

Multilocus Genotyping Assays for Single Nucleotide Polymorphism-Based Subtyping of *Listeria monocytogenes* Isolates^{∇†}

Todd J. Ward,^{1*} Thomas F. Ducey,^{1‡} Thomas Usgaard,¹
Katherine A. Dunn,² and Joseph P. Bielawski²

Microbial Genomics and Bioprocessing Research Unit, Agricultural Research Service, U.S. Department of Agriculture, 1815 North University Street, Peoria, Illinois 61604,¹ and Department of Biology, Dalhousie University, Halifax, Nova Scotia B3H 4J1, Canada²

Received 19 May 2008/Accepted 9 October 2008

Listeria monocytogenes is responsible for serious invasive illness associated with consumption of contaminated food and places a significant burden on public health and the agricultural economy. We recently developed a multilocus genotyping (MLGT) assay for high-throughput subtype determination of *L. monocytogenes* lineage I isolates based on interrogation of single nucleotide polymorphisms (SNPs) via multiplexed primer extension reactions. Here we report the development and validation of two additional MLGT assays that address the need for comprehensive DNA sequence-based subtyping of *L. monocytogenes*. The first of these novel MLGT assays targeted variation segregating within lineage II, while the second assay combined probes for lineage III strains with probes for strains representing a recently characterized fourth evolutionary lineage (IV) of *L. monocytogenes*. These assays were based on nucleotide variation identified in >3.8 Mb of comparative DNA sequence and consisted of 115 total probes that differentiated 93% of the 100 haplotypes defined by the multilocus sequence data. MLGT reproducibly typed the 173 isolates used in SNP discovery, and the 10,448 genotypes derived from MLGT analysis of these isolates were consistent with DNA sequence data. Application of the MLGT assays to assess subtype prevalence among isolates from ready-to-eat foods and food-processing facilities indicated a low frequency (6.3%) of epidemic clone subtypes and a substantial population of isolates (>30%) harboring mutations in *inlA* associated with attenuated virulence in cell culture and animal models. These mutations were restricted to serogroup 1/2 isolates, which may explain the overrepresentation of serotype 4b isolates in human listeriosis cases.

Listeria monocytogenes is the causative agent of listeriosis, a food-borne disease with clinical presentations that include febrile gastroenteritis, encephalitis, meningitis, septicemia, and spontaneous abortion (7). Listeriosis infections are associated with high hospitalization (92%) and mortality (20 to 30%) rates and account for over one-quarter of all deaths attributable to known food-borne pathogens (12, 24). *L. monocytogenes* is widely distributed in the environment, forms biofilms, grows at refrigeration temperatures, and is relatively resistant to acid and high salt concentrations (23, 48). These characteristics enable *L. monocytogenes* to persist for extended periods in food-processing environments and make *L. monocytogenes* contamination of ready-to-eat (RTE) foods a significant concern. Accordingly, regulatory agencies have applied a zero tolerance policy for *L. monocytogenes* contamination in RTE foods, and *L. monocytogenes* has been a leading cause of food recalls due to microbial adulteration.

Molecular subtyping is a critical component of *L. monocytogenes*

outbreak detection and epidemiological investigations, which are complicated by the long incubation time for invasive listeriosis and the difficulty in identifying appropriate controls for case-control studies (44). Pulsed-field gel electrophoresis (PFGE) is the current gold standard for subtyping most food-borne bacterial pathogens and has been used successfully to subtype *L. monocytogenes* isolates as part of the PulseNet system (11, 43). However, PFGE is labor-intensive, results in complex patterns which are not always easy to interpret, is unable to target specific polymorphisms, and does not always discriminate between related but distinct strains (4, 5, 11). In addition, PFGE patterns may be affected by unstable genetic elements that could interfere with outbreak detection and long-term epidemiological tracking. As a result, there has been significant interest in the development of DNA sequence-based subtyping assays for food-borne pathogens (5, 11, 16, 37, 42, 43, 50). Toward that end, we recently described a single-well DNA sequence-based subtyping method utilizing multilocus genotyping (MLGT) of single-nucleotide polymorphisms (SNPs) segregating within *L. monocytogenes* lineage I that provides high discriminatory power and good epidemiological concordance (5).

While we previously focused on subtyping lineage I isolates because of their association with epidemic listeriosis, four major evolutionary lineages have been described for *L. monocytogenes* (22, 33, 36, 38, 46, 47). *L. monocytogenes* isolates from lineage II are common contaminants of RTE and other food products (13, 46), contribute significantly to sporadic listeriosis in humans (18), and include a previously described epidemic

* Corresponding author. Mailing address: Microbial Genomics and Bioprocessing Research Unit, National Center for Agricultural Utilization Research, Agricultural Research Service, USDA, Peoria, IL 61604. Phone: (309) 681-6394. Fax: (309) 681-6672. E-mail: todd.ward@ars.usda.gov.

‡ Current address: Coastal Plains Soil, Water and Plant Research Center, Agricultural Research Service, USDA, 2611 W. Lucas Street, Florence, SC 29501.

† Supplemental material for this article may be found at <http://aem.asm.org/>.

∇ Published ahead of print on 17 October 2008.

clonal lineage (epidemic clone III [ECIII]) responsible for a multistate outbreak in 2000 linked to turkey products (20, 31). Lineage III strains are primarily associated with animal listeriosis cases; however, approximately 1% of human sporadic listeriosis cases are attributable to lineage III (18). Several recent phylogenetic studies have demonstrated that isolates classified as subgroup IIIB represent a distinct fourth lineage of *L. monocytogenes* (22, 33, 38). In the present study, we refer to this group as lineage IV in order to make clear that this unique lineage is not part of, or even closely related to, a monophyletic lineage III. Fewer than a dozen isolates from lineage IV have been characterized to date, but these include human, animal, and food isolates (22, 33, 38). In order to address the need for comprehensive subtyping of all *L. monocytogenes* isolates, we describe the development and validation of a modular set of single-well assays for rapid SNP-based subtyping of lineage II, III, and IV isolates which complement our previously published lineage I assay (5).

MATERIALS AND METHODS

Bacterial strains and DNA sequence analysis. A diverse panel of 173 *L. monocytogenes* isolates was used in the design and validation of the MLGT assays (Table 1). An additional 20 *L. monocytogenes* isolates from sporadic listeriosis cases were used in a comparative subtyping analysis (Table 2). Isolates were cultured in brain heart infusion broth or tryptic soy agar supplemented with 0.6% (wt/vol) yeast extract (Difco) at 37°C and can be acquired from the Agricultural Research Service Culture Collection (NCAUR, Peoria, IL). With the exception of lineage IV isolates, which were characterized by phylogenetic analyses of DNA sequence data, the lineage identity of individual isolates was determined using a multiplex PCR assay (46). PCR serogroups for lineage II isolates were determined following the method of Doumith et al. (2). This method does not permit discrimination of serotypes within lineages III or IV. Therefore, previously published serotype data are reported (1, 10, 46).

In order to identify SNPs and other genetic polymorphisms segregating within lineage II, we sequenced 24,852 bp of DNA from each of 125 isolates (Table 1). This comparative DNA sequence database encompassed 12 regions of the *L. monocytogenes* genome and included genes responsible for virulence, stress response, and housekeeping functions (Table 3). Eight of these regions corresponded to those used in the lineage I MLGT assay (5), although VGC sequences for lineage II isolates were collected in two nonoverlapping segments (VGCa and VGCb). Three additional regions (ARO, PUR, and TRU) were included in the SNP discovery based on their high levels of sequence divergence in comparisons of EGD-e (GenBank accession number NC_003210) and F6854 (GenBank accession number AADQ00000000) genome sequences. The TRP region was included in the SNP discovery due to variation identified while characterizing a putative small noncoding RNA (sRNA) in the intergenic region between *trpE* and *lmo1634*. Due to high levels of sequence divergence observed for lineage III and IV isolates, SNP discovery for these lineages focused on the three most informative regions from the lineage I assay (5) and included 14,881 bp of DNA sequence (Table 3) from each of 48 isolates (Table 1). DNA isolation and sequencing were performed as previously described (46). PCR amplification protocols were previously described (5, 46) or are provided in the supplemental material.

DNA sequences were edited and aligned with Sequencher (version 4.1.2; Gene Codes), variable sites were identified with MEGA (version 4.0), and unique multilocus haplotypes were identified with DAMBE (49). Uncorrected genetic distance values (p-distances) were estimated in MEGA. Prior to phylogenetic or haplotype analyses, ambiguously aligned characters were removed from the data set and insertion or deletion (indel) polymorphisms that defined unique haplotypes were coded as single events. Distance analyses were performed using the neighbor-joining algorithm and the Kimura two-parameter model as implemented in MEGA. Maximum parsimony analyses were conducted using the tree bisection and reconnection method of branch swapping and the heuristic search algorithm of PAUP* (version 4.0B; Sinauer Associates). Relative support for individual nodes was assessed by bootstrap analysis with 2,000 replications (8, 35).

