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Disparate effects of simvastatin on angiogenesis during hypoxia and inflammation

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Abstract

Aims—Studies have shown that some of statin's pleiotropic effects were achieved by either promotion or inhibition of angiogenesis, depending on the underlying disease. This study tested the hypothesis that the angiogenic potential of simvastatin is related to the microenvironmental conditions.

Main methods—Human umbilical vein endothelial cells (HUVEC) were studied after exposure to hypoxia or the inflammatory factors tumor necrosis factor (TNF)- α , with or without co-incubation with simvastatin (1 μ mol/L) and mevalonate. HUVEC angiogenesis was evaluated by tube formation, migration, and proliferation assays. Hypoxia inducible factor (HIF)-1 α , vascular endothelial growth factor (VEGF), Akt, endothelium nitric oxide synthase (e-NOS), and oxidative stress were evaluated.

Key findings—HUVEC angiogenesis increased during hypoxia (tube length 14.7 \pm 0.5 vs. 7.8 \pm 0.6 mm, $p < 0.05$) and further enhanced by simvastatin (19.3 \pm 1.1 mm, $p < 0.05$ vs. hypoxia alone), which downregulated the expression of the HIF-1 inhibitor PHD2 and upregulated HIF-1 α , VEGF, and Akt, without changing oxidative stress or eNOS. Incubation with TNF- α promoted HUVEC angiogenesis (7.4 \pm 0.2 vs. 6.5 \pm 0.2 mm, $p < 0.05$) with increased oxidative stress. However, simvastatin inhibited this promotion (2.5 \pm 0.3 mm, $p < 0.001$ vs. TNF- α alone) by decreasing oxidative stress, VEGF, Akt, and eNOS.

Significance—We conclude that at the same dosage, simvastatin can either promote or inhibit angiogenesis, possibly by activating upstream regulators of HIF-1 α in hypoxia, but conversely interfering with angiogenic signaling downstream to inflammation. These opposing angiogenic effects should be considered in the therapeutic strategies with statins.

Keywords

Simvastatin; hypoxia; angiogenesis; inflammation

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Introduction

Pathological angiogenesis is implicated in the pathogenesis of cancer and atherosclerosis (Virmani, et al.,2005), and often mediated by hypoxia. Hypoxia changes expression of angiogenic genes, like hypoxia inducible factor (HIF), which is one of the main regulators for the major angiogenic mediator, vascular endothelial growth factor (VEGF)(Semenza, et al., 2000). Alternatively, angiogenesis can be induced by inflammatory cytokines like tumor necrosis factor (TNF)- α (Vanderslice, et al.,1998), and both processes have been implicated in atherosclerotic plaque angiogenesis(Virmani, et al.,2005). Clinical studies indicate that 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitors (statins) possess cardiovascular protective properties that compliment their lipid lowering effect(Byington, et al.,2001,Guptha,1995,Sever, et al.,2003). Among other effects, the ability of statins to stabilize plaques(Luan, et al.,2003) and inhibit cancer(Moyad, et al.,2006) may be related to attenuation of angiogenesis(Dulak and Jozkowicz,2005). Indeed, high doses of statins have been shown to inhibit endothelial cell migration and angiogenesis(Urbich, et al.,2002,Weis, et al.,2002).

In contrast, studies have also shown that statins protect against cardiac ischemia-reperfusion injury of the heart(Wayman, et al.,2003,Wolfrum, et al.,2004) and stimulate the growth of new blood vessels in ischemic limbs(Kureishi, et al.,2000) or kidney(Chade, et al.,2006) of normocholesterolemic animals. This effect may be achieved in part by activation of the serine/threonine protein kinase Akt that regulates multiple angiogenic processes in endothelial cells (Wolfrum, et al.,2004), including the generation of nitric oxide (NO) and reactive oxidative species (ROS).

The disparate effects of statins on angiogenesis under different conditions may partly depend on their dose (Urbich, et al.,2002,Weis, et al.,2002) or on the endothelial cell type used in the experiment(Frick, et al.,2003). However, we have also shown in swine models that the same dose simvastatin enhanced angiogenesis in the ischemic kidney(Chade, et al.,2006), but decreased pathologic angiogenesis in early coronary atherosclerosis(Wilson, et al.,2002). Similarly, Sata et al(Sata, et al.,2004) have shown that in the same animal (and therefore the same dose) statins can augment collateral growth (arteriogenesis) in the ischemic hind limb without affecting tumor capillary density (neovascularization). However, the mechanisms underlying these incongruent effects of statins are not completely understood. Thus, the purpose of this study was to assess mechanisms of these disparate effects observed in vivo, and test the hypothesis that the angiogenic potential of statins depends partly on the microenvironmental conditions, and would therefore vary in angiogenesis induced by inflammation compared to hypoxia.

Methods

Human umbilical vein endothelial cells (HUVEC, PromoCell, Heidelberg, Germany) were cultured at 37°C in endothelial culture media. The use of HUVEC in a matrigel system is a well-established and commercially available experimental platform for angiogenesis-related studies (Morales, et al.,1995,Shimpuku, et al.,2000,Soeda, et al.,2000).

