ORIGINAL ARTICLE

Features associated with germline CDKN2A mutations: a GenoMEL study of melanoma-prone families from three continents

Alisa M Goldstein, May Chan, Mark Harland, Nicholas K Hayward, Florence Demenais, D Timothy Bishop, Esther Azizi, Wilma Bergman, Giovanna Bianchi-Scarra, William Bruno, Donato Calista, Lisa A Cannon Albright, Valerie Chaudru, Agnes Chompret, Francisco Cuellar, David E Elder, Paola Ghiorzo, Elizabeth M Gillanders, Nelleke A Gruis, Johan Hansson, David Hogg, Elizabeth A Holland, Peter A Kanetsky, Richard F Kefford, Maria Teresa Landi, Julie Lang, Sancy A Leachman, Rona M MacKie, Veronica Magnusson, Graham J Mann, Julia Newton Bishop, Jane M Palmer, Susana Puig, Joan A Puig-Butille, Mitchell Stark, Hensin Tsao, Margaret A Tucker, Linda Whitaker, Emanuel Yakobson, The Lund Melanoma Study Group, and the Melanoma Genetics Consortium (GenoMEL)

J Med Genet 2007;44:99-106. doi: 10.1136/jmg.2006.043802

See end of article for authors' affiliations

Correspondence to: Dr A M Goldstein, Genetic Epidemiology Branch/NCI/ NIH/DHHS, Executive Plaza South, Room 7004, 6120 Executive Blvd. MSC 7236, Bethesda, MD 20892-7236, USA; goldstea@ exchange.nih.gov

Received 10 May 2006 Revised 10 July 2006 Accepted 12 July 2006 **Published Online First** 11 August 2006 **Background:** The major factors individually reported to be associated with an increased frequency of CDKN2A mutations are increased number of patients with melanoma in a family, early age at melanoma diagnosis, and family members with multiple primary melanomas (MPM) or pancreatic cancer.

Methods: These four features were examined in 385 families with ≥3 patients with melanoma pooled by 17 GenoMEL groups, and these attributes were compared across continents.

Results: Overall, 39% of families had CDKN2A mutations ranging from 20% (32/162) in Australia to 45% (29/65) in North America to 57% (89/157) in Europe. All four features in each group, except pancreatic cancer in Australia (p = 0.38), individually showed significant associations with CDKN2A mutations, but the effects varied widely across continents. Multivariate examination also showed different predictors of mutation risk across continents. In Australian families, \geqslant 2 patients with MPM, median age at melanoma diagnosis \leqslant 40 years and \geqslant 6 patients with melanoma in a family jointly predicted the mutation risk. In European families, all four factors concurrently predicted the risk, but with less stringent criteria than in Australia. In North American families, only \geqslant 1 patient with MPM and age at diagnosis \leqslant 40 years simultaneously predicted the mutation risk.

Conclusions: The variation in CDKN2A mutations for the four features across continents is consistent with the lower melanoma incidence rates in Europe and higher rates of sporadic melanoma in Australia. The lack of a pancreatic cancer–CDKN2A mutation relationship in Australia probably reflects the divergent spectrum of mutations in families from Australia versus those from North America and Europe. GenoMEL is exploring candidate host, genetic and/or environmental risk factors to better understand the variation observed.

■he CDKN2A (MIM# 600160) gene is the major known high-risk cutaneous malignant melanoma (CMM) susceptibility gene. CDKN2A encodes two distinct proteins translated, in alternate reading frames (ARFs), from alternatively spliced transcripts. The α transcript encodes the p16 protein; the smaller β transcript specifies the alternative product p14ARF. Germline CDKN2A mutations have been observed in approximately 20-40% of melanoma-prone families from around the world.1 Several variables have individually been reported to be associated with an increased frequency of CDKN2A mutations, including increased number of patients with melanoma, early median age at melanoma diagnosis, the occurrence of pancreatic cancer in a family and the occurrence of multiple melanoma tumours in a patient.2-17 These four features have been assessed separately. However, it has not been previously possible to compare the four factors across geographical regions, nor to examine them simultaneously. Such an evaluation would facilitate interpretation of risks by clinicians from different continents and give a view on putative relationships between genotype and latitude of residence.

The Melanoma Genetics Consortium GenoMEL (http://www.genomel.org), comprising major familial melanoma

research groups from North America, Europe, Australia and the Middle East, conducted a study to explore the relationship between these four attributes and the presence of a CDKN2A mutation. All GenoMEL member groups with eligible families participated in this study. The resultant sample of 385 families, 150 of which had CDKN2A mutations, allowed simultaneous evaluation of the four features, as well as inspection of differences by continent.

METHODS

Patients and design

Seventeen GenoMEL centres participated in this study. Families with at least three confirmed patients with melanoma who were screened for mutations in *CDKN2A* (exons 1α , 2 and 3) were eligible for the study. Mutation evaluation, predominantly sequencing or denaturing high performance liquid chromatography followed by sequencing, was conducted at each centre. All families (n = 385) with and without identified mutations

Abbreviations: ARF, alternate reading frame; CMM, cutaneous malignant melanoma; MPM, multiple primary melanoma

100 Goldstein, Chan, Harland, et al

were included. CDKN2A mutations that altered the p16 protein were included in the evaluation; a subset of these mutations also influenced p14ARF.

Table 1 presents the number of families and total number of patients with melanoma by study centre. For all centres, written informed consent was obtained from the subjects before participation in the study under Institutional Review Board-approved protocols. The precise methods of ascertainment for the eligible families differed between groups. Details of the participating families from each centre are described elsewhere (see table 1 for references). All melanoma diagnoses were confirmed by review of histological materials, pathology reports, medical records or death certificates. Only patients with confirmed melanomas (invasive or in situ) were included. This restriction may have resulted in differences for some centres in reported numbers of families presented previously. For each family, the absence/presence and type of CDKN2A mutation were reported. Other variables included number of patients with CMM in each family, age at first melanoma diagnosis for each patient, whether or not a patient with CMM had one versus multiple melanoma tumours, and the number of patients with melanoma or first-degree relatives of patients with melanoma in each family who had pancreatic cancer (81% confirmed).