Multilocus genotyping assays. In the present study, we developed and validated two MLGT assays using the approach and platform previously described

for the lineage I assay (5). The first of these novel MLGT assays specifically targeted SNP variation segregating within lineage II, while we combined probes for lineages III and IV in a single MLGT assay. In order to provide for simultaneous interrogation of strain-discriminating polymorphisms identified in the comparative sequence analysis, multiple genomic regions were coamplified via an assay-specific multiplex PCR, which utilized seven sets of amplification primers for the lineage II assay and five sets of amplification primers for the lineage III and IV assay (Table 4). Multiplex amplifications were performed in 50- μ l reaction mixtures and included 2 mM MgSO₄, 100 μ M deoxynucleoside triphosphate, 300 nM primer, 1.5 U Platinum Taq DNA polymerase High Fidelity (Invitrogen Life Technologies), and 100 ng of genomic DNA. PCR consisted of an initial denaturation of 90 s at 96°C, followed by 40 cycles of 30 s at 94°C, 45 s at 50°C, and 150 s at 68°C. Amplification products were purified using Montage PCR cleanup filter plates (Millipore) and served as templates for allele-specific primer extension (ASPE) reactions.

We performed ASPE in multiplex reactions containing 64 (lineage II assay) or 51 (lineage III and IV assay) probes (Table 5). ASPE probes were designed with unique 5' sequence tags corresponding to individual sets of Luminex xMAP fluorescent polystyrene microspheres (Luminex Corporation). These probes served as allele-specific primers in extension reactions producing single-stranded DNA extension products from template DNA containing the probe sequence. Extensions were performed in 20- μ l reaction mixtures and included 2 mM MgCl₂, 5 μ M biotin-dCTP, 5 μ M dATP/dGTP/dTTP, 25 nM primer, 0.75 U Platinum GenoTYPE Tsp DNA polymerase (Invitrogen Life Technologies). Five μ l of purified multiplex PCR amplicon was added as template for extension reactions performed with an initial denaturation of 120 s at 96°C, followed by 40 cycles of 30 s at 94°C, 60 s at 52°C, and 120 s at 74°C.

Biotinylated extension products from ASPE reactions were sorted via hybridization with fluorescent microspheres. Microspheres were coated with anti-tag sequences specific to the 5' sequence tags appended to the extension probes. Hybridization reactions were performed in 50- μ l volumes with 1 \times TM buffer (0.2 M NaCl, 0.1 M Tris, 0.08% Triton X-100; pH 8.0), 10 μ l of extension product, and 625 microspheres from each set. The samples were incubated for 90 s at 96°C, followed by 45 min at 37°C. Microspheres were twice pelleted by centrifugation (3 min at 3,700 \times g) and resuspended in 70 μ l 1 \times TM buffer. Following these washes, microspheres were pelleted and resuspended in 70 μ l 1 \times TM buffer containing 2 μ g/ml streptavidin R-phycoerythrin. Samples were incubated for 15 min at 37°C prior to detecting the microsphere complexes with a Luminex 100 flow cytometer (Luminex Corporation).

The median fluorescence intensity (MFI) from biotinylated extension products attached to 100 polystyrene microspheres was adjusted to account for background fluorescence by subtracting the average MFI of three template-free controls from the MFI for each probe. Probe performance was assessed via the index of discrimination, defined as the ratio of the lowest target MFI to the highest nontarget MFI obtained in two independent runs of the sequenced isolate panels. Probes with a ratio less than 2.0 were redesigned. Critical values for discriminating between positive and negative alleles were set halfway between the lowest target MFI and the highest nontarget MFI observed among the sequenced isolates. In addition, positive control probes were designed to confirm the presence of each of the amplicons in the multiplex PCR. Minimum thresholds for positive control probes were set at 75% of the lowest MFI observed in the MLGT analysis of sequenced isolates.

Comparative subtyping. The discriminatory power of MLGT was compared with PFGE and two recently described multilocus sequence typing methods (37, 50) using a panel of 20 human sporadic listeriosis isolates provided by L. Graves (Centers for Disease Control and Prevention [CDC]). This independent panel of isolates was not used in SNP discovery or the design of MLGT probes. PulseNet pattern identifiers for AscI and ApaI were provided with the isolates. Multilocus sequencing was carried out as previously described (37, 50). DNA sequences were edited and analyzed as described above. Simpson's discrimination index (SDI) values were calculated as previously described (15). Comparative subtyping analysis for the lineage III and IV assay was not performed due to the rarity of these isolates.

Nucleotide sequence accession numbers. DNA sequences generated for this study were deposited in the GenBank database under accession numbers FJ024725 to FJ025060 and FJ041118 to FJ041147.

RESULTS AND DISCUSSION

DNA sequence analysis. A database of 24,852 bp of aligned DNA sequence from each of 125 lineage II isolates (Table 1) was developed for SNP discovery. This database consisted of

TABLE 1. *L. monocytogenes* isolates used in development of multilocus genotyping assays

NRRL ^a	Equivalent	Origin	Year	Lineage	Serotype ^b	PCR serogroup ^c	ST ^d	MLGT ^e
33002	RM2212	Food		II	1/2a	1/2a complex	ST2-1	Lm2.1
33009	RM2388	Food		II	1/2a	1/2a complex	ST2-2	Lm2.2
33014	12443	Animal		II	1/2a	1/2a complex	ST2-3	Lm2.3
33022	DSM20600	Animal		II	1/2a	1/2a complex	ST2-4	Lm2.4
33027		Frankfurters	2000	II	1/2a	1/2a complex	ST2-5	Lm2.5 (T700)
33029		Hot dogs	2000	II	1/2c	1/2c complex	ST2-6	Lm2.6
33031		Frankfurters	1999	II	1/2a	1/2a complex	ST2-7	Lm2.7
33034		Corn beef	2000	II	1/2a	1/2a complex	ST2-8	Lm2.8 (T189)
33035		Sausage	2000	II	1/2a	1/2a complex	ST2-9	Lm2.9
33039		Beef jerkey	2000	II	1/2c	1/2c complex	ST2-10	Lm2.10
33040		Chicken salad	2000	II	1/2a	1/2a complex	ST2-9	Lm2.9
33041		Red hots	2000	II	1/2a	1/2a complex	ST2-9	Lm2.9
33043		Lunch meat		II	1/2a	1/2a complex	ST2-11	Lm2.11
33044		Hot dogs		II	1/2a	1/2a complex	ST2-11	Lm2.11
33064	2064	Animal		II	1/2a	1/2a complex	ST2-12	Lm2.12
33069	2070	Milk		II	1/2a	1/2a complex	ST2-12	Lm2.12
33100	2612	Animal		II	1/2a	N/T	ST2-13	Lm2.13
33106	2420	Milk	1983	II	1/2a	1/2a complex	ST2-14	Lm2.14
33127	2063	Animal		II	1/2a	1/2a complex	ST2-12	Lm2.12
33128	2153	Potato		II	1/2a	1/2a complex	ST2-15	Lm2.15
33152	2072	Milk		II	1/2a	1/2a complex	ST2-12	Lm2.12
33167	2362	Environment		II	1/2a	1/2a complex	ST2-16	Lm2.16
33169	SE106	NA ^f		II	1/2a	1/2a complex	ST2-17	Lm2.17
33171	H6900	Human		II	1/2a	1/2a complex	ST2-53	Lm2.12
33180	41966-97	Animal		II	1/2a	1/2a complex	ST2-53	Lm2.12
33189	32285-01	Animal		II	1/2a	1/2a complex	ST2-18	Lm2.18
33215	LMB0027	Milk		II	1/2a	1/2a complex	ST2-12	Lm2.12
33216	LMB0033	Milk		II	1/2a	1/2a complex	ST2-54	Lm2.12
33219	LMB0340	Environment		II	1/2a	1/2a complex	ST2-19	Lm2.19 (T700)
33223	H9333	Human		II	1/2c	1/2c complex	ST2-10	Lm2.10
33225	J0095	NA		II	3a	1/2a complex	ST2-20	Lm2.20 (T492)
33226	J0096	NA		II	3c	1/2c complex	ST2-21	Lm2.21 (T685)
33235		Frankfurters	2000	II		1/2a complex	ST2-5	Lm2.5 (T700)
33236		Frankfurters	2000	II		1/2a complex	ST2-22	Lm2.22
33238		Beef jerkey	2000	II		1/2c complex	ST2-10	Lm2.10
33241		Sausage	2000	II		1/2a complex	ST2-5	Lm2.5 (T700)
33243		Beef	2000	II		1/2a complex	ST2-11	Lm2.11
33244		NA	2000	II		1/2a complex	ST2-23	Lm2.23
33246		Chicken salad	2000	II		1/2a complex	ST2-9	Lm2.9
33247		Roast beef	2000	II		1/2a complex	ST2-9	Lm2.9
33249		NA	2000	II		1/2a complex	ST2-55	Lm2.12
33253		Ham	2001	II		1/2a complex	ST2-9	Lm2.9
33255		Sausage	2001	II		1/2a complex	ST2-7	Lm2.7
33256		Sausage	2001	II		1/2a complex	ST2-7	Lm2.7
33257		Roast beef	2001	II		1/2a complex	ST2-9	Lm2.9
33259		Chicken	2001	II		1/2c complex	ST2-10	Lm2.10
33260		Sausage	2001	II		1/2c complex	ST2-24	Lm2.24 (T577)
33264		Beef	2001	II		1/2c complex	ST2-10	Lm2.10
33271		NA	2001	II		1/2a complex	ST2-11	Lm2.11
33274		NA	2001	II		1/2a complex	ST2-11	Lm2.11
33275		NA	2001	II		1/2a complex	ST2-25	Lm2.25
33276		Chicken	2001	II		1/2a complex	ST2-56	Lm2.12
33277		NA	2001	II		1/2c complex	ST2-10	Lm2.10
33278		NA	2001	II		1/2c complex	ST2-10	Lm2.10
33279		NA	2001	II		1/2c complex	ST2-10	Lm2.10
33280		NA	2001	II		1/2c complex	ST2-10	Lm2.10
33281		Sausage	2001	II		1/2c complex	ST2-26	Lm2.26
33282		Duck breast	2001	II		1/2a complex	ST2-5	Lm2.5 (T700)
33283		Chicken base	2001	II		1/2a complex	ST2-5	Lm2.5 (T700)
33285		Turkey breast	2001	II		1/2c complex	ST2-10	Lm2.10
33286		Sausage	2001	II		1/2a complex	ST2-27	Lm2.27
33288		Beef	2001	II		1/2c complex	ST2-10	Lm2.10
33290		Beef quesadilla	2001	II		1/2a complex	ST2-28	Lm2.28
33292		Turkey breast	2001	II		1/2c complex	ST2-10	Lm2.10
33295		Chorizo	2001	II		1/2c complex	ST2-10	Lm2.10
33297		Sausage	2001	II		1/2c complex	ST2-26	Lm2.26
33298		Sausage	2001	II		1/2a complex	ST2-27	Lm2.27