HUVEC were studied under normal conditions, as well as during hypoxia or during exposure to inflammatory mediators. Hypoxia was achieved through incubation at 37°C for 24h in a Modular Incubator Chamber (Billups-Rothenberg; Del Mar, CA) filled with 95% N₂ +5% CO₂. For the hypoxia study, HUVEC were divided into following groups: Normal (normal media), Hypoxia, Hypoxia+simvastatin (0.1, 1, and 10 μ mol/L, active sodium salt, Calbiochem, Germany)(Veillard, et al.,2006), Hypoxia+simvastatin+mevalonate (500 μ mol/L, Sigma) (Veillard, et al.,2006). Each group consisted of 3-6 replicates. Statins exert their effect by inhibiting HMG-CoA reductase, thereby decreasing mevalonate. Co-incubating with

mevalonate may therefore by-pass the inhibition created by statins, and reverse those effects which are mediated by this pathway. The concentration of simvastatin of 1 $\mu\text{mol/L}$ may be somewhat comparable to therapeutic plasma levels in patients (Barrett, et al., 2006), and one lower and one higher concentrations (0.1 and 10 $\mu\text{mol/L}$, respectively) of simvastatin were also tested to evaluate the dose response of the drug. The effects of simvastatin and mevalonate on HUVEC under normal condition were also evaluated. To simulate inflammatory conditions, HUVEC were studied after incubation with TNF- α (1 ng/ml, Cell Science, Canton, MA) at a clinically relevant concentration (Danis, et al., 1992, Fajardo, et al., 1992), concurrent TNF- α + simvastatin (1 $\mu\text{mol/L}$), or TNF- α + simvastatin + mevalonate (500 $\mu\text{mol/L}$). Angiogenesis was evaluated by HUVEC tube formation in matrigel, HUVEC migration, and proliferation, and by the expression of angiogenic factors like VEGF, its major regulator, HIF-1 α , and the regulators of HIF-1 α , prolyl hydroxylases domain (PHD), von Hippel-Lindau (VHL), and 26S proteasome. As an index of oxidative stress, in situ superoxide anion was assessed using dihydroethidium (DHE) staining. Angiogenesis-related signals were also evaluated by Akt, endothelium nitric oxide synthase (eNOS), and matrix metalloproteinase (MMP-9).

HUVEC Tube formation

The BD BioCoat™ angiogenesis system (96 wells, BD Biosciences, Bedford, MA) was used to evaluate HUVEC tube formation, as we have previously shown (Daghini, et al., 2007). The tube number and length were measured using a computer-aided image-analysis program (MetaMorph, Meta Imaging Series 6.3).

HUVEC Migration Assay

HUVEC migratory function was examined using a modified Boyden chamber technique, as we have shown before (Daghini, et al., 2007). The magnitude of HUVEC migration was evaluated by counting the migrated cells in 4 random ($\times 40$) microscope fields.

In vitro proliferation assay

The HUVEC proliferative activity was determined by a tetrazolium compound MTS assay (CellTiter 96 Non-Radioactive Cell Proliferation Assay; Promega, Madison, WI), which monitors the number of viable cells, as we have previously shown (Daghini, et al., 2007).

In situ superoxide anion production

Generation of superoxide anion in HUVEC was measured with DHE staining and fluorescence microscopy, following manufacturer's instructions (Daghini, et al., 2007). Briefly, RPMI 1640 without phenol red media containing DHE (2 $\mu\text{mol/L}$) was applied onto each plate, incubated for 30 minutes in a light-protected humidified chamber at 37°C, washed once by PBS, and evaluated by inverted fluorescence microscopy.

Western blotting

HUVEC were homogenized at 4°C, and standard Western blotting protocols were used, with specific antibodies against HIF-1 α , VEGF, PHD2, VHL (all 1:200, Santa Cruz, CA), 26S proteasome (1:500, Calbiochem), Akt (total and phosphorylated form, 1:1000, Cell Signaling), phospho-eNOS, total eNOS, p47phox (all 1:200, Santa Cruz, CA), MMP-9 (1:500, Chemicon International, CA), and actin (1:1000, Sigma) as loading control. The intensities of the protein bands were determined using densitometry. Protein expression was assessed relative to non-phospho form or actin, and expressed as ratio.

Statistical Analysis

Continuous data are expressed as mean \pm SEM. Multiple group comparisons utilized one-way analysis-of-variance followed by the Tukey Test. Statistical significance was accepted if $p \leq 0.05$.

Results

Simvastatin enhances hypoxia-induced angiogenesis by upregulating angiogenic factors

HUVEC tube formation and migration were increased after exposure to hypoxia, and further enhanced during co-incubation with simvastatin, an increase that was blocked by mevalonate (Figure 1). HUVEC proliferation was not affected by co-incubation with simvastatin (Figure 1). The two lower doses of simvastatin (0.1 and 1 μ mol/L) elicited similar enhancements of tube formation, while the highest dose (10 μ mol/L) abolished angiogenesis induced by hypoxia. Simvastatin and mevalonate had no effect on HUVEC tube formation under normoxic conditions. The enhancing pro-angiogenic effect of simvastatin in hypoxia was accompanied by upregulation of the expression of HIF-1 α and VEGF (Figure 2), along with further suppression of the HIF inhibitor PHD2. Interestingly, VHL, which also mediates HIF-1 α destabilization and degradation, was indeed downregulated by hypoxia, but restored by simvastatin. On the other hand, the 26s proteasome, which degrades HIF-1 α , was upregulated in hypoxia (possibly due to feedback mechanism), but normalized by simvastatin (Figure 2). In addition, hypoxic conditions increased p-Akt expression, which was further enhanced by simvastatin (Figure 2). There were no significant changes in p-eNOS expression, and MMP9 was similarly increased in HUVEC during hypoxia and unaffected by simvastatin (Figure 2). Furthermore, under hypoxic conditions there was no change of p47phox expression (Figure 2) or DHE fluorescence (Figure 3), indicating that oxidative stress may not be involved in the mechanism of hypoxia-induced angiogenesis.

