

# The physiological costs of reproduction in small mammals

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Life-history trade-offs between components of fitness arise because reproduction entails both gains and costs. Costs of reproduction can be divided into ecological and physiological costs. The latter have been rarely studied yet are probably a dominant component of the effect. A deeper understanding of life-history evolution will only come about once these physiological costs are better understood. Physiological costs may be direct or indirect. Direct costs include the energy and nutrient demands of the reproductive event, and the morphological changes that are necessary to facilitate achieving these demands. Indirect costs may be optional 'compensatory costs' whereby the animal chooses to reduce investment in some other aspect of its physiology to maximize the input of resource to reproduction. Such costs may be distinguished from consequential costs that are an inescapable consequence of the reproductive event. In small mammals, the direct costs of reproduction involve increased energy, protein and calcium demands during pregnancy, but most particularly during lactation. Organ remodelling is necessary to achieve the high demands of lactation and involves growth of the alimentary tract and associated organs such as the liver and pancreas. Compensatory indirect costs include reductions in thermogenesis, immune function and physical activity. Obligatory consequential costs include hyperthermia, bone loss, disruption of sleep patterns and oxidative stress. This is unlikely to be a complete list. Our knowledge of these physiological costs is currently at best described as rudimentary. For some, we do not even know whether they are compensatory or obligatory. For almost all of them, we have no idea of exact mechanisms or how these costs translate into fitness trade-offs.

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## 1. INTRODUCTION

The theory of evolution by natural selection assumes that an individual's attributes result from their historical attempts to maximize fitness—their genetic contribution to the future gene pool. Fitness, however, is not a singular concept, but rather consists of a number of components. The life history of an organism is a reflection of how the animal blends together these components into an integrated life-history 'strategy'. A fundamental axiom of our understanding and interpretation of the evolution of life-history strategies is that trade-offs exist between the various life-history components ([Fisher 1930](#page-18-0)). For example, an increase in current reproductive output by maximizing fecundity might occur only at the expense of future survivorship or future reproductive output ([Williams 1966](#page-23-0); [Stearns](#page-22-0) [1992](#page-22-0); [Charnov 1993](#page-17-0); [Charlesworth 1994](#page-17-0)). We assume that animals cannot combine together the fitness components in a strategy that simultaneously maximizes them all, and must therefore trade-off the components against each other (but see [Johnston](#page-19-0) et al. [2006\)](#page-19-0).

The idea that there must be trade-offs in fitness components stems from the idea that reproductive

events generate fitness benefits in terms of viable offspring, but that reproduction also entails costs as well. These costs drive the nature of the trade-off between the fitness components ([Reznick](#page-21-0) et al. 1990; [Roff 1992\)](#page-21-0). There are several different types of trade-off among components of current reproductive effort, for example, trading off the size against number of offspring ([Smith & Fretwell 1974\)](#page-22-0) and also trade-offs between current reproduction and components of future fitness: either survival or fecundity. [Stearns](#page-22-0) [\(1992\)](#page-22-0) identified 10 different fitness components or life-history 'traits' and detailed a matrix of potential trade-offs between them. For each trade-off, there must be a mechanism that links the fitness components together. This mechanism is the 'cost' that is attached to the reproductive event. Previous investigators have identified that these costs can be further partitioned into physiological costs, and costs that are mediated via ecology (e.g. [Zera & Harshman 2001\)](#page-23-0). An example of an ecological cost is the increased risk of predation that is associated with foraging while acquiring energy and nutrients, like protein and calcium, for the reproductive event. Historically, the study of life histories has been rooted in the determination of the magnitude of trade-offs with little concern for their mechanistic basis. However, it is becoming increasingly recognized that distinguishing the importance of ecological and physiological mechanisms is a key goal if we are to understand the evolution of life-history patterns

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[\(Zera & Harshman 2001;](#page-23-0) [McNamara & Houston](#page-20-0) [2008](#page-20-0)). Nevertheless, it should be borne in mind that ecological and physiological components strongly interact and cannot be completely isolated from each other. Despite the fact that it is widely recognized that physiological costs underpin all life-history trade-offs (e.g. [Stearns 1992](#page-22-0); [Ricklefs & Wikelski 2002\)](#page-21-0), the physiological contribution to most, if not all, trade-offs remains very poorly studied. In this paper, I propose to review our understanding of some of the physiological costs that are attached to the process of reproduction. I have focused the review on small mammals where there is at least some depth of study, but I also confess that this betrays a bias towards a group of animals that I have studied personally.

Physiological costs of reproduction can be divided into two different types. The first is direct costs that stem from satisfying the demands of the reproductive event itself. These demands at their simplest level are the energy and nutrients that the parental animal needs to acquire to successfully reproduce. In the previous paragraph, I identified the predation risk that attends the extra foraging required to collect the energy or nutrients required for reproduction as an ecological cost. The direct physiological cost of reproduction is that energy and nutrient demand. It is probable that these direct costs have deeper layers of complexity to them. Animals not only require energy and the major macronutrients (like protein) but also essential amino and fatty acids, as well as intakes of vitamins and micronutrients. Our knowledge of many of these physiological requirements is almost non-existent. I have also included in this 'direct physiological cost' the physiological and anatomical modifications that are necessary for the animal to achieve these demands. For example, if an animal needs to double or treble its rate of food intake to satisfy the demand for energy during reproduction, it cannot simply do that without some modifications to its alimentary tract and the machinery that processes ingested food. These necessary modifications are a second form of direct cost.

The second type of cost of reproduction is indirect. Like direct costs, we can also subdivide the indirect costs into two sub-types. First, to meet the direct physiological costs of reproduction, an animal may cut corners in other components of its physiology. Many animals probably inhabit environments where it is simply not possible for their intake to be increased adequately to cover their reproductive demands. In these circumstances, the animals may make compensatory adjustments in aspects of their physiology to save energy that can then be devoted to reproduction. Even where food is sufficiently available, there may be ecological costs involved in obtaining it (e.g. predation risks), which are greater than the physiological costs of cutting down the expenditure on other components of physiological functioning. These compensatory physiological costs are an indirect cost of the reproductive event. A distinguishing feature of these costs is that they have a degree of optionality about them. The animal is choosing to modify its physiology to invest more in reproduction, and it can choose to make this allocation of resource or not. The costs derive from the fact that resource is limited and animals must therefore choose

how to allocate this limited resource between competing functions. We can distinguish these optional 'compensatory costs' from other indirect costs that are an inevitable physiological consequence of the reproductive event. For example, reproduction might involve the production of a toxic by-product that elevates the risk of the animal developing a fatal disease like cancer. This would be a cost of reproduction that would not be optional. The animal could not choose to reproduce without this negative consequence.

## 2. DIRECT COSTS

## (a) Increased demand for energy and nutrients (i) Energy

Probably the most detailed data available for the energy demands of reproduction are derived from domesticated rodents. This is primarily because these animals are easily kept, and even quite invasive measurements can be made on them without the risk that they will desert their offspring. The time course of food intake, at 21°C, throughout reproduction in a strain of laboratory mouse (MF1) that we have been studying is shown in [figure 1](#page-2-0)a ([Johnson](#page-19-0) et al. 2001a). Food intake increased during pregnancy to approximately 8 g, compared with 5.5 g in the non-breeding females prior to reproduction. Although the pattern of foetal growth is essentially exponential between conception and birth, the pattern of food intake does not mirror this, but rather rises to a peak approximately 2–3 days earlier and then declines slightly before the day of parturition. The reason for this pattern is uncertain, but one hypothesis is that the expanding foetal mass competes for space with the alimentary tract in the abdomen, limiting the food intake. This may suggest that resources to support the gestation become limited in late pregnancy. Under such limitation, competing demands may have detrimental effects on the success of the pregnancy. One such competing demand may be the level of basal energy requirements (basal metabolic rate, BMR). We have recently shown in another strain of mouse (the C57BL/6 black mouse) that mice with higher BMR have a greater likelihood of mass anomalies occurring during pregnancy. These mass anomalies probably reflect foetal resorption events. Because BMR is correlated with body mass, this relationship might only be an artefact of both resorption rate and BMR being affected by mass, but the effect of BMR is also evident if the effect of mass is removed statistically ([Johnston](#page-19-0) et al. 2007). This effect supports the hypothesis that resource intake in late pregnancy may be limited, perhaps by space competition in the abdomen between the alimentary tract and the developing foetuses.

The most dramatic increase in food intake occurred during lactation. During the initial 10 days, this increase was linear, but then it reached a plateau at approximately 23 g of food per day. Food intake at the plateau (between days 10 and 18) was related to litter size. Small litters reached plateau intakes of less than 23 g (figure  $1b$ ), but as the litter size increased food intake also increased to a plateau at approximately 23 g of food per day. The mice seemed to reach a limit in

<span id="page-2-0"></span>

Figure 1. (a) Food intake each day throughout pre-breeding, pregnancy and lactation phases for MF1 mice. Data averaged across 71 litters. (b) Daily food intake averaged over the last days of lactation (10–18) plotted against litter size in MF1 mice. (c) Pup mass in relation to litter size in MF1 mice.  $(d)$  Histogram showing the frequency at birth of different litter sizes across 71 litters of MF1 mice.

their food intake at this level. Although litter sizes increased, food intake did not increase in parallel.

We have shown that this limit in food intake is not mediated via the aspects of the cage the animals live in [\(Speakman & Krol 2005\)](#page-22-0), and therefore appears to be a physiological feature intrinsic to the animals themselves. This apparent physiological limit in the capacity of the mouse to ingest food at peak lactation may underpin an important life-history trait (maximum litter size) and an important life-history trade-off. Since

the maximum asymptotic food intake at peak lactation is fixed, the energy that can be devoted to milk is also fixed. As the litter size increases, this milk must be divided between more and more offspring and, consequently, the pups wean at progressively smaller body masses (figure 1c; [Johnson](#page-19-0) et al. 2001a). A physiological limit in the capacity to ingest food therefore appears to underpin the life-history trade-off between the number and size of offspring. Presumably, there is a minimum size of offspring at weaning, which would stand any chance of survival. Where the declining relationship between pup size and litter size intersects, this theoretical minimum viable pup size may define the maximum litter size. A distribution of natural litter sizes in this mouse strain is shown in figure 1d. It is noticeable that the asymptotic intakes for the mice that raised 14 and 15 pups were actually higher than the average 23 g limit (figure  $1d$ ). Perhaps these females were only able to raise such large litters because they were individuals that were capable of above-average intakes, and therefore still able to wean their pups at a pup mass of 7.5 g. Natural litter sizes of 16 and above in this strain may not be feasible owing to the interaction between the limit on maximal intake rate and the minimum viable pup size (this minimum viable pup size at weaning may also be physiologically mediated).

Understanding the physiological basis of this maximal intake limit is therefore of critical importance. Two hypotheses concerning the nature of this limit were proposed in the early 1990s ([Peterson](#page-21-0) et al. 1990; [Weiner 1992;](#page-23-0) [Hammond & Diamond 1997](#page-18-0)). The first hypothesis was that the limit was imposed by the capacity of the alimentary tract to absorb food (energy) and process it into a form for mobilization. This was called the 'central limit hypothesis'. The second hypothesis was that the limit is imposed at the peripheral site where the centrally supplied energy is used: the mammary glands. This was called the peripheral limit hypothesis.

[Hammond & Diamond \(1992\)](#page-18-0) manipulated litters of Swiss Webster mice and found that females did not elevate their food intake when given up to 23 pups. Similarly, we have also shown that MF1 mice given up to 19 pups also could not breach the 23 g limit that they reached during unmanipulated lactations with litters of greater than 10 offspring ([Johnson](#page-19-0) et al.  $2001a$ ). Faced with this problem of a litter size that is 'too large', females will often cull their offspring rather than eat more food ([Johnson](#page-19-0) et al. 2001a; [Gandelman & Simon](#page-18-0) [1978\)](#page-18-0). In a separate experiment, [Hammond &](#page-18-0) [Diamond \(1994\)](#page-18-0) prevented pups from weaning at their normal weaning age and hence their increasing growth demands needed to be supplied by the mother until the pups were 24 days old. In these conditions, the mothers also could not upregulate their food intake. The absence of an increase in the intake when pup numbers are increased or lactation extended confirms there is a physiologically imposed limit, but does not separate between the peripheral and central limits hypotheses.

Giving female mice additional energy-demanding tasks during late lactation can better separate the alternative hypotheses. The basis of these experiments

is that, if the system is centrally limited, the total energy available will be fixed. Using some of this energy to perform another task will reduce the amount available to support milk production and the consequence will be a diminution of reproductive output. Alternatively, if the system is peripherally limited, the animals will increase their intakes to meet the combined demands, and reproductive output will remain unaffected. Three manipulations have been performed to test these ideas: lactating mice have been forced to exercise to obtain their food; they have been made simultaneously pregnant, combining the demands detailed in [figure 1](#page-2-0)a, and they have been exposed to cold.

[Perrigo \(1987\)](#page-21-0) compared the reproductive strategies of house mice Mus domesticus and deer mice Peromyscus maniculatus by forcing females to run a preset number of revolutions (between 75 and 275) on a wheel to obtain each pellet of food. Despite the combined demands of lactation and locomotor activity, neither house nor deer mice exceeded the upper limit of food intake compared with unmanipulated mothers, given free access to food. As a result of the decreased amount of energy available for reproduction, the wheel-running house mice routinely killed some of the offspring throughout the first 12 days of lactation, whereas deer mice extended lactation well beyond normal weaning age.

[Johnson](#page-19-0) et al. (2001b) followed the intakes of mice that had been mated immediately post-partum and found that the mice concurrently lactating and pregnant did not respond to the increased energy burden by elevating their food intake. Instead, they delayed implantation at the start of the second pregnancy and the length of this delay was directly related to the numbers of pups. The animals therefore 'avoided' overlapping their energy demands, perhaps because they could not elevate their total intake at peak lactation to accommodate both. Similar observations were made in Rockland-Swiss mice [\(Biggerstaff & Mann 1992\)](#page-16-0), and in rats (Rattus norvegicus: [Koiter](#page-19-0) et al. 1999), when food intake in late lactation was actually reduced in those rats that carried a simultaneous pregnancy, relative to rats just lactating. Together, these activity and pregnancy studies indicate that the limits are centrally, rather than peripherally, mediated; although, in case of concurrent pregnancy, the evidence is less strong as the animals avoided the problem.

However, when [Hammond](#page-18-0) et al. (1994) exposed lactating Swiss Webster mice to  $8^{\circ}$ C (approx. 22 $^{\circ}$ C below the lower critical temperature), they found that food intake increased dramatically beyond the supposed centrally imposed limit. Similar observations have since been made in deer mice ([Hammond &](#page-18-0) [Kristan 2000](#page-18-0)), MF1 mice ([Johnson & Speakman](#page-19-0) [2001](#page-19-0)) and cotton rats (Sigmodon hispidus; [Rogowitz](#page-21-0) [1998](#page-21-0)). The capacity of the mice to elevate their food intake in the cold was completely at odds with the central limitation hypothesis. To test whether the limit was imposed peripherally, [Hammond](#page-18-0) et al. (1996) experimentally manipulated female mice by surgically removing some of their mammary glands during late lactation. The rationale behind this experiment was that if the capacity of the mammary glands was limited, then, if the mammary tissue was halved in size, the remaining tissue would be unable to compensate by

elevating its milk production. However, if the capacity of the tissue was flexible and limited only by the centrally controlled supply of energy, then it would respond to the absence of half the tissue by expanding its capacity. They found that productivity in the halved glands did not increase, suggesting that the mammary gland was indeed the point at which the system was peripherally limited.

The above experiments suggested that there is a physiological limit in the capacity of the mammary tissue to secrete milk which underlies the asymptotic food intake in late lactation, and thus the trade-off between litter size and pup size and potentially defines the maximal litter size. When animals were manipulated at room temperature by giving them more pups to raise, food intake did not increase because milk production was limited by the capacity of the mammary glands. When exposed to cold conditions, food intake did increase (demonstrating a lack of central limitation) owing to the combined demands for milk production (at maximal capacity) and increased thermogenesis.

Several other studies have been performed, which also indicate that the limits to intake at peak lactation are not centrally mediated. These studies involve manipulation of the energy density of the food. If the energy density of food is decreased, but animals have a central processing capacity limit, they should be unable to upregulate their intake to compensate. [Speakman](#page-22-0) et al. (2001) fed MF1 mice a diet that provided 25% less digestible energy than their normal food and then mated them. Food intake in the mice fed on the low energy density food increased at peak lactation on average of 3.8 g (from 23.1 to 26.9 g per day). Similar data are available for brown hares (Lepus europaeus; [Hacklander](#page-18-0) et al. 2002). When fed on a diet with lower energy content, asymptotic food intake in late lactation increased from 230–250 to 280–300 g per day. Consequently, milk production was stable across the dietary treatments at approximately 35 g per day for females raising single offspring and 70 g per day for females raising twins. Conversely, when energy density was manipulated in the opposite direction, there was no indication that energy intake was elevated. In another example, rats fed a high energy density diet during lactation decreased their food intake to sustain energy intake constant ([Denis](#page-17-0) et al. 2003).

An important aspect of the peripheral limitation idea is that milk production levels remain constant across the different manipulations—reflecting the fact that mammary glands are working at capacity. Several studies have measured milk production and support this prediction. [Drummond](#page-17-0) et al. (2000) studied milk production in rabbits (Oryctolagus cuniculus) and observed that, following natural deaths of some offspring, the flow of milk was unaltered. Fink [et al.](#page-18-0) [\(2001\)](#page-18-0) studied lactation in captive mink (Mustela vision) and showed that in mothers raising litters of three, six and nine offspring, milk production did not increase when litters increased from six to nine offspring. [Rogowitz \(1998\)](#page-21-0) demonstrated in cotton rats that levels of milk production in rats at 21 and  $8^{\circ}$ C were similar—consistent with the mammary glands

working at maximal capacity. However, other studies have failed to find this consistency ([Johnson &](#page-19-0) [Speakman 2001](#page-19-0); Król & Speakman 2003a,[b](#page-19-0); Król et al.  $2003a,b$  $2003a,b$ ). We have now studied food intake and milk production in mice at peak lactation at three different temperatures: 30, 21 and  $8^{\circ}$ C. As predicted by the peripheral limitation hypothesis, food intake increased across these measurements as temperature declined. However, unexpectedly, milk production and, consequently, pup growth were not constant across the different temperatures, but followed the pattern of food intake. Mice exported 88 kJ energy in milk per day at 30 $\degree$ C, 167 kJ at 21 $\degree$ C and 288 kJ at 8 $\degree$ C (Król & Speakman 2003b). Moreover, the weaning masses of pups at 30, 21 and  $8^{\circ}$ C averaged 6.1, 7.0 and 7.3 g, respectively (Król & Speakman 2003a). Consequently, the colder it got the more food the mice ate, the more milk they produced and the heavier the pups were at weaning.