Continued on following page

TABLE 1—Continued

NRRL ^a	Equivalent	Origin	Year	Lineage	Serotype ^b	PCR serogroup ^c	ST ^d	MLGT ^e
33299		Sausage	2001	II		1/2a complex	ST2-27	Lm2.27
33307		Chicken breast	2001	II		1/2a complex	ST2-5	Lm2.5 (T700)
33310		Ham	2001	II		1/2c complex	ST2-29	Lm2.29
33311		Chicken quesadilla	2001	II		1/2a complex	ST2-30	Lm2.30 (T189)
33316		Roast beef	2001	II		1/2a complex	ST2-31	Lm2.31
33317		Deli turkey	2001	II		1/2a complex	ST2-52	Lm2.11
33318		Turkey, cheese, deli	2001	II		1/2a complex	ST2-52	Lm2.11
33319		Frankfurters	2001	II		1/2a complex	ST2-11	Lm2.11
33321		Roast duckling	2002	II		1/2a complex	ST2-32	Lm2.32
33324		Pork spring rolls	2002	II		1/2a complex	ST2-33	Lm2.33 (T700)
33326		Salami	2002	II		1/2a complex	ST2-34	Lm2.34 (T492)
33332		Sausage	2001	II		1/2a complex	ST2-5	Lm2.5 (T700)
33335		Pork chops	2002	II		1/2a complex	ST2-35	Lm2.35
33336		Pork chops	2002	II		1/2a complex	ST2-5	Lm2.5 (T700)
33338		Pork	2002	II		1/2a complex	ST2-9	Lm2.9
33339		Chicken egg roll	2002	II		1/2a complex	ST2-33	Lm2.33 (T700)
33340		Soujouk	2002	II		1/2c complex	ST2-10	Lm2.10
33342		Turkey pastrami	2002	II		1/2a complex	ST2-11	Lm2.11
33344		Sausage	2002	II		1/2c complex	ST2-26	Lm2.26
33348		Bologna	2003	II		1/2a complex	ST2-36	Lm2.36
33349		Pork chops	2003	II		1/2a complex	ST2-37	Lm2.37
33350		Sausage	2003	II		1/2a complex	ST2-51	Lm2.9
33353		Ham	2003	II		1/2a complex	ST2-38	Lm2.38 (T492)
33357		Chicken salad	2003	II		1/2a complex	ST2-5	Lm2.5 (T700)
33380		Animal	2004	II		1/2a complex	ST2-39	Lm2.39
33387	FSL J2-020	Animal	1986	II	1/2a	1/2a complex	ST2-40	Lm2.40
33395	FSL C1-056	Human	1998	II	1/2a	1/2a complex	ST2-53	Lm2.12
33396	FSL J2-054	Animal	1993	II	1/2a	1/2a complex	ST2-41	Lm2.41
33397	FSL M1-004	Human	1997	II		1/2a complex	ST2-11	Lm2.11
33398	FSL J2-031	Animal	1996	II	1/2a	1/2a complex	ST2-42	Lm2.42
33399	FSL J2-066	Animal	1994	II	1/2a	1/2a complex	ST2-43	Lm2.43
33402	FSL C1-115	Human	1998	II	3a	1/2a complex	ST2-44	Lm2.44
33417	F6854	Hot dog	2000	II	1/2a	1/2a complex	ST2-1	Lm2.1
33419	J0161	Human	2000	II	1/2a	1/2a complex	ST2-1	Lm2.1
33427		Environment	2004	II		1/2a complex	ST2-45	Lm2.45
33428		Turkey	2003	II		1/2a complex	ST2-46	Lm2.46
33431		Turkey	2003	II		1/2a complex	ST2-46	Lm2.46
33433		Salami	2003	II		1/2a complex	ST2-20	Lm2.20 (T492)
33435		Environment	2003	II		1/2a complex	ST2-11	Lm2.11
33436		Environment	2003	II		1/2a complex	ST2-11	Lm2.11
33437		Environment	2003	II		1/2a complex	ST2-11	Lm2.11
33438		Pork	2003	II		1/2a complex	ST2-11	Lm2.11
33439		Pork	2003	II		1/2a complex	ST2-11	Lm2.11
33440		Pork	2003	II		1/2a complex	ST2-11	Lm2.11
33441		Pork	2003	II		1/2a complex	ST2-11	Lm2.11
33443		Roast beef	2003	II		1/2a complex	ST2-30	Lm2.30 (T189)
33446		Chicken breast	2003	II		1/2a complex	ST2-9	Lm2.9
33447		Food	2003	II		1/2a complex	ST2-47	Lm2.47
33448		Ham	2003	II		1/2a complex	ST2-7	Lm2.7
33449		Prosciutto	2003	II		1/2a complex	ST2-5	Lm2.5 (T700)
33450		Prosciutto	2003	II		1/2c complex	ST2-24	Lm2.24 (T577)
33452		Chicken burrito	2004	II		1/2a complex	ST2-1	Lm2.1
33454		Environment	2004	II		1/2a complex	ST2-50	Lm2.5 (T700)
33455		Beef	2004	II		1/2a complex	ST2-5	Lm2.5 (T700)
33456		Ham	2004	II		1/2a complex	ST2-5	Lm2.5 (T700)
33460		Environment	2004	II		1/2a complex	ST2-48	Lm2.48
33467		Environment	2004	II		1/2a complex	ST2-49	Lm2.49
33468		Pork	2004	II		1/2a complex	ST2-5	Lm2.5 (T700)
33077	7035	Animal		III	4b		ST3-1	Lm3.1
33092	7678	Animal		III	4b		ST3-2	Lm3.2
33105	7676	Animal		III	4b		ST3-3	Lm3.3
33115	3890	Animal		III	4c		ST3-16	Lm3.16
33177	28838-95	Animal		III	4c		ST3-4	Lm3.4
33181	1709-98	Animal		III	4b		ST3-5	Lm3.5
33182	7259-98	Animal		III	4c		ST3-6	Lm3.6
33183	20842-98	Animal		III	4b		ST3-7	Lm3.7
33184	11466-01	Animal		III	4c		ST3-8	Lm3.8

Continued on following page

TABLE 1—Continued

NRRL ^a	Equivalent	Origin	Year	Lineage	Serotype ^b	PCR serogroup ^c	ST ^d	MLGT ^e
33185	12459-01	Animal		III	4b		ST3-9	Lm3.9
33187	22409-01	Animal		III	4b		ST3-10	Lm3.10
33188	23594-01	Animal		III	4c		ST3-11	Lm3.11
33190	36087-01	Animal		III	4b		ST3-12	Lm3.12
33191	50301-01	Animal		III	4b		ST3-13	Lm3.13
33227	J0099	NA		III	4c		ST3-14	Lm3.14
33229	ILSI19	Human		III	4c		ST3-38	Lm3.38
33230	LMB0291	Food		III	4b		ST3-15	Lm3.15
33231	ATCC19116	Food		III	4c		ST3-16	Lm3.16
33330		Liquid whole egg	2002	III			ST3-17	Lm3.17
33360		Human		III			ST3-18	Lm3.18
33362		Animal	2002	III			ST3-27	Lm3.27
33363		Animal	2002	III			ST3-19	Lm3.19
33364		Animal	2002	III			ST3-21	Lm3.21
33365		Animal	2002	III			ST3-20	Lm3.20
33366		Animal	2002	III			ST3-21	Lm3.21
33367		Animal	2002	III			ST3-22	Lm3.22
33368		Animal	2003	III			ST3-23	Lm3.23
33370		Animal	2003	III			ST3-24	Lm3.24
33371		Animal	2003	III			ST3-25	Lm3.25
33372		Animal	2004	III			ST3-26	Lm3.26
33373		Animal	2004	III			ST3-27	Lm3.27
33374		Animal	2004	III			ST3-28	Lm3.28
33375		Animal	2004	III			ST3-29	Lm3.29
33376		Animal	2004	III			ST3-30	Lm3.30
33377		Animal	2004	III			ST3-31	Lm3.31
33378		Animal	2004	III			ST3-32	Lm3.32
33379		Animal	2004	III			ST3-33	Lm3.33
33381		Animal	2004	III			ST3-34	Lm3.34
33382		Animal	2004	III			ST3-35	Lm3.35
33383		Animal	2004	III			ST3-36	Lm3.36
33384		Animal	2004	III			ST3-37	Lm3.37
33403	FSL J1-031	Human		III	4a		ST3-38	Lm3.38
33407	FSL W1-110	NA		III	4c		ST3-39	Lm3.39
33425		Animal		III			ST3-40	Lm3.40
33426	FDA 3365	Food		III	4b		ST3-41	Lm3.41
33405	FSL W1-111	NA		IV	4c		ST4-1	Lm4.1
33406	FSL W1-112	NA		IV	4a		ST4-2	Lm4.2
33408	FSL J1-158	Animal	1997	IV	4b		ST4-3	Lm4.3

^a Isolates are identified with NRRL-B numbers from the U.S. Department of Agriculture ARS Culture Collection, Peoria, IL. Additional strain history information is available from the ARS Culture Collection website (<http://nrri.ncaur.usda.gov/cgi-bin/usda>).