Simvastatin inhibits TNF- α induced angiogenesis by attenuating oxidative stress

Incubation of HUVEC with TNF- α at a clinically relevant concentration (1 ng/ml) also promoted tube formation, migration and proliferation. Both tube formation and HUVEC proliferation were inhibited by incubation with simvastatin (Figure 4). The lower doses of simvastatin (0.1 and 1 μ mol/L) exerted similar inhibition of the tube formation, and the highest dose (10 μ mol/L) further inhibited it. TNF- α -promoted HUVEC tube formation was accompanied by increased VEGF, p47phox expression (Figure 5), and superoxide production (Figure 3), while HIF-1 α , PHD-2, and VHL remained unchanged (Figure 5). Simvastatin inhibited TNF- α -induced HUVEC angiogenesis (Figure 4) and decreased oxidative stress (Figure 3), although p47 expression remained elevated (Figure 5). Moreover, simvastatin decreased p-Akt and p-eNOS expression, and restored 26s expression that decreased by TNF- α , while MMP9 expression was not increased by TNF- α and remained unchanged in all the groups (Figure 5). The effect of simvastatin on tube formation, HUVEC proliferation, HUVEC migration, and p-Akt expression was reversed by mevalonate, indicating Akt-dependent mechanism involved in angiogenesis under inflammatory conditions.

Discussion

This study demonstrates that at the same dosage, simvastatin can either promote or inhibit angiogenesis, by modulating intrinsic mechanisms activated by the specific angiogenic trigger. This is likely achieved by activating upstream regulators of HIF-1 α in hypoxia, but conversely interfering with angiogenic signaling downstream to inflammation, such as oxidative stress. These results suggest that the angiogenic qualities of the statins depend on the underlying angiogenic stimulus, mechanisms, and microenvironmental conditions.

Both animal and clinical studies demonstrated the vascular protection afforded by statins beyond their lipid lowering effects(Callahan,2003,Napoli, et al.,2005,Susic, et al.,2003). Our previous studies demonstrated that simvastatin decreases injury in both hypercholesterolemia (Wilson, et al.,2003) and in the ischemic kidney in vivo(Chade, et al.,2006). The current study extends our previous observations and demonstrates that simvastatin also promotes angiogenesis when induced by hypoxia in vitro, but inhibits angiogenesis stimulated by the inflammatory factor TNF- α . Hence, the effect of simvastatin on angiogenesis appears to be microenvironment-dependent. This disparate effect might be important for design of therapeutic strategies in individual subjects.

Atorvastatin has been reported to exert pro-angiogenic effects at low concentrations, but inhibit blood vessel formation at high concentrations(Urbich, et al.,2002,Weis, et al.,2002). Our results are in agreement with these previous observations, and showed that the highest concentration of simvastatin used in this study (10 μ mol/L) inhibited angiogenesis induced by both hypoxia and inflammation, possibly due to cytotoxic effect of simvastatin at this very high concentration. However, in the present study we observed that using the lower dose simvastatin might exert either pro- or anti-angiogenic effect, depending on the angiogenic stimuli. This may imply that administration of a lower dose of simvastatin should be considered for ischemic disease, while a higher dose could be considered in subjects with tumor, diabetes, or atherosclerotic plaque angiogenesis. Interestingly, under hypoxic conditions atorvastatin at micromolar concentrations enhanced the transcriptional activity of the VEGF promoter, but neither VEGF mRNA nor protein(Loboda, et al.,2006), while we observed that a 1 μ M concentration of simvastatin up-regulated hypoxia-induced VEGF protein expression. The differential effect on VEGF expression might be due to the different statin preparation, cell types, and degree of the hypoxia, since HIF-1 binding activity increases only two-fold between 6-20% O₂ but ten-fold between 0.5-6%, and its maximal response is observed at 0.5% oxygen (Jiang, et al.,1996).

Hypoxia is involved in several vascular pathologies including cardiac and peripheral muscle ischemia, tumor development, or imbalanced wound healing. It increases the expression of many genes, particularly VEGF, a major angiogenic factor essential for neovascularization. We observed that simvastatin enhanced hypoxia-induced angiogenesis mainly by manipulating the regulation of HIF-1 expression, and thereby increasing VEGF expression. HIF-1 is regulated by oxygen sensors like PHD, which in normoxia hydroxylates the HIF-1 α subunits, thereby targeting them for proteasomal degradation. In contrast, acute hypoxia inhibits PHD, leading to HIF-1 α stabilization(Takeda, et al.,2007). Similarly, the important negative regulator VHL (Maxwell, et al.,2001) was also downregulated by hypoxia. VHL participates in the ubiquitin E3 ligase complex, and leads to HIF-1 α ubiquitination and degradation by the 26S proteasome(Brahimi-Horn and Pouyssegur,2005). The 26S proteasome binds polyubiquitin chains, unfolds the target protein, and feeds it into the proteolytic chamber that degrades it(Brahimi-Horn and Pouyssegur,2005). In the current study, protein expression of the 26s proteasome was increased in hypoxia, probably due to feedback mechanism, but not sufficiently to suppress the elevation in HIF-1 α protein. Interestingly, we observed that simvastatin restored VHL expression under hypoxic condition, yet enhanced HIF-1 α and VEGF expression, possibly by further suppressing PHD and by downregulating the 26s proteasome, and thereby overall inhibiting HIF-1 α degradation. Thus, simvastatin promotes angiogenesis under hypoxia mainly through regulating PHD and thereby HIF-1 and VEGF expression.