Statistical analysis

Using the data provided, we derived four factors for analysis: number of patients with CMM in a family (#CMM/family), number of patients with CMM in a family with multiple primary melanoma (MPM) tumours, median age at CMM diagnosis in a family (MedAge), and number of patients with pancreatic cancer in a family, combining the reports of pancreatic cancer in patients with melanoma and in first-degree relatives of patients with melanoma. These four factors were evaluated across all 17 GenoMEL groups (total) and within specific geographical regions defined by continent (Australia, Europe and North America). In addition, the four attributes were also individually assessed in the European groups, for which there were sufficient numbers of families (ie, Sweden (Lund+Stockholm), Mediterranean Europe (Barcelona+Genoa+Emilia-Romagna), UK (Leeds+Glasgow),

The Netherlands (Leiden) and France (Paris)). The nonparametric Wilcoxon-Mann-Whitney test, Fisher's exact, Jonckheere–Terpstra or Kruskal–Wallis tests, as implemented in the computer programme StatXact (StatXact-4, V.4.0.1), were used to evaluate each of the four factors individually. Multivariate evaluation of the four factors in all families (total) and in families from Australia, Europe or North America was conducted using unconditional logistic regression as implemented in the program Stata (Stata 8.2, V.8.2). It was not possible to evaluate the four factors jointly in groups smaller than that in a continent. For the logistic regression analysis, the presence/absence of a CDKN2A mutation was the dependent variable. We thus measured the association between the "risk" of a CDKN2A mutation and the four factors using backward and forward stepwise logistic regression analyses. In addition, the final models were evaluated using likelihood ratio tests. All statistical tests were two sided.

RESULTS

Table 1 presents the number of families, number of patients with melanoma and number of families with CDKN2A mutations by participating centre. There were 385 families with 1720 patients with CMM in this study. Overall, 39% of families (n = 150) had mutations. The frequency of mutations ranged from 20% (32/162) in Australia to 45% (29/65) in North America to 57% (89/157) in Europe. Table 2 shows the CDKN2A mutations by continent and whether the mutation altered p14ARF.

Figures 1–4 show the percentage of families with CDKN2A mutations by number of patients with CMM/family (fig 1), MPM (fig 2), pancreatic cancer (fig 3) and MedAge (fig 4) for each of the four groups (total, North America, Australia and Europe). The frequency of mutations increased significantly as the number of CMM/family increased in each of the four comparison groups (fig 1, p values ranging from p<0.001 to p=0.004). In addition, among the mutation-positive families, there were significant differences in the distribution of the number of patients with melanoma per family across continents (p=0.002). In North American and Australian families, the largest increase in mutation frequency was from those families with 5 patients with CMM/family to those with \geq 6 CMM/family

Table 1 Number of families, patients with cutaneous malignant melanoma and families with CDKN2A mutations by study centre and summarised by country/continent and overall (total)

Centre	Country/continent	Total number of families	Total number of patients with CMM	Number of families with CDKN2A mutations	References for family ascertainment
Boston	USA/North America	3	9	0	18
Leeds	USA/North America	1	3	0	29
NCI	USA/North America	38	192	19	19 20
Philadelphia	USA/North America	5	18	1	
Utah	USA/North America	11	150	5	21
Toronto	Canada/North America	7	22	4	
North America	·	65	394	29	
Barcelona	Spain/Europe	11	38	5	22–25
Emilia-Romagna	Italy/Europe	8	27	i	26
Genoa	Italy/Europe	8	31	5	11
Glasgow	Scotland/Europe	10	31	6	9 27
Leeds	England/Europe	29	107	22	28-30
Leiden	The Netherlands/Europe	26	154	19	31-32
Paris	France/Europe	40	133	19	6 33-34
Lund	Sweden/Europe	15	60	8	12 35
Stockholm	Sweden/Europe	10	36	4	36-38
Europe	2.1.2.2.1, 22.2,62	157	617	89	
Brisbane	Australia	100	446	18	39-42
Sydney	Australia	62	258	14	10 28 43-44
Australia		162	704	32	
Tel-Aviv	Israel/Asia	1	5	0	
Total	•	385	1720	150	

CMM, cutaneous malignant melanoma.

Location of mutation	p16 amino acid or CDKN2A base change	p14ARF amino acid change	Europe (n = 89)	Australia (n = 32)	North America (n = 29)
5' UTR	c34G→T	None	1	1	<u>5</u>
Exon					
1α	c.9_32del24	None		1	
1α	c.18_19ins6	None	1	'	
1α	c.32_33ins9-32	None	2	2	2
1α	p.W15X	None	2	1	2
				i	
1α	c.46delC	None	1		
1α	p.L16P	None	1	2	
1α	p.L16R	None	_		1
1α	c.52_57dup6	None	1		
1α	p.G23D	None	1		
1α	p.G23R	None	1		
1α	p.R24P	None	<u>5</u> T	3	
1α	p.L32P	None	Ī	3	
1α	p.G35A	None	1	3 3 1	
1α	p.A36P	None		i	
1α	p.P38R	None	1		
1α	p.P48L	None	i		
1α	p.P48T	None	i		
			'	1	
1α	p.149S	None		1	
1α	p.Q50R	None		1	
2	p.V51F	p.G65V		1	
2	p.M53I	p.D68H	<u>6</u>	<u>5</u>	<u>3</u> 1
2	c.167–197del31	p.1-70p14:70-156p16	_	_	
2	p.R58X	p.P72L			1
2	p.V59G	p.S73R	2		
2	p.L62P	p.A76A	1		
2	p.L63P	p.A77A	1		
2	p.L65P	p.A79A	i		
2	p.G67R	p.R81P	2		
2			1	1	
	p.G67S	p.R81Q	i	1	
2	p.A68L	p.R82L	•		
2	p.E69G	p.G83G	1		
2	p.N71K	p.L86M	2		
2	p.N71S	p.Q85Q		1	1
2	c.225_243del19	p.1-90p14:82-156p16	<u>18</u>	1	1
2	c.240_253del14	p.T95fs	T		1
2	p.R87P	p.P101P			1
2	p.L97R	p.P111P	1		
2	p.R99P	p.P113P	i		
2	p.G101W	p.R11 <i>5</i> L	10		4
2	c.307_308del2	p.A117fs			4 T
2				1	
	p.D108N	p.R122Q	1		
2	p.R112G	p.P126R	1	1	
2	p.R112_L113insR	p.S127-A128insS	11		
2	p.L113L / p.P114S	p.A128T/ p.A128A	Ţ		
2	p.A118T	p.G132D	1		
2	c.358delG	None	1		
2	p.V126D	None	1		4
2	p.D153spl	p.1-65p14:154-156p16			<u>4</u> 1
Intron					
2	c.IVS2-105A→G	None	<u>6</u>	<u>3</u>	1
2	c.IVS2+1G→T	p.1-65p14:154-156p16	-	-	i

(from 4/9 (44%) to 12/15 (80%) and from 6/24 (25%) to 12/25 patients with (48%), respectively). By contrast, in Europe, the greatest increase in mutation frequency was among families with \geq 4 patients with CMM/family (77%) relative to those with 3 patients with CMM/family (42%). Although all European groups showed patterns consistent with the overall Europe result, only families from Sweden showed a significant association between a mutation and number of patients with melanoma per family (p = 0.001).