^b Serotypes were previously reported (1, 10, 46) or were previously determined by a reference laboratory.

^c Determined using the method of Doumith et al., which was not designed to discriminate serotypes within lineages III or IV (2).

^d ST, sequence type.

^e The amino acid positions of premature stop codons in *inlA* relative to the 800-amino-acid protein predicted for strain EGD-e (NC_003210) are indicated parenthetically following the MLGT designation.

^f NA, not available.

sequences from 12 regions of the *L. monocytogenes* genome and contained 1,419 SNPs (Table 3), as well as indel variation, which defined 56 unique multilocus sequence types (Table 1). Six sequence types were observed among the 22 isolates assigned via PCR analysis to serogroup 1/2c (includes serotype 1/2c and 3c isolates), while 50 unique sequence types were observed among the 103 isolates from serogroup 1/2a (includes serotype 1/2a and 3a isolates) (Table 1). There were no sequence types in common between the serogroup 1/2c and 1/2a isolates in Table 1, and phylogenetic analyses of the multilocus sequence data placed all serogroup 1/2c isolates within a single clade (Fig. 1). However, analysis of sequences from the reference strain EGD-e revealed that this serotype 1/2a strain had the same sequence type (ST2-10) as 14 isolates from serogroup 1/2c (Table 1). In addition, phylogenetic analyses placed NRRL B-33467, a serogroup 1/2a isolate, within the 1/2c clade

(Fig. 1). Previous analyses using macroarrays revealed that EGD-e is an atypical strain of serotype 1/2a with a genome more similar to serotype 1/2c strains than to other 1/2a strains (2, 3). Our results indicate that NRRL B-33467 is a similarly atypical strain and suggest that the 1/2a serotype has multiple evolutionary origins or that the 1/2a serotype has been acquired by some strains with a 1/2c genomic background via genetic exchange. Excluding EGD-e and NRRL B-33467, the serogroup 1/2c isolates formed a monophyletic clade with 100% bootstrap support, while the serogroup 1/2a isolates constituted a highly diverse paraphyletic assemblage of strains that were genetically distinct from serogroup 1/2c strains (Fig. 1).

The 125 lineage II isolates in Table 1 included two representatives of the previously described ECIII, which is the only lineage II epidemic clone described to date (20). NRRL B-33417 was implicated in a human listeriosis case associated

TABLE 2. Comparative subtype data for sporadic human listeriosis isolates

NRRL ^a	Equivalent	Serotype ^b	MLGT ^c	AscI pattern	ApaI pattern	MVLST ^d	MLST ^e
33555	J0221	1/2a	Lm2.46	GX6A16.0170	GX6A12.0316	2	2
33556	J0809	1/2a	Lm2.50	GX6A16.0401	GX6A12.0519	1	1
33557	J0834	1/2a	Lm2.11	GX6A16.0018	GX6A12.0022	5	5
33558	J0846	1/2a	Lm2.11	GX6A16.0034	GX6A12.0022	5	5
33559	J0847	1/2a	Lm2.46	GX6A16.0233	GX6A12.0467	2	2
33560	J1038	1/2a	Lm2.9	GX6A16.0010	GX6A12.0005	3	3
33561	J1108	1/2a	Lm2.28	GX6A16.0458	GX6A12.0559	5	5
33562	J1219	1/2a	Lm2.11	GX6A16.0018	GX6A12.0022	5	5
33563	J1769	1/2a	Lm2.44	GX6A16.0055	GX6A12.0708	4	4
33564	J2061	1/2a	Lm2.11	GX6A16.0018	GX6A12.0022	5	5
33565	J2342	1/2a	Lm2.51	GX6A16.0675	GX6A12.0716	6	6
33566	J2692	1/2a	Lm2.12	GX6A16.0083	GX6A26.0598	10	11
33567	J2980	1/2a	Lm2.52	GX6A16.0347	GX6A12.0031	7	7
33568	J3560	1/2a	Lm2.15	GX6A16.0849	GX6A12.0451	8	8
33575	J1109	1/2c	Lm2.10	GX6A16.0057	GX6A12.0085	9	10
33576	J1131	1/2c	Lm2.10	GX6A16.0279	GX6A12.0356	9	9
33577	J2252	1/2c	Lm2.10	GX6A16.0082	GX6A12.0705	9	10
33578	J2283	1/2c	Lm2.24	GX6A16.0057	GX6A12.0085	9	10
33579	J2315	1/2c	Lm2.10	GX6A16.0082	GX6A12.0679	9	10
33580	J0845	3a	Lm2.12	GX6A16.0609	GX6A12.0797	10	11

^a Isolates are identified with NRRL-B numbers from the U.S. Department of Agriculture ARS Culture Collection, Peoria, IL. Additional strain history information is available from the ARS Culture Collection website (<http://nrri.ncaur.usda.gov/cgi-bin/usda>).

^b Serotype data were provided by Lewis Graves (CDC).

^c Lm2.50 to 2.52 were not observed in the original set of sequenced isolates (Table 1).

^d Multi-virulence locus sequence typing was performed as described by Zhang et al. (50).

^e Multilocus sequence typing was performed as described by Revazishvili et al. (37).

with turkey franks in 1988, and NRRL B-33419 was associated with a multistate outbreak of listeriosis due to contaminated turkey deli meat in 2000. These isolates have an identical sequence type (ST2-1), which is consistent with previous evidence indicating they represent a strain that persisted in the same processing facility for more than a decade with minimal genetic change (20). Two additional food isolates (NRRL B-33002 and NRRL B-33452) with ST2-1 were identified among the 125 isolates examined. In addition, NRRL B-33275 had a sequence type (ST2-25) that differed from ST2-1 by only four substitutions out of the 24,852 bp of sequence analyzed (p -distance = 2×10^{-4}), and it was considered a representative of the ECIII lineage on the basis of phylogenetic analyses

(Fig. 1) and the phylogenetic definition of clonal lineages described by Ducey et al. (5).

Five unique mutations leading to premature stop codons in *inlA* were observed among 27 of the 125 lineage II isolates sequenced for SNP discovery (Tables 1 and 6). The *inlA* gene encodes a membrane-anchored invasion protein that interacts directly with host cell receptors to facilitate *L. monocytogenes* entry into nonprofessional phagocytes (14, 21). Previous studies documented that strains carrying mutations resulting in *InlA* truncations 5' to the C-terminal LPXTG membrane-anchoring motif display virulence-attenuated phenotypes in animal models and a significantly reduced ability to invade human intestinal epithelial cells (17, 19, 25, 26, 29, 30, 39). All

TABLE 3. Single nucleotide polymorphism and haplotype variations among 173 *L. monocytogenes* isolates

Lineage(s)	Region	Position ^a	Genes or loci ^b	No. of SNPs	No. of haplotypes
II	VGCa	205,814–208,259	<i>hly</i> , <i>mpl</i>	43	28
II	VGCb	209,573–211,447	<i>actA</i> , <i>plcB</i>	92	25
II	LMO	325,093–327,031	<i>lmo0298</i> , <i>lmo0299</i> , <i>lmo0300</i>	160	26
II	INL	454,789–458,788	<i>inlA</i> , <i>inlB</i>	139	31
II	SIG	930,314–931,861	<i>rsbV</i> , <i>rsbW</i> , <i>sigB</i>	16	14
II	PDH	1,079,263–1,081,952	<i>pdhA</i> , <i>pdhB</i> , <i>pdhC</i>	106	22
II	TRU	1,356,824–1,357,931	<i>truB</i> , <i>ribC</i>	202	18
II	AMI	1,553,548–1,555,355	<i>ami</i> , <i>hisS</i>	27	16
II	ACC	1,612,612–1,614,874	<i>accD</i> , <i>dnaE</i>	143	25
II	TRP	1,676,679–1,678,787	<i>lmo1634</i>	69	23
II	PUR	1,838,571–1,840,142	<i>purN</i> , <i>purM</i> , <i>purF</i>	237	24
II	ARO	1,998,007–1,999,495	<i>tyrA</i> , <i>aroE</i> , <i>lmo1922</i>	185	25
III & IV	VGC	203,659–212,465	<i>prfA</i> , <i>plcA</i> , <i>hly</i> , <i>mpl</i> , <i>actA</i> , <i>plcB</i> , <i>lmo0206</i>	876	42
III & IV	LMO	325,093–327,032	<i>lmo0298</i> , <i>lmo0299</i> , <i>lmo0300</i>	228	39
III & IV	INL	454,708–458,828	<i>inlA</i> , <i>inlB</i>	510	40

^a Corresponding nucleotide positions in the complete genome sequence of *L. monocytogenes* strain EGD-e (NC_003210).

^b Gene designations follow the annotated EGD-e genome (NC_003210).