The current study also shows that simvastatin promoted angiogenesis in response to hypoxia by increasing Akt expression, which is an important mediator of the angiogenic activity of VEGF. Akt signaling axis is activated by a variety of stimuli (mainly growth factors like VEGF) in endothelial cells and regulates multiple critical steps in angiogenesis, including endothelial

cell survival, migration, and capillary-like structure formation. Furthermore, this signaling pathway also regulates cardiovascular homeostasis and vessel integrity at least in part by controlling nitric oxide synthesis(Shiojima and Walsh,2002). Nitric oxide also enhances the endothelial migration in vitro(Morbidelli, et al.,1996). Furthermore, growth factor activation of angiogenesis is dependent on proper endothelial cell–extracellular matrix attachment (Shiojima and Walsh,2002). Statin may also induce Akt-mediated phosphorylation of eNOS (Kureishi, et al.,2000), although phospho-eNOS expression was not upregulated under our experimental condition. Moreover, increased expression of MMP-9 under hypoxic condition may also contribute to HUVEC migration and proliferation(Romanic, et al.,2001), but did not appear to be involved in the effect of simvastatin.

In contrast to the findings during hypoxia, we observed that simvastatin inhibited angiogenesis in a pro-inflammatory environment achieved by TNF- α . TNF- α can directly induce angiogenesis(Fajardo, et al.,1992,Vanderslice, et al.,1998), and increases the expression of adhesion molecules and chemokines(Rajashekhar, et al.,2006) that can also promote angiogenesis(Hong, et al.,2005). Furthermore, under normoxic conditions TNF- α may also increase HIF-1 α and VEGF expression by activation of NF κ B (Jung, et al.,2003). Indeed, we observed that HUVEC incubation with TNF- α increased VEGF expression, as well as HUVEC tube formation, migration and proliferation, but had no effect on HIF-1 α , PHD, and VHL expression, indicating that TNF- α induced angiogenesis does not involve this pathway. Instead, TNF- α may induce angiogenesis through generation of ROS(Eligini, et al.,2005), which are important second messengers of angiogenic stimuli, underscoring our observation of increased superoxide anion generation, likely derived from NAD(P)H oxidase, since the expression of its p47phox subunit was increased. Simvastatin inhibited NAD(P)H oxidase derived ROS, and therefore may attenuate tube formation by its antioxidant effects(Haendeler, et al.,2004). In contrast to simvastatin-enhanced angiogenesis that involves activating Akt in the ischemic limb (Kureishi, et al.,2000) (as well as in this study under hypoxic conditions), simvastatin down-regulated Akt and eNOS under inflammatory conditions, probably by inhibiting the geranylgeranylation of the small GTP-binding protein Rho, leading to their inactivation(Park, et al.,2002).

The pleiotropic effects of simvastatin in our study involved activating Akt, HIF-1 α , and VEGF under hypoxic conditions, and down-regulated Akt, VEGF, and eNOS under inflammatory conditions. Some of these effects may be mediated by inhibiting the geranylgeranylation of the small GTP-binding protein Rho(Park, et al.,2002). Such effects are clinically relevant, as many of the effects of statins in humans exceed their capacity to reduce LDL cholesterol (Bonetti, et al.,2003). In addition, the effects of simvastatin on the expression of some proteins, such as p-eNOS and VEGF under inflammatory conditions, and VHL and 26s under hypoxic conditions, were not fully reversed by mevalonate. These results suggest that under those experimental conditions, the pleiotropic effects of statins on these proteins may be at least partly mediated through alternative routes, such as the PI3-kinase(Wolfrum, et al.,2004) or leukocyte function antigen-1(Weitz-Schmidt, et al.,2001) pathways. The observation that simvastatin did not increase eNOS is also underscored by a recent study which demonstrated that statin exerted eNOS-independent cardiac and renal protective actions(Yagi, et al.,2008).

Collectively, this study provides evidence that at low physiological concentration simvastatin promotes angiogenesis in response to hypoxia, likely by regulating the expression of HIF-1 and blunting its degradation. In contrast, under inflammatory conditions induced by TNF- α , simvastatin inhibits angiogenesis through oxidative stress-sensitive pathways. These data therefore provide new insight into the mechanisms by which simvastatin might be beneficial under different pathological situations.