The frequency of mutations increased significantly as the number of patients with MPM in a family increased for all groups (fig 2, p<0.001). We also found significant differences in MPM distribution across continents in mutation-positive families (p=0.012). The presence of one patient with MPM produced a dramatic increase in mutation frequency for North

American (0 patients with MPM: 3/30, $10\% \ v \ge 1$ patient with MPM: 26/35, 74%) and European families (0 patients with MPM: 18/63, 29% $v \ge 1$ patient with MPM: 71/94, 76%) The MPM patterns observed in individual European groups were all similar, but significant associations between MPM and mutations were observed only for families from Sweden (p<0.001), France (p = 0.003) and Mediterranean Europe (p = 0.006). By contrast, for Australian families, a striking increase in the mutation frequency required ≥ 2 patients with MPM (≤ 1 patient with MPM: 13/122, $11\% \ v \ge 2$ patients with MPM: 19/40, 48%).

The relationship between pancreatic cancer and CDKN2A mutations showed differences across groups (fig 3). Overall (total), 72% of families with one reported patient with pancreatic cancer had mutations (31/43) and 81% of families with \geq 2 patients with pancreatic cancer had mutations (13/16).

102 Goldstein, Chan, Harland, et al

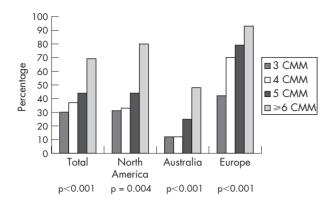


Figure 1 Percentage of families with CDKN2A mutations by number of patients with cutaneous malignant melanoma per family (no. of CMM/family: 3, 4, 5 and ≥6) for each of the four groups (total, North America, Australia and Europe). p Values are shown for each of the four groups.

There was a significant association between pancreatic cancer and mutations in families from the total (p<0.001), North America (p = 0.02) and Europe (p<0.001). However, pancreatic cancer was not associated with mutations in Australia (p = 0.38). Only three of nine Australian families (33%) with pancreatic cancer had mutations. By contrast, for North American and European families, \geq 75% of families with at least one patient with pancreatic cancer had mutations (9/12 and 32/38 families, respectively). In the European families, the strongest evidence for an association between pancreatic cancer and CDKN2A mutations came from The Netherlands (p = 0.006), France (p = 0.007) and Sweden (p = 0.04).

Figure 4 shows the relationship between median age at melanoma diagnosis in a family (MedAge) and CDKN2A mutations using quartiles (<34, 34–40, >40–50 and >50 years) defined among the total. All groups showed a significant decrease in the frequency of mutations as MedAge increased. Again, in the families positive for CDKN2A mutation, there was a significant difference in the distribution of MedAge across continents (p = 0.002). The significance resulted from an older median age at melanoma diagnosis in European mutationpositive families compared with either Australian (p = 0.002) or North American (p = 0.03) families. For European families, a mutation frequency >60% was observed when MedAge was ≤50 years. For Australian families, however, a high mutation frequency (>35%) required an earlier median age at melanoma diagnosis (ie, ≤40 years). North American families showed a pattern intermediate between Europe and Australia, with a step-function decrease in mutation frequency as MedAge increased.

We compared the distributions of #CMM/family, MPM, pancreatic cancer and MedAge across the most frequent CDKN2A mutations, defined as mutations that occurred in ≥5 families (c.225 243del19, p.M53I, p.G101W, p.R112-L113insR, c.IVS2-105A>G, p.R24P, c.-34G>T, c.32 33ins9-32 and p.V126D). At least 70% of families with each of these mutations had ≥1 patient with MPM. For seven of the nine mutations, most of the families had early MedAge. Only p.R112–L113insR and c.-34G>T had <50% of families with MedAge ≤40 years. The percentage of families with ≥5 patients with CMM/family ranged from 20% to 80%, with five of nine frequent mutations having <40% of families with ≥5 patients with CMM/family. However, there were no significant differences in #CMM/family (p = 0.14), MPM (p = 0.40) or MedAge (p = 0.14) across the nine most frequent mutations. By contrast, the distribution of pancreatic cancer differed significantly across these mutations (p<0.001). No families with *p.M531* (0/14), *c.IVS2-105A>G* (0/10) or

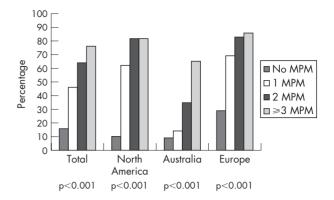


Figure 2 Percentage of families with CDKN2A mutations by number of patients with multiple primary melanoma (MPM: 0, 1, 2 and ≥3) in a family for each of the four groups (total, North America, Australia and Europe). p Values are shown for each of the four groups.

c.32_33ins9-32 (0/6) mutations had pancreatic cancer; 25–36% of families with p.R24P (2/8), c.-34G>T (2/7) or p.G101W (5/14) mutations had pancreatic cancer; finally, pancreatic cancer was observed in ≥60% of families with p.R112-L113insR (7/11), c.225_243del19 (12/20) or p.V126D (3/5) mutations. The distribution of pancreatic cancer in families with the most frequent CDKN2A mutations suggested an association with p14ARF (41% of families with a mutation that altered p14ARF had pancreatic cancer v 19% of families with a mutation that did not affect p14ARF). However, the observed pattern was not completely consistent. p.M53I, a mutation that alters p14ARF had no pancreatic cancer in 14 families; similarly, 60% of families with p.V126D had pancreatic cancer, yet this mutation does not alter p14ARF. Additional studies are needed to further evaluate this relationship.

Table 3 shows the multivariate predictors of a CDKN2A mutation for all families (total) and by continent. Given the significant differences observed between continents, we did not constrain the exposed and unexposed/referent categories in the multivariate analysis of the four features to be identical in the continent-specific analyses. There were substantial differences observed across continents. In all groups (total), MedAge ≤ 50 years, ≥ 1 patient with MPM and ≥ 1 patient with pancreatic cancer in a family were significant joint predictors of a CDKN2A mutation. However, #CMM/family was not an independent predictor for mutation risk. For the Australian families, ≥ 6 patients with melanoma in a family, ≥ 2 patients with MPM and MedAge ≤ 40 years simultaneously

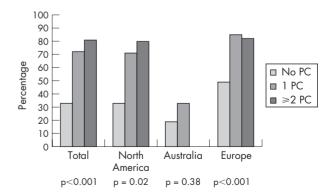


Figure 3 Percentage of families with CDKN2A mutations by number of patients with pancreatic cancer $(0, 1 \text{ and } \ge 2)$ in a family for each of the four groups (total, North America, Australia and Europe). p Values are shown for each of the four groups.