These data are fundamentally inconsistent with both the suggestion that the limits are imposed by the capacity of the alimentary tract to process ingested energy and the suggestion that the limits reside in the milk production capacity of the mammary gland. There are a number of new ideas about the nature of the limit in food intake during lactation, which have been recently summarized ([Speakman & Krol 2005](#page-22-0)). For example, one of these is that the limits are imposed by aspects of the neuroendocrinological system that regulates food intake rates—the 'neural saturation hypothesis'. An alternative, suggested by (Król  $\&$ Speakman  $2003a,b$  $2003a,b$ , is that the capacity to expend energy during lactation at  $21^{\circ}$ C might be limited by the ability of the female mouse to dissipate heat. Hence, manipulations at  $21^{\circ}$ C—which aim to stimulate both food intake and milk production—notably increasing litter size ([Hammond & Diamond 1992\)](#page-18-0), extending lactation ([Hammond & Diamond 1994\)](#page-18-0), making them simultaneously pregnant ([Biggerstaff &](#page-16-0) [Mann 1992](#page-16-0); [Koiter](#page-19-0) et al. 1999; [Johnson](#page-19-0) et al. 2001b) and making them exercise ([Perrigo 1987](#page-21-0))—all failed to increase either food intake or milk production, because the animals could not increase their heat production without risking fatal hyperthermia (see below). In effect, this hypothesis is a central limitation idea, but is focused around the ability to dissipate heat rather than assimilation from the gut. Under the heat dissipation limit hypothesis, when mice are exposed to the cold, this is not an additional demand, but a relaxation of the heat dissipation limit, allowing the animals to elevate not only their food intake but also their milk production and thus the size of their offspring. Similarly, when mice were placed in the hot, this reduced their capacity to dissipate heat, restricted their food intake and milk production, and led to smaller pups being weaned.

There are two putative mechanisms for how the capacity to dissipate heat may influence lactation performance. At high ambient temperatures, lactating mice may continuously face difficulties dissipating heat. This would lead to a perpetually elevated body temperature (see below) which might influence the regulation of milk production. It is well established that endogenous opioids in the preoptic anterior

hypothalamus are of key importance in the regulation of body temperature, and are reduced during lactation (Kim et al[. 1997](#page-19-0)). Projections from this area terminate in the paraventricular nucleus (PVN), where the magnocellular cells synthesize oxytocin. [Rayner](#page-21-0) et al. (1988) showed that intracerebroventricular (ICV) administration of morphine inhibits oxytocin production in the PVN. Thus, elevated endogenous opioids under hyperthermia might directly reduce oxytocin secretion. Morphine administration also disrupts maternal suckling behaviour (Cox [et al.](#page-17-0) 1976; [Bridges & Grimm 1982\)](#page-17-0). In addition, thyroid hormone, which may be regulated by differences in body temperature, is an important modulator of prolactin production. Continual maternal hyperthermia in relation to external ambient temperatures (or differential capacity to dissipate body heat) may therefore directly inhibit oxytocin and prolactin secretion, thereby reducing milk production. Finally, there may be a more direct link between hyperthermia and milk production. A response to hyperthermia might be to direct blood flow away from the mammary glands to other peripheral areas to dissipate heat by vasodilatation [\(Black](#page-16-0) et al. 1993). Blood flow in the mammary glands has been shown to have a direct effect on milk production ([Vernon & Flint 1983](#page-23-0)).

A second mechanism is that suckling schedules of the mice may be influenced by the development of maternal hyperthermia in the nest. Although pups act as heat sinks early in their development ([Scribner &](#page-22-0) Wynne-Edwards  $1994a,b$  $1994a,b$ , in late lactation offspring are capable of considerable heat production. The suckling unit of mother and pups, therefore, may generate heat that leads to maternal hyperthermia, ultimately forcing the female to discontinue suckling [\(Croskerry](#page-17-0) et al. 1978). The progress to hyperthermia would be more rapid as the capacity to dissipate heat declines. The sucking stimulus is one of the primary factors stimulating oxytocin release and milk let down, and also feeds back onto prolactin release, thereby regulating milk production. Continual disruption of suckling, due to intermittent hyperthermia, would be a second mechanism linking heat dissipation capacity to lactation performance.

Many studies have examined the pattern of change in food intake across the reproductive cycle among other small non-domesticated mammals. Some examples of these measurements are in [table 1.](#page-5-0) This is not an exhaustive review. Using the data for rodents and converting estimated food intakes into energy for those species where only food intakes are reported in the original papers, there was a significant effect of body mass ( $F=42.6$ ,  $p<.001$ ) and a significant effect of reproductive status  $(F=30.6, p<.001)$ , but no significant interaction between these two variables  $(F=1.55, p=0.225;$  [figure 2](#page-7-0)a). Using pairwise Tukey comparisons, the non-breeding energy demands did not differ significantly from those in pregnancy, but the demands in lactation were significantly elevated above both the non-breeding and pregnancy demands. This analysis reveals that the energy costs of pregnancy are relatively trivial. This does not mean that demands in pregnancy do not impose limits on reproduction

<span id="page-5-0"></span>Table 1. Some examples of peak energy intakes of non-domesticated small (less than 1 kg) mammals at different phases of the reproduction cycle. LS is litter size. Under P this is litter size at birth; under L, this is litter size at weaning. Where original paper does not state which, it is entered under both. Status: NB, non-breeding; P, pregnant; L, lactating.  $\times$  NB is the intake expressed as a multiple of the non-breeding intake. In all cases (except \*), the animals were studied in the laboratory at standard room temperatures (21–24°C) with ad libitum access to food.  $*$  animals kept at 5°C  $*$  animals kept at 10°C. In some cases, the original data were quoted relative to body mass<sup>0.75</sup>, and in these instances the actual intakes have been recalculated using the cited body mass. In other papers, only mass of food ingested is cited and these have been converted assuming a dry mass energy content for the food of 20.9 kJ  $g^{-1}$ , unless a different value was cited in the paper. In yet other studies, average daily metabolic rate (ADMR) is quoted. I have converted this to energy intake assuming a digestive efficiency of 80%. Genera: Rodents  $C = \text{Clethrionomys}$ ,  $P. =$  Peromyscus, M. = Microtus, Me. = Meriones, A. = Acomys, Ca. = Cavia, S. = Sigmodon, Sc. = Sciurus. Insectivores, S. = Sorex, C. = Crocidura, Ec. = Echinops, E. = Erinaceus. Bats P. = Plecotus, T. = Tadarida, E. = Eptesicus, M. = Myotis,  $R. =$  Rousettus.  $\wedge$  estimates relative to NB calculated using the NB in reference 7. \$ estimates expressed as difference to unquoted NB level in original ms.



(Continued.)





<sup>a</sup> References: 1. [Glazier \(1985\)](#page-18-0), 2. Innes & Millar [\(1981,](#page-19-0) [1985\),](#page-20-0) 3. [Millar \(1985\),](#page-20-0) [Millar & Innes \(1985\),](#page-20-0) 4. [Kaczmarski \(1966\)](#page-19-0), 5. [Hammond &](#page-18-0) [Kristan \(2000\)](#page-18-0), 6. [Migula \(1969\)](#page-20-0), 7. [Millar \(1978\)](#page-20-0), 8. [Weiner \(1987\),](#page-23-0) 9. [Sikes \(1995\)](#page-22-0) 27, 10. Degen et al. [\(2002\),](#page-17-0) 11. Liu et al. [\(2003\)](#page-20-0), 12. [Kam](#page-19-0) et al. [\(2003\),](#page-19-0) 13. [Randolph](#page-21-0) et al. (1977), 14. [Mattingly & McClure \(1982\)](#page-20-0), 15. [Rogowitz \(1998\),](#page-21-0) 16. Künkele (2000), 17 [Havera \(1979\),](#page-18-0) 18 [Nievergelt & Martin \(1999\)](#page-21-0), 19. [Genoud & Vogel \(1990\),](#page-18-0) 20. [Stephenson & Racey \(1993](#page-22-0)b), 21. [Poppitt](#page-21-0) et al. (1993), 22. Poppitt et al. (1994), 23. Kro<sup>1</sup> (1985), 24. Kurta et al. [\(1989\),](#page-20-0) 25. [Mclean & Speakman \(1999\)](#page-20-0), 26. Kunz et al. [\(1995\)](#page-20-0), 27. Kurta et al. [\(1990\),](#page-20-0) 28. [Korine](#page-19-0) et al. (2004), 29. [Atramentowicz \(1992\)](#page-16-0).

(see above arguments regarding foetal volume competing for abdominal space with the alimentary tract). Nevertheless, the pattern observed in laboratory mice

and rats that the costs of reproduction are substantially greater during lactation than in pregnancy appears to be very broadly applicable.

Table 1 (Continued.)

<span id="page-7-0"></span>

Figure 2. (a) Peak energy intake of female small rodents, when not breeding (black), pregnant (red) and lactating (green). There was no significant elevation in intake during pregnancy, but intake during lactation was significantly elevated compared with both non-breeders and pregnant individuals. (b) Residual energy intake in lactation was significantly associated with litter size. Data from small rodents are given in [table 1.](#page-5-0)

There was a wide variation in the energy demands at peak lactation ([table 1;](#page-5-0) figure  $2a$ ). The residual variation, once the effects of body mass had been removed, was significantly associated with litter size (figure 2b), which explained  $49.3\%$  of the residual variance ( $p=0.0103$ ). Although litter size and body size explain much of the variation in energy intake at peak lactation, there is still considerable variation in the energy intakes across species. One factor that may be of importance in this variation is the developmental strategy. Studies of the patterns of development in eutherian mammals have identified two separate developmental strategies (reviewed in [Martin &](#page-20-0) [MacLarnon 1985;](#page-20-0) [Martin 1989\)](#page-20-0), with some species having prolonged periods of gestation followed by birth of relatively well-developed offspring that rapidly become independent of the mother and are capable of feeding themselves relatively quickly after birth (precocial development). In contrast, other species have a relatively short gestation that is followed by a more protracted period of lactation during which they are completely dependent on nutrient supply from the mother (altricial development). The use of these strategies is not independent of body mass with

mammals weighing more than 100 kg using only the precocial strategy, while species weighing less than 100 g use the altricial strategy almost exclusively, although some species follow what has been called an 'intermediate' strategy (defined in [Martin &](#page-20-0) [MacLarnon 1985\)](#page-20-0), e.g. Acomys caharinus ([Degen](#page-17-0) et al. [2004](#page-17-0)). As anticipated from their size, mice follow the altricial strategy. Between 100 g and 100 kg species are found that follow either precocial or altricial strategies with a few 'intermediates' ([Martin &](#page-20-0) [MacLarnon 1985](#page-20-0); [Martin 1989\)](#page-20-0).

Clearly, this dichotomy in developmental strategy may impact on the levels of energy investment by the mother during lactation ([Oftedal 1984](#page-21-0); [Hill 1992](#page-19-0); [Kam](#page-19-0) et al. [2006\)](#page-19-0). Since it is the rarer strategy, relatively few studies have addressed the levels of food intake of small rodents following the clearly precocial strategy (but see, for example, the studies of the guinea pig Cavia porcellus; [Kunkele & Trillmich 1997](#page-19-0)) with many more studies following the patterns of investment during lactation of species raising clearly altricial or intermediate offspring. Gestation period (days) reflecting position on the altricial–precocial continuum was not an additional significant factor influencing peak energy demands ( $p=0.13$ ). The absence of an effect of this dichotomy might be expected given the selection of species that were either altricial or intermediate strategists. The only rodent with clear precocial development in the data reviewed in [table 1](#page-5-0) was the guinea pig ([Krebs 1950](#page-19-0)), but the manner in which the data were presented in that paper relative to unstated levels in non-breeding animals precluded its inclusion in the data analysis.

The residual variation once the effects of litter size and body mass were removed to an extent follow different strategies for coping with the direct costs of reproduction. Hence, the low values in the cotton rat (Sigmodon hispidus) and the hamster (Phodopus sungorus) reflect the fact that these animals deposit fat stores during the early phase of reproduction, which are withdrawn in lactation and reduce the need to supply all the energy from food intake. Low demands in other species may reflect the use of compensatory mechanisms that reduce expenditure on other components. These strategies will be discussed in detail below.

Expressing the peak lactation energy intake as a multiple of the intake of non-breeding animals, the average across all the rodents in [table 1](#page-5-0) was 2.1 (s.d.  $=$ 0.72,  $n=21$ ). This is considerably lower than the ratio in MF1 mice at  $21^{\circ}$ C, which equalled 4.2. However, the intake of the MF1 mice at this temperature is within the 95% confidence limits of the prediction from the fitted regression equation from the nondomesticated rodents, given the body size and litter size for this animal.

The importance of the differences in magnitude between the peak levels of intake in wild rodents and laboratory mice and rats remain uncertain. In most of the species studied in [table 1,](#page-5-0) and in other species, peak intake of food during lactation is dependent on litter size. Generally, however, these patterns do not reach an asymptotic level as observed in the MF1 mice [\(figure 1](#page-2-0)b). Two examples are shown in [figure 3.](#page-8-0) However, despite this absence of an asymptote in both

<span id="page-8-0"></span>

Figure 3. (a) Food intake at peak lactation (grams over 3 days) in relation to litter size in Peromyscus leucopus (drawn from tabulated data in [Millar 1978\)](#page-20-0). (b) Energy intake during lactation (kJ d<sup>-1</sup>) in relation to litter size in Sigmodon hispidus (drawn from tabulated data in [Rogowitz 1998](#page-21-0)).

species (and others), there is still an inverse relationship between the litter size and the mean offspring size, indicating that increases in investment were insufficient to match elevated demand. These data suggest that the trade-off between litter size and pup mass is more complex than the simple model of the mother reaching an asymptotic intake at which point a fixed investment is divided between increasing numbers of offspring. Part of this equation, however, may be capacity limits in the offspring themselves and how these depend on litter size. Pup–pup competition may be a key element of the impact of litter size on their growth efficiency.

## (ii) Protein and calcium

Previous discussion of limits on lactational performance has focused almost entirely on energy. Yet growing offspring also require large amounts of protein, calcium and other micronutrients. At weaning, an MF1 mouse has produced on average 12 pups each weighing 7.5 g: a total of 96 g of wet tissue. Of this tissue, approximately 13.5 g is protein and 1.8 g is calcium, both of which must be supplied by the mother. The diet we feed our mice on contains 22.45% crude protein (RM3 breeder diet SDS diet services, Witham Essex). Thus, over the last 8 days of lactation when they are taking in 23.2 g each day, their total protein intake amounts to 41.6 g of protein—substantially more than ultimately appears in the pups. This rough balance suggests that in the lactating MF1 mouse system, protein supply is unlikely to exert limits on the animals.



Figure 4. Mean mass at weaning of male (grey) and female (white) pups of  $(a)$  rats and  $(b)$  mice when fed on diets of varying protein content (from 16.3% to 9.4% protein). Drawn from data in [Goettsch \(1960\).](#page-18-0)

Several studies have been performed, however, where the level of protein in the diet is much lower.

[Goettsch \(1960\)](#page-18-0) examined the effects of feeding nursing rats and mice on four diets containing 16.3, 13.6, 11.4 and 9.4% protein. Food intake in lactation was broadly independent of the protein content of the diets, except that it was reduced in mice feeding on the lowest protein diet and 'highly variable' in rats on this diet. However, the growth of offspring was directly affected in both rats and mice, with offspring size at weaning being positively related to the protein content of the diet (figure  $4a,b$ ). Maternal mean protein intake in these mice throughout lactation averaged only 0.5 g per day in the lowest protein group which even accounting for the mean litter size difference between the strains (12 in our work and 7.1–7.5 in [Goettsch](#page-18-0) [1960](#page-18-0)) is still substantially lower than that for our mice (5.2 g/day at peak lactation). There are several additional studies which show that low maternal dietary protein levels affect pup growth. The mothers appear constrained, however, owing to the maximal amounts of energy that they can process—perhaps defined by the heat dissipation limit. Hence, this energetic limit prevents them from simply eating more food to meet their protein demands. These data indicate that energy limitation imposes a primary constraint on the animals under low protein supply.

If low protein in the maternal diet imposes a restriction on the growth of offspring then, perhaps, protein content of the diet is the key factor that links food intake to pup growth. When we exposed lactating females to hot and cold conditions, we may have manipulated the constraint on their total energy intake that was imposed by their capacity to dissipate heat ([Krol & Speakman 2003](#page-19-0) $a,b$  $a,b$ ). However, the impact on pup growth may have been mediated via the consequences of these different levels of energy intake for levels of protein intake—indeed the mice at  $8^{\circ}$ C were ingesting approximately 7.2 g of protein each day, those at  $21^{\circ}$ C only 5.2 g per day and those at 30°C only 2.91 g per day. Perhaps the trade-off between litter size and pup size (figure  $1c$ ) may move depending on dietary protein contents—to the right when protein in the diet is elevated and to the left when it is reduced. Supporting these ideas, [Hitchcock \(1927\)](#page-19-0) fed nursing rats a base diet that contained 29% protein, and additionally supplemented some of the animals with raw meat. Litter size in both groups was the same yet the offspring from the mothers fed meat daily weighed 61.8 g at weaning, compared with only 47.3 g in the rats not given the meat ration.

A similar situation probably pertains to calcium supply. The diet we use in our studies of mouse lactation contains 1.24% Ca. Thus, in the last 8 days of lactation, the mice ingest 2.30 g of calcium, compared with an estimated calcium content of the pups of 1.8 g. Hence, as long as calcium and protein levels in diets are above a certain limit, the constraint on maximal energy intake is unlikely to severely impact offspring in terms of their protein and calcium status.

For wild animals, the situation where calcium and protein contents of the diet are below the critical point, where the energy limit becomes restrictive, may be more routinely breached. One situation that has attracted considerable attention is in microchiropteran bats. Because microchiropteran bats are almost exclusively nocturnal, a constraint probably imposed by ecological factors such as predation risk and competition [\(Speakman 1990](#page-22-0); [1991](#page-22-0)a,[b](#page-22-0); [Rydell &](#page-22-0) [Speakman 1995](#page-22-0); [Speakman](#page-22-0) et al. 2000), they do not routinely get any exposure to sunlight. This may influence their production of vitamin D ([Cavaleros](#page-17-0) et al. [2003](#page-17-0)), which is an essential component of calcium physiology. Moreover, most microchiropteran species are predominantly insectivores and these prey are very low in calcium content. Their inability to manoeuvre effectively on the ground restricts them from finding additional mineral sources of calcium that may be available to other animals. Insectivorous bats have very low litter sizes for their body sizes ([Kurta & Kunz](#page-20-0) [1987](#page-20-0)) and one interpretation of these low rates of productivity is that they are primarily constrained by their capacity to obtain calcium ([Barclay 1994](#page-16-0)).

## (b) Organ remodelling to meet these demands

The alimentary tract has a more limited capacity to process energy than the mouth has capability to supply it. This is why the gut includes storage organs like the crop and stomach at the entry to the tract to take on board the intake at this elevated rate. If an animal requires to increase its food intake from approximately 5 g per day, prior to breeding, to 23 g per day at peak lactation, it could probably achieve this intake immediately, at the level of the mouth. There are no obvious changes in the morphology of the mouth as the animals progress through reproduction. However, while a nonbreeding mouse might be capable of eating 23 g each day, its alimentary tract would probably be unable to process the intake. Hence, animals need to modify their internal architecture to cope with the altered demands.

That lactating animals modify their tracts and associated organs in this manner has been known for at least 45 years. During lactation, there is an increase in the sizes of both the liver ([Kennedy](#page-19-0) et al. 1958) and the pancreas ([Jolicoeur](#page-19-0) et al. 1980). The most dramatic changes however are in the alimentary tract itself, which involve major morphological increases in the absorptive surface of the intestinal mucosa and also growth in the length of the tract [\(Boyne](#page-17-0) et al. 1966; [Fell](#page-18-0) [et al.](#page-18-0) 1963; [Campbell & Fell 1964](#page-17-0); [Craft 1970;](#page-17-0) [Cripps & Williams 1975;](#page-17-0) [Burdett & Reek 1979](#page-17-0); [Prieto](#page-21-0) et al. [1994;](#page-21-0) [Speakman & McQueenie 1996;](#page-22-0) [Hammond](#page-18-0) [1997](#page-18-0)). These changes are paralleled by alterations in the transport capacity at the cellular level ([Larradale](#page-20-0) [et al.](#page-20-0) 1966; Dugas et al[. 1970\)](#page-17-0). The growth of the mucosal layer of the alimentary tract in lactation includes both hypertrophic (cell expansion) and hyperplastic (cell proliferation) responses (Fell [et al](#page-18-0). [1963](#page-18-0); [Cairnie & Bentley 1967](#page-17-0); [Prieto](#page-21-0) et al. 1994). These changes effectively allow the lactating female to continue to extract the same amount  $(\%)$  of energy from the ingested food independent of the rate of intake (e.g. [Campbell & Fell 1964](#page-17-0); [Hammond](#page-18-0) et al. 1994).