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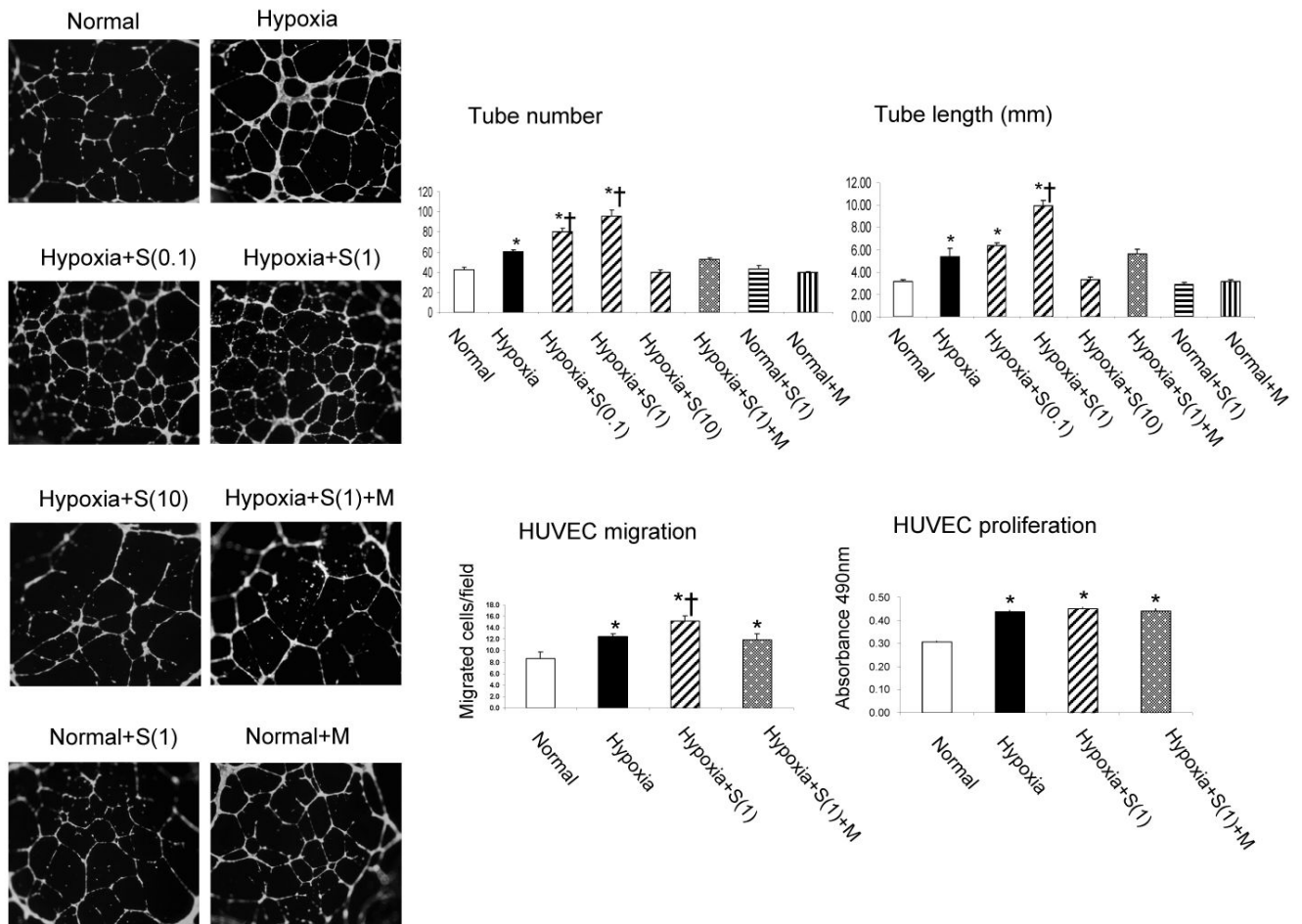


Figure 1. Angiogenesis of human umbilical vein endothelial cells (HUVEC) exposed to hypoxia, and the dose-dependent effects of simvastatin (S at 0.1, 1, and 10 μ mol/L). Left: Representative tube formation images (4 \times). Right: HUVEC tube formation (at 0.1, 1, and 10 μ mol/L), migration, and proliferation (at 1 μ mol/L) quantification. The increase in HUVEC tube formation suggests that low and medium doses of S (0.1 and 1 μ mol/L) further enhance angiogenesis triggered by hypoxia, while the higher concentration (10 μ mol/L) abolish this enhancement. Simvastatin and mevalonate (M) alone had no effect on HUVEC tube formation. * p <0.05 vs. normal, † p <0.05 vs. hypoxia,