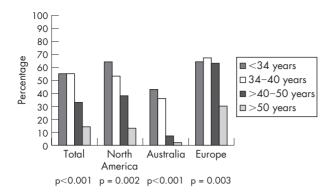


Figure 4 Percentage of families with CDKN2A mutations by median age (in years) at melanoma diagnosis (MedAge: <34, 34–40, >40–50 and >50 years) in a family for each of the four groups (total, North America, Australia and Europe). p Values are shown for each of the four groups.

predicted the risk of a mutation. As was observed in the univariate analyses, pancreatic cancer in a family did not influence the risk for a CDKN2A mutation. For North American families with the smallest sample size of 65 families, only MedAge \leq 40 years and \geq 1 patient with MPM were concurrent predictors of a CDKN2A mutation. Finally, for European families, all four factors jointly predicted the mutation risk (table 3). However, the levels for MedAge (\leq 50 years) and #CMM/family (\geq 4) were less stringent than that seen in the other comparison groups.

DISCUSSION

GenoMEL explored the relationship between select risk factors and presence of a CDKN2A mutation in 385 families with at least three confirmed patients with CMM. Individual examination of each of the four attributes showed that for all comparison groups examined (total, North America, Australia and Europe), there were significant associations between CDKN2A mutations and number of patients with melanoma in a family, the occurrence of multiple primary melanoma tumours and median age at melanoma diagnosis in a family. For pancreatic cancer, all groups except Australia showed significant associations with mutations. The frequencies of mutations and the effects of the four attributes, however, varied significantly across continents. Similarly, joint evaluation of the four features showed different predictors of mutation risk across the geographical areas.

Melanoma incidence rates vary widely among Caucasian populations around the world. The incidence rates are highest in Australia (38.5/100 000 for men; 29.5/100 000 for women), intermediate in North America (16.4/100 000 for men; 11.7/ 100 000 for women) and generally lowest in Western Europe (7.3/100 000 for men; 10/100 000 for women; CANCERMondial: http://www-dep.iarc.fr).45 Differences in the amount of exposure to ultraviolet radiation, the predominant environmental risk factor for melanoma and variation in host characteristics (eg, hair colour, eye colour, melanocytic nevi, freckling and skin type) may contribute to the wide geographical variation in melanoma incidence rates. In this study, the lowest frequency of CDKN2A mutations was observed in Australia (20%), the area with the highest incidence rates. By contrast, the highest frequency of mutations was observed in Europe (57%), the region with the lowest incidence rates. The underlying difference in incidence rates for melanoma between Australia and Europe is further reflected by the categorisation of the four features in the multivariate analyses. In Australian families (n = 162) in which higher rates of sporadic melanoma occur and fair skin and intense sun exposure predominate, ≥ 2 patients with MPM, MedAge ≤ 40 years and ≥6 patients with melanoma in a family were needed to concurrently predict CDKN2A mutation risk. For the 157 European families, the factor levels were less stringent (ie, broader), consistent with the lower melanoma incidence rates. Specifically, for the European families, only one patient with MPM, only ≥4 patients with melanoma in a family and a median age at melanoma diagnosis up to 50 years, in addition to ≥1 patient with pancreatic cancer, jointly predicted the risk of a CDKN2A mutation. The variation in frequency of CDKN2A mutations by number of patients with melanoma in a family, presence of patients with multiple primary melanoma tumours or pancreatic cancer, and median age at melanoma diagnosis across geographical regions may reflect different distributions of host, genetic or environmental risk factors.

In the univariate analyses, pancreatic cancer was significantly associated with CDKN2A mutations in all regions studied except Australia. As smoking is a well-known risk factor for pancreatic cancer,⁴⁶ reported differences in smoking patterns with lower rates in Australia compared with most of Western Europe⁴⁷ may contribute to some of the differences seen. However, the rates of smoking in Sweden are lower than that observed in Australia, and pancreatic cancer is strongly associated with CDKN2A mutations in families from Sweden.¹² In addition, there is little difference in pancreatic cancer incidence rates across the regions involved in the current study.

Table 3 Predictors of a CDKN2A mutation from simultaneous evaluation of the four factors (MedAge, multiple primary melanomas, pancreatic cancer, no of cutaneous malignant melanomas/family) across the groups (Total, Australia, North America and Europe)

	Level of variable [OR (95% CI)]					
Variables in final model	Total	Australia	North America	Europe		
MedAge (years)	≤ 50	≤ 40	≤ 40	≤ 50		
MPM	[7.42 (3.44 to 16.02)] ≥1	[19.62 (5.31 to 72.46)] ≥2	[4.04 (1.06 to 15.32)] ≥1	[5.15 (1.74 to 15.21)] ≥1		
PC	[5.73 (3.40 to 9.68)]	[6.23 (2.20 to 17.60)]	[5.30 (2.42 to 11.65)]	[6.24 (2.72 to 14.32)] ≥1		
rc	≥1 [7.54 (3.42 to 16.67)]	_	_	[8.21 (2.39 to 28.24)]		
#CMM/family	_	≥6 [6.65 (1.76 to 25.06)]	_	≥4 [2.44 (1.05 to 5.64)]		
Number of families	385	162	65	157		

CMM, cutaneous malignant melanoma; #CMM/family, number of patients with CMM per family; MPM, multiple primary melanomas; PC, pancreatic cancer; dash indicates that the variable did not appear in the final model.

Only the levels of the features and corresponding OR and 95% CI that were significantly associated with the risk of mutation are shown for each group (final model).

The age-standardised incidence rates across the regions range from 6-8/100 000 for men to 5-7/100 000 for women.⁴⁵ Alternatively, different distributions of mutations observed in Australia versus other areas may influence the association with pancreatic cancer. Most of the common mutations observed in families from Australia (p.M531, c.IVS2-105A>G, p.R24P or p.L32P) have generally shown low frequencies of pancreatic cancer even in non-Australian populations. In the current study, no Australian families with p.M53I (n = 5), c.IVS2-105A>G (n = 3) or *p.R24P* (n = 3) mutations had pancreatic cancer. Conversely, two of three families with p.L32P had pancreatic cancer. Further evaluation of the relationship between these common mutations and pancreatic cancer showed that no non-Australian families with p.M53I (0/9) or c.IVS2-105A>G (0/7) had pancreatic cancer. Yet, 40% (2/5) of families with p.R24P from the UK (2/3) and France (0/2) and 1/1 family from the UK with p.L32P had pancreatic cancer. In addition, the CDKN2A mutations (p.R112-L113insR, c.225 243del19, p.V126D and p.G101W) showing the strongest associations with pancreatic cancer in this and previous studies (summarised in Goldstein⁵) were essentially absent from Australia. Only one Australian family in the current study had any of these four mutations. Thus, the lack of a pancreatic cancer-CDKN2A mutation relationship in Australian families may reflect the spectrum of mutations observed in Australia versus Europe and North America. This finding needs to be confirmed.