Studies of wild rodents reveal similar patterns of change, although generally including more modest increases (e.g. [Myrcha 1964;](#page-20-0) Chlethrionomys glareolus; [Myrcha 1965](#page-20-0) Apodemus flavicollis; [Gebczynska &](#page-18-0) [Gebczynski 1971](#page-18-0) Clethrionomys oeconomus) reflective of the lower level of increase in food intake between non-breeding and lactation states ([table 1\)](#page-5-0). This is consistent with the fact that the extent of modification of the intestine is related to inter-individual variations in litter size in mice.

Several hypotheses have been advanced about how the growth in the gut is stimulated. The first is that the hormones that underpin the increase in food intake during lactation (reviewed in [Speakman & Krol 2005\)](#page-22-0), including elevated prolactin and reduced leptin, directly stimulate the changes in the alimentary organs. Alternatively, the food intake itself may result in the production of local growth factors stimulating tissue proliferation ([Datta](#page-17-0) et al. 1995). Finally, hormones linked to milk production, e.g. oxytocin, may stimulate the growth. Unfortunately, separating these effects is difficult. Injection of hormones, like prolactin, affect food intake ([Noel & Woodside 1993\)](#page-21-0), so eliminating a secondary local effect stimulated by the elevated food intake would need animals to be pair-fed with noninjected controls, and these critical experiments have not yet been performed. [Hammond \(1997\)](#page-18-0), however, noted that hypertrophy of the gut is a generalized response to elevated nutritional demands (such as cold exposure) and the hormonal profile in these circumstances is completely different from that in lactation, favouring the hypothesis that changes are primarily stimulated by a local response, stimulated directly by the food intake.

Many other morphological changes occur during pregnancy and lactation, not least of which is the growth of mammary tissue during late pregnancy to facilitate milk production during the subsequent lactation. As might be expected, considerable attention has been paid to this growth process and its hormonal basis, particularly the roles of sex steroids progesterone

and 17*b*-oestradiol (reviewed in [Lamote](#page-20-0) et al. 2004). Not all changes in the lactating animal, however, include expansion of tissue sizes. In particular, there is a large reduction in the size of the adipose tissue stores ([Speakman & McQueenie 1996](#page-22-0); [Vernon & Pond](#page-23-0) [1997](#page-23-0)). These reductions in the size of adipose tissue stores have generally been interpreted as withdrawal of stored energy to support energy delivery during the lactation event. Certainly, in some species (like the cotton rat: [Randolph](#page-21-0) et al. 1977 and the Siberian hamster: [Weiner 1987\)](#page-23-0), there is accumulation of fat during pregnancy that is withdrawn later in lactation and this strategy appears to reduce the peak food intake demands in lactation (see above and [table 1\)](#page-5-0). However, in other animals, the contribution of fat to the overall energy budget is relatively trivial. In mice, for example, the fat stores decline by approximately 2 g during lactation ([Johnson](#page-19-0) et al. 2001 $c$ ) equivalent to approximately 80 kJ of energy, compared with a metabolizable energy intake over the last 10 days of lactation of approximately 2600 kJ.

The increasing recognition that adipose tissue is not only an energy store but also an endocrine regulator suggests an alternative explanation of the reduction in fat mass may be that this reduces production of leptin (and possibly other adipokines) which then act to stimulate food intake. That low levels of leptin during lactation are involved in stimulating food intake has been demonstrated by repleting leptin levels using miniosmotic pumps. This provision of exogenous leptin blunts the level of increase in food intake during lactation ([Stocker](#page-22-0) et al. 2004). Decreased leptin levels may also be important in signalling reductions in the UCP-1 gene expression in brown adipose tissue of lactating rodents, the significance of which will be addressed below. Elevated levels of circulating adipokines are often inferred to underlie the links between obesity and disease. For example, high leptin levels, independent of adiposity, are an increased risk factor for the development of the metabolic syndrome in humans ([Franks](#page-18-0) *et al.* 2005). By reducing the levels of production of these compounds, lactating animals may actually derive an advantage relative to non-lactating individuals.

It is often suggested that a direct impact of this remodelling of the internal architecture during lactation is an increase in the resting metabolic rate relative to non-breeding animals. This occurs in theory because the animal expands the size of tissues that have high rates of metabolism (e.g. the liver and alimentary tract; Field et al[. 1939](#page-18-0); [Krebs 1950\)](#page-19-0), while simultaneously decreasing the size of other tissues with low energy demands. This is the direct energy cost linked to the lactation process of making these morphological modifications. When comparisons are made of the resting or basal energy expenditure in lactating small mammals compared with non-breeding individuals, many studies have reported that the RMR of lactating animals is higher [\(Camas](#page-17-0) et al. 1982; [Stephenson &](#page-22-0) [Racey 1993](#page-22-0)a,[b](#page-22-0); [Garton](#page-18-0) et al. 1994; [Poppitt](#page-21-0) et al. 1994; [Harder](#page-18-0) et al. 1996; [Speakman & McQueenie 1996](#page-22-0); [McLean & Speakman 2000;](#page-20-0) [Johnson](#page-19-0) et al. 2001b; [Zenuto](#page-23-0) et al. 2002; Krol et al[. 2003](#page-19-0)a,[b](#page-19-0)). Although other studies, particularly in non-domestic species, have

reported minor or even no changes (e.g. [Dryden](#page-17-0) et al. [1974](#page-17-0); [Turbill & Geiser 2006\)](#page-23-0), this might be anticipated in those groups where the increase in food intake in lactation is more modest [\(table 1](#page-5-0)) and thus requires correspondingly minor adjustments in morphology. However, if the theoretical basis of the effect is that organs such as the alimentary tract and liver require greater maintenance costs (Field et al. [1939,](#page-18-0) [Krebs](#page-19-0) [1950\)](#page-19-0), then one would anticipate that differences between lactating individuals in the sizes of these organs would have impacts on metabolism, and corresponding impacts on food intake and lactation performance. However, when such associations have been sought, they have generally not been found (reviewed in [Speakman](#page-22-0) et al. 2004). It remains uncertain why these expected trends are not observed, but this lack of an association calls into question the cost of the tissue remodelling.

#### 3. INDIRECT COSTS

#### (a) Optional compensatory costs

## (i) Thermoregulatory demands

During lactation, many species of small mammals experience large morphological and biochemical changes in their interscapular brown adipose tissue (BAT). These modifications include reductions in the amount of BAT in mice, rats, ground squirrels and hamsters ([Agius & Williamson 1980](#page-16-0); [Wade](#page-23-0) et al. 1986 [Johnson](#page-19-0) et al. 2001b). In addition to reductions in overall tissue mass, there are also reductions in BAT mitochondrial mass ([Trayhurn](#page-22-0) et al. 1982; [Trayhurn &](#page-22-0) [Jennings 1987](#page-22-0)*a*,*[b](#page-22-0)*). In late lactation, mitochondriaspecific content of uncoupling protein  $1$  (UCP-1) is reduced to only 8% of the level found in non-breeding mice (Trayhurn & Jennings [1987](#page-22-0)a,[b](#page-22-0), [1988](#page-22-0)) and to 26% in ground squirrels ([Nizielski](#page-21-0) et al. 1993). GDP binding, which is a measure of mitochondrial thermogenic capacity, is also reduced in mice and rats ([Trayhurn](#page-22-0) et al. 1982) and ground squirrels ([Nizielski](#page-21-0) et al[. 1993](#page-21-0)), but not in hamsters ([Wade](#page-23-0) et al. 1986). Brown adipose tissue is the key thermogenic organ in small rodents [\(Cannon & Nedergaard 2004\)](#page-17-0) and the changes observed in lactating rodents mediate a reduction in the noradrenaline-induced non-shivering thermogenesis ([Trayhurn](#page-22-0) et al. 1982; [Trayhurn 1983\)](#page-22-0), which is rapidly reversed upon weaning (Trayhurn & Jennings [1987](#page-22-0) $a,b$  $a,b$ , [1988\)](#page-22-0). In rats, the extent of decrease in thermogenic capacity is related to litter size ([Isler](#page-19-0) et al. [1984\)](#page-19-0), but this does not appear to be the case in mice ([Trayhurn & Wusterman 1987](#page-22-0) $a,b$  $a,b$ ). These morphological, physiological and biochemical changes in BATappear to be controlled by reduced sympathetic activity in lactation ([Trayhurn & Wusterman 1987](#page-22-0) $a,b$  $a,b$ ), which may be responsive to elevated corticosteroid levels ([Vernon & Flint 1983](#page-23-0)). Treatment of lactating rats with exogenous leptin completely reversed the downregulation of UCP-1 gene expression in BAT (Xiao et al. [2004](#page-23-0)). Changes in other aspects of BAT physiology are also apparent during lactation, including reduction in the activity of iodothyronine 5'-deiodinase, which catalyses conversion of thyroxine (T4) to triiodothyronine ([Giralt](#page-18-0) et al. 1986). All these changes are consistent with small lactating animals attempting

to reduce obligatory heat production from BAT. [Trayhurn \(1989\)](#page-22-0) interpreted this reduction as an energy-saving mechanism that increased the efficiency of milk production. However, in the light of our studies of limits to food intake in lactating mice described above, an alternative interpretation is that this downregulation does not save energy which can be used for milk production, but rather reduces the heat burden on the animal, allowing it to elevate milk production.

In recent years, a number of additional uncoupling proteins have been described (UCP-2 to UCP-5). These uncoupling proteins have different tissue distributions, with UCP-2 being very widespread, UCP-3 being restricted to BAT and skeletal muscle, and both UCP-4 and UCP-5 being found only in the brain. The role of these UCPs in resting and thermogenic heat production has been an issue of debate (e.g. Erlanson-Albertsson [2002,](#page-18-0) [2003\)](#page-18-0). Studies of the UCP-1 knockout mouse indicate that the other UCPs cannot reverse the lack of thermogenic capacity brought about by the absence of UCP-1 (Enerbäck et al. 1997; [Golozoubova](#page-18-0) et al. 2001; [Nedergaard](#page-20-0) et al. 2001). Although UCP-3 cannot be facultatively upregulated to replace the function of UCP-1, when it is transgenically overexpressed, the resultant mice have elevated resting metabolism ([Clapham](#page-17-0) et al. 2000). Surprisingly, given the suggested absence of any role for natural levels of UCP-3 in thermogenesis, it has recently been shown that UCP-3 is also downregulated enormously in BAT during lactation, and this is reflected in reduced protein levels as well (Pedraza et al. [2000,](#page-21-0) [2001](#page-21-0); Xiao [et al.](#page-23-0) 2004), but UCP-2 is unchanged ([Pedraza](#page-21-0) et al. 2001). Levels of UCP-3 gene expression in muscle are also decreased in lactation (Xiao [et al.](#page-23-0) 2004). These effects appear to reflect circulating levels of free fatty acids ([Pedraza](#page-21-0) et al. [2000](#page-21-0)). Possibly, UCP-3 is thermogenically insignificant but it is incidentally downregulated because the mechanisms for regulating UCP-1 and UCP-3 are similar. There may be consequences of this downregulation, which the animal cannot avoid ([Cadenas](#page-17-0) [et al.](#page-17-0) 2002), and will be discussed below under 'consequential costs'.

Many small mammals are able to make profound savings of energy by relaxing thermoregulation at normothermic levels and entering periods of torpor, during which body temperature is commonly reduced to levels just above ambient [\(Heldmaier](#page-19-0) et al. 2004). While bringing significant benefits in terms of energy saving, torpor is fundamentally incompatible with some of the processes of reproduction as was first demonstrated in a series of elegant experiments in the early 1970s, where bats were forced into torpor for varying periods with the consequence that gestation period was extended by the exact same period that the mice had been made to spend in torpor ([Racey 1973\)](#page-21-0). Foetal growth therefore halts when bats enter torpor. The significance of this effect was demonstrated in wild populations of bats during the early 1980s, when it was shown that adverse weather conditions during early pregnancy slowed foetal growth in wild bat populations resulting in significant year-to-year differences in the duration of pregnancy ([Racey & Swift 1981\)](#page-21-0). These findings seem to be at odds with the fact that hibernating bears gestate and lactate. However, studies of body temperature in bears during lactation reveal that while metabolism is downregulated to a similar extent as in small mammals, body temperatures only cool to approximately  $30-32^{\circ}$ C. If the interruption of foetal growth is temperature mediated, this difference in body temperatures during torpor may explain how bears are able to combine their reproductive functions with hibernation. Despite these apparently negative impacts of torpor in pregnancy, it is used frequently in some species of marsupial (e.g. dunnarts (Sminthopsis macroura): [Geiser](#page-18-0) et al. 2005; and mulgaras (Dasycercus cristicauda): [Geiser & Masters 1994](#page-18-0)), although in these animals the body temperature is still defended at approximately  $14^{\circ}$ C.

The impact of torpor on lactation is less clear. Many species appear to use periodic entry into torpor as an energy-saving mechanism during lactation ([Geiser](#page-18-0) [1994;](#page-18-0) Turbill et al[. 2003\)](#page-23-0) and energy budgeting indicates that this effect may be sufficiently great that the species concerned only need to make minor increases in food intake during lactation to achieve an overall energy balance. For example, the mean dry food consumption of non-reproductive brown long-eared bats averaged 1.8 g/day (48 kJ d<sup>-1</sup>) while lactating bats ate 2.0 g each day (53 kJ), yet the bats in lactation were able to export 22 kJ  $d^{-1}$  of this intake as milk ([Mclean & Speakman 1999](#page-20-0)) while deriving only on average 1.2 kJ from stored fat. Physiological compensation of energy budgets in this manner, so that food intake requirements are unaltered, may clearly have profound effects on the ecological costs of reproduction. Brown long-eared bats in the wild do not increase their flight times between pregnancy and lactation ([Entwistle](#page-17-0) et al. 1996), a pattern repeated in other species such as the common pipistrelle (Pipistrellus pipistrellus (= pygmaeus); [Swift 1980\)](#page-22-0) and long-tailed bats (Chalinolobus tuberculatus; [O'Donnell 2002](#page-21-0)). If predation risk during reproduction is a simple function of time spent flying, then for these animals there may be no ecological cost at all. This pattern however may be peculiar to bats under particular energy stress (e.g. Turbill et al[. 2003](#page-23-0)) or at the margins of their distributions ([Speakman](#page-22-0) et al. 1991), since studies of other bat species more centrally in their distributions yield different patterns. Big brown bats (Eptesicus fuscus) and little brown bats (Myotis lucifugus), for example, have increased food intake during lactation (Kurta et al. [1989,](#page-20-0) [1990\)](#page-20-0) and a decreased tendency to enter torpor during lactation [\(Audet & Fenton 1988;](#page-16-0) [Hamilton & Barclay 1994;](#page-18-0) [Grinevitch](#page-18-0) et al. 1995; [Lausen & Barclay 2003\)](#page-20-0). Flight time in northern bats in Sweden is almost doubled in lactation compared with non-breeding individuals ([Rydell 1993](#page-21-0)).

The fact that not all bats use torpor during lactation suggests that there are significant costs associated with its use. One possibility is that torpor is incompatible with milk synthesis. Studies using explants of mammary tissue from bats confirm that they have no special protection from reduced milk synthesis as temperature declines compared with explants of rodent mammary tissue (Wilde et al[. 1999](#page-23-0)). Using torpor in lactation by bats at the margins of their distributions may be a necessity



Figure 5. (a) Food intake and (b) wheel-running activity of rats during pregnancy and lactation. Days are expressed relative to the day of parturition (day  $= 0$ ) also indicated by the dotted line. Data are split between those raising large ( $n>6$ ; open squares) and those raising small ( $n<6$ ; filled squares) litters. Note how food intake in the larger litters increases dramatically a few days before weaning probably due to intake by the offspring (plots from data presented in tables 2 and 3 of [Slonaker 1924](#page-22-0)).

forced on them by the short duration of the night which constrains their capacity to extend flight times.

In spite of the energetic benefits and widespread use of torpor by bats in lactation, some groups that are otherwise capable of torpor appear to avoid using it during lactation. For example, longitudinal records of body temperature in free-ranging female short-beaked echidna (Tachyglossus aculeatus) indicate that they defend a continuous but low body temperature throughout the summer unless they lose their offspring ([Nicol](#page-21-0) et al. 2005). Although many species of rodent are known to hibernate and use daily torpor, no species of rodent has ever been reported to use torpor during lactation. Whether torpor and lactation are mutually incompatible in rodents was addressed by [Stamper](#page-22-0) et al[. \(1998\)](#page-22-0) who administered 2-deoxy-D-glucose (2-DG), a glucose analogue that interferes with cellular glycolysis, to lactating and non-lactating Siberian hamsters. 2-DG induced torpor in both groups, although the duration of torpor tended to be shorter in lactating animals. Evidence suggested that pups were able to obtain milk from torpid mothers, but whether any milk synthesis occurred was uncertain. Although apparently physiologically capable of entering torpor females that were subjected either to a combination of brief food deprivation and food restriction or just food restriction failed to display torpor, but instead killed one or more of their pups [\(Stamper](#page-22-0) et al. 1998).

#### (ii) Physical activity

In a classic paper, [Slonaker \(1924\)](#page-22-0) detailed the changes in physical activity that occurred in female

rats during reproduction. Plots of some of the meticulously reported data are shown in figure 5. These data show that rats have similar patterns of food intake to mice during pregnancy and lactation, and that wheel-running activity is dramatically suppressed during lactation to approximately onethird to one-half of that in pregnancy. The suppression of activity however was not proportional to litter size—since rats with small litters  $(n < 6$  offspring) ran about the same amount as those with large litters  $(n>6$  offspring) (figure 5). Mice (MF1) also show similar reductions in spontaneous physical activity during lactation [\(Speakman](#page-22-0) et al. 2001). Rabbits also increase the time spent resting as lactation progresses [\(Fernandez-Carmona](#page-18-0) et al. 2005).