TABLE 4. Primers used in multiplex amplifications for the MLGT assays

Lineage(s)	Region	Primer	Sequence (5'-3') ^a
II-III-IV	VGCa	hly-F2	GAAGGAGAGTGAACCCATG
II-III-IV	VGCa	mpl-R	CTCGCAACTTCYGGCTCAGC
II-III-IV	VGCb	actA-F	CCAAATGCATTACGATTAACC
II-III-IV	VGCb	plcB-R	CAAGCACATACCTAGAACCAC
II-III-IV	INLa	inlA-F2	ACGAAYGTAACAGACACGGTC
II	INLa	inlA-R2	CTACTTCTATTTACTAGCACG
III-IV	INLa	inlA-R3	TCTAGCTCTTYACACTACTTC
II-III-IV	INLb	inlB-F2	AATCAAGGAGAGGATAGTGTG
II-III-IV	INLb	inlB-R	CTACCGGRACCTTATAGTAYG
II	LMO	lmo0300-F	GATTGGATTAAGTACGAGC
II	LMO	lmo0298-R	CTCTTATCAATGTGAAGGTGC
III-IV	LMO	lmo0300-F2	ATGTRGCGAGCGATCACTACC
III-IV	LMO	lmo0300-R2	AATCTTCGTGRTAGTGTTCG
II	TRP	adhE-F	TGGAAGTTTGAACCATTGCAT
II	TRP	adhE-R	AGGACGAGAGACCCGTGGTA
II	ACC	dnaE-F	GGATTTTCDCTTGAGAAAGCAG
II	ACC	dnaE-R	CRCCGACAACAGARCCCATAC

^a See IUPAC codes for definitions of degenerate bases.

of the InlA truncation mutations observed in this study occurred 5' to the LPXTG motif, and four of these five mutations were previously characterized (Table 6). However, we identified a novel mutation (T189) (Table 6) in three RTE meat isolates, which resulted in a predicted protein of only 189 amino acids. While the InlA truncation mutations were observed in strains from different parts of the lineage II phylogeny, little sequence variation was observed between isolates with the same truncation mutation (p -distance $< 1.2 \times 10^{-4}$), indicating that each of these mutations had a recent evolutionary origin (Fig. 1). Interestingly, strains carrying T189 or T492 mutations formed a monophyletic clade that was closely aligned with ECIII strains in the multigene phylogeny (Fig. 1), suggesting that closely related clonal lineages might have different virulence properties.

A comparative sequence database consisting of 14,881 bp of DNA sequence from 48 isolates was developed for SNP discovery within the rare lineages III and IV. The regions sequenced (VGC, LMO, and INL) contained 1,614 SNPs (Table 3), as well as indel variation that defined 44 multilocus sequence types among the 48 lineage III and IV isolates examined (Table 1). With the exception of a mutation leading to a single amino acid reduction in the predicted length of InlA among two of the lineage IV strains, there were no premature stop codons observed within the lineage III and IV sequences.

Phylogenetic analysis of VGC, LMO, and INL sequences confirmed previous reports (22, 33, 38) that lineage IV isolates are not part of a monophyletic lineage III and indicated that they represent an early divergence within *L. monocytogenes* (Fig. 2). While lineages III and IV are both polymorphic for serotypes 4a, 4c, and 4b, the results of the current analysis (Fig. 2) and previous phylogenetic analyses (33, 38) demonstrate that this shared serotype polymorphism is not the result of an exclusive shared ancestry. In addition, mapping serotype data onto the multigene phylogeny (Fig. 2) indicated that in contrast to lineages I and II, genetic variation within lineage III is not strongly partitioned in accordance with serotype divisions. Taken together, these results support the hypothesis that serotype polymorphism shared by lineages III and IV was derived independently from a serogroup 4 ancestor and is not a reliable indicator of evolutionary relatedness for these lin-

eages. Alternatively, these results could be interpreted as indicating an inability of serotyping techniques to precisely differentiate among serotypes 4a, 4c, and 4b, as has been reported previously for serotypes 4b, 4d, and 4e within lineage I (34).

Design and validation of the MLGT assays. In order to develop single-well assays for the rapid and accurate discrimination of lineage II, III, and IV subtypes, variation identified in the comparative sequence analysis (Table 3) was used to design multilocus genotyping probes from the seven genomic regions listed in Table 4. All three of the regions sequenced for SNP discovery within lineages III and IV (Table 3) were included in a combined lineage III and IV MLGT assay (Table 4), which identified all 44 unique haplotypes among the 48 lineage III and IV isolates sequenced. However, in order to optimize the performance of the multiplex PCR while providing maximal haplotype discrimination, the lineage II MLGT was focused on the seven amplicons listed in Table 4. This single-well assay differentiated 49 unique haplotypes among the 125 lineage II isolates in Table 1. Each of the regions listed in Table 3 that were excluded from the lineage II MLGT assay differentiated no more than one additional haplotype, with the exception of PDH, which differentiated two novel haplotypes.

The lineage II MLGT assay consisted of 64 probes, which included seven positive control probes designed to confirm the presence of each of the amplicons in the multiplex PCR (Table 4). The MLGT assay for lineages III and IV consisted of 51 probes, which included positive control probes for each of five amplicons (Table 4). Probe sequences and probe performance data are provided in Table 5. The index of discrimination values for the MLGT probes ranged from 2.4 to 73.7 with a mean of 14.0. This means that the fluorescence intensities (MFI values) for isolates with a positive allele were always more than twice as high, and on average 14 times as high, as background values for isolates with a negative allele. Replicate runs of the 173 sequenced isolates produced identical results, demonstrating the reproducibility of MLGT data and the ability of the MLGT assays to type all of the tested isolates. In addition, the single-well MLGT assays recovered 93% of the 100 haplotypes defined by the full DNA sequence data set (>3.8 Mb) described in Table 3 and 100% of the haplotypes defined by more than 2.5 Mb of comparative DNA sequence data from the regions incorporated into the assays (Table 4). Probe patterns for unique haplotypes are provided in Table S1 of the supplemental material.

The 10,448 MLGT genotypes derived from the 115 total probes utilized in the assays were consistent with DNA sequence data for the 173 sequenced isolates. Isolates from the previously linked listeriosis case associated with turkey franks in 1988 and a multistate outbreak in 2000 had identical MLGT patterns (Lm2.1), demonstrating good epidemiological concordance. In addition, MLGT types were not shared in common between serogroup 1/2a and 1/2c isolates that we tested, indicating that MLGT type could provide accurate information about major serogroup identity for most lineage II isolates. Serotype prediction based on MLGT may not be possible among lineage III isolates or in rare instances in which serotype 1/2a strains have a predominately 1/2c genome (Fig. 1). However, this results from a poor correlation between genomic and antigenic variation and does not reflect a failure of MLGT