The current study had several limitations. The ascertainment and sampling of families at most of the GenoMEL centres was not population-based, and each centre obtained data on extended family members to different degrees. Additionally, the study sample was restricted to families with at least three patients with melanoma, thereby selecting for families at higher risk of melanoma compared with either populationbased cases or unselected case series.17 48 The divergent ascertainment across study sites could, in consequence, have produced variation in the distribution of the four factors of interest. Nonetheless, in the families without CDKN2A mutations, there were no significant differences in the distribution of median age at melanoma diagnosis (p = 0.54), patients with multiple primary melanoma tumours (p = 0.39) or pancreatic cancer (p = 0.24) across the families from North America, Europe or Australia. However, the number of patients with melanoma in a family varied, with European families having significantly fewer patients with melanoma than Australian (p<0.001) and North American (p = 0.009) families. This difference probably reflects the lower incidence rates of melanoma in Western Europe, and consequently fewer families with large numbers of patients with melanoma. Moreover, it was not possible to fully evaluate the four attributes in areas smaller than a continent. Although continent was selected as the geographical region under study, there is variation in melanoma incidence rates, particularly in the European centres that were part of this study. As such, a continent may not adequately separate families with different risks of melanoma and hence, distributions of risk factors. Finally, as the study was restricted to melanoma-prone families with at least three confirmed patients with CMM, the results from the study may not be applicable to families with only one or two patients with

Additional studies are needed to expand the findings from the current study to a wider spectrum of patients with melanoma by evaluating simultaneously the four features examined here in patients with melanoma unselected for a positive family history. A clinic-based study from Boston and Toronto¹⁶ assessed the predictors of a CDKN2A mutation in unselected patients with melanoma, and showed that number of patients with melanoma in a family, median age at

melanoma diagnosis and multiple melanoma tumours in a patient were the most important predictors for a mutation. These findings are generally consistent with those observed in the current study. However, given the different joint predictors of mutation risk observed in families from each of the three continents, larger sample sizes and greater geographical diversity are needed to extend these findings to multiple geographical regions. Furthermore, comparison of results from multiple-case families (eg, the current study) and geographically comparable population-based or unselected series of patients could extend the interpretation of risks to different geographical regions and advance the understanding of the relationships between genotype and latitude of residence.

By working together, GenoMEL has produced a dataset to allow simultaneous evaluation of four factors previously individually associated with an increased frequency of CDKN2A mutations. Identification of such features enhances our ability to understand the differential risks that contribute to melanoma. The analyses showed differences in the frequency of CDKN2A mutations according to the number of patients with melanoma in a family, the occurrence of multiple CMM tumours in a patient, early median age at melanoma diagnosis in a family, and the occurrence of pancreatic cancer in a family across geographical regions. The differences may reflect distinct host, genetic or environmental risk factors. GenoMEL is exploring candidate risk factors to better understand the variation observed.

ACKNOWLEDGEMENTS

We thank the participating families, whose generosity and cooperation have made this study possible. We also thank the nurses, doctors, scientists and other health professionals who referred or evaluated patients with melanoma and families for this study.

Authors' affiliations

Alisa M Goldstein, Maria Teresa Landi, Margaret A Tucker, Genetic Epidemiology Branch, Division of Cancer Epidemiology and Genetics, National Cancer Institute, NIH, DHHS, Bethesda, Maryland, USA May Chan, Mark Harland, D Timothy Bishop, Julia Newton Bishop, Linda Whitaker, Genetic Epidemiology Division, Cancer Research UK Clinical Centre, Leeds, UK

Nicholas K Hayward, Jane M Palmer, Mitchell Stark, Queensland Institute of Medical Research, Brisbane, Queensland, Australia

Florence Demenais, Valerie Chaudru, INSERM, U794, Université d'Evry, Evry, France

Esther Azizi, Department of Dermatology, Sheba Medical Center, Tel Aviv University, Tel Aviv, Israel

Wilma Bergman, Nelleke A Gruis, Department of Dermatology, Leiden University Medical Center, Leiden, The Netherlands

Giovanna Bianchi-Scarra, William Bruno, Paola Ghiorzo, Department of Oncology, Biology, and Genetics, University of Genova, Genova, Italy Donato Calista, Dermatology Unit, Maurizio Bufalini Hospital, Cesena, Italy

Lisa A Cannon Albright, Department of Medical Informatics, University of Utah School of Medicine, Salt Lake City, Utah, USA

Agnes Chompret, Service de Génétique, Institut Gustave Roussy, Villejuif, France

Francisco Cuellar, Susana Puig, Dermatology Department, Melanoma Unit, Hospital Clínic de Barcelona, IDIBAPS (Institut de Investigació Biomèdica August Pi Suñe), Barcelona, Spain

David E Elder, Department of Pathology and Laboratory Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, USA

Elizabeth M Gillanders, Inherited Disease Research Branch, National Human Genome Research Institute, National Institutes of Health, DHHS, Baltimore, Maryland, USA

Johan Hansson, Veronica Magnusson, Department of Oncology-Pathology, Karolinska Institute and Karolinska University Hospital Solna, Stockholm, Sweden

David Hogg, Departments of Medicine and Medical Biophysics, University of Toronto, Toronto, Ontaria, Canada

Elizabeth A Holland, Richard F Kefford, Graham J Mann, Westmead

Institute for Cancer Research, University of Sydney at Westmead Millennium Institute, Sydney, New South Wales, Australia

Peter A Kanetsky, Department of Biostatistics and Epidemiology and Center for Clinical Epidemiology and Biostatistics, University of Pennsylvania, Philadelphia, Pennsylvania, USA

Julie Lang, Rona M MacKie, Department of Medical Genetics, University of Glasgow, Glasgow, UK