Equivalent data on physical activity for nondomestic animals are scarce. This is probably because most activity in wild animals concerns foraging activity, which is generally increased during lactation owing to the elevated food intake requirements (see above). In domestic animals, physical activity and food intake are not inescapably linked, so modulations of activity without affecting food intake are feasible. However, animals engage in other activities that are not associated with foraging, e.g. grooming. We have monitored the behaviour of bats during lactation and found that brown long-eared bats (Plecotus auritus) significantly reduce the amount of time spent grooming when compared with non-breeding females occupying the same roost. On average non-lactating bats were observed grooming in 26.9% of the behaviour records, while lactating bats were only observed grooming in 11.1% of records. Similar high levels of time spent grooming in non-lactating bats have been observed in some other species ([Shen & Lee 2000](#page-22-0); [Fleming](#page-18-0) et al. [1998](#page-18-0)). However, such high levels are not universal ([Winchell & Kunz 1996](#page-23-0)) and reductions in grooming during lactation are also not observed in these species ([Winchell & Kunz 1996\)](#page-23-0). Female long-eared bats groom their offspring during lactation but this time allocation was relatively small and even when this was taken into account, the lactating females we studied still spent less than half of the time spent grooming by nonbreeding individuals ([McLean & Speakman 1997\)](#page-20-0). Grooming behaviour by bats is energetically expensive [\(Giorgi](#page-18-0) et al. 2001) and plays a role in the removal of ectoparasites. By reducing the time spent grooming, lactating bats may release significant amounts of energy for lactation, but may pay a price in terms of elevated ectoparasite burden. In the wild, in the mouse-eared bat (Myotis myotis), ectoparasite burden increases during pregnancy and lactation [\(Christe](#page-17-0) et al. 2000) when it is also related to body mass—suggesting a role for energy balance in driving ectoparasite burden. However, potential interactions here are complex. [Neuhaus \(2003\)](#page-20-0) experimentally removed ectoparasites (mainly fleas) from female Columbian ground squirrels using a commercially available insecticide. Removing parasites led to an increase in female body condition during lactation and at weaning and an increase in weaned litter size. Hence, removing ectoparasites may be costly, but not removing them may impose other costs (see also [Khokhlova](#page-19-0) et al. 2002).

## (iii) Immune system

The adaptive interplay between immune function and reproduction has been extensively studied in birds (Cichoń et al. 2001; [Martin](#page-20-0) et al. 2008). Relatively little attention has been paid to this trade-off in small mammals. There is some evidence that particularly energetically costly phases of reproduction in small mammals lead to impaired immune function. This effect can be magnified if the reproductive event coincides with a period of food shortage. If rats are experimentally malnourished, for example, during lactation there is a marked increase in the numbers of binucleate lymphocytes in the maternal spleen ([Ortiz](#page-21-0) et al[. 1995\)](#page-21-0), increasing from 11.5% of cells in nonlactating controls to 21.5% in the malnourished lactating animals. Since the spleen is an important lymphopoetic organ in rodents, this direct cell damage during energy restriction in lactation may have important negative impacts on the mother's immune system. Direct evidence of this impact of restriction was the experimental observation that malnourished rats in lactation had an impaired response to cholera toxin (Fló et al[. 1994](#page-18-0)).

Even during normal pregnancy, there are significant effects on the immune system. Committed B-lymphocyte precursors in the bone marrow of pregnant mice decline by 6.5 days into the pregnancy and by parturition they are down to only 10% of control levels [\(Medina](#page-20-0) et al. 1993). Newly formed B cells were also reduced, but cells carrying myeloid and erythroid markers were not. This indicates the production and export of B cells during pregnancy is much reduced. These effects could be mimicked by oestrogen treatment—suggesting an endocrine sex steroid-based mechanism of control [\(Medina](#page-20-0) et al. 1993).

However, the reasons for this effect are unclear, because the energy demands of maintaining the immune system are generally regarded to be trivial ([Derting & Compton 2003\)](#page-17-0). One possibility is that the reduction in body fatness at peak lactation itself mediates reduced immunity, since studies in voles and hamsters have shown that surgical removal of body fat decreases immune capability ([Demas](#page-17-0) et al. 2003; [Demas 2004](#page-17-0)). A mechanism for this effect might involve changes in circulating levels of leptin, as it has direct effects on immune cells stimulating T-cell immunity, phagocytosis, cytokine production and haemopoiesis, resulting in attenuated susceptibility to infectious insults ([Ingvartsen & Boisclair 2001\)](#page-19-0). In that case, the immunocompetence cost of reproduction may not be a compensatory cost but an obligatory consequential cost of the reduced fat content, which may itself be part of the mechanism of endocrine control of lactational food intake or manipulation of heat dissipation limits by effects on UCPs (see above).

## (b) Obligatory consequential costs

## (iii) Hyperthermia

It has been known for at least 50 years that high ambient temperatures and humidity, or exposure to solar radiation, all negatively affect milk production in dairy cows (Bos taurus; [Cobble & Herman 1951;](#page-17-0) [Brody](#page-17-0) et al. [1958\)](#page-17-0). Similar negative effects of high ambient temperatures have been reported in other large domestic



Figure 6. Body temperatures of a single female MF1 mouse measured at 1-minute intervals using an implanted body temperature transmitter (e-mitter Minimitter Inc) for 7 days prior to breeding and for 7 days at peak lactation. Data are screened to include only times when the animals are at rest, and then averaged over 30-minute periods within each day. The error bars refer to the s.d. for day-to-day variability  $(n=7)$ for each data set). During lactation, the body temperature is consistently elevated approximately  $1.5^{\circ}$ C higher than that prior to breeding during the light phase and much of the dark phase. Dark bar indicates period when lights out (J. R. Speakman, K. Drerrer, F. J. Munro & C. T. Troup 2005, unpublished data).

animals such as pigs (Sus scrofa: [Black](#page-16-0) et al. 1993; [Quiniou & Noblet 1999](#page-21-0); [Renaudeau & Noblet 2001;](#page-21-0) [Renaudeau](#page-21-0) et al. 2003) and sheep (Ovis aries: [Abdalla](#page-16-0) et al. [1993](#page-16-0)). Many of these larger animals also exhibit chronic hyperthermia during lactation (e.g. sows, [Ulmershakibaei & Plonait 1992\)](#page-23-0). The risk of hyperthermia in lactation therefore appears to be a significant obligatory cost in larger mammals. Heat dissipation difficulties found in large domestic animals may be far less significant in small mammals owing to their more favourable surface-to-volume ratios. Nevertheless, direct measurements of maternal body temperatures (figure 6) confirm that lactating mice are continuously hotter than their non-reproducing equivalents. Similar effects have been observed in rats [\(Croskerry](#page-17-0) et al. 1978; Leon et al. [1978](#page-20-0), [1985;](#page-20-0) [Kittrell & Satinoff 1988\)](#page-19-0) and Siberian hamsters Phodopus sungorus [\(Scribner &](#page-22-0) [Wynne Edwards 1994](#page-22-0)a). Rabbits also show negative effects of high temperatures on food intake and pup growth ([Marai](#page-20-0) et al. 2001).

Studies of blood flow during lactation indicate that there is no major redirection of blood away from the mammary glands to facilitate heat loss (cattle, [Lough](#page-20-0) et al[. 1990](#page-20-0); goats Capra hircus, Sano et al. [1985;](#page-22-0) rabbits, [Lublin & Wolfenson 1996\)](#page-20-0). There is a substantial body of behavioural literature following the pioneering work of Leon et al. [\(1978\)](#page-20-0), which suggests that suckling behaviour of small mammals may be influenced by the risks of maternal hyperthermia. These studies included observations that rats terminated suckling bouts when their body temperatures rose. Similar effects have been reported in larger lactating animals that have contact with their offspring such as sows ([Renaudeau & Noblet](#page-21-0) [2001](#page-21-0)), where elevated temperature led to reduced durations and greater frequencies of suckling bouts coupled with lowered milk production and piglet growth. Perhaps the best evidence comes from direct experimental manipulations of body temperature and examination of the subsequent effects on suckling behaviour. These experiments have only been performed on rats, but have involved two separate manipulations. Direct heating of the preoptic area of the brain, which led to premature termination of suckling bouts ([Woodside & Leon 1980](#page-23-0)), and injection of rats with morphine elevates body temperature and causes disruption of maternal suckling behaviour [\(Bridges & Grimm 1982\)](#page-17-0). However, rats are an order of magnitude larger than mice, so perhaps of greater relevance are the studies of Siberian hamsters, which show that during the daytime, the time spent with the litter in late lactation may be constrained by ambient temperature ([Scribner & Wynne Edwards 1994](#page-22-0)b). Interestingly, the two subspecies of Siberian hamster  $(P.$  sungorus sungorus and  $P.$  sungorus campbelli) differ in the rates at which the offspring develop their own thermoregulatory capacities—faster in  $P$ . s. campbelli ([Newkirk](#page-20-0) et al. 1998). This difference is reflected in greater and earlier problems in maintaining body temperature during suckling bouts by female  *s.* campbelli ([Scribner & Wynne Edwards 1994](#page-22-0)b) and consequent negative effects on pup growth in this subspecies ([Newkirk](#page-20-0) et al. 1998).

Despite a large literature indicating that there are direct effects of the litter on maternal hyperthermia, there are some contradictory data. For example, the body temperatures at which rats discontinue suckling are generally lower than the levels they tolerate while exercising outside the nest ([Kittrell & Satinoff 1988\)](#page-19-0). Perhaps more important are experimental inductions of reduced body temperature using sodium salicylate, which did not extend suckling bouts [\(Bates](#page-16-0) *et al.* 1985). Although treatment of lactating rats with morphine simultaneously elevates body temperature and disrupts maternal suckling (above), the negative effect of morphine appears to be independent of its effects on body temperature. This is shown by experiments in which morphine was administered with naloxone, an opioid receptor antagonist. Blocking morphine with naloxone reversed the negative effects on maternal behaviour [\(Bridges & Grimm 1982\)](#page-17-0) but not maternal hyperthermia (Cox et al. [1976\)](#page-17-0). Rats with experimentally induced increases in body temperature using morphine and naloxone combinations ([Stern & Azzara](#page-22-0) [2002](#page-22-0)) did not shorten their suckling bouts. However, this latter manipulation was performed only 7 days into lactation, and it would be instructive to know whether a similar absence of the effects of combined morphine/ naloxone treatment was apparent later in lactation when pup heat stress is more profound. Finally, rats fed low-quality protein in their diets did not show reduced performance when raised at  $30^{\circ}$ C compared with  $20^{\circ}$ C ([Jansen & Binard 1991\)](#page-19-0), even though their litters probably generated similar heat dissipation problems. Overall, there is a definite negative effect of ambient temperature on lactation performance of large animals, but the significance of these effects in smaller animals like mice remains uncertain.

## (iv) Disruption of sleep patterns

Anyone who has had a child of their own is aware that one of the key features of sharing ones life with a small neonate is the disruption of adult sleep patterns to accommodate suckling demands of the offspring

[\(Shinkoda](#page-22-0) et al. 1999). Only approximately 23% of infants sleep through entire nights (defined as from 0000 to 0500 h) by eight weeks of age ([Pinilla & Birch](#page-21-0) [1993\)](#page-21-0). Breast feeding mothers exhibit stronger delta and theta power spectra during NREM sleep ([Nishihara](#page-21-0) et al. 2004), but this pattern is the same on all nights whether sleep is disrupted or not to attend to their offspring, suggesting that there is an association between the sleep patterns and the hormonal changes in lactation. This is supported by observations that slow wave sleep duration is approximately three times longer in females who breast feed compared with agematched and body condition-matched individuals who bottle fed their offspring [\(Blyton](#page-16-0) et al. 2002). Effects of disruption of sleep patterns as a physiological cost of lactation in animals has been virtually unstudied. In black tufted ear marmosets (Callithrix kuhlii), the presence of young infants significantly impacted the sleep of mothers who were awake more than three times as often as mothers without infants (Fite et al[. 2003\)](#page-18-0). Surprisingly, there was no impact on the sleep patterns of fathers—despite the fact that callitrichid mothers relinquish the care of their offspring to fathers and alloparents very early in infant life. The impact of the disruption of sleep on lactating female animals is unknown.

#### (v) Bone loss

During pregnancy and lactation, the female animal must deliver sufficient calcium to her offspring to ensure adequate bone development. During pregnancy, females increase their absorption of calcium in the alimentary tract ([Halloran & DeLuca 1980\)](#page-18-0). Although urinary losses also increase [\(Omi & Ezawa](#page-21-0) [2001](#page-21-0)), the elevated absorption leads to elevated calcium levels in the blood (hypercalciuria), which persist throughout reproduction (Sarli et al. [2005\)](#page-22-0). In the first trimester of pregnancy, this extra uptake may actually exceed the foetal growth requirements. Consequently, there is a phase of bone accretion in the earliest part of pregnancy (Zeni et al[. 1999](#page-23-0); [Omi &](#page-21-0) [Ezawa 2001\)](#page-21-0), which is characterized by increases in markers of bone turnover (like osteocalcin and alkaline phosphatase levels; [Sengupta](#page-22-0) et al. 2005). This bone accretion appears to act as a temporary storage of calcium that can be withdrawn as the pregnancy proceeds and the foetal demands for calcium increase. By the end of pregnancy, most studies report significantly reduced bone mineral densities relative to controls, although Leopold et al[. \(1995\)](#page-20-0) indicated that levels were still elevated at day 20 in rats.

In lactation, the requirements for calcium are much greater, in line with the much larger offspring (as is also the case for energy and protein requirements). Lactating mothers reduce their levels of urinary calcium excretion (Sarli et al[. 2005\)](#page-22-0) to facilitate secretion into the milk. Although food intake in lactation is greatly elevated, lactating animals derive a large amount of the calcium that is exported in milk from their own skeletons. This leads to a substantial reduction in the thickness of bones from across the whole body [\(Miller & Bowman 2004](#page-20-0)). An additional direct effect of the calcium withdrawal from bone is a reduction in dentine and enamel mineralization in the

teeth of lactating rats [\(Ozbek](#page-21-0) et al. 2004). On the face of it, this use of skeletal calcium to provide the requirements for pup development may appear to indicate a shortfall in the available calcium from the diet, relative to the energy intake. However, three factors suggest that this is not the case. First, simple budgets of calcium availability relative to how much appears in the offspring (see above) indicate that it should not be limiting on most unsupplemented diets. Second, supplementing the dietary intake with calcium has no impact on the extent of calcium withdrawal from bone ([Weisstaub](#page-23-0) et al. 2003; [Kovacs & Fuleihan 2006\)](#page-19-0). Finally, in rats the extent of withdrawal is independent of litter size (at least comparing two to six pups), and hence neonatal calcium requirements [\(Sengupta](#page-22-0) et al. [2005](#page-22-0)), although Tojo et al[. \(1998\)](#page-22-0) did suggest that litter size correlated with the extent of withdrawal when data for non-lactating animals (litter size $=0$ ) were included in the regression. In spite of this, the diet supplementation data strongly suggest that the loss of calcium from bone is not driven by a shortfall in dietary intake.

Although supplementing calcium in the diet does not impact on bone resorption, providing less dietary calcium increases the rate of bone turnover [\(Gruber &](#page-18-0) [Stover 1994](#page-18-0)). This is indicated indirectly by increases in the markers of turnover ([Sengupta](#page-22-0) et al. 2005) and also directly from labelling studies. For example, by pre-labelling bone tissue of lactating female rats with  $45$ Ca, it was shown that under a calcium-deficient diet, 40% of the label was lost from the whole skeleton over the course of a normal pregnancy and lactation cycle with losses in some bones like the femur being 75% (Wang et al[. 1994\)](#page-23-0). Under situations where the diet naturally contains low calcium levels (e.g. fruit and insects), lactating and pregnant individuals may seek out other sources to keep their calcium supply high (e.g. fruit bats, [Ruby](#page-21-0) et al. 2000, [Nelson](#page-20-0) et al. 2005 and insectivorous bats, [Adams](#page-16-0) et al. 2003).

A primary hypothesis therefore is that the endocrine mechanisms that promote the secretion of calcium into the milk are incompatible with the retention of calcium in bone. Several lines of evidence support this hypothesis. First, prolactin, which is one of the key hormones necessary to support successful milk production in the lactating female, and the prolactin receptor, are also found in the synovial fluid [\(Ogueta](#page-21-0) et al. 2002). Studies of the impact of prolactin on mesenchymal stem cells strongly implicate it as playing a role in bone and cartilage formation and repair processes. Prolactin receptor KO mice (Kelly *et al.* 2001) have greatly elevated prolactin concentrations and reduced bone formation. Prolactin secretion to support lactation may therefore influence bone resorption.

Ovariectomized rats also lose bone mass ([Miller &](#page-20-0) [Wronski 1993\)](#page-20-0). Consequently, absence of oestrogen may be a contributory factor in lactation [\(Miller &](#page-20-0) [Bowman 1998](#page-20-0)). This is supported by observations that oestrogen treatment of ovariectomized lactating rats improved bone retention in lactation by 15% [\(Garner](#page-18-0) et al[. 1991\)](#page-18-0), but the mechanism here is confused because these mothers also ate less and their pups grew less—so presumably other hormones related to lactation (e.g. prolactin) were also at lower levels. Another hormone in addition to prolactin that appears

important in the supply of calcium to the foetuses is the parathyroid hormone-related peptide (PTHrP; [Yamamoto](#page-23-0) et al. 1991). PTHrP has strong sequence similarities to parathyroid hormone (PTH) and both seem to interact with parathyroid hormone receptors located on osteoclasts, which they inhibit. PTHrP increases during late pregnancy and remains high throughout lactation (Sarli et al[. 2005](#page-22-0)) where it is produced in mammary epithelial cells. This promotes maternal bone resorption allowing calcium release for export in the milk. Production of mammary PTHrP appears to be sensitive to extracellular calcium levels acting through the calcium-sensing receptor. Involvement of PTH itself in the calcium withdrawal process seems unlikely since parathyroidectomy had no impact on the extent of bone resorption in lactation [\(Hodnett](#page-19-0) et al. 1992).

In addition to hormones that support milk production having impacts on bone, there is also evidence of actions working in the reverse direction. For example, the osteoprotegerin ligand (OPGL) is a known osteoclast differentiation and activation factor that is essential for successful bone remodelling. Transgenic mice lacking OPGL (or its receptor) fail to develop lobular alveolar structures in their mammary glands resulting in an inability to secrete milk successfully. This effect can be reversed by recombinant OPGL treatment ([Fata](#page-18-0) et al. [2000](#page-18-0)). Mammary and bone function therefore appears to be intimately associated and this association, which may not necessarily be functional, leads to the obligatory bone loss during lactation. Following weaning, there is a rapid increase in osteoblast proliferation and cancellous bone deposition ([Miller](#page-20-0) et al. 2005). This osteoprogenitor pool may be primed during the late phase of lactation for this post-weaning activity. Peak bone formation in rats occurs approximately four weeks after the end of lactation ([Miller & Bowman 2004\)](#page-20-0), restoring the bone lost during the lactation event ([Nishiwaki](#page-21-0) et al. 1999; [Bowman](#page-16-0) et al. 2002) and its strength ([Vajda](#page-23-0) et al. 2001).

The physiological mechanism of lactation therefore exposes the lactating female to an elevated risk of osteoporosis, which may extend for at least twice the duration of the actual lactation event before levels are normalized. The direct effect of this calcium withdrawal from bone is a reduction in bone strength and this increases the risks of bone fracture. Direct measurements of bone physical properties support this interpretation. By the end of pregnancy, rat tibiae were not significantly different in their strength from controls, but by the end of lactation, bone strength was substantially reduced in both cortical and cancellous bone, paralleled by the decline in bone volume ([Vajda](#page-23-0) *et al.* 2001).

There are no direct measures of the increased risk of fracture during lactation in small mammals. However, in humans, there are reports of case studies where lactation-induced osteoporosis did lead to limb fracture [\(Clemetson](#page-17-0) et al. 2004) suggesting that its effects may be significant. Paradoxically repeated bouts of lactation appear to have a protective effect in humans on later life risks offracture attached to osteoporosis ([Hoffman](#page-19-0) et al. [1993](#page-19-0); [Jacobsen](#page-19-0) et al. 1998; [Michaelsson](#page-20-0) et al. 2001; [Petersen](#page-21-0) et al. 2002; [Naves](#page-20-0) et al. 2005). This suggests that in the post-weaning period regrowth of bone is

<span id="page-16-0"></span>overcompensated. There is some direct evidence to support this suggestion (Akesson *et al.* 2004).