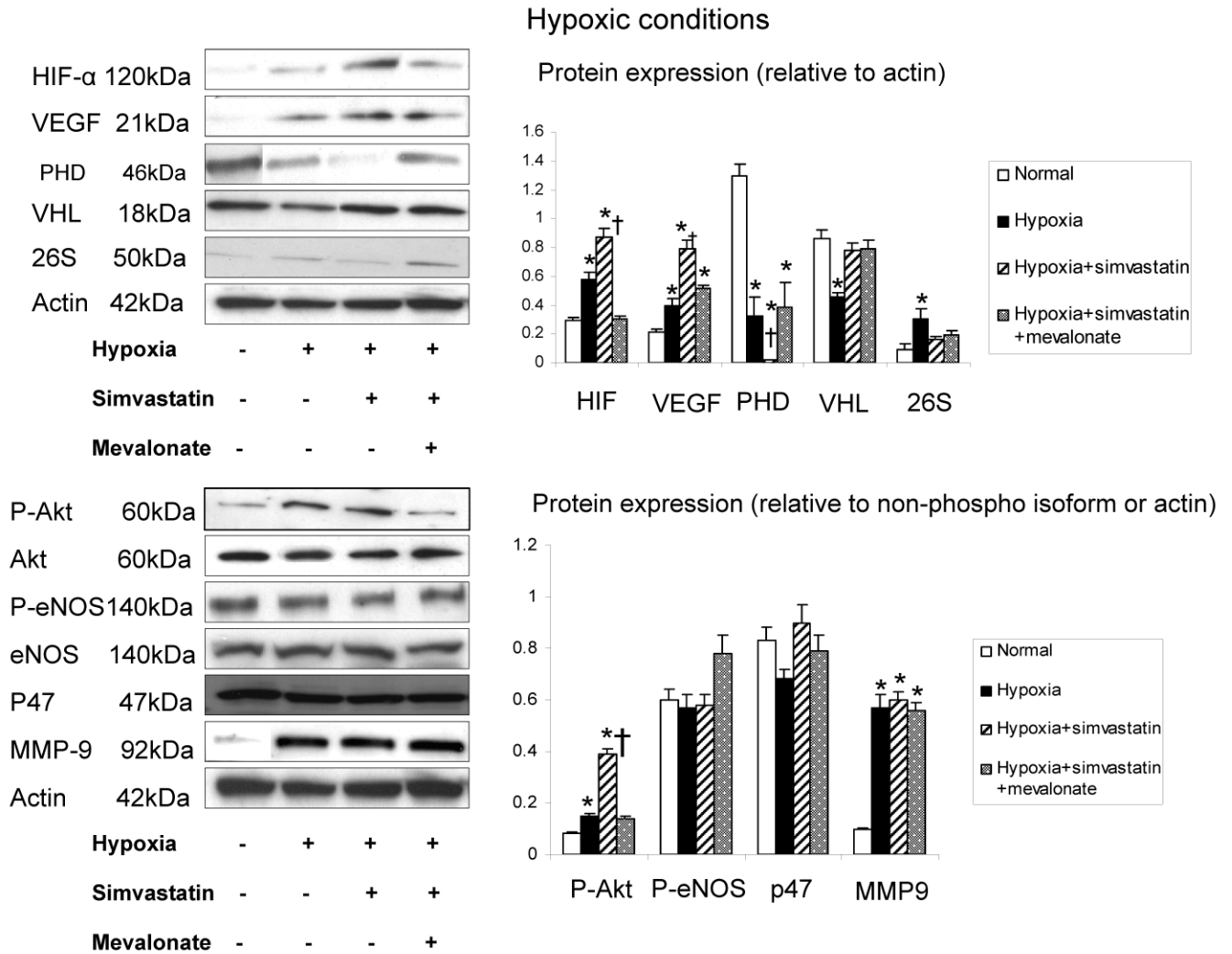


Figure 2. Densitometry and representative immunoblots of hypoxia inducible factor (HIF)-1 α , vascular endothelial growth factor (VEGF), prolyl hydroxylases domain (PHD)-2, VHL, 26s proteasome, phospho- and total Akt, NAD(P)H subunit p47phox, phospho- and total eNOS, and MMP-9 in HUVEC exposed to hypoxia with and without simvastatin (1 μ mol/L). Expression of HIF-1 α , VEGF, and p-Akt were increased under hypoxia and further enhanced by simvastatin. The changes of VHL and 26s under hypoxia were restored by simvastatin, while p47 and eNOS expression remained unchanged among groups. P-Akt and p-eNOS expression was assessed relative to their respective total proteins, while the others were normalized to actin, and all were expressed as ratio. *p<0.05 vs. normal, †p<0.05 vs. hypoxia