Sancy A Leachman, Department of Dermatology and Huntsman Cancer Institute, University of Utah School of Medicine, Salt Lake City, Utah, USA Joan A Puig-Butille, Genetics Service, Melanoma Unit, Hospital Clínic de Barcelona, IDIBAPS (Institut de Investigació Biomèdica August Pi Suñe), Barcelona, Spain

Hensin Tsao, Wellman Center for Photomedicine, Department of Dermatology and Cancer Center, Massachusetts General Hospital, Boston, Massachusetts. USA

Emanuel Yakobson, Molecular Cell Biology Laboratory, Department of Internal Medicine C, Sheba Medical Center, Sackler Faculty of Medicine, Tel-Aviv University, Tel-Aviv, Israel

The Lund Melanoma Study Group, Lund Cancer Center Department of Oncology, University Hospital, Lund, Sweden

Funding: This research was supported in part by the intramural research programme of the National Institutes of Health, NCI, DCEG and NHGRI. This work was also partially supported by research grants 1 RO1 CA83115-01A2 ''Genetic epidemiology of melanoma'' from the National Cancer Institute (DEE); RO1 CA88363 from NCI (NH), National Health and Medical Research Council of Australia (Brisbane group); R01 CA102422 from NCI (LACA), the Tom C Mathews Jr Familial Melanoma Research Clinic, the Huntsman Cancer Foundation (Utah group); Cancer Research UK (Leeds group); grants 01/1546 and 03/0019 from Fondo de Investigaciones Sanitarias, V2003-REDC03/03 and /07, a personal grant to Francisco Cuellar CONACYT, Personal Scholarship 152256/158706, Mexico City, Mexico (Barcelona group); FIRB RBNE0149752001 and COFIN 2004/061840_003 (GB-S; Genoa group); The Swedish Cancer Society: research grants 4860-B05-03XAC, 5012-B05-01PAF, The Swedish Research Council: research grant K2006-74X-20141-01-3 and grants from The Radiumhemmet Research Funds (JH; Stockholm group); Programme Hospitalier de Recherche Clinique Régional d'Ile de France grant AP-HP AOR 01 091 (Marie-Francoise Avril); Research grants and scholarships of the Australian National Health and Medical Research Council (G J M and R F K), The Cancer Councils of Australia, the Australian Cancer Research Foundation, the Melanoma Foundation and Cancer Research Fund of the University of Sydney (Sydney group). P A K was supported by a National Cancer Institute Preventive Oncology Academic Award (K07 CA80700). Some data collection for this publication was assisted by the Utah Cancer Registry supported by NIH Contract NO1-PC-35141, SEER Program, with additional support from the Utah Department of Health and the University of Utah. Partial support for all datasets within the Utah Population Database (UPDB) was provided by the University of Utah Huntsman Cancer Institute.

Competing interests: None.

Authors from Lund Melanoma Study Group:

Lund Cancer Center Department of Oncology, University Hospital, Lund, Sweden (Christian Ingvar, Ake Borg, Johan Westerdahl, Anna Masback, Hakan Olsson)

Additional authors from GenoMEL listed by group/centre:

Barcelona: Dermatology Department (Josep Malvehy) and Genetics Service (Celia Badenas, Remedios Cervera), Melanoma Unit, Hospital Clínic de Barcelona, IDIBAPS, Barcelona, Spain; Dermatology Department, Hospital Arnau de Vilanova, Universitat de Lleida, Lleida, Spain (Rosa Martí); Genetic Counseling of Cancer, Hospital Sant Joan, Universitat Rovira i Virgili, Reus, Spain and ICO, Hospital Trueta, Girona, Spain (Joan Brunet-Vidal); Boston: Massachusetts General Hospital, Boston, MA (Guang Yang); Brisbane: Queensland Institute of Medical Research (Nicholas Martin, David Whiteman, Adele Green) and Queensland Cancer Fund (Joanne Aitken), Brisbane, Australia; Emilia-Romagna: Dermatology Unit, Maurizio Bufalini Hospital, Cesena, Italy (Paola Minghetti); Genoa: Dipartimento di Oncologia, Biologia e Genetica, Universita degli Studi di Genova, Vle Benedetto XV, 6, 16132 Genova, Italy (Michela Mantelli, Lorenza Pastorino, Sabina Nasti, Sara Gargiulo), Istituto Nazionale per la Ricerca sul Cancro (IST), Largo Rosanna Benzi, 10, 16132 Genova, Italy (Sara Gliori); Leeds: Genetic Epidemiology Division, Cancer Research UK Clinical Centre, Leeds (Sushila Mistry, Juliette Randerson-Moor); Leiden: Leiden University Medical Center (Femke A de Snoo, Jeanet AC ter Huurne,

Jasper van der Rhee, Leny van Mourik, Frans van Nieuwpoort), STOET, The Netherlands Foundation for the Detection of Hereditary Tumors (Clasine van der Drift), Leiden, The Netherlands; Paris: Service de Génétique, Institut Gustave Roussy, Villejuif (Brigitte Bressac-de Paillerets), AP-HP-Hopital Cochin, Université Réné Descartes, Paris (Marie-Francoise Avril), French Hereditary Melanoma Study Group: Drs F Grange, B Sassolas, F Boitier, J Chevrant-Breton, C Lasset, C Dugast, P Vabres, France; Philadelphia: Department of Genetics (Arupa Ganguly), Department of Dermatology (Michael Ming), Department of Pathology and Laboratory Medicine (Patricia Van Belle), University of Pennsylvania, Philadelphia, PA, USA;

Stockholm: Department of Oncology-Pathology, Karolinska Institute and Karolinska University Hospital Solna, Stockholm, Sweden (Anton Platz, Suzanne Egyhazi, Rainer Tuominen, Diana Linden); Sydney: Westmead Institute for Cancer Research, Sydney, Australia (Helen Schmid);

Tel-Aviv: Department of Dermatology (Alon Scope, Felix Pavlotsky), The Susanne Levy Oncogenetics Unit (Eitan Friedman), Sheba Medical Center, Sackler Faculty of Medicine, Tel-Aviv University, Israel; Utah: Department of Dermatology and Huntsman Cancer Institute, University of Utah School of Medicine, Salt Lake City, UT (Mark Eliason).

Copyright Title 17 U.SC § 105 states that copyright protection is not available for any work of the United States Government. Since my authorship contribution was done as part of my official duties as a National Institutes of Health (NIH) employee, my work is a work of the United States Government and as such is in the public domain. If the Publisher intends to disseminate the work in foreign countries, the Publisher may secure copyright to the extent authorized under the domestic laws of those foreign countries. The copyright will be subject to a paid-up, nonexclusive, irrevocable worldwide license to the United States in the manuscript of such copyrighted work to reproduce, prepare derivative works, distribute copies to the public and perform publicly and display publicly the work, and to permit others to do so. The Publisher should not pay royalty income for work done by Federal employees as part of their official duties.