## (vi) Oxidative stress

Animals during lactation have high rates of metabolism, which may result in elevated generation of free radicals. These free radicals may cause oxidative damage to macromolecules if the mice do not simultaneously upregulate their oxidative defence and repair mechanisms. [Wiersma](#page-23-0) et al. (2004) showed that birds feeding nestlings, which is the most energetically expensive phase in avian reproduction, do not upregulate the levels of superoxide dismutase, catalase and glutathione peroxidase, the main defences against radical oxygen species. However, that study did not include measures of oxidative damage—so the lack of elevation of defence mechanisms may have been because there was no increase in free radical production to defend against. Indeed, the links between radical oxygen species production and metabolic rate are not straightforward [\(Speakman](#page-22-0) et al. 2002; [Speakman](#page-22-0) [2003](#page-22-0)), and in some cases increased metabolism may actually reduce oxidative stress [\(Speakman](#page-22-0) et al. 2004). However, the reduction in levels of UCP-1 and UCP-3 in lactation (above) would suggest that mitochondria in lactating animals are more closely coupled—supporting the suggestion that this may be a time of elevated oxidative stress ([Demin](#page-17-0) et al. 1998a,[b](#page-17-0); [Brand 2000](#page-17-0); [Speakman 2003](#page-22-0)). Studies of antioxidant protection levels and damage during reproduction are needed.

## 4. CONCLUSION

The physiological costs of reproduction are probably the most significant component underlying life-history trade-offs. In this review, I have summarized some of the work that has been performed to measure physiological costs in small mammals. Physiological costs can be divided into two different aspects. The first is direct costs of the reproductive event. These include the energy and nutrient costs of reproduction and the changes in organ morphology necessary to meet these demands. Considerable progress in elucidating the limits on energy budgets in small domesticated rodents have been made in the last 15 years or so. However, even in this area of intensive activity, we are still far short of being able to translate this knowledge into an understanding of a simple life-history trade-off between the number and size of offspring. Knowledge of these demands among non-domesticated species is very sparse, physiological information gleaned in a wild context is virtually non-existent. To these direct costs must be added indirect costs of reproduction. These include optional compensatory adjustments that animals make to divert more resource into reproduction. Such compensations include reductions in thermoregulation, physical activity and immunocompetence. In some species, these adjustments can be sufficient that total energy requirements during reproduction are no greater than experienced by nonbreeding animals. This may annul any ecological impact of reproduction mediated, for example, via increased predation risk due to extra time spent foraging. Their long-term impacts have however

never been explored. To these compensatory costs must be added obligatory costs, which are an inevitable side effect of reproduction that cannot be avoided. For small mammals these include hyperthermia, bone loss, disruption of sleep patterns and oxidative stress. Knowledge of all these effects is presently rudimentary. Indeed, whether some changes are optional or obligatory (like immunocompetence changes) is uncertain, and the list of costs presented here is unlikely to be complete.

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## REFERENCES

- Abdalla, E. B., Kotby, E. A. & Johnson, H. D. 1993 Physiological responses to heat-induced hyperthermia of pregnant and lactating ewes. Small Ruminant Res. 11, 125–134. [\(doi:10.1016/0921-4488\(93\)90145-8\)](http://dx.doi.org/doi:10.1016/0921-4488(93)90145-8)
- Adams, R. A., Pedersen, S. C., Thibault, K. M., Jadin, J. & Petru, B. 2003 Calcium as a limiting resource to insectivorous bats: can water holes provide a supplemental mineral source? *J. Zool.* 260, 189-194. ([doi:10.1017/](http://dx.doi.org/doi:10.1017/S0952836903003613) [S0952836903003613\)](http://dx.doi.org/doi:10.1017/S0952836903003613)
- Akesson, A., Vahter, M., Berglund, M., Eklof, T., Bremme, K. & Bjellerup, P. 2004 Bone turnover from early pregnancy to postweaning. Acta Obstet. Gyn. Scan. 83, 1049–1055. [\(doi:10.1111/j.0001-6349.2004.00428.x\)](http://dx.doi.org/doi:10.1111/j.0001-6349.2004.00428.x)
- Agius, L. & Williamson, D. H. 1980 Lipogenesis in interscapular brown adipose tissue of virgin, pregnant and lactating rats. The effects of intragastric feeding. Biochem. J. 190, 477-480.
- Atramentowicz, M. 1992 Optimal litter size—does it cost more to raise a large litter in Caluromys philander? Can. J. Zool. 70, 1511–1515.
- Audet, D. & Fenton, M. B. 1988 Heterothermy and the use of torpor by the bat Eptesicus fuscus (Chiroptera: Vespertilionidae): a field study. Physiol. Zool. 61, 197–204.
- Barclay, R. M. R. 1994 Constraints on reproduction by flying vertebrates—energy and calcium. Am. Nat. 144, 1021–1031. [\(doi:10.1086/285723\)](http://dx.doi.org/doi:10.1086/285723)
- Bates, A., Adels, L. & Leon, M. 1985 Thermal control of maternal contact bouts: the interbout interval. Physiol. Behav. 34, 835–837. [\(doi:10.1016/0031-9384\(85\)90386-5](http://dx.doi.org/doi:10.1016/0031-9384(85)90386-5))
- Biggerstaff, S. & Mann, M. 1992 Consummatory behaviors and weight regulation in pregnant, lactating, and pregnant-lactating mice. Physiol. Behav. 52, 485-491. ([doi:10.](http://dx.doi.org/doi:10.1016/0031-9384(92)90335-Y) [1016/0031-9384\(92\)90335-Y\)](http://dx.doi.org/doi:10.1016/0031-9384(92)90335-Y)
- Black, J. L., Mullan, B. P., Lorschy, M. L. & Giles, L. R. 1993 Lactation in the sow during heat-stress. Livest. Prod. Sci. 35, 153–170. ([doi:10.1016/0301-6226\(93\)90188-N\)](http://dx.doi.org/doi:10.1016/0301-6226(93)90188-N)
- Blyton, D. M., Sullivan, C. E. & Edwards, N. 2002 Lactation is associated with an increase in slow-wave sleep in women. *J. Sleep Res.* 11, 297-303. ([doi:10.1046/j.1365-](http://dx.doi.org/doi:10.1046/j.1365-2869.2002.00315.x) [2869.2002.00315.x\)](http://dx.doi.org/doi:10.1046/j.1365-2869.2002.00315.x)
- Bowman, B. M., Siska, C. C. & Miller, S. C. 2002 Greatly increased cancellous bone formation with rapid

<span id="page-17-0"></span>improvements in bone structure in the rat maternal skeleton after lactation. J. Bone Mineral Res. 17, 1954–1960. [\(doi:10.1359/jbmr.2002.17.11.1954](http://dx.doi.org/doi:10.1359/jbmr.2002.17.11.1954))

