3 yielded two different sulfated polysaccharides. These polysaccharides showed comparable antiviral activities. Detailed performances of these polysaccharides and their characterization were reported in Wang *et al.* (2007b). Some seaweed polysaccharides, e.g., dextran sulfate and heparin, have been shown to inhibit the replication of various enveloped viruses, including HSV and HIV (Witvrouw and de Clercq, 1997). The modes of action of most of these polysaccharides have often been attributed to a blockade of the early stages of the virus replication cycle (Bourne *et al.*, 1999). The polysaccharides isolated from *H. clathratus* could have the

same anti-HSV action as that shown by dextran sulfate, but more details of their antiviral mechanism are still being evaluated.

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