This agreement shall be governed and construed in accordance with Federal law as interpreted by the Federal courts in the District of Columbia.

REFERENCES

- Hayward NK. Genetics of melanoma predisposition. Oncogene 2003;22:3053-62.
- 2 Bergman W, Watson P, deJong J, Lynch HT, Fusaro RM. Systemic cancer and the FAMMM syndrome. Br J Cancer 1990;61:932–6.
- 3 Bergman W, Gruis N. Correspondence: familial melanoma and pancreatic cancer. N Engl J Med 1996;334:471.
- 4 Goldstein AM, Fraser MC, Struewing JP, Hussussian CJ, Ranade K, Zametkin DP, Fontaine LS, Organic SM, Dracopoli NC, Clark WH Jr, Tucker MA. Increased risk of pancreatic cancer in melanoma-prone kindreds with p16^{INKA} mutations. N Engl J Med 1995;333:970–4.
- 5 Goldstein AM. Familial melanoma, pancreatic cancer and germline CDKN2A mutations. Hum Mut 2004;23:630 [Mutation in Brief #718].
- 6 Soufir N, Avril M-F, Chompret A, Demenais F, Bombled J, Spatz A, Stoppa-Lyonnet D, the French Familial Melanoma Study Group, Benard J, Bressac-de Paillerets B. Prevalence of p16 and CDK4 germline mutations in 48 melanomaprone families in France. Hum Mol Genet 1998;7:209–16.
- 7 Kefford RF, Newton-Bishop JA, Bergman W, Tucker MA on behalf of the Melanoma Genetics Consortium. Counseling and DNA testing for individuals perceived to be genetically predisposed to melanoma: a consensus statement of the Melanoma Genetics Consortium. J Clin Oncol 1999;17:3245–51.
- Monzon J, Liu L, Brill H, Goldstein AM, Tucker MA, From L, McLaughlin J, Hogg D, Lassam NJ. CDKN2A mutations in multiple primary melanomas. N Engl J Med 1998;338:879–87.
- MacKie RM, Andrew N, Lanyon WG, Connor JM. CDKN2A germline mutations in U.K. patients with familial melanoma and multiple primary melanomas. J Invest Dermatol 1998;111:269–72.
- 10 Holland EA, Schmid H, Kefford RF, Mann GJ. CDKN2A. (p16INK4a) and CDK4 mutation analysis in 131 Australian melanoma probands: effect of family history and multiple primary melanomas, Genes Chromosomes Cancer, 1999;25:1–10.
- 11 Ghiorzo P, Ciotti P, Mantelli M, Heouaine A, Queirolo P, Rainero ML, Ferrari C, Santi PL, De Marchi R, Farris A, Ajmar F, Bruzzi P, Bianchi-Scarra G. Characterization of Ligurian melanoma families and risk of occurrence of other neoplasia. *Int J Cancer* 1999;83:441–8.
- 12 Borg A, Sandberg T, Nilsson K, Johannsson O, Klinker M, Masback A, Westerdahl J, Olsson H, Ingvar C. High frequency of multiple melanomas and breast and pancreas carcinomas in CDKN2A mutation-positive melanoma families. J Natl Cancer Inst 2000;92:1260–6.
- 13 Vasen HFA, Gruis NA, Frants RR, van der Velden PA, Hille ETM, Bergman W. Risk of developing pancreatic cancer in families with familial atypical multiple mole melanoma associated with a specific 19 deletion of p16 (p16-Leiden). Int J Cancer 2000;87:809–11.

- 14 Parker J, Florell SR, Alexander A, DiSario JA, Shami PJ, Leachman SA Pancreatic carcinoma surveillance in patients with familial melanoma. Arch Dermatol 2003:139:1019-25.
- 15 Puig S, Malvehy J, Badenas C, Ruiz A, Jimenez D, Cuellar F, Azon A González U, Castel T, Campoy A, Herrero J, Martí R, Brunet-Vidal J, Milà M. Role of the CDKN2A locus in patients with multiple primary melanomas. J Clin Oncol 2005:**23**:3043-51.
- 16 Niendorf K, Goggins W, Yang G, Tsai K, Shennan M, Bell D, Sober A, Hogg D, Tsao H. MELPREDICT: a logistic regression model to estimate CDKN2A carrier probability. J Med Genet 2006;43:501–6.
- Begg CB, Orlow I, Hummer AJ, Armstrong BK, Kricker A, Marrett LD, Millikan RC, Gruber SB, Anton-Culver H, Zanetti R, Gallagher RP, Dwyer T, Rebbeck TR, Mitra N, Busam K, From L, Berwick M for the Genes Environment and Melanoma (GEM) Study Group. Lifetime risk of melanoma in CDKN2A mutation carriers in a population-based sample. J Natl Cancer Inst 2005;97:1507-15.
- Yang G, Niendorf KB, Tsao H. A novel methionine-53-valine mutation of p16 in a hereditary melanoma kindred. J Invest Dermatol 2004;123:574-5.
- Hussussian CJ, Struewing JP, Goldstein AM, Higgins PAT, Ally DS, Sheahan MD, Clark WH Jr, Tucker MA, Dracopoli NC. Germline p16 mutations in familial melanoma. *Nat Genet* 1994;8:15–21.
- Goldstein AM, Struewing JP, Chidambaram A, Fraser MC, Tucker MA. Genotype-phenotype relationships in American melanoma-prone families with CDKN2A and CDK4 mutations. J Natl Cancer Inst 2000;92:1006–10.
- 21 Eliason MJ, Larson AA, Florell SR, Zone JJ, Cannon-Albright LA, Samlowski WE, Leachman SA. Population-based prevalence of CDKN2A mutations in Utah
- melanoma families. *J Invest Dermatol* 2006;1**26**:660–6.