- Boyne, R., Fell, B. F. & Robb, I. 1966 The surface area of the intestinal mucosa in the lactating rat. *J. Physiol.* 183, 570–576.
- Brand, M. D. 2000 Uncoupling to survive? The role of mitochondrial inefficiency in ageing. Exp. Gerontol. 35, 811–820. [\(doi:10.1016/S0531-5565\(00\)00135-2](http://dx.doi.org/doi:10.1016/S0531-5565(00)00135-2))
- Bridges, R. S. & Grimm, C. T. 1982 Reversal of morphine disruption of maternal behavior by concurrent treatment with the opiate antagonist naloxone. Science 218, 166-168. ([doi:10.1126/science.7123227\)](http://dx.doi.org/doi:10.1126/science.7123227)
- Brody, S., Ragsdale, A. C., Yeck, R. G. & Worstell, D. 1958 Environmental physiology and shelter engineering with special reference to domestic animals XXXIII. Milk production, feed and water consumption and body weight of Jersey and Holstein cows in relation to several diurnal temperature rhythms. Mo AES Res. Bull. 1958, 578.
- Burdett, K. & Reek, C. 1979 Adaptation of the small intestine during pregnancy and lactation in the rat. Biochem.  $\ddot{f}$ . 184, 245–251.
- Cadenas, S. et al. 2002 The basal proton conductance of skeletal muscle mitochondria from transgenic mice overexpressing or lacking uncoupling protein-3. *J. Biol. Chem.* 277, 2773–2778. [\(doi:10.1074/jbc.M109736200](http://dx.doi.org/doi:10.1074/jbc.M109736200))
- Cairnie, A. B. & Bentley, R. E. 1967 Cell proliferation studies in the intestinal epithelium of the rat. Hyperplasia during lactation. Exper. Cell Res. 46, 428–440. ([doi:10.1016/](http://dx.doi.org/doi:10.1016/0014-4827(67)90079-1) [0014-4827\(67\)90079-1\)](http://dx.doi.org/doi:10.1016/0014-4827(67)90079-1)
- Camas, R., Romero, V. J. & Baldwin, R. L. 1982 Maintenance energy requirements during lactation in rats. *J. Nutr.* 112, 1876-1880.
- Campbell, R. M. S. & Fell, B. F. 1964 Gastro-intestinal hypertrophy in the lactating rat in relation to its food intake. *J. Physiol.* **171**, 90–97.
- Cannon, B. & Nedergaard, J. 2004 Brown adipose tissue: function and physiological significance. Physiol. Rev. 84, 277–359. [\(doi:10.1152/physrev.00015.2003\)](http://dx.doi.org/doi:10.1152/physrev.00015.2003)
- Cavaleros, M., Buffenstein, R., Ross, F. P. & Pettifor, J. M. 2003 Vitamin D metabolism in a frugivorous nocturnal mammal, the Egyptian fruit bat (Rousettus aegyptiacus). Gen. Comp. Endocrinol. 133, 109–117. ([doi:10.1016/](http://dx.doi.org/doi:10.1016/S0016-6480(03)00150-3) [S0016-6480\(03\)00150-3](http://dx.doi.org/doi:10.1016/S0016-6480(03)00150-3))
- Charlesworth, B. C. 1994 Evolution in age-structured populations, 2nd edn. Cambridge, UK: Cambridge University Press.
- Charnov, E. L. 1993 Life history invariants: some explorations of symmetry in evolutionary ecology. Oxford, UK: Oxford University Press.
- Christe, P., Arlettaz, R. & Vogel, P. 2000 Variation in intensity of a parasitic mite (Spinturnix myoti) in relation to the reproductive cycle and immunocompetence of its bat host (Myotis myotis). Ecol. Lett. 3, 207–212. ([doi:10.1046/](http://dx.doi.org/doi:10.1046/j.1461-0248.2000.00142.x) [j.1461-0248.2000.00142.x\)](http://dx.doi.org/doi:10.1046/j.1461-0248.2000.00142.x)
- Cichoń, M., Dubiec, A. & Chadzińska, M. 2001 The effect of elevated reproductive effort on humoral immune function in collared flycatcher females. Acta Oecol. 22, 71–76. ([doi:10.1016/S1146-609X\(00\)01094-8\)](http://dx.doi.org/doi:10.1016/S1146-609X(00)01094-8)
- Clapham, J. C. et al. 2000 Mice overexpressing human uncoupling protein-3 in skeletal muscle are hyperphagic and lean. Nature 406, 415–418. ([doi:10.1038/35019082](http://dx.doi.org/doi:10.1038/35019082))
- Clemetson, I. A., Popp, A., Lippuner, K., Ballmer, F. & Anderson, S. E. 2004 Postpartum osteoporosis associated with proximal tibial stress fracture. Skeletal Radiol. 33, 96–98. [\(doi:10.1007/s00256-003-0721-2\)](http://dx.doi.org/doi:10.1007/s00256-003-0721-2)
- Cobble, J. W. & Herman, H. A. 1951 The influence of environmental temperature on the composition of milk of the dairy cow. Mo AES Res. Bull., 485.
- Cox, B., Ary, M., Chesarkek, W. & Lomax, P. 1976 Morphine hyperthermia in the rat: an action on the central thermostats. Eur. J. Pharmacol. 36, 33-39. ([doi:10.1016/](http://dx.doi.org/doi:10.1016/0014-2999(76)90253-3) [0014-2999\(76\)90253-3](http://dx.doi.org/doi:10.1016/0014-2999(76)90253-3))
- Craft, I. L. 1970 The influence of pregnancy and lactation on the morphology and absorptive capacity of the rat small intestine. Clin. Sci. 38, 287–295.
- Cripps, A. W. & Williams, V. J. 1975 The effect of pregnancy and lactation on food intake, gastrointestinal anatomy and the absorptive capacity of the small intestine of the albino rat. Br. J. Nutr. 33, 17–32.
- Croskerry, P. G., Smith, G. K. & Leon, M. 1978 Thermoregulation and the maternal behaviour of the rat. Nature 273, 299–300. [\(doi:10.1038/273299a0\)](http://dx.doi.org/doi:10.1038/273299a0)
- Datta, U. K., Datta, A. N. & Mukherjee, S. 1995 Role of hyperphagia in structural changes of small intestine during lactation. Indian J. Physiol. Pharmacol. 39, 259-262.
- Degen, A. A., Khokhlova, I. S., Kam, M. & Snider, I. 2002 Energy requirements during reproduction in female common spiny mice (Acomys cahirinus). J. Mammal. 83, 645-651. [\(doi:10.1644/1545-1542\(2002\)083](http://dx.doi.org/doi:10.1644/1545-1542(2002)083%3C0645:ERDRIF%3E2.0.CO;2)<0645:ERD  $RIF > 2.0$ .CO;2)
- Degen, A. A., Khokhlova, I. S., Kam, M. & Snider, I. 2004 Water budget during reproduction in female common spiny mice (Acomys cahirinus). J. Mammal. 85, 1106–1110. [\(doi:10.1644/BRG-211.1\)](http://dx.doi.org/doi:10.1644/BRG-211.1)
- Demas, G. E. 2004 The energetics of immunity: a neuroendocrine link between energy balance and immune function. Horm. Behav. 45, 173–180. ([doi:10.1016/](http://dx.doi.org/doi:10.1016/j.yhbeh.2003.11.002) [j.yhbeh.2003.11.002](http://dx.doi.org/doi:10.1016/j.yhbeh.2003.11.002))
- Demas, G. E., Drazen, D. L. & Nelson, R. J. 2003 Reductions in total body fat decrease humoral immunity.Proc. R. Soc. B 270, 905–911. ([doi:10.1098/rspb.2003.2341](http://dx.doi.org/doi:10.1098/rspb.2003.2341))
- Demin, O. V., Kholodenko, B. N. & Skulachev, V. P. 1998a A model of  $O_2$  generation in the complex III of the electron transport chain. Mol. Cell. Biochem. 184, 21–33. ([doi:10.](http://dx.doi.org/doi:10.1023/A:1006849920918) [1023/A:1006849920918](http://dx.doi.org/doi:10.1023/A:1006849920918))
- Demin, O. V., Westerhoff, H. V. & Kholodenko, B. N. 1998b Mathematical modelling of superoxide generation with the bc(1) complex of mitochondria. Biochemistry (Mosc) 63, 634–649.
- Denis, R. G., Williams, G. & Vernon, R. G. 2003 Regulation of serum leptin and its role in the hyperphagia of lactation in the rat. *J. Endocrinol.* 176, 193-203. ([doi:10.1677/](http://dx.doi.org/doi:10.1677/joe.0.1760193) [joe.0.1760193\)](http://dx.doi.org/doi:10.1677/joe.0.1760193)
- Derting, T. L. & Compton, S. 2003 Immune response, not immune maintenance, is energetically costly in wild whitefooted mice (Peromyscus leucopus). Physiol. Biochem. Zool. 76, 744–752. ([doi:10.1086/375662](http://dx.doi.org/doi:10.1086/375662))
- Dryden, G. L., Gebczynski, M. & Douglas, E. L. 1974 Oxygen consumption by nursling and adult musk shrews. Acta Theriol. 19, 453–461.
- Drummond, H., Vazquez, E., Sanchez-Colon, S., Martinez-Gomez, M. & Hudson, R. 2000 Competition for milk in the domestic rabbit: survivors benefit from littermate deaths. Ethology 106, 511–526. [\(doi:10.1046/j.1439-0310.](http://dx.doi.org/doi:10.1046/j.1439-0310.2000.00554.x) [2000.00554.x\)](http://dx.doi.org/doi:10.1046/j.1439-0310.2000.00554.x)
- Dugas, M. C., Hazelwood, R. L. & Lawrence, A. L. 1970 Influence of pregnancy and/or exercise on intestinal transport of amino acids in rats. Proc. Soc. Exptl Biol. Med. 135, 127–128.
- Enerback, S., Jacobsson, A., Simpson, E. M., Guerra, C., Yamashita, H., Harper, M. E. & Kozak, L. P. 1997 Mice lacking mitochondrial uncoupling protein are coldsensitive but not obese. Nature 387, 90-94. ([doi:10.](http://dx.doi.org/doi:10.1038/387090a0) [1038/387090a0\)](http://dx.doi.org/doi:10.1038/387090a0)
- Entwistle, A. C., Racey, P. A. & Speakman, J. R. 1996 Habitat exploitation by a gleaning bat, Plecotus auritus. Phil. Trans. R. Soc. B 351, 921–931. ([doi:10.1098/rstb.1996.0085](http://dx.doi.org/doi:10.1098/rstb.1996.0085))
- <span id="page-18-0"></span>Erlanson-Albertsson, C. 2002 Uncoupling proteins—a new family of proteins with unknown function. Nutr. Neurosci. 5, 1–11. [\(doi:10.1080/10284150290007038](http://dx.doi.org/doi:10.1080/10284150290007038))
- Erlanson-Albertsson, C. 2003 The role of uncoupling proteins in the regulation of metabolism. Acta Physiol. Scand. 178, 405–412. ([doi:10.1046/j.1365-201X.2003.01159.x](http://dx.doi.org/doi:10.1046/j.1365-201X.2003.01159.x))
- Fata, J. E. et al. 2000 The osteoclast differentiation factor osteoprotegerin-ligand is essential for mammary gland development. Cell 103, 41–50. [\(doi:10.1016/S0092-](http://dx.doi.org/doi:10.1016/S0092-8674(00)00103-3) [8674\(00\)00103-3\)](http://dx.doi.org/doi:10.1016/S0092-8674(00)00103-3)
- Fell, B. F., Smith, A. & Campbell, R. M. 1963 Hypertrophic and hyperplastic changes in the alimentary canal of the lactating rat. J. Pathol. Bact. 85, 179–188. ([doi:10.1002/](http://dx.doi.org/doi:10.1002/path.1700850117) [path.1700850117\)](http://dx.doi.org/doi:10.1002/path.1700850117)
- Fernandez-Carmona, J. et al. 2005 The behaviour of farm rabbit does around parturition and during lactation. World Rabbit Sci. 13, 253–277.
- Field 2nd, J., Belding, H. S. & Martin, A. W. 1939 An analysis of the relation between basal metabolism and summated tissue respiration in the rat. I. The postpubertal albino rat. *J. Cell. Comp. Physiol.* 14, 143-157. ([doi:10.1002/jcp.1030140202\)](http://dx.doi.org/doi:10.1002/jcp.1030140202)
- Fink, R., Tauson, A. H., Hansen, K. B., Wamberg, S. & Kristensen, N. B. 2001 Energy intake and milk production in mink (Mustela vison)—effect of litter size. Arch. Anim. Nutr. 55, 221–242.
- Fisher, R. A. 1930 The genetical theory of natural selection. Oxford, UK: Clarendon Press.
- Fite, J. E., French, J. A., Patera, K. J., Hopkins, E. C., Rukstalis, M., Jensen, H. A. & Ross, C. N. 2003 Nighttime wakefulness associated with infant rearing in Callithrix kuhlii. Int. J. Primatol. 24, 1267-1280. ([doi:10.](http://dx.doi.org/doi:10.1023/B:IJOP.0000005992.72026.e6) [1023/B:IJOP.0000005992.72026.e6\)](http://dx.doi.org/doi:10.1023/B:IJOP.0000005992.72026.e6)
- Fleming, T. H., Nelson, A. A. & Dalton, V. M. 1998 Roosting behavior of the lesser long-nosed bat, Leptonycteris curasoae. J. Mammal. 79, 147–155. [\(doi:10.2307/1382849\)](http://dx.doi.org/doi:10.2307/1382849)
- Fló, J., Elías, F., Massouh, E. & Roux, M. E. 1994 Impairment of B and T cell maturation in gut associated lymphoid tissues due to malnutrition during lactation. Devel. Comparat. Immunol. 18, 543–555. ([doi:10.1016/](http://dx.doi.org/doi:10.1016/S0145-305X(06)80008-X) [S0145-305X\(06\)80008-X](http://dx.doi.org/doi:10.1016/S0145-305X(06)80008-X))
- Franks, P. W., Brage, S., Luan, J., Ekelund, U., Rahman, M., Farooqi, I. S., Halsall, I., O' Rahilly, S. & Wareham, N. J. 2005 Leptin predicts a worsening of the features of the metabolic syndrome independently of obesity. Obes. Res. 13, 1476–1484.
- Gandelman, R. & Simon, N. G. 1978 Spontaneous pupkilling by mice in response to large litters. Dev. Psychobiol. 11, 235–241. ([doi:10.1002/dev.420110307\)](http://dx.doi.org/doi:10.1002/dev.420110307)
- Garner, S. C., Anderson, J. J. B., Mar, M.-H. & Parikh, I. 1991 Estrogens reduce bone loss in the ovariectomized, lactating rat model. Bone Mineral 15, 19–31. ([doi:10.1016/](http://dx.doi.org/doi:10.1016/0169-6009(91)90108-C) [0169-6009\(91\)90108-C\)](http://dx.doi.org/doi:10.1016/0169-6009(91)90108-C)
- Garton, D. W., Hsu, M. J. & Harder, J. D. 1994 Environmental temperature and metabolic rates during gestation and lactation in golden-hamsters (Mesocricetus auratus). Physiol. Zool. 67, 497–514.
- Gebczynska, Z. & Gebczynski, M. 1971 Length and weight of the alimentary tract of the root vole. Acta Theriol. 26, 359–369.
- Geiser, F. 1994 Hibernation and daily torpor in marsupials—a review. Aust. J. Zool. 42, 1-16. [\(doi:10.1071/ZO9940001](http://dx.doi.org/doi:10.1071/ZO9940001))
- Geiser, F. & Masters, P. 1994 Torpor in relation to reproduction in the mulgara, Dasycercus cristicauda (Dasyuridae: Marsupialia). *J. Therm. Biol.* 19, 33-40. ([doi:10.1016/0306-4565\(94\)90007-8](http://dx.doi.org/doi:10.1016/0306-4565(94)90007-8))
- Geiser, F., McAllan, B. M. & Brigham, R. 2005 Daily torpor in a pregnant dunnart (Sminthopsis macroura Dasyuridae: Marsupialia). Mamm. Biol. 70, 117–121. ([doi:10.1016/](http://dx.doi.org/doi:10.1016/j.mambio.2004.06.003) [j.mambio.2004.06.003](http://dx.doi.org/doi:10.1016/j.mambio.2004.06.003))
- Genoud, M. & Vogel, P. 1990 Energy-requirements during reproduction and reproductive effort in shrews (Soricidae).  $J. Zool. 220, 41–60.$
- Giorgi, M. S., Arlettaz, R., Christe, P. & Vogel, P. 2001 The energetic grooming costs imposed by a parasitic mite (Spinturnix myoti) upon its bat host (Myotis myotis). Proc. R. Soc. B 268, 2071–2075. [\(doi:10.1098/rspb.2001.1686](http://dx.doi.org/doi:10.1098/rspb.2001.1686))
- Giralt, M., Villarroya, F., Mampel, T. & Iglesias, R. 1986 Impaired basal and noradrenaline induced isodothyrionine 5'-deiodinase activity in brown adipose tissue form pregnant and lactating rats. Biochem. Biophys. Res. Commun. 138, 1315–1321. [\(doi:10.1016/S0006-291X](http://dx.doi.org/doi:10.1016/S0006-291X(86)80426-0) [\(86\)80426-0](http://dx.doi.org/doi:10.1016/S0006-291X(86)80426-0))
- Glazier, D. S. 1985 Energetics of litter size in five species of peromyscus with generalisations for other mammals. J. Mamm. 66, 629–642. ([doi:10.2307/1380789](http://dx.doi.org/doi:10.2307/1380789))
- Goettsch, M. 1960 Comparative protein requirement of the rat and mouse for growth, reproduction and lactation using casein diets.  $\frac{4}{7}$ . Nutr. 70, 307–312.
- Golozoubova, V., Hohtola, E., Matthias, A., Jacobsson, A., Cannon, B. & Nedergaard, J. 2001 Only UCP1 can mediate adaptive nonshivering thermogenesis in the cold. FASEB 7. 15, U327-U340.
- Grinevitch, L., Holroyd, S. L. & Barclay, R. M. R. 1995 Sexdifferences in the use of daily torpor and foraging time by big brown bats (Eptesicus fuscus) during the reproductive season. *J. Zool.* 235, 301-309.
- Gruber, H. E. & Stover, S. J. 1994 Maternal and weanling bone—the influence of lowered calcium intake and maternal dietary history. Bone 15, 167–176. ([doi:10.](http://dx.doi.org/doi:10.1016/8756-3282(94)90704-8) [1016/8756-3282\(94\)90704-8](http://dx.doi.org/doi:10.1016/8756-3282(94)90704-8))
- Hacklander, K., Tataruch, F. & Ruf, T. 2002 The effect of dietary fat content on lactation energetics in the European hare (Lepus europaeus). Physiol. Biochem. Zool. 75, 19–28. ([doi:10.1086/324770](http://dx.doi.org/doi:10.1086/324770))
- Halloran, B. P. & DeLuca, H. F. 1980 Calcium transport in small intestine during pregnancy and lactation. Am. J. Physiol. 239, E64–E68.
- Hamilton, I. M. & Barclay, R. M. R. 1994 Patterns of daily torpor and day-roost selection by male and female big brown bats (Eptesicus fuscus). Can. J. Zool. 72, 744-749.
- Hammond, K. A. 1997 Adaptation of the maternal intestine during lactation. J. Mammary Gland Biol. Neoplasia 2, 243–252. [\(doi:10.1023/A:1026332304435](http://dx.doi.org/doi:10.1023/A:1026332304435))
- Hammond, K. A. & Diamond, J. 1992 An experimental test for a ceiling on sustained metabolic rate in lactating mice. Physiol. Zool. 65, 952–977.
- Hammond, K. A. & Diamond, J. 1994 Limits to dietary nutrient uptake and intestinal nutrient uptake in lactating mice. Physiol. Zool. 67, 282–303.
- Hammond, K. A. & Diamond, J. 1997 Maximal sustained energy budgets in humans and animals. Nature 386, 457–462. [\(doi:10.1038/386457a0\)](http://dx.doi.org/doi:10.1038/386457a0)
- Hammond, K. A. & Kristan, D. M. 2000 Responses to lactation and cold exposure by deer mice (Peromyscus maniculatus). Physiol. Biochem. Zool. 73, 547–556. ([doi:10.](http://dx.doi.org/doi:10.1086/317757) [1086/317757](http://dx.doi.org/doi:10.1086/317757))
- Hammond, K. A., Konarzewski, M., Torres, R. M. & Diamond, J. 1994 Metabolic ceilings under a combination of peak energy demands. Physiol. Zool. 67, 1479–1506.
- Hammond, K. A., Lloyd, K. C. K. & Diamond, J. 1996 Is mammary output capacity limiting to lactational performance in mice? *J. Exp. Biol.* **199**, 337–349.
- Harder, J. D., Hsu, M. J. & Garton, D. W. 1996 Metabolic rates and body temperature of the gray short-tailed opossum (Monodelphis domestica) during gestation and lactation. Physiol. Zool. 69, 317–339.
- Havera, S. P. 1979 Energy and nutrient cost of lactation in fox squirrels. *J. Wildlife Manage*. 43, 958-965. ([doi:10.2307/](http://dx.doi.org/doi:10.2307/3808281) [3808281\)](http://dx.doi.org/doi:10.2307/3808281)
- <span id="page-19-0"></span>Heldmaier, G., Ortmann, S. & Elvert, R. 2004 Natural hypometabolism during hibernation and daily torpor in mammals. Resp. Physiol. Neurobiol. 141, 317–329. ([doi:10.](http://dx.doi.org/doi:10.1016/j.resp.2004.03.014) [1016/j.resp.2004.03.014\)](http://dx.doi.org/doi:10.1016/j.resp.2004.03.014)
- Hill, R. W. 1992 The altricial/precocial contrast in the thermal relations and energetics of small mammals. In Mammalian energetics: interdiciplinary views of metabolism and reproduction (eds T. E. Tomasi & T. H. Horton), ch. 7, pp. 122–159. Ithaca, New York, NY: Cornell University Press.
- Hitchcock, F. A. 1927 The total energy requirement of the albino rat for growth and activity. Am.  $\tilde{f}$ . Physiol. 83, 28–36.
- Hodnett, D. W., Deluca, H. F. & Jorgensen, N. A. 1992 Bonemineral loss during lactation occurs in absence of parathyroid tissue. Am. J. Physiol. 262, E230-E233.
- Hoffman, S., Grisso, J. A., Kelsey, J. L., Gammon, M. D. & O'Brien, L. A. 1993 Parity, lactation and hip fracture. Osteoporosis Int. 3, 171–176. [\(doi:10.1007/BF01623672](http://dx.doi.org/doi:10.1007/BF01623672))
- Ingvartsen, K. L. & Boisclair, Y. R. 2001 Leptin and the regulation of food intake, energy homeostasis and immunity with special focus on the periparturient ruminants. Domest. Anim. Endocrinol. 21, 215–250. ([doi:10.1016/S0739-7240\(02\)00119-4\)](http://dx.doi.org/doi:10.1016/S0739-7240(02)00119-4)
- Innes, D. G. & Millar, J. S. 1981 Body weight, litter size and energetics of reproduction in Clethrionomys gapperi and Microtus pennsylvanicus. Can. J. Zool. 59, 785-789.
- Isler, D., Trayhurn, P. & Lunn, P. G. 1984 Brown adiposetissue metabolism in lactating rats—the effect of litter size. Ann. Nutr. Metab. 28, 101–109.
- Jacobsen, B. K., Nilssen, S., Heuch, I. & Kvale, G. 1998 Reproductive factors and fatal hip fractures. A Norwegian prospective study of 63 000 women. *J. Epidemiol*. Commun. Health 52, 645–650.
- Jansen, G. R. & Binard, R. 1991 Effect of dietary protein and environmental factors on lactation performance in rats. Physiol. Behav. 50, 297–304. ([doi:10.1016/0031-9384\(91\)](http://dx.doi.org/doi:10.1016/0031-9384(91)90069-Z) [90069-Z](http://dx.doi.org/doi:10.1016/0031-9384(91)90069-Z))
- Johnson, M. S. & Speakman, J. R. 2001 Limits to sustained energy intake V. Effect of cold exposure during lactation in Mus musculus. J. Exp. Biol. 204, 1967–1977.
- Johnson, M. S., Thomson, S. C. & Speakman, J. R. 2001a Limits to sustained energy intake I. Lactation in the laboratory mouse Mus musculus. J. Exp. Biol. 204, 1925–1935.
- Johnson, M. S., Thomson, S. C. & Speakman, J. R. 2001b Limits to sustained energy intake III. Effects of concurrent pregnancy and lactation in Mus musculus. J. Exp. Biol. 204, 1947–1956.
- Johnson, M. S., Thomson, S. C. & Speakman, J. R.  $2001c$ Limits to sustained energy intake II. Inter-relationships between resting metabolic rate, life-history traits and morphology in Mus musculus. J. Exp. Biol. 204, 1937–1946.
- Johnston, S. L. et al. 2006 Having it all: historical energy intakes do not generate the anticipated trade-offs in fecundity. Proc. R. Soc. B 273, 1369–1374. ([doi:10.1098/](http://dx.doi.org/doi:10.1098/rspb.2005.3456) [rspb.2005.3456\)](http://dx.doi.org/doi:10.1098/rspb.2005.3456)
- Johnston, S. L., Souter, D. M., Erwin, S. S., Tolkamp, B. J., Yearsley, J. M., Gordon, I. J., Illius, A. W., Kyriazakis, I. & Speakman, J. R. 2007 Associations between basal metabolic rate and reproductive performance in C57BL/6J mice. *J. Exp. Biol.* 210, 65-74. ([doi:10.1242/](http://dx.doi.org/doi:10.1242/jeb.02625) [jeb.02625\)](http://dx.doi.org/doi:10.1242/jeb.02625)
- Jolicoeur, L., Asselin, J. & Morisset, J. 1980 Trophic effects of gestation and lactation on rat pancreas. Biomed. Res. 1, 482–488.

Kaczmarski, F. 1966 Bioenergetics of pregnancy and lactation in the bank vole. Acta Theriol. XI 19, 409–417.

Kam, M., Cohen-Gross, S., Khokhlova, I. S., Degen, A. A. & Geffen, E. 2003 Average daily metabolic rate, reproduction and energy allocation during lactation in the Sundevall jird Meriones crassus. Funct. Ecol. 17, 496–503. [\(doi:10.1046/j.1365-2435.2003.00754.x\)](http://dx.doi.org/doi:10.1046/j.1365-2435.2003.00754.x)