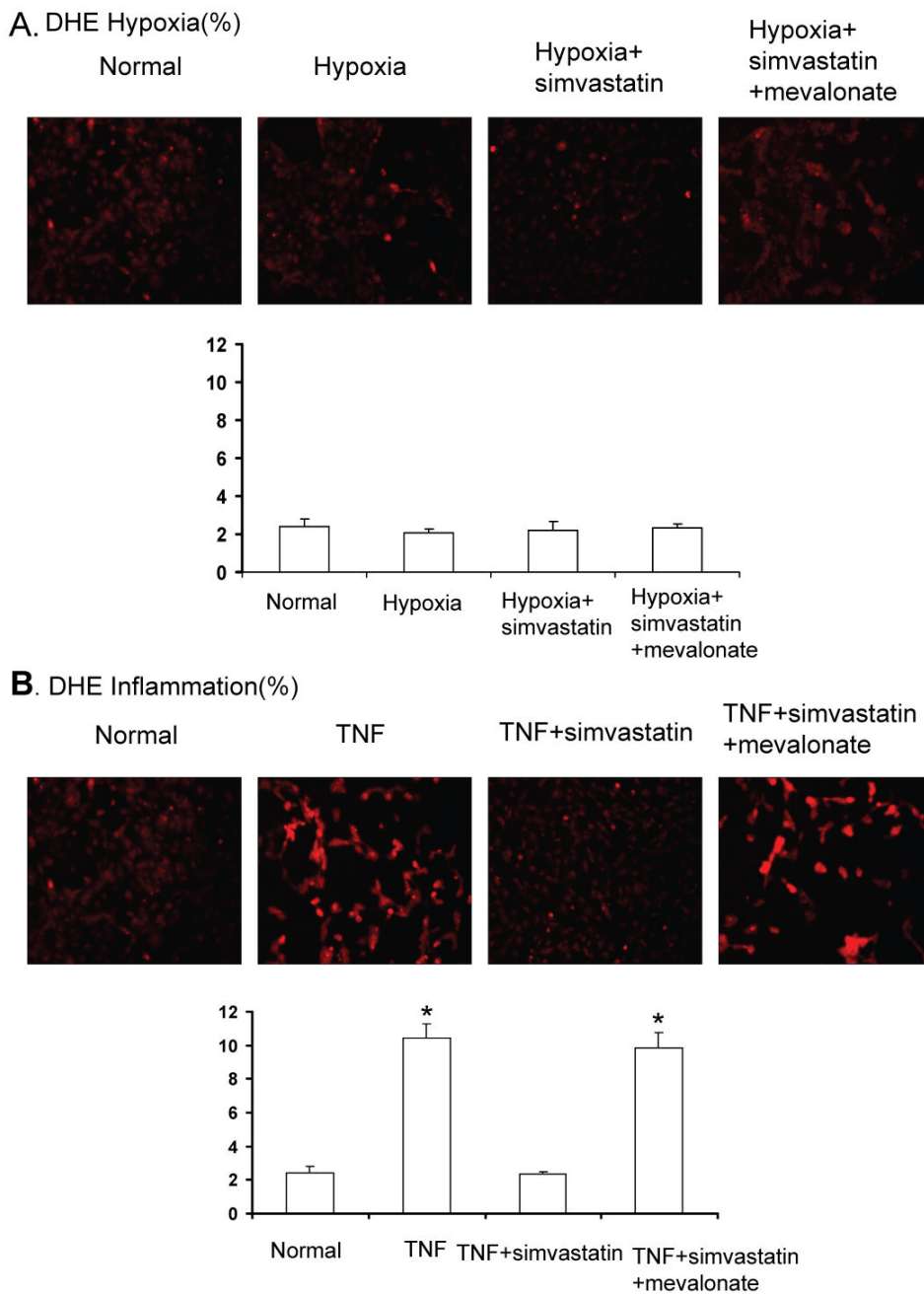


Figure 3.

A: Representative images and quantification of in situ superoxide generation (DHE staining) in HUVEC exposed to hypoxia, showing simvastatin (1 μ mol/L) did not affect DHE in any hypoxic groups. **B:** Representative images and quantification of DHE staining in HUVEC exposed to TNF- α , showing that simvastatin (1 μ mol/L) normalized oxidative stress in TNF- α treated HUVEC. * p <0.05 vs. normal

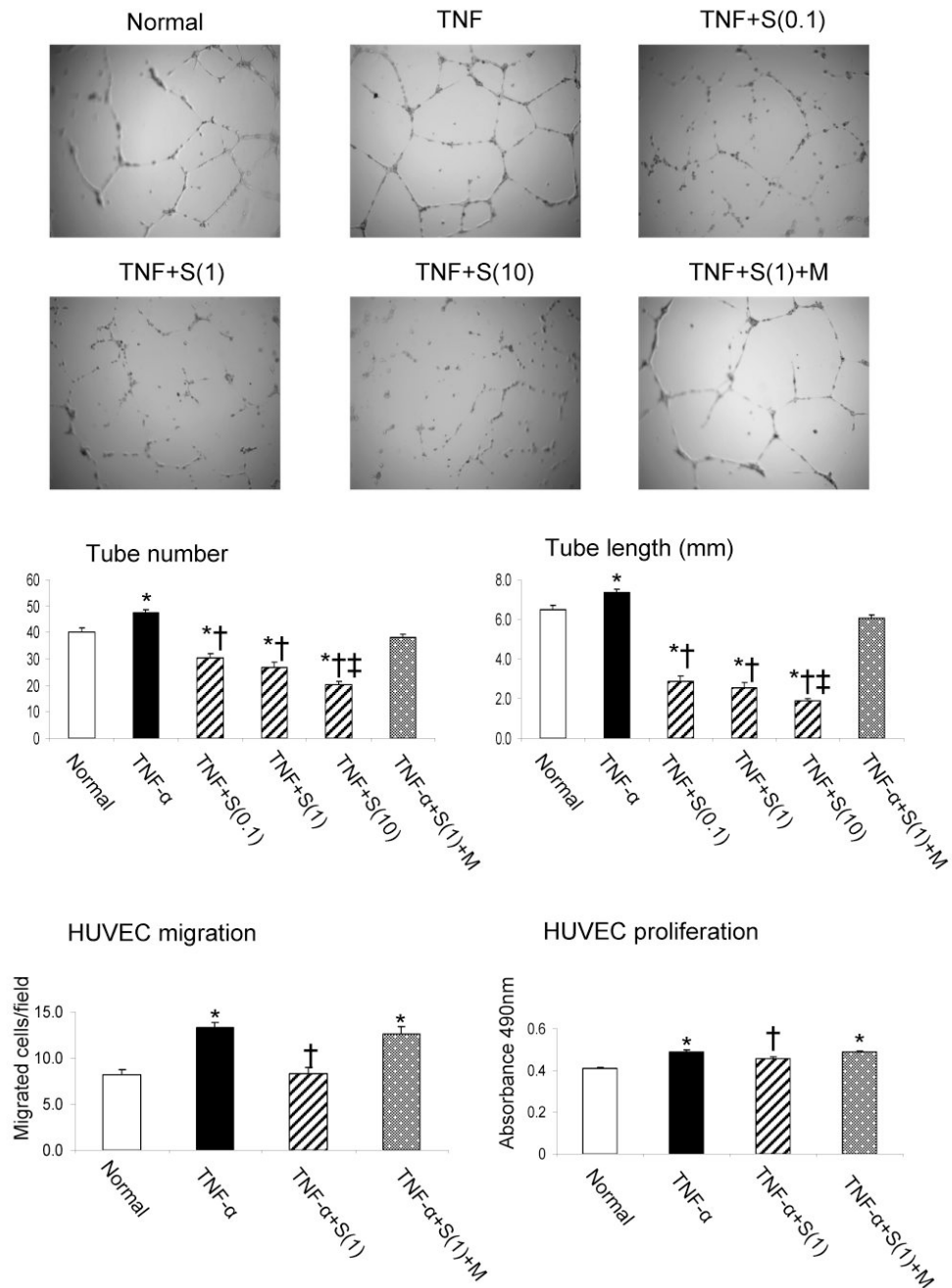


Figure 4.