 22 **Puig S**, Ruiz A, Castel T, Volpini V, Malvehy J, Cardellach F, Lynch M, Mascaro JM, Estivill X, Inherited susceptibility to several cancers but absence of linkage between dysplastic nevus syndrome and CDKN2A in a melanoma family with a mutation in the CDKN2A (p16INK4A) gene. Human Genet 1997;101:359-64.
- 23 Ruiz A, Puig S, Malvehy J, Lazaro C, Lynch M, Gimenez-Arnau AM, Puig LI, Sanchez-Conejo J, Estivill X, Castel T. CDKN2A mutations in Spanish cutaneous malignant melanoma families and patients with multiple melanomas and other neoplasia. J Med Genet 1999;36:490-4.
- Rizos H, Puig S, Badenas C, Malvehy J, Darmanian AP, Jimenez L, Mila M, Kefford RF. A melanoma-associated germline mutation in exon 1β inactivates p14ARF. Oncogene 2001;**20**:5543–7.
- Yakobson E, Eisenberg S, Isacson R, Halle D, Levy-Lahad E, Catane R, Safro M, Sobolev V, Huot T, Peters G, Ruiz A, Malvehy J, Puig S, Chompret A, Avril MF, Shafir R, Peretz H, Paillerets BB. A single Mediterranean, possibly Jewish, origin for the Val59Gly CDKN2A mutation in four melanoma-prone families. Eur J Hum Genet 2003;11:288–96.
- 26 Landi MT, Goldstein AM, Tsang S, Munroe D, Modi W, Ter-Minassian M, Steighner R, Dean M, Metheny N, Staats B, Agatep R, Hogg D, Calista D. Genetic susceptibility in familial melanoma from North Eastern Italy. J Med Genet 2004;41:557-66
- Lang J, Boxer M, MacKie RM. CDKN2A. mutations in Scottish families with cutaneous melanoma: results from 32 newly identified families, Br J Dermatol, 2005:153:1121-5
- 28 Harland M, Holland EA, Ghiorzo P, Mantelli M, Bianchi-Scarra G, Goldstein AM, Tucker MA, Ponder BAJ, Mann GJ, Bishop DT, Bishop JN. Mutation screening of the CDKN2A promoter in melanoma families. Genes Chromosomes Cancer 2000;**28**:45–57
- Newton Bishop JA, Harland M, Bennett DC, Bataille V, Goldstein AM, Tucker MA, Ponder BAJ, Cuzick J, Selby P, Bishop DT. Mutation testing melanoma families: INK4A, CDK4, and INK4D. Br J Cancer 1999;80:295-300.

- 30 Newton Bishop JA, Wachsmuth RC, Harland M, Bataille V, Pinney E, Mack P, Baglietto L, Cuzick J, Bishop DT. Genotype/phenotype and penetrance studies in melanoma families with germline CDKN2A mutations. J Invest Dermatol 2000:114:28-33.
- 31 Bergman W, Gruis NA, Frants RR. The Dutch FAMMM family material Clinical and genetic data. Cytogenet Cell Genet 1992;59:161–4.

 Gruis NA, van der Velden PA, Sandkuijl LA, Prins DE, Weaver-Feldhaus J,
- Kamb A, Bergman W, Frants RR. Homozygotes for CDKN2 (p16) germline mutation in Dutch familial melanoma kindreds. Nat Genet 1995;10:351-3.
- 33 Kannegiesser C, Avril M-F, Spatz A, Laud K, Lenoir GM, Bressac-de Paillerets. CDKN2A as a uveal and cutaneous melanoma susceptibility gene. Genes Chromosomes Cancer 2003;38:265-8.
- 34 **Chaudru V**, Chompret A, Bressac-de Paillerets B, Spatz A, Avril M-F, Demenais F. Influence of genes, nevi, and sun sensitivity on melanoma risk in a family sample unselected by family history and in melanoma-prone families. J Natl Cancer Inst 2004:96:785-95.
- Borg A, Johannsson U, Johannson O, Hakansson S, Westerdahl J, Masback A, Olsson H, Ingvar C. Novel germline p16 mutation in familial malignant melanoma in southern Sweden. *Cancer Res* 1996;**56**:2497–500.
- 36 Platz A, Hansson J, Mansson-Brahme E, Lagerlof B, Linder S, Lundqvist E, Sevigny P, Inganas M, Ringborg U. Screening of germline mutations in the CDKN2A and CDKN2B genes in Swedish families with hereditary cutaneous melanoma. J Natl Cancer Inst 1997;89:697-702.
- 37 Platz A, Hansson J, Ringborg U. Screening of germline mutations in the CDK4, CDKN2C, and TP53 genes in familial melanoma: a clinic-based population study. Int J Cancer 1998;78:13-15.
- 38 Hashemi J, Linder S, Platz A, Hansson J. Melanoma development in relation to non-functional p16/INK4A protein and dysplastic nevus syndrome in Swedish melanoma kindreds. Melanoma Res 1999;9:21–30.
- Walker GJ, Hussussian CJ, Flores JF, Glendening JM, Haluska FG, Dracopoli NC, Hayward NK, Fountain JW. Mutations of the CDKN2/p16INK4 gene in Australian melanoma kindreds. Hum Molec Genet 1995;**4**:1845–52.
- 40 Flores JF, Pollock PM, Walker GJ, Glendening JM, Lin AHT, Palmer JM Walters MK, Hayward NK, Fountain JW. Analysis of the CDKN2A, CDKN2B and CDK4 genes in 48 Australian melanoma kindreds. *Oncogene* 1997;15:2999–3005.
- Whiteman DC, Milligan A, Welch J, Green AC, Hayward NK. Germline CDKN2A mutations in childhood melanoma. J Natl Cancer Inst 1997;89:1460.
- Aitken J, Welch J, Duffy D, Milligan A, Green A, Martin N, Hayward N. CDKN2A variants in a population-based sample of queensland families with nelanoma. J Natl Cancer Inst 1999;**91**:446–52.
- 43 Holland EA, Beaton SC, Becker TM, Grulet OMC, Peters BA, Rizos H, Kefford RF, Mann GJ. Analysis of the p16 gene, CDKN2, in 17 Australian melanoma kindreds. Oncogene 1995;11:2289–94.
 44 Rizos H, Becker TM, Holland EA, Kefford RF, Mann GJ. Differential expression of
- p16(INK4a) and p16 beta transcripts in B-lymphoblastoid cells from members of hereditary melanoma families without CDKN2A exon mutations. Oncogene 1997:**15**:515-23
- 45 Parkin DM, Whelan SL, Ferlay J, Storm H. Cancer Incidence in Five Continents. Vol I to VIII. CANCERMondial, http://www-dep.iarc.fr; Lyon, IARC CancerBase No, 7, 2005;(France).
- Schottenfeld D, Fraumeni JF, Jr. Cancer epidemiology and prevention, 2nd edn., New York, NY: Oxford University Press, 1996.
 Shafey O, Dolwick S, Guindon GE, eds. Tobacco control country Profiles, 2003.
- Atlanta, GA: American Cancer Society, 2003.
- 48 Goldstein AM, Tucker MA. Editorial: a piece of the melanoma puzzle. J Natl Cancer Inst 2005:97:1486-7.