- Kam, M., Khokhlova, I. S. & Degen, A. A. 2006 Partitioning of metabolizable energy intake in sucking altricial and precocial rodent pups. *J. Zool.* 269, 502-505. ([doi:10.](http://dx.doi.org/doi:10.1111/j.1469-7998.2006.00088.x) [1111/j.1469-7998.2006.00088.x](http://dx.doi.org/doi:10.1111/j.1469-7998.2006.00088.x))
- Kelly, P. A., Binart, N., Freemark, M., Lucas, B., Goffin, V. & Bouchard, B. 2001 Prolactin receptor signal transduction pathways and actions determined in prolactin receptor knockout mice. Biochem. Soc. Trans. 29, 48–52. ([doi:10.](http://dx.doi.org/doi:10.1042/BST0290048) [1042/BST0290048\)](http://dx.doi.org/doi:10.1042/BST0290048)
- Kennedy, G. C., Pearce, K. M. & Parrott, D. M. V. 1958 Liver growth in the lactating rat. J. Endocrinol. 17, 158–160.
- Khokhlova, I. S., Krasnov, B. R., Kam, M., Burdelova, N. & Degen, A. A. 2002 Energy cost of ectoparasitism: the flea Xenopsylla ramesis on the desert gerbil Gerbillus dasyurus. J. Zool. 258, 349–354. [\(doi:10.1017/S0952836902001498](http://dx.doi.org/doi:10.1017/S0952836902001498))
- Kim, E. M., Kotz, C. M., Welch, C. C., Grace, M. K., Billington, C. J. & Levine, A. S. 1997 Lactation decreases mRNA levels of opioid peptides in the arcuate nucleus of the rat. Brain Res. 769, 303–308. [\(doi:10.1016/S0006-](http://dx.doi.org/doi:10.1016/S0006-8993(97)00722-1) [8993\(97\)00722-1\)](http://dx.doi.org/doi:10.1016/S0006-8993(97)00722-1)
- Kittrell, E. M. W. & Satinoff, E. 1988 Diurnal rhythms of body temperature, drinking and activity over reproductive cycles. Physiol. Behav. 42, 477–484. ([doi:10.1016/0031-](http://dx.doi.org/doi:10.1016/0031-9384(88)90180-1) [9384\(88\)90180-1\)](http://dx.doi.org/doi:10.1016/0031-9384(88)90180-1)
- Koiter, T. R., Moes, H., Valkhof, N. & Wijkstra, S. 1999 Interaction of late pregnancy and lactation in rats. *J. Reprod.* Fertil. 115, 341–347. [\(doi:10.1530/jrf.0.1150341](http://dx.doi.org/doi:10.1530/jrf.0.1150341))
- Korine, C., Speakman, J. R. & Arad, Z. 2004 Reproductive energetics of captive and free-ranging Egyptian fruit bats (Rousettus aegyptiacus). Ecology 85, 220–230. ([doi:10.](http://dx.doi.org/doi:10.1890/02-0632) [1890/02-0632\)](http://dx.doi.org/doi:10.1890/02-0632)
- Kovacs, C. S. & Fuleihan, G. E.-H. 2006 Calcium and bone disorders during pregnancy and lactation. Endocrinol. Metabol. Clinics North America 35, 21–27. ([doi:10.1016/](http://dx.doi.org/doi:10.1016/j.ecl.2005.09.004) [j.ecl.2005.09.004\)](http://dx.doi.org/doi:10.1016/j.ecl.2005.09.004)
- Krebs, H. A. 1950 Body size and tissue respiration. Biochim. Biophys. Acta 4, 249–269. ([doi:10.1016/0006-3002\(50\)](http://dx.doi.org/doi:10.1016/0006-3002(50)90032-1) [90032-1](http://dx.doi.org/doi:10.1016/0006-3002(50)90032-1))
- Król, E. 1985 Reproductive energy budgets of hedgehogs during lactation. Zesz. Nauk. Filii UW Biol. 10, 105-117.
- Król, E. & Speakman, J. R. 2003a Limits to sustained energy intake VI. Energetics of lactation in laboratory mice at thermoneutrality. *J. Exp. Biol.* 206, 4255-4266. ([doi:10.](http://dx.doi.org/doi:10.1242/jeb.00674) [1242/jeb.00674](http://dx.doi.org/doi:10.1242/jeb.00674))
- Król, E. & Speakman, J. R. 2003b Limits to sustained energy intake VII. Milk energy output in laboratory mice at thermoneutrality. *J. Exp. Biol.* 206, 4267-4281. ([doi:10.](http://dx.doi.org/doi:10.1242/jeb.00675) [1242/jeb.00675](http://dx.doi.org/doi:10.1242/jeb.00675))
- Król, E., Johnson, M. & Speakman, J. R. 2003a Limits to sustained energy intake VII. Resting metabolic rate and organ morphology of laboratory mice lactating at thermoneutrality. *J. Exp. Biol.* 206, 4267-4281. ([doi:10.1242/jeb.](http://dx.doi.org/doi:10.1242/jeb.00674) [00674](http://dx.doi.org/doi:10.1242/jeb.00674))
- Król, E., Johnson, M. S. & Speakman, J. R. 2003b Limits to sustained energy intake VIII. Resting metabolic rate and organ morphology of laboratory mice lactating at thermoneutrality. *J. Exp. Biol.* 206, 4283-4291. ([doi:10.1242/jeb.](http://dx.doi.org/doi:10.1242/jeb.00676) [00676](http://dx.doi.org/doi:10.1242/jeb.00676))
- Künkele, J. 2000 Effects of litter size on the energetics of reproduction in a highly precocial rodent, the guinea pig. J. Mammal. 81, 691–700. [\(doi:10.1644/1545-1542\(2000\)](http://dx.doi.org/doi:10.1644/1545-1542(2000)081%3C0691:EOLSOT%3E2.3.CO;2) 081<[0691:EOLSOT](http://dx.doi.org/doi:10.1644/1545-1542(2000)081%3C0691:EOLSOT%3E2.3.CO;2)>2.3.CO;2)
- Kunkele, J. & Trillmich, F. 1997 Are precocial young cheaper? Lactation energetics in the guinea pig. Physiol. Zool. 70, 589–596.
- <span id="page-20-0"></span>Kunz, T. H., Whitaker, J. O. & Wadanoli, M. D. 1995 Dietary energetics of the insectivorous Mexican free-tailed bat (Tadarida brasiliensis) during pregnancy and lactation. Oecologia 101, 407–415. [\(doi:10.1007/BF00329419\)](http://dx.doi.org/doi:10.1007/BF00329419)
- Kurta, A. & Kunz, T. H. 1987 Size of bats at birth and maternal investment during pregnancy. Symp. Zool. Soc. 57, 79–107.
- Kurta, A., Bell, G. P., Nagy, K. A. & Kunz, T. H. 1989 Energetics of pregnancy and lactation in free-ranging little brown bats (Myotis lucifugus). Physiol. Zool. 62, 804–818.
- Kurta, A., Kunz, T. H. & Nagy, K. A. 1990 Energetics and water flux of free-ranging big brown bats (Eptesicus fuscus) during pregnancy and lactation. *J. Mammal.* 71, 59-65. ([doi:10.2307/1381316](http://dx.doi.org/doi:10.2307/1381316))
- Lamote, I., Meyer, E., Massart-Leen, A. M. & Burvenich, C. 2004 Sex steroids and growth factors in the regulation of mammary gland proliferation, differentiation, and involution. Steroids 69, 145–159. ([doi:10.1016/j.steroids.2003.](http://dx.doi.org/doi:10.1016/j.steroids.2003.12.008) [12.008\)](http://dx.doi.org/doi:10.1016/j.steroids.2003.12.008)
- Larradale, J., Fernandez-Ortero, P. & Gonzalez, M. 1966 Increased active transport of glucose through the intestine during pregnancy. Nature 209, 1356–1357. ([doi:10.1038/](http://dx.doi.org/doi:10.1038/2091356b0) [2091356b0\)](http://dx.doi.org/doi:10.1038/2091356b0)
- Lausen, C. L. & Barclay, R. M. R. 2003 Thermoregulation and roost selection by reproductive female big brown bats (Eptesicus fuscus) roosting in rock crevices. J. Zool. 260, 235–244. [\(doi:10.1017/S0952836903003686\)](http://dx.doi.org/doi:10.1017/S0952836903003686)
- Leon, M., Croskerry, P. G. & Smith, G. 1978 Thermal control of mother–young contact in rats. Physiol. Behav. 21, 793–811. ([doi:10.1016/0031-9384\(78\)90021-5](http://dx.doi.org/doi:10.1016/0031-9384(78)90021-5))
- Leon, M., Adels, L. & Coppersmith, R. 1985 Thermal limitation of mother-young contact in Norway rats. Dev. Psychobiol. 18, 85–105. [\(doi:10.1002/dev.420180202](http://dx.doi.org/doi:10.1002/dev.420180202))
- Leopold, S. S., Boskey, A. L., Doty, S. B., Gertner, J. M., Peterson, M. G. E. & Torzilli, P. A. 1995 Diminished material properties and altered bone structure in rat femora during pregnancy. *J. Orthopaed. Res.* 13, 41-49. ([doi:10.1002/jor.1100130108](http://dx.doi.org/doi:10.1002/jor.1100130108))
- Liu, H., Wang, D. H. & Wang, Z. W. 2003 Energy requirements during reproduction in female Brandt's voles (Microtus brandtii). J. Mammal. 84, 1410–1416. ([doi:10.1644/BRG-030\)](http://dx.doi.org/doi:10.1644/BRG-030)
- Lough, D. S., Beede, D. L. & Wilcox, C. J. 1990 Effects of feed intake and thermal stress on mammary blood-flow and other physiological measurements in lactating dairy cows. *J. Dairy Sci.* 73, 325-332.
- Lublin, A. & Wolfenson, D. 1996 Lactation and pregnancy effects on blood flow to mammary and reproductive systems in heat-stressed rabbits. Comp. Biochem. Physiol. A 115, 277–285. ([doi:10.1016/S0300-9629\(96\)](http://dx.doi.org/doi:10.1016/S0300-9629(96)00060-6) [00060-6](http://dx.doi.org/doi:10.1016/S0300-9629(96)00060-6))
- McNamara, J. M. & Houston, A. I. 2008 Optimal annual routines: behaviour in the context of physiology and ecology. Phil. Trans. R. Soc. 363, 301–319. ([doi:10.1098/](http://dx.doi.org/doi:10.1098/rstb.2007.2141) [rstb.2007.2141\)](http://dx.doi.org/doi:10.1098/rstb.2007.2141).
- Marai, I. F. M., Ayyat, M. S. & Abd El-Monem, U. M. 2001 Growth performance and reproductive traits at first parity of New Zealand White female rabbits as affected by heat stress and its alleviation under Egyptian conditions. Trop. Anim. Health Prod. 33, 451–462.
- Martin, L. B., Weil, Z. M. & Nelson, R. J. 2008 Seasonal changes in vertebrate immune activity: mediation by physiological trade-offs. Phil. Trans. R. Soc. B 363, 321–339. [\(doi:10.1098/rstb.2007.2142\)](http://dx.doi.org/doi:10.1098/rstb.2007.2142)
- Martin, R. D. 1989 Size, shape and evolution. In *Evolutionary* studies—a centenary celebration of the life of Julian Huxley (ed. M. Keynes), pp. 96–141. London, UK: Eugenics Society.
- Martin, R. D. & MacLarnon, A. M. 1985 Gestation period, neonatal size and maternal investment in placental mammals. Nature 313, 220–223. [\(doi:10.1038/313220a0\)](http://dx.doi.org/doi:10.1038/313220a0)
- Mattingly, D. K. & McClure, P. A. 1982 Energetics of reproduction in large littered cotton rats (Sigmodon hispidus). Ecology 63, 183-195. [\(doi:10.2307/1937043\)](http://dx.doi.org/doi:10.2307/1937043)
- Mclean, J. A. & Speakman, J. R. 1997 Non-nutritional maternal support in the brown long-eared bat. Anim. Behav. 54, 1193–1204.
- Mclean, J. A. & Speakman, J. R. 1999 Energy budgets of lactating and non-reproductive brown long-eared bats (Plecotus auritus) suggest females use compensation in lactation. Funct. Ecol. 13, 360–372. ([doi:10.1046/j.1365-](http://dx.doi.org/doi:10.1046/j.1365-2435.1999.00321.x) [2435.1999.00321.x\)](http://dx.doi.org/doi:10.1046/j.1365-2435.1999.00321.x)
- McLean, J. A. & Speakman, J. R. 2000 Effects of body mass and reproduction on the basal metabolic rate of brown long-eared bats (Plecotus auritus). Physiol. Biochem. Zool. 73, 112–121. ([doi:10.1086/316715](http://dx.doi.org/doi:10.1086/316715))
- Medina, K. L., Smithson, G. & Kincade, P. W. 1993 Supression of B lymphopoiesis during normal pregnancy. J. Exp. Med. 178, 1507–1515. ([doi:10.1084/jem.178.5.](http://dx.doi.org/doi:10.1084/jem.178.5.1507) [1507](http://dx.doi.org/doi:10.1084/jem.178.5.1507))
- Michaelsson, K., Baron, J. A., Farahmand, B. Y. & Ljunghall, S. 2001 Influence of parity and lactation on hip fracture risk. Am. J. Epidemiol. 153, 1166-1172. [\(doi:10.1093/aje/](http://dx.doi.org/doi:10.1093/aje/153.12.1166) [153.12.1166](http://dx.doi.org/doi:10.1093/aje/153.12.1166))
- Migula, P. 1969 Bioenergetics of pregnancy and lactation in the common vole. Acta Theriol. 13, 167–179.
- Millar, J. S. 1978 Energetics of reproduction in Peromyscus leucopus: the cost of lactation. Ecology 59, 1055-1061. ([doi:10.2307/1938558](http://dx.doi.org/doi:10.2307/1938558))
- Millar, J. S. 1985 Life cycle characteristics of Peromyscus maniculatus nebrascensis. Can. J. Zool. 63, 1280-1284.
- Millar, J. S. & Innes, D. G. 1985 Breeding by Peromyscus maniculatus over an elevational gradient. Can. J. Zool. 63, 124–129.
- Miller, S. C. & Wronski, T. J. 1993 Long-term osteopenic changes in cancellous bone structure in ovariectomized rats. Anat. Rec. 236, 433–441. ([doi:10.1002/ar.109236](http://dx.doi.org/doi:10.1002/ar.1092360303) [0303](http://dx.doi.org/doi:10.1002/ar.1092360303))
- Miller, S. C. & Bowman, B. M. 2004 Rapid improvements in cortical bone dynamics and structure after lactation in established breeder rats. Anat. Rec. A—Disc. Molec. Cell. Evol. Biol. 276A, 143–149. ([doi:10.1002/ar.a.10138\)](http://dx.doi.org/doi:10.1002/ar.a.10138)
- Miller, S. C., Anderson, B. L. & Bowman, B. M. 2005 Weaning initiates a rapid and powerful anabolic phase in the rat maternal skeleton. Biol. Reprod. 73, 156–162. ([doi:10.1095/biolreprod.105.039610](http://dx.doi.org/doi:10.1095/biolreprod.105.039610))
- Myrcha, A. 1964 Variations in the length and weight of the alimentary tract of Clethrionomys glareolus (Schreber, 1780). Acta Theriol. IX **10**, 139-148.
- Myrcha, A. 1965 Length and weight of the alimentary tract of Apodemus flavicollis. Acta Theriol. 10, 225–228.
- Naves, M., Diaz-Lopez, J. B., Gomez, C., Rodriguez-Rebollar, A. & Cannata-Andia, J. B. 2005 Determinants of incidence of osteoporotic fractures in the female Spanish population older than 50. Osteoporosis Int. 16, 2013–2017. [\(doi:10.1007/s00198-005-1983-4\)](http://dx.doi.org/doi:10.1007/s00198-005-1983-4)
- Nedergaard, J., Golozoubova, V., Matthias, A., Shabalina, I., Ohba, K. I., Ohlson, K., Jacobsson, A. & Cannon, B. 2001 Life without UCPI: mitochondrial, cellular and organismal characteristics of the UCP1-ablated mice. Biochem. Soc. Trans. **29**, 756-763. [\(doi:10.1042/BST0290756\)](http://dx.doi.org/doi:10.1042/BST0290756)
- Nelson, S. L., Kunz, T. H. & Humphrey, S. R. 2005 Folivory in fruit bats: leaves provide a natural source of calicum. J. Chem. Ecol. 31, 1683–1691. ([doi:10.1007/s10886-005-](http://dx.doi.org/doi:10.1007/s10886-005-5920-y) [5920-y\)](http://dx.doi.org/doi:10.1007/s10886-005-5920-y)
- Neuhaus, P. 2003 Parasite removal and its impact on litter size and body condition in Columbian ground squirrels (Spermophilus columbianus). Proc. R. Soc. B 270(Suppl.), S213–S215. ([doi:10.1098/rsbl.2003.0073](http://dx.doi.org/doi:10.1098/rsbl.2003.0073))
- Newkirk, K. D., Cheung, B. L. W., Scribner, S. J. & Wynne-Edwards, K. E. 1998 Earlier thermoregulation and

<span id="page-21-0"></span>consequences for pup growth in the Siberian versus Djungarian dwarf hamster (Phodopus). Physiol. Behav. 63, 435–443. ([doi:10.1016/S0031-9384\(97\)00465-4\)](http://dx.doi.org/doi:10.1016/S0031-9384(97)00465-4)