TABLE 5. ASPE probes and probe performance data

Probe ^b	Probe sequence ^c	Adjusted MFI (range)		Target N ^d	ID ^e
		Nontarget	Target		
Lm2.ACC1 (94)	CTTCTATCTTTCTACTCAATAATccgttatgtttccggaatgct	7–183	1,943–2,340	1	10.6
Lm2.ACC2 (11)	TACAAATCATCAATCACTTTAATCggtattatgacaaaatgctcg	44–158	2,379–3,043	14	15.1
Lm2.ACC3 (67)	TCATTTACTCAACAATTACAAATCcaagaagaaagcggatgtcctg	104–231	2,916–2,935	1	12.6
Lm2.ACC4 (63)	CTACTTCATATACTTTATACTACAggggatagtaagggtatcgtt	–3 to 193	3,424–4,285	4	17.8
Lm2.ACC5 (55)	TATATACACTTCTCAATAACTAACgacgattttggcaactattgatg	48–208	3,949–3,974	1	19.0
Lm2.ACCx (69)	CTATAAACATATTACATTCACATCggttccagcayccatgacg	N/A ^a	3,118–4,842	125	N/A
Lm2.INLa1 (5)	CAATTCAAATCACAATAATCAATCaccacctccgcaaatatag	266–545	5,880–6,371	1	10.8
Lm2.INLa10 (13)	CAATAAACTATACTTCTTCACTAAaacagatggkaaaaaatg	60–63	1,399–3,002	124	22.3
Lm2.INLa11 (98)	AATCATACTCAACTAATCATTCAAaaacaggyggaactaaatggrat	56–73	320–1,569	123	4.4
Lm2.INLa12 (8)	AATCCTTTTACATTCATTACTTACgcaaatgacattacgctgtag	16–165	2,063–2,806	17	12.5
Lm2.INLa13 (59)	TCATCAATCAATCTTTTTCACTTTgtgctgcttcagggttaactagtctat	95–254	2,390–2,882	3	9.4
Lm2.INLa14 (65)	CTTTTCATCAATAATCTTACCTTTTcatttagggaactgtgacgt	22–127	2,172–2,508	4	17.2
Lm2.INLa15 (97)	AATCTCATAATCTACATACACTATaaacaggyggaactaaatggtat	27–97	1,981–2,380	2	20.5
Lm2.INLa2(54)	CTTTTTCATCACTTTCAATTCATCgcaaatgacattacgctgtag	143–307	2,436–2,645	1	7.9
Lm2.INLa3 (28)	CTACAAAACAAAACAACTTATCAAcgaaaaaacagatgggaaaaaata	17–133	981–1,048	1	7.4
Lm2.INLa4 (23)	TTCAATCATTCAAATCTCAACTTTCcttatatgcccaatatagcgt	41–246	4,533–4,839	1	18.4
Lm2.INLa5 (74)	TACACATCTTCAAACAAATTTCAcggatataaacgccactaaag	20–108	852–1,417	39	7.9
Lm2.INLa6 (12)	TACATTTTCTTTTCTTTCTTTTaaactaatctacagcaatatt	–13 to 72	487–582	1	6.7
Lm2.INLa7 (100)	CTATCTTTAAACTACAAATCTAACgcaaatgayattacgctgtac	54–100	1,194–2,157	108	12.0
Lm2.INLa8 (70)	ATACCAATAATCCAATTCATATCAgtgctgcttcagggttaaytartetac	52–82	753–1,542	122	9.2
Lm2.INLa9 (43)	CTTTCAATTACAATACTCATTACacatttagtggaaacygtgacgc	39–59	1,103–2,309	121	18.6
Lm2.INLax (62)	TCAATCATAAATCTCATAATCCAATCccaccactgacacatcagtc	N/A	906–1,751	125	N/A
Lm2.INLb1 (29)	AATCTTACTACAAATCCTTTCTTTgatgttgatggaacggttaatt	47–202	2,129–2,331	1	10.5
Lm2.INLb2 (60)	AATCTACAAATCCAATAATCTCATggatagtgtaagaaagacgcg	18–125	678–838	1	5.4
Lm2.INLb3 (20)	CTTTTACAATACTTCAATACAATCcttccagcagaagataactca	72–197	4,568–4,847	1	23.1
Lm2.INLb4 (73)	ATCAAATCTCATCAATTTCAACAATCcaagaaagcagctcaacttaact	66–185	4,930–5,110	1	26.6
Lm2.INLb5 (72)	TCATTTACCTTTAATCCAATAATCcaagcggagactataccgctgt	45–115	2,249–2,804	22	19.5
Lm2.INLb6 (18)	TCAAAATCTCAAATACTCAAATCAcgtatctgctgcaagccaaaacg	128–242	2,391–3,030	14	9.9
Lm2.INLbx (24)	TCAATTACCTTTTCAATACAATAcgtacaagcggagactatacacc	N/A	1,315–2,072	125	N/A
Lm2.LMO1 (26)	TTACTCAAATCTACACTTTTTTCAactcggattgctctataagcat	68–269	5,230–5,319	1	19.5
Lm2.LMO10 (86)	CTAATTACTAACATCACTAACAATCgaatgacgcacacaacgctcg	1–314	6,231–7,806	20	19.8
Lm2.LMO11 (22)	AATCCTTTTTACTCAATTCATCAcgtatctagcaagttttgatag	–28 to 826	8,534–10,203	8	10.3
Lm2.LMO12 (14)	CTACTATACATCTTACTATACTTTTggcgttgcgtgctaagttta	32–106	1,458–2,039	23	13.8
Lm2.LMO2 (3)	TACATTTTATCAAATCTTACAATCgtgttgcgcaagcggcttca	44–179	1,945–2,045	1	10.9
Lm2.LMO3 (30)	TTACCTTTTATACCTTTCTTTTATCctgtgctgagactatgctgtct	61–401	8,661–9,449	1	21.6
Lm2.LMO4 (76)	AATCTAACAAACTCATCTAAATACcacaacggctcaatatacaa	30–126	2,612–2,864	1	20.8
Lm2.LMO5 (6)	TCAACAATCTTTTACAATCAAATCaggcaagcaggaaagccatta	46–217	4,267–4,554	1	19.6
Lm2.LMO6 (16)	AATCAATCTTCAATCAAATCATCAgagaccaagcgtatgagccg	153–311	5,100–7,093	39	16.4
Lm2.LMO7 (19)	TCAATCAATTACTTACTCAAATACggcagcagtgacataaaggcg	113–242	1,417–1,572	1	5.9
Lm2.LMO8 (9)	TAATCTTCTATATCAACATCTTACTcctgagctggcaayaggtca	27–388	2,770–7,807	12	7.1
Lm2.LMO9 (37)	CTTTTCATCTTTTCACTTTTCAATcacaaccagcagcattgctgta	46–168	2,866–3,809	9	17.1
Lm2.LMOx (56)	CAATTTACTCATATACATCACTTTTcgattaagccatcttgaagcc	N/A	2,368–4,315	125	N/A
Lm2.TRP1 (77)	CAATTAACATACATAACAATACATACgcaaatatcccaatctcat	219–781	3,431–5,879	3	4.4
Lm2.TRP2 (88)	TTACTTCACTTTTCTATTTACAATCgtgacatttataacggcttcatca	524–930	10,830–11,262	1	11.6
Lm2.TRP3 (93)	CTTTCTATTCATCTAAATACAAACcaagtaagaaaaagaactaccgg	26–122	3,373–4,622	17	27.7
Lm2.TRP4 (89)	TATACTAACAATCAACAATATcaaatgataatcgctaaatct	28–141	3,133–3,359	1	22.3
Lm2.TRP5 (91)	TTCAATAACATCAATCATAACTTACgcaacaagcctaattttttggg	100–234	2,937–3,000	1	12.6
Lm2.TRPx (78)	CTATCTATCTAACTATCTATATCAacatcatgctcgcctactgac	N/A	742–1,506	125	N/A
Lm2.VGCa1 (80)	CTAACTAAACAATAATCTAACTAACggagagtgaaaaccatgaaaaat	24–101	528–544	1	5.2
Lm2.VGCa2 (96)	ATACTAACTCAACTAACTTTTAAACctgaagcaaaagatgcatctgt	14–118	770–931	1	6.5
Lm2.VGCa3 (44)	TCATTTACCAATCTTTCTTTTATACgctttgatgctgctgcaagt	60–134	1,935–2,033	1	14.4
Lm2.VGCa4 (2)	CTTTTACAATACATACTACAATCAgagaacaccagggattcccg	166–322	3,646–3,873	1	11.3
Lm2.VGCa5 (21)	AATCCTTTCTTTAATCTCAAATCAgagagatgagtgacaaatggtcca	39–140	1,322–1,420	1	9.4
Lm2.VGCa6 (51)	TCATTTCAATCAATCATCAACAATgcaaaaactataatttagttct	25–105	1,870–2,066	2	17.8
Lm2.VGCa7 (66)	TAACTATACAATATACTACTACTACgcttgggaatgggtggagaaa	152–271	998–1,185	3	3.7
Lm2.VGCa8 (57)	CAATATCATCATCTTTTATCATTACagttgttaggtgcttaaacct	15–82	365–587	18	4.4
Lm2.VGCa9 (17)	CTTTAATCCTTTTATCACTTTATCAgaccggaacctaccacttga	53–171	2,816–2,952	1	16.5
Lm2.VGCax (40)	CTTTTCACTATTTCACAACATTAactcagcattgatttggccgg	N/A	721–2,016	125	N/A
Lm2.VGcb1 (41)	TTACTACACAATATACTACTCAATTTgtccagagacaccgatgca	17–199	4,435–4,499	1	22.3
Lm2.VGcb2 (10)	ATCATACATACATACAAATCTACAgacataactaaaaagcgcattc	24–129	3,518–4,056	1	27.3
Lm2.VGcb3 (35)	CAATTCATCATTCATTCATTTCAcggaccgaccagctatacaagta	15–362	2,783–3,117	1	7.7
Lm2.VGcb4 (1)	CTTTAATCTCAATCAATCAAATCgcaaacaggaaatggtgctac	6–180	1,549–2,654	15	8.6
Lm2.VGcb5 (85)	ATACTACATCATAATCAAACATCAcacaagcagtagcagaactccta	84–450	4,210–5,031	4	9.4
Lm2.VGcbx (7)	CAATTCATTTACCAATTTACCAATgcaagcagcagatagcgaaga	N/A	1,241–3,935	125	N/A
Lm3-4.INLa1 (76)	AATCTAACAAACTCATCTAAATACgcaccaccaacaactggagga	74–477	3,026–3,861	2	6.3
Lm3-4.INLa2 (19)	TCAATCAATTACTTACTCAAATACgctcaatatagcccaatgctt	33–199	2,294–2,484	1	11.5