Angiogenesis of human umbilical vein endothelial cell (HUVEC) exposed to TNF- α (1 ng/ml), and the dose-dependent effects of simvastatin (S, 0.1, 1, and 10 μ mol/L). Top: representative tube formation images (5 \times). Bottom: Quantification of tube formation (0.1, 1, and 10 μ mol/L), migration, and proliferation (1 μ mol/L) in HUVEC exposed to TNF- α and simvastatin. The inhibitory effect of simvastatin on the tube formation induced by TNF- α was similar at 0.1 and 1 μ mol/L, but further inhibited in 10 μ mol/L. This effect was reversed by mevalonate (M, co-incubated with 1 μ mol/L). * p <0.05 vs. Normal, † p <0.05 vs. TNF- α , ‡ p <0.05 vs. simvastatin at 0.1 and 1 μ mol/L

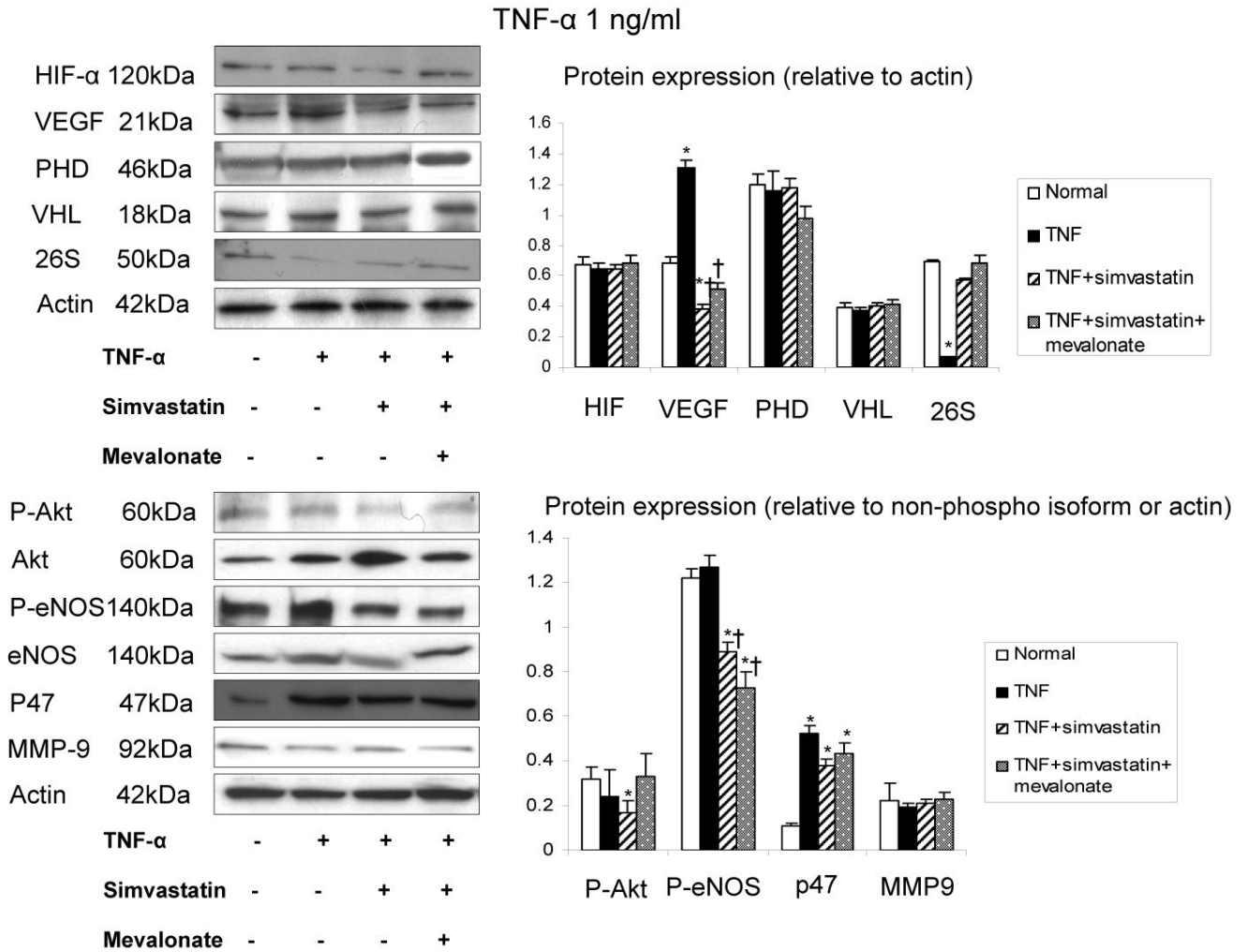


Figure 5. Densitometry and representative immunoblots of hypoxia inducible factor (HIF)-1 α , vascular endothelial growth factor (VEGF), prolyl hydroxylases domain (PHD)-2, VHL, phospho- and total Akt, NAD(P)H subunit p47phox, phospho- and total eNOS in HUVEC exposed to TNF- α (1 ng/ml) with and without simvastatin. Simvastatin decreased the expression of VEGF, p-Akt, and p-eNOS, restored 26s proteasome, and had no effect on HIF, VHL, and MMP-9 expression. *p<0.05 vs. normal, †p<0.05 vs. TNF- α .