- Nicol, S., Vedel-Smith, C. & Andersen, N. A. 2005 Behaviour, body temperature and hibernation in Tasmanian echidnas (Tachyglossus aculeatus). In Life in the cold. Evolution, mechanisms, adaptation and application. Twelfth international hibernation symposium, (eds B. M. Barnes & H. V. Carey). Biological papers of the University of Alaska, number 27. Fairbanks, Alaska USA, Institute of Arctic Biology, University of Alaska.
- Nievergelt, C. M. & Martin, R. D. 1999 Energy intake during reproduction in captive common marmosets (Callithrix jacchus). Physiol. Behav. 65, 849–854. ([doi:10.1016/](http://dx.doi.org/doi:10.1016/S0031-9384(98)00249-2) [S0031-9384\(98\)00249-2](http://dx.doi.org/doi:10.1016/S0031-9384(98)00249-2))
- Nishihara, K., Horiuchi, S., Eto, H., Uchida, S. & Honda, M. 2004 Delta and theta power spectra of night sleep EEG are higher in breast-feeding mothers than in non-pregnant women. Neurosci. Lett. 368, 216–2202. ([doi:10.1016/](http://dx.doi.org/doi:10.1016/j.neulet.2004.07.021) [j.neulet.2004.07.021\)](http://dx.doi.org/doi:10.1016/j.neulet.2004.07.021)
- Nishiwaki, M., Yasumizu, T., Hoshi, K. & Ushijima, H. 1999 Effect of pregnancy, lactation and weaning on bone mineral density in rats as determined by dual-energy X-ray absorptiometry. Endocr. 7. 46, 711-716.
- Nizielski, S. E., Billington, C. J. & Levine, A. S. 1993 Bat thermogenic activity and capacity are reduced during lactation in ground squirrels. Am. J. Physiol. Regul. Integr. Comp. Physiol. 264, R16–R21.
- Noel, M. B. & Woodside, B. 1993 Effects of systemic and central prolactin injections on food intake, weight gain, and estrous cyclicity in female rats. Physiol. Behav. 54, 151–154. [\(doi:10.1016/0031-9384\(93\)90057-M\)](http://dx.doi.org/doi:10.1016/0031-9384(93)90057-M)
- O'Donnell, C. F. J. 2002 Influence of sex and reproductive status on nocturnal activity of long-tailed bats (Chalinolobus tuberculatus). *J. Mammal.* 83, 794-803. [\(doi:10.1644/1545-](http://dx.doi.org/doi:10.1644/1545-1542(2002)083%3C0794:IOSARS%3E2.0.CO;2) [1542\(2002\)083](http://dx.doi.org/doi:10.1644/1545-1542(2002)083%3C0794:IOSARS%3E2.0.CO;2)<0794:IOSARS>2.0.CO;2)
- Oftedal, O. T. 1984 Body size and reproductive strategy as correlates of milk energy output in lactating mammals. Acta Zool. Fenn. 171, 183–186.
- Ogueta, S., Muñoz, J., Obregon, E., Delgado-Baeza, E. & García-Ruiz, J. P. 2002 Prolactin is a component of the human synovial liquid and modulates the growth and chondrogenic differentiation of bone marrow-derived mesenchymal stem cells. Mol. Cell. Endocrinol. 190, 51–63. [\(doi:10.1016/S0303-7207\(02\)00013-8](http://dx.doi.org/doi:10.1016/S0303-7207(02)00013-8))
- Omi, N. & Ezawa, I. 2001 Change in calcium balance and bone mineral density during pregnancy in female rats. J. Nutr. Sci. Vitaminol. 47, 195–200.
- Ortiz, R., Cortés, E., González, C., Pérez, L. & Betancourt, M. 1995 Micronucleus frequency in spleen lymphocytes from severely malnourished rats during lactation. Env. Mol. Mutagen. 26, 55–59. ([doi:10.1002/em.2850260108\)](http://dx.doi.org/doi:10.1002/em.2850260108)
- Ozbek, M., Kanli, A., Dural, S., Sahin, I., Gonen, E. & Tulunoglu, I. 2004 Effects of pregnancy and lactation on the microhardness of rat incisor dentine and enamel. Arch. Oral Biol. 49, 607–612. ([doi:10.1016/j.archoralbio.2004.](http://dx.doi.org/doi:10.1016/j.archoralbio.2004.02.003) [02.003\)](http://dx.doi.org/doi:10.1016/j.archoralbio.2004.02.003)
- Pedraza, N., Solanes, G., Carmona, M. C., Iglesias, R., Vinas, O., Mampel, T., Vazquez, M., Giralt, M. & Villarroya, F. 2000 Impaired expression of the uncoupling protein-3 gene in skeletal muscle during lactation: Fibrates and troglitazone reverse lactation-induced downregulation of the uncoupling protein-3 gene. Diabetes 49, 1224–1230. ([doi:10.2337/diabetes.49.7.1224\)](http://dx.doi.org/doi:10.2337/diabetes.49.7.1224)
- Pedraza, N., Solanes, G., Iglesias, R., Vazquez, M., Giralt, M. & Villarroya, F. 2001 Differential regulation of expression of genes encoding uncoupling proteins 2 and 3 in brown adipose tissue during lactation in mice. Biochem. J. 355, 105–111. [\(doi:10.1042/0264-6021:3550105](http://dx.doi.org/doi:10.1042/0264-6021:3550105))
- Perrigo, G. 1987 Breeding and feeding strategies in deer mice and house mice when females are challenged to work for their food. Anim. Behav. 35, 1298–1316. ([doi:10.1016/](http://dx.doi.org/doi:10.1016/S0003-3472(87)80002-7) [S0003-3472\(87\)80002-7](http://dx.doi.org/doi:10.1016/S0003-3472(87)80002-7))
- Petersen, H. C., Jeune, B., Vaupel, J. W. & Christensen, K. 2002 Reproduction life history and hip fractures. Ann. Epidemiol. 12, 257–263. [\(doi:10.1016/S1047-](http://dx.doi.org/doi:10.1016/S1047-2797(01)00275-7) [2797\(01\)00275-7\)](http://dx.doi.org/doi:10.1016/S1047-2797(01)00275-7)
- Peterson, C. C., Nagy, K. A. & Diamond, J. 1990 Sustained metabolic scope. Proc. Natl Acad. Sci. USA 87, 2324–2328. [\(doi:10.1073/pnas.87.6.2324](http://dx.doi.org/doi:10.1073/pnas.87.6.2324))
- Prieto, R. M. S., Ferrer, M., Fe, J. M., Rayo, J. M. & Tur, J. 1994 Morphological adaptative changes of small intestinal tract regions due to pregnancy and lactation in rats. Ann. Nutr. Metab. 38, 295–300.
- Pinilla, T. & Birch, L. L. 1993 Help me make it through the night—behavioral entrainment of breast-fed infants sleep patterns. Pediatrics 91, 436–444.
- Poppitt, S. D., Speakman, J. R. & Racey, P. A. 1993 The energetics of reproduction in the common shrew (Sorex araneus)—a comparison of indirect calorimetry and the doubly labeled water method. Physiol. Zool. 66, 964–982.
- Poppitt, S. D., Speakman, J. R. & Racey, P. A. 1994 Energetics of reproduction in the lesser hedgehog tenrec, Echinops telfairi (Martin). Physiol. Zool. 67, 976–994.
- Quiniou, N. & Noblet, J. 1999 Influence of high ambient temperatures on performance of multiparous lactating sows. J. Anim. Sci. 77, 2124–2134.
- Racey, P. A. 1973 Environmental factors influencing the length of gestation in heterothermic bats. J. Reprod. Fertil. Supp. 19, 175-189.
- Racey, P. A. & Swift, S. M. 1981 Variations in gestation length in a colony of pipistrelle bats (Pipistrellus pipistrellus) from year to year. *J. Reprod. Fertil.* **61**, 123-129. ([doi:10.](http://dx.doi.org/doi:10.1530/jrf.0.0610123) [1530/jrf.0.0610123](http://dx.doi.org/doi:10.1530/jrf.0.0610123))
- Randolph, P. A., Randolph, J. C., Mattingly, K. & Foster, M. M. 1977 Energy costs of reproduction in the cotton rat (Sigmodon hispidus). Ecology 58, 31–45. ([doi:10.2307/](http://dx.doi.org/doi:10.2307/1935106) [1935106](http://dx.doi.org/doi:10.2307/1935106))
- Rayner, V. C., Robinson, I. C. A. F. & Russell, J. A. 1988 Chronic intracerebroventricular morphine and lactation in rats—dependence and tolerance in relation to oxytocin neurons. *J. Physiol.* **396**, 319–347.
- Renaudeau, D. & Noblet, J. 2001 Effects of exposure to high ambient temperature and dietary protein level on sow milk production and performance of piglets. *J. Anim. Sci.* 79, 1540–1548.
- Renaudeau, D., Noblet, J. & Dourmad, J. Y. 2003 Effect of ambient temperature on mammary gland metabolism in lactating sows. *J. Anim. Sci.* 81, 217-231.
- Reznick, D. N., Bryga, H. & Endler, J. A. 1990 Experimentally induced life history evolution in natural population. Nature 346, 357–359. [\(doi:10.1038/346357a0\)](http://dx.doi.org/doi:10.1038/346357a0)
- Ricklefs, R. E. & Wikelski, M. 2002 The physiology–life history nexus. Trends Ecol. Evol. 17, 462-468. ([doi:10.](http://dx.doi.org/doi:10.1016/S0169-5347(02)02578-8) [1016/S0169-5347\(02\)02578-8\)](http://dx.doi.org/doi:10.1016/S0169-5347(02)02578-8)
- Roff, D. A. 1992 The evolution of life histories: theory and analysis. New York, NY: Chapman & Hall.
- Rogowitz, G. L. 1998 Limits to milk flow and energy allocation during lactation in the hispod cotton rat (Sigmodon hispidus). Physiol. Zool. 71, 312–320.
- Ruby, J., Nathan, P. T., Balasingh, J. & Kunz, T. H. 2000 Chemical composition of fruits and leaves eaten by shortnosed fruit bat, Cynopterus sphinx. J. Chem. Ecol. 26, 2825–2841. [\(doi:10.1023/A:1026446011693](http://dx.doi.org/doi:10.1023/A:1026446011693))
- Rydell, J. 1993 Variation in foraging activity of an aerial insectivorous bat during reproduction. *J. Mammal.* 74, 503–509. [\(doi:10.2307/1382411\)](http://dx.doi.org/doi:10.2307/1382411)
- <span id="page-22-0"></span>Rydell, J. & Speakman, J. R. 1995 Evolution of nocturnality in bats—potential competitors and predators during their early history. Biol. J. Linn. Soc. 54, 183-191. ([doi:10.](http://dx.doi.org/doi:10.1006/bijl.1995.0011) [1006/bijl.1995.0011\)](http://dx.doi.org/doi:10.1006/bijl.1995.0011)
- Sano, H., Ambo, K. & Tsuda, T. 1985 Blood glucose kinetics in whole body and mammary gland of lactating goats exposed to heat. J. Dairy Sci. 68, 2557-2564.
- Sarli, M., Hakim, C., Rey, P. & Zanchetta, J. 2005 Osteoporosis during pregnancy and lactation. Medicina— Buenos Aires 65, 533–540.
- Scribner, S. J. & Wynne Edwards, K. E. 1994a Disruption of body temperature and behavior rhythms during reproduction in dwarf hamster (Phodopus). Physiol. Behav. 55, 361–369. [\(doi:10.1016/0031-9384\(94\)90147-3\)](http://dx.doi.org/doi:10.1016/0031-9384(94)90147-3)
- Scribner, S. J. & Wynne Edwards, K. E. 1994b Thermal constraints on maternal behavior during reproduction in dwarf hamsters (Phodopus). Physiol. Behav. 55, 897–903. ([doi:10.1016/0031-9384\(94\)90077-9](http://dx.doi.org/doi:10.1016/0031-9384(94)90077-9))
- Sengupta, S., Arshad, M., Sharma, S., Dubey, M. & Singh, M. M. 2005 Attainment of peak bone mass and bone turnover rate in relation to estrous cycle, pregnancy and lactation in colony-bred Sprague–Dawley rats: suitability for studies on pathophysiology of bone and therapeutic measures for its management. *J. Steroid Biochem. Mol.* Biol. 94, 421–429. [\(doi:10.1016/j.jsbmb.2004.12.039\)](http://dx.doi.org/doi:10.1016/j.jsbmb.2004.12.039)
- Shen, H.-P. & Lee, L.-L. 2000 Mother–young interactions in a maternity colony of Myotis formosus. J. Mammol. 81, 726-733. ([doi:10.1644/1545-1542\(2000\)081](http://dx.doi.org/doi:10.1644/1545-1542(2000)081%3C0726:MYIIAM%3E2.3.CO;2)<0726:  $MYIIAM > 2.3.CO;2)$  $MYIIAM > 2.3.CO;2)$
- Shinkoda, H., Matsumoto, K. & Park, Y. M. 1999 Changes in sleep–wake cycle during the period from late pregnancy to puerperium identified through the wrist actigraph and sleep logs. Psychiatry Clin. Neurosci. 53, 133-135. ([doi:10.](http://dx.doi.org/doi:10.1046/j.1440-1819.1999.00518.x) [1046/j.1440-1819.1999.00518.x](http://dx.doi.org/doi:10.1046/j.1440-1819.1999.00518.x))
- Sikes, R. S. 1995 Costs of lactation and optimal litter size in northern grasshopper mice (Onychomys leucogaster). J. Mammal. 76, 348–357.
- Slonaker, J. R. 1924 The effect of copulation, pregnancy, pseudopregnancy and lactation on the voluntary activity and food consumption of the albino rat. Am.  $\tilde{f}$ . Physiol. 71, 362–394.
- Smith, C. C. & Fretwell, S. D. 1974 The optimal balance between size and number of offspring. Am. Nat. 108, 499–506. [\(doi:10.1086/282929\)](http://dx.doi.org/doi:10.1086/282929)
- Speakman, J. R. 1990 The function of daylight flying in British bats. *J. Zool.* 220, 101-113.
- Speakman, J. R. 1991a Why do insectivorous bats in Britain not fly in daylight more frequently. Funct. Ecol. 5, 518–524. [\(doi:10.2307/2389634\)](http://dx.doi.org/doi:10.2307/2389634)
- Speakman, J. R. 1991b The impact of predation by birds on bat populations in the British-isles. Mamm. Rev. 21, 123–142.
- Speakman, J. R. 2003 Oxidative phosphorylation, mitochondrial proton cycling, free-radical production and aging. In Energy metabolism and lifespan determination. (ed. M. P. Mattson). Elsevier series advances in cell aging and gerontology, pp. 35–68. Amsterdam, The Netherlands: Elsevier.
- Speakman, J. R. & McQueenie, J. 1996 Limits to sustained metabolic rate: the link between food intake, basal metabolic rate, and morphology in reproducing mice, Mus musculus. Physiol. Zool. 69, 746–769.
- Speakman, J. R. & Krol, E. 2005 Limits to sustained energy intake IX: a review of hypotheses. *J. Comp. Physiol. B* 175, 375–394. [\(doi:10.1007/s00360-005-0013-3\)](http://dx.doi.org/doi:10.1007/s00360-005-0013-3)
- Speakman, J. R., Racey, P. A., Catto, C. M. C., Webb, P. I., Swift, S. M. & Burnett, A. M. 1991 Minimum summer population and densities of bats in N.E. Scotland near the northern borders of their distributions. *J. Zool.* 225, 327–345.
- Speakman, J. R., Rydell, J., Webb, P. I., Hayes, J. P., Hays, G. C., Hulbert, I. A. R. & McDevitt, R. M. 2000 Activity patterns of insectivorous bats and birds in northern Scandinavia (69 degrees N), during continuous midsummer daylight. Oikos 88, 75–86. ([doi:10.1034/j.1600-](http://dx.doi.org/doi:10.1034/j.1600-0706.2000.880109.x) [0706.2000.880109.x\)](http://dx.doi.org/doi:10.1034/j.1600-0706.2000.880109.x)
- Speakman, J. R., Gidney, A., Bett, J., Mitchell, I. P. & Johnson, M. S. 2001 Limits to sustained energy intake IV. Effect of variation in food quality on lactating mice Mus musculus. J. Exp. Biol. 204, 1957–1965.
- Speakman, J. R., Selman, C., McLaren, J. S. & Harper, E. J. 2002 Living fast, dying when? The link between aging and energetics. *J. Nutr.* 132, 1583S-1597S.
- Speakman, J. R. et al. 2004 Uncoupled and surviving: individual mice with high metabolism have greater mitochondrial uncoupling and live longer. Aging Cell 3, 87–95. [\(doi:10.1111/j.1474-9728.2004.00097.x\)](http://dx.doi.org/doi:10.1111/j.1474-9728.2004.00097.x)
- Stamper, J. L., Zucker, I., Lewis, D. A. & Dark, J. 1998 Torpor in lactating Siberian hamsters subjected to glucoprivation. Am. J. Physiol.—Reg. Int. Compar. Physiol. 274, R46–R51.
- Stearns, S. C. 1992 The evolution of life histories. Oxford, UK: Oxford University Press.
- Stephenson, P. J. & Racey, P. A. 1993a Reproductive energetics of the Tenrecidae (Mammalia: Insectivora). II. The shrew-tenrecs, Microgale spp. Physiol. Zool. 66, 664–685.
- Stephenson, P. J. & Racey, P. A. 1993b Reproductive energetics of the Tenrecidae (Mammalia: Insectivora). I. The large-eared tenrec, Geogale aurita. Physiol. Zool. 66, 643–663.
- Stern, J. M. & Azzara, A. V. 2002 Thermal control of mother– young contact revisited: Hyperthermic rats nurse normally. Physiol. Behav. 77, 11–18. [\(doi:10.1016/S0031-](http://dx.doi.org/doi:10.1016/S0031-9384(02)00798-9) [9384\(02\)00798-9\)](http://dx.doi.org/doi:10.1016/S0031-9384(02)00798-9)
- Stocker, C. et al. 2004 Modulation of susceptibility to weight gain and insulin resistance in low birthweight rats by treatment of their mothers with leptin during pregnancy and lactation. Int. *J. Obesity* 28, 129-136. [\(doi:10.1038/sj.](http://dx.doi.org/doi:10.1038/sj.ijo.0802476) [ijo.0802476\)](http://dx.doi.org/doi:10.1038/sj.ijo.0802476)
- Swift, S. M. 1980 Activity patterns of pipistrelle bats (Pipistrellus pipistrellus) in north-east scotland. J. Zool. 190, 285–295.
- Tojo, Y., Kurabayashi, T., Honda, A., Yamamoto, Y., Yahata, T., Takakuwa, K. & Tanaka, K. 1998 Bone structural and metabolic changes at the end of pregnancy and lactation in rats. Am. J. Obstetrics Gynecol. 178, 180-185. ([doi:10.](http://dx.doi.org/doi:10.1016/S0002-9378(98)70649-0) [1016/S0002-9378\(98\)70649-0\)](http://dx.doi.org/doi:10.1016/S0002-9378(98)70649-0)
- Trayhurn, P. 1983 Decreased capacity for non-shivering thermogenesis during lactation in mice. Pflügers Archiv. 398, 264–265. ([doi:10.1007/BF00657164](http://dx.doi.org/doi:10.1007/BF00657164))
- Trayhurn, P. 1989 Thermogenesis and the energetics of pregnancy and lactation. Can. J. Physiol. Pharmacol. 67, 370–375.
- Trayhurn, P. & Jennings, G. 1987a Functional atrophy of brown adipose-tissue during lactation in mice—effects of lactation and weaning on mitochondrial gdp binding and uncoupling protein. Biochem. *J.* 248, 273-276.
- Trayhurn, P. & Wusterman, M. C. 1987b Sympathetic activity in brown adipose tissue during lactation in mice. Proc. Nutr. Soc. 46, A27.
- Trayhurn, P. & Jennings, G. 1988 Non-shivering thermogenesis and the thermogenic capacity of brown fat in fasted/re-fed mice. Am. J. Physiol. Regul. Integr. Comp. Physiol. 254, R11–R16.
- Trayhurn, P., Douglas, J. B. & McGuckin, M. M. 1982 Brown adipose tissue thermogenesis is suppressed during lactation in mice. Nature 298, 59–60. ([doi:10.1038/](http://dx.doi.org/doi:10.1038/298059a0) [298059a0](http://dx.doi.org/doi:10.1038/298059a0))
- <span id="page-23-0"></span>Turbill, C. & Geiser, F. 2006 Thermal physiology of pregnant and lactating female and male long-eared bats, Nyctophilus geoffroyi and N. gouldi. J. Comp. Physiol. B 176, 165-172. ([doi:10.1007/s00360-005-0038-7](http://dx.doi.org/doi:10.1007/s00360-005-0038-7))
- Turbill, C., Kortner, G. & Geiser, F. 2003 Natural use of heterothermy by a small, tree-roosting bat during Summer. Physiol. Biochem. Zool. 76, 868–876. ([doi:10.](http://dx.doi.org/doi:10.1086/378915) [1086/378915](http://dx.doi.org/doi:10.1086/378915))
- Ulmershakibaei, C. & Plonait, H. 1992 Studies of lactational hyperthermia in sows. Tierarztl. Umschau 47, 605–611.
- Vajda, E. G., Bowman, B. M. & Miller, S. C. 2001 Cancellous and cortical bone mechanical properties and tissue dynamics during pregnancy, lactation, and postlactation in the rat. Biol. Reprod. 65, 689–695. ([doi:10.](http://dx.doi.org/doi:10.1095/biolreprod65.3.689) [1095/biolreprod65.3.689](http://dx.doi.org/doi:10.1095/biolreprod65.3.689))
- Vernon, R. G. & Flint, D. G. 1983 Control of fatty acid synthesis during lactation. Proc. Nutr. Soc. 42, 315-331. ([doi:10.1079/PNS19830035\)](http://dx.doi.org/doi:10.1079/PNS19830035)
- Vernon, R. L. & Pond, C. M. 1997 Adaptations of maternal adipose tissue to lactation. *J. Mam. Gland Biol. Neoplasia* 2, 231–241. [\(doi:10.1023/A:1026380220364](http://dx.doi.org/doi:10.1023/A:1026380220364))
- Wade, G. N., Jennings, G. & Trayhurn, P. 1986 Energy balance and brown adipose tissue thermogenesis during pregnancy in Syrian hamsters. Am. J. Physiol. 250, R845-R850.
- Wang, C. H., Brown, S. & Bhattacharyya, M. H. 1994 Effect of cadmium on bone calcium and 45Ca in mouse dams on a calcium-deficient diet: evidence of Itai-Itai-like syndrome. Toxicol. Appl. Pharmacol. 127, 320-330. ([doi:10.1006/taap.1994.1168\)](http://dx.doi.org/doi:10.1006/taap.1994.1168)
- Weiner, J. 1987 Limits to energy budget and tactics in energy investments during reproduction in the Djungarian hamster (Phodopus sungorus sungorus Pallas 1770). Symp. Zool. Soc. Lond. 57, 167–187.
- Weiner, J. 1992 Physiological limits to sustainable energy budgets in birds and mammals: ecological implications. Trends Ecol. Evol. 7, 384–388. ([doi:10.1016/0169-5347](http://dx.doi.org/doi:10.1016/0169-5347(92)90009-Z) [\(92\)90009-Z](http://dx.doi.org/doi:10.1016/0169-5347(92)90009-Z))
- Weisstaub, A., de Ferrer, P. R., Zeni, S. & de Portela, M. L. 2003 Influence of low dietary calcium during pregnancy

and lactation on zinc levels in maternal blood and bone in rats. J. Trace Elements Med. Biol. 17, 27–32. ([doi:10.1016/](http://dx.doi.org/doi:10.1016/S0946-672X(03)80042-1) [S0946-672X\(03\)80042-1](http://dx.doi.org/doi:10.1016/S0946-672X(03)80042-1))

- Wiersma, P., Selman, C., Speakman, J. R. & Verhulst, S. 2004 Birds sacrifice oxidative protection for reproduction. Proc. R. Soc. B 271, S360-S363. [\(doi:10.1098/rsbl.2004.](http://dx.doi.org/doi:10.1098/rsbl.2004.0171) [0171\)](http://dx.doi.org/doi:10.1098/rsbl.2004.0171)
- Wilde, C. J., Knight, C. R. & Racey, P. A. 1999 Influence of torpor on milk protein composition and secretion in lactating bats. *J. Exp. Zool.* 284, 35-41. ([doi:10.1002/](http://dx.doi.org/doi:10.1002/(SICI)1097-010X(19990615)284:1%3C35::AID-JEZ6%3E3.0.CO;2-Z)  $(SICI)1097-010X(19990615)284:1 < 35::AID-JEZ6 > 3.$  $(SICI)1097-010X(19990615)284:1 < 35::AID-JEZ6 > 3.$ [0.CO;2-Z](http://dx.doi.org/doi:10.1002/(SICI)1097-010X(19990615)284:1%3C35::AID-JEZ6%3E3.0.CO;2-Z))
- Williams, G. C. 1966 Adaptation and natural selection. Princeton, NJ: Princeton University Press.
- Winchell, J. M. & Kunz, T. H. 1996 Day-roosting activity budgets of the Eastern pipistrelle bat, Pipistrellus subflavus (Chiroptera: Vespertilionidae). Can. J. Zool. 74, 431-441.
- Woodside, B. & Leon, M. 1980 Thermoendocrine influences on maternal nesting behavior in rats. *J. Comp. Physiol.* Psychol. 94, 41–60. ([doi:10.1037/h0077652](http://dx.doi.org/doi:10.1037/h0077652))
- Xiao, X. Q., Grove, K. L., Grayson, B. E. & Smith, M. S. 2004 Inhibition of uncoupling protein expression during lactation: Role of leptin. Endocrinology 145, 830-838. ([doi:10.1210/en.2003-0836](http://dx.doi.org/doi:10.1210/en.2003-0836))
- Yamamoto, M. et al. 1991 Suckling-mediated increases in urinary phosphate and  $3^{\prime},5^{\prime}$ -cyclic adenosinemonophosphate excretion in lactating rats—possible systemic effects of parathyroid hormone-related protein. Endocrinology 129, 2614–2622.
- Zeni, S. N., Di Gregorio, S. & Mautalen, C. 1999 Bone mass changes during pregnancy and lactation in the rat. Bone 25, 681–685. ([doi:10.1016/S8756-3282\(99\)00228-8\)](http://dx.doi.org/doi:10.1016/S8756-3282(99)00228-8)
- Zenuto, R. R., Antinuchi, C. D. & Busch, C. 2002 Bioenergetics of reproduction and pup development in a subterranean rodent (Ctenomys talarum). Physiol. Biochem. Zool. 75, 469–478. [\(doi:10.1086/344739\)](http://dx.doi.org/doi:10.1086/344739)
- Zera, A. J. & Harshman, L. G. 2001 The physiology of life history trade-offs in animals. Ann. Rev. Ecol. Syst. 32, 95–126. [\(doi:10.1146/annurev.ecolsys.32.081501.114006\)](http://dx.doi.org/doi:10.1146/annurev.ecolsys.32.081501.114006)