Continued on following page

TABLE 5—Continued

Probe ^b	Probe sequence ^c	Adjusted MFI (range)		Target N ^d	ID ^e
		Nontarget	Target		
Lm3-4.INLa3 (74)	TACACATCTTACAAACTAATTTCAgcaacgttgataatgacggtt	55–252	2,932–3,403	1	11.6
Lm3-4.INLa4 (88)	TTACTTCACTTTCTATTTACAATCgcaaccttgatggtgatggg	357–683	5,316–8,397	15	7.8
Lm3-4.INLa5 (95)	TACTTTAAACTTACTACTACTAAgcaaccttgatggtgatgga	458–1,026	2,506–4,353	33	2.4
Lm3-4.INLa6 (94)	CTTTCTATCTTTCTACTCAATAATgacatcgatttatatgcgct	6–122	3,325–3,468	1	27.4
Lm3-4.INLa7 (43)	CTTCAATTACAATACTACTTACATAaatatcgcttagaactatcc	1–108	1,802–1,865	1	16.8
Lm3-4.INLa8 (68)	TCATAATCTCAACAATCTTTCTTTgcccctaacaggtctaact	45–217	2,217–2,333	1	10.2
Lm3-4.INLa9 (51)	TCATTTCAATCAATCATCAACAATcagctacacagcaacgttta	18–150	1,402–1,521	1	9.4
Lm3-4.INLax (12)	TACACTTTCTTTCTTTCTTTCTTTgtagatttagtgcgtttcagg	N/A ^a	2,634–5,197	48	N/A
Lm3-4.INLb1 (17)	CTTTAATCCTTTATCACTTTATCAgcaacgtttatcggtgagat	153–481	3,418–4,144	1	7.1
Lm3-4.INLb2 (29)	AATCTTACTACAATCCTTTCTTTcttcaacatcacagcgct	–37 to 129	9,517–10,444	1	73.7
Lm3-4.INLb3 (32)	ATTATTCACCTTCAAATAATCTACgaaatgacaaaggtgttact	42–211	2,417–2,589	1	11.5
Lm3-4.INLb4 (72)	TCATTTACCTTTAATCCAATAATCcaattgtgaaacaagcttct	27–144	3,308–3,515	1	23.1
Lm3-4.INLb5 (6)	TCAACAATCTTTACAATCAAATCgaggcaggagcgcgaataat	179–445	2,074–2,270	1	4.7
Lm3-4.INLb6 (37)	CTTTTCATCTTTTCACTTTTCAATggaatcaacatattacc	–5 to 97	3,654–3,740	1	37.7
Lm3-4.INLb7 (62)	TCAATCATAATCTCATAATCCAATctcgcaccgctgtaaagctc	24–163	2,506–2,515	1	15.3
Lm3-4.INLb8 (73)	ATCAAACTCTCATCAATCAACAATaaccggggcgaagtaacaagcc	49–177	3,231–3,515	1	18.2
Lm3-4.INLbx (100)	CTATCTTTAAACTACAATCTAACTatgtagaataatgctgcaagcc	N/A	2,051–3,476	48	N/A
Lm3-4.LMO1 (20)	CTTTTACAATACTTCAATACAATACTcattgccaatcacagccacc	28–540	4,538–6,339	5	8.4
Lm3-4.LMO2 (2)	CTTTTACAATACTACTACAATCAgcccgaacatcacaccagct	56–989	4,372–9,565	9	4.4
Lm3-4.LMOx (59)	TCATCAATCAATCTTTTCACTTTcgattgaaacatctgtgaaagcc	N/A	2,238–6,590	48	N/A
Lm3-4.VGCa1 (3)	TACACTTTATCAAATCTTACAATCcaaggatgtagaataaagc	62–277	3,464–3,693	1	12.5
Lm3-4.VGCa2 (99)	AATCTTACATAACAATTTCAATAACgccaatcaatccaggtgca	33–178	2,823–3,013	1	15.9
Lm3-4.VGCa3 (91)	TTCATAACATCAATCATAACTTACctttatccgaaatagataagc	17–331	2,417–2,718	1	7.3
Lm3-4.VGCa4 (70)	ATACCAATAATCCAATTCATATCAgcatatcccaagtttaagt	83–227	4,419–5,184	1	19.4
Lm3-4.VGCa5 (48)	AAACAACTTCACATCTCAATAATgtaaaagaatgctgcttga	32–414	4,849–5,369	2	11.7
Lm3-4.VGCa6 (64)	TACATATTCAAATACTACTTACcattgatgaccggaactaccg	42–554	4,879–6,285	3	8.8
Lm3-4.VGCa7 (42)	CTATCTTCATATTTCACTATAAAcagccggaactaccactgtga	52–309	3,452–3,651	1	11.2
Lm3-4.VGCa8 (49)	TCATCAATCTTTCAATTTACTTACgaggtcccattgtctatacaacc	98–262	5,326–5,495	1	20.4
Lm3-4.VGCa9 (26)	TTACTCAAAATCTACACTTTTTCAGgaggtcagttgctcaattcaat	620–1,652	9,110–9,702	1	5.5
Lm3-4.VGCax (28)	CTACAAACAACAACATTTCAATCAAgatataaccagggatgagtgagc	N/A	498–1,304	48	N/A
Lm3-4.VGcb1 (77)	CAATTAACATACATAACAATACATAccaatcccaacagaagaaggg	54–912	8,267–9,471	1	9.1
Lm3-4.VGcb2 (14)	CTACTATACATCTTACTATACCTTctgctatccaggatgccaatca	30–968	7,661–8,679	1	7.9
Lm3-4.VGcb3 (57)	CAATTCATCATCTTTATCATTACgcttgaagccttacttactctgt	16–383	2,970–3,421	1	7.8
Lm3-4.VGcb4 (35)	CAATTCATCATCTTACTTACTTACggtttactgacgtccggttct	5–139	2,991–3,505	3	21.5
Lm3-4.VGcb5 (44)	TCATTTACCAATCTTTCTTTTATACggtgtgttctctttaggggct	97–336	2,475–2,997	2	7.4
Lm3-4.VGcb6 (18)	TCAAAATCTCAAATACTCAAATCAgcaaaataagccaccggtt	–10 to 147	2,658–3,136	1	18.1
Lm3-4.VGcb7 (97)	AATCTCATAAATCTACATACACTATcaattgtgataaaagtgcaggga	134–504	1,859–2,196	2	3.7
Lm3-4.VGcb8 (30)	TTACCTTTTATACCTTTCTTTTTCatgcttcggactttccacct	59–1,071	10,699–11,430	1	10.0
Lm3-4.VGcb9 (24)	TCAATFACCTTTTCAATACAATAACgcatgcccagcctaccat	89–459	4,224–4,322	1	9.2
Lm3-4.VGcb10 (5)	CAATTCAAATCACAATAATCAATCgagcagccaagcgaggtaaac	66–255	1,415–1,507	1	5.6
Lm3-4.VGcb11 (16)	AATCAATCTTTCATTCAAATCATCAagcgaggtaaacacgggaccg	–117 to 199	4,627–5,741	3	23.3
Lm3-4.VGcb12 (46)	TACATCAACAATTCATTCAATACTactagctgatttaagagatag	275–561	5,167–5,742	3	9.2
Lm3-4.VGcb13 (80)	CTAACTAACAATAATCAATAACgcaagaactgagaatgcccggc	12–182	6,160–7,025	1	33.9
Lm3-4.VGcb14 (60)	AATCTACAATCCAATAATCTCATcttctcatcacgaaagcagc	22–208	2,986–3,392	1	14.3
Lm3-4.VGcb15 (33)	TCAATFACTTCACTTTAATCCTTTtagactgtcttgcagagagacc	97–612	5,032–5,259	1	8.2
Lm3-4.VGcb16 (96)	ATACTAACTCAACTTAACTTAAACcataatattgcaacgacagataa	37–195	2,375–2,671	1	12.2
Lm3-4.VGcb17 (55)	TATATACTTCTCAATAACTAAACcagacgagatggaagaaggg	103–315	4,468–4,926	1	14.2
Lm3-4.VGcb18 (86)	CTAATFACTAACATCACTAACAATctggtataaaccaaaaagcaac	7–148	3,477–3,936	1	23.6
Lm3-4.VGcbx (50)	CAATATACCAATATCATCTTTACgacttagattctagcatgacgtc	N/A	1,484–4,209	48	N/A

^a N/A, not applicable for positive control probes.

^b The 64 probes used in the lineage II assay are prefaced with Lm2, while the 51 probes used in the lineage III and IV assay are prefaced with Lm3-4. Luminex microsphere sets (Luminex Corporation) used for hybridization reactions are indicated in parentheses.

^c The 5' sequence tag portions of extension probes are capitalized.

^d Target sample size among the 125 lineage II and 48 lineage III and IV strains sequenced.

^e ID, index of discrimination.

to accurately characterize the genomic background of these strains.

Phylogenetic trees inferred via analyses of genotype data from the lineage II and the lineage III and IV MLGT assays (not shown) were largely congruent with phylogenies based on multilocus DNA sequence data (Fig. 1 and 2). Interior branches in the MLGT trees were not strongly supported by bootstrap analysis, and additional probe design may be re-

quired to improve phylogenetic resolution. However, because there is a direct connection between MLGT subtypes and the underlying DNA sequence data, MLGT data can be mapped onto the multilocus DNA sequence tree (Fig. 1), allowing the phylogenetic information provided by the large comparative sequence databases to be integrated with MLGT results.

Subtype prevalence in RTE food. Of the 173 lineage II, III, and IV isolates used in the design and validation of the MLGT

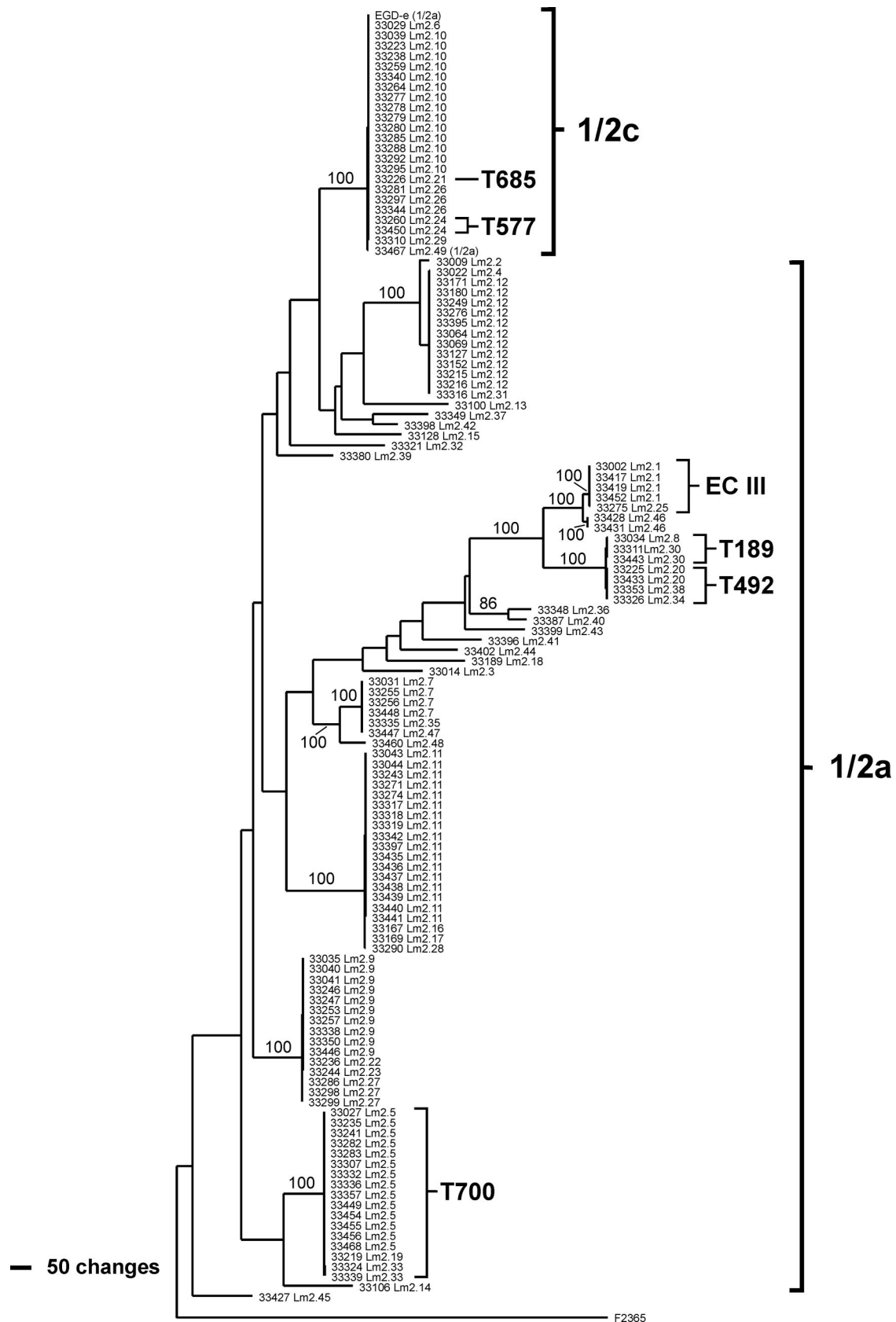


FIG. 1. One of 20 most parsimonious trees inferred from analysis of the combined sequence data for the 125 lineage II isolates used in the SNP discovery (Table 1). Sequence data for strain EGD-e (NC_003210) were included for comparison, and the tree was rooted with sequence data from the lineage I strain F2365 (NC_002973). The frequency (percentage) with which a given branch was recovered in 2,000 bootstrap replications is shown above branches recovered in more than 70% of bootstrap replicates. With the exception of F2365 and EGD-e, strains are identified by NRRL-B numbers followed by MLGT subtype designations. Similar results were obtained with a neighbor-joining analysis.

TABLE 6. Mutations leading to truncated forms of InlA among 125 sequenced isolates from *L. monocytogenes* lineage II

Mutation ^a	Equivalent ^b	MLGT probe(s) ^c	MLGT haplotype(s)	RTE (%) ^d
T189	H1	Lm2.INLa8, Lm2.INLa13	Lm2.8, Lm2.30	3.9
T492		Lm2.INLa9, Lm2.INLa14	Lm2.20, Lm2.34, Lm2.38	3.9
T577	LO28	Lm2.INLa11, Lm2.INLa15	Lm2.24	2.6
T685	NV5	Lm2.INLa10	Lm2.21	0.0
T700	PMSC3	Lm2.INLa7, Lm2.INLa12	Lm2.5, Lm2.19, Lm2.33	20.8

^a Mutation nomenclature reflects the amino acid position of premature stop codons relative to the 800-amino-acid protein predicted for strain EGD-e (NC_003210).

^b Strain names or previous designations for equivalent mutations described by Jonquieres et al. (19), Olier et al. (29), Rousseaux et al. (39), and Nightingale et al. (26).

^c Four of the five truncation mutations were targeted by pairs of reciprocal probes, one of which was specific to the nucleotide character state responsible for the truncation, while the other probe was specific to the alternate form of the *inlA* allele. After multiple attempts, only a single acceptable probe was identified for the T685 mutation.

^d Frequency of InlA truncation mutations among 77 lineage II isolates collected by USDA-FSIS from RTE products and processing facilities.

assays (Table 1), 94 were collected by the USDA Food Safety and Inspection Service (FSIS) from RTE foods and food-processing facilities. We excluded isolates collected on the same day from the same state, as well as isolates lacking information on date of isolation in order to minimize duplicate sampling. This produced a set of 78 FSIS isolates with which we evaluated subtype frequencies (see Table S2 in the supplemental material). Only one lineage III isolate was observed in this panel, while the remaining 77 isolates were identified as lineage II. Thirty unique MLGT haplotypes were observed among these 77 lineage II isolates, indicating substantial diversity within the FSIS panel. Of these, 21 MLGT types from 59 (76.6%) lineage II isolates corresponded to serogroup 1/2a, and 5 MLGT types from 18 (23.4%) isolates corresponded to serogroup 1/2c. The FSIS isolate NRRL B-33452 had the same MLGT type (Lm2.1) as the strain responsible for the multi-state outbreak of listeriosis in 2000. This was the only isolate with an ECIII-associated MLGT type among the FSIS isolate panel. Combining these results with data used to determine the frequency of lineage I epidemic clones (5) indicates that strains from epidemic clone groups were rare (6.3% of 144 isolates) in RTE food products and processing facilities sampled by FSIS.

Conversely, MLGT analysis demonstrated that strains with mutations leading to truncated forms of the key virulence determinant, InlA, comprised at least 31.2% of the lineage II isolates recovered from RTE food and food-processing facilities monitored by FSIS. The lineage II MLGT contained probes for each of the InlA truncation mutations identified among the sequenced isolates, and we observed four of these five mutation types among the FSIS isolate panel (Table 6). The most common truncation mutation (T700) was identified in 16 (20.8%) of the FSIS isolates, while the other truncation types were observed at less than 4% frequency (Table 6). The results for T700 are consistent with a previous estimate of the frequency (11.2%) of ribotype patterns associated with this mutation among food isolates (26). However, frequency estimates for the other lineage II truncation types among food isolates have not been previously investigated. Despite the fact

that each of the InlA truncation mutations examined had a single and recent evolutionary origin (Fig. 1), three of the five InlA truncation types in Table 6 were differentiated into multiple MLGT types (Table 6), and at least two mutation types are associated with multiple serotypes from the same major serogroup (Table 1). Therefore, it is clear that some level of genetic and antigenic divergence or exchange has occurred since these truncation mutations were introduced into their respective clonal lineages.

Combining information from the FSIS panel with previously published data from a similar panel of lineage I isolates (5), we estimate that at least 30% of all *L. monocytogenes* isolates obtained from FSIS surveillance of RTE foods and food-processing facilities carry mutations leading to truncated forms of InlA. In addition, this is likely to represent a conservative estimate of the InlA truncation frequency among food isolates because it is based only on the mutations that were observed in our SNP discovery. These results support and extend previous hypotheses that a substantial fraction of *L. monocytogenes* isolates from foods have reduced abilities to cause systemic listeriosis (5, 26). However, this does not imply that strains with InlA truncation mutations pose no threat to public health. While rare, such strains have been obtained from human and animal listeriosis cases (13, 26, 27), and different InlA truncation types have been associated with different levels of virulence attenuation in animal models (29, 30). In addition, the precise contribution of InlA truncations to attenuated virulence phenotypes needs to be more conclusively defined (5, 26, 28).

The fact that multiple independent mutations leading to InlA truncations have been identified within lineages I and II led to the suggestion that positive selection has driven the loss of functional or cell wall-anchored InlA, particularly among isolates from food environments (26, 32). However, no specific adaptive advantage responsible for the hypothesized positive selection has been identified (32). In addition, the observations regarding InlA truncation frequencies among food isolates could result from food and food-processing facilities providing permissive environments in which there should be little or no selective constraint to eliminate mutations that disrupt the function or cell wall association of an invasion protein such as InlA. Ecological success of *inlA* alleles carrying truncation mutations could be indirect evidence of an adaptive advantage (26). However, many of the InlA truncation mutations were found at low frequencies in surveys of food isolates (Table 6) (5, 26), and it is not clear if the mutations leading to truncated forms of InlA represent the cause or the effect of successful colonization of food-processing environments. This is not to suggest that the positive selection hypothesis should be rejected but highlights the need to consider environment-specific relaxation of constraints as an alternative explanation.

Regardless of the nature of selection acting on *inlA* alleles harboring truncation mutations, there is clearly an association between subtype prevalence in food environments and the subtype distribution of InlA truncation mutations. The InlA truncation mutations identified to date are restricted to serogroups 1/2a and 1/2c from lineage II and serogroup 1/2b from lineage I (5, 19, 26, 29, 32, 39). These are the most common subtypes isolated from foods and food-processing environments (5, 13, 46). As such, it is reasonable to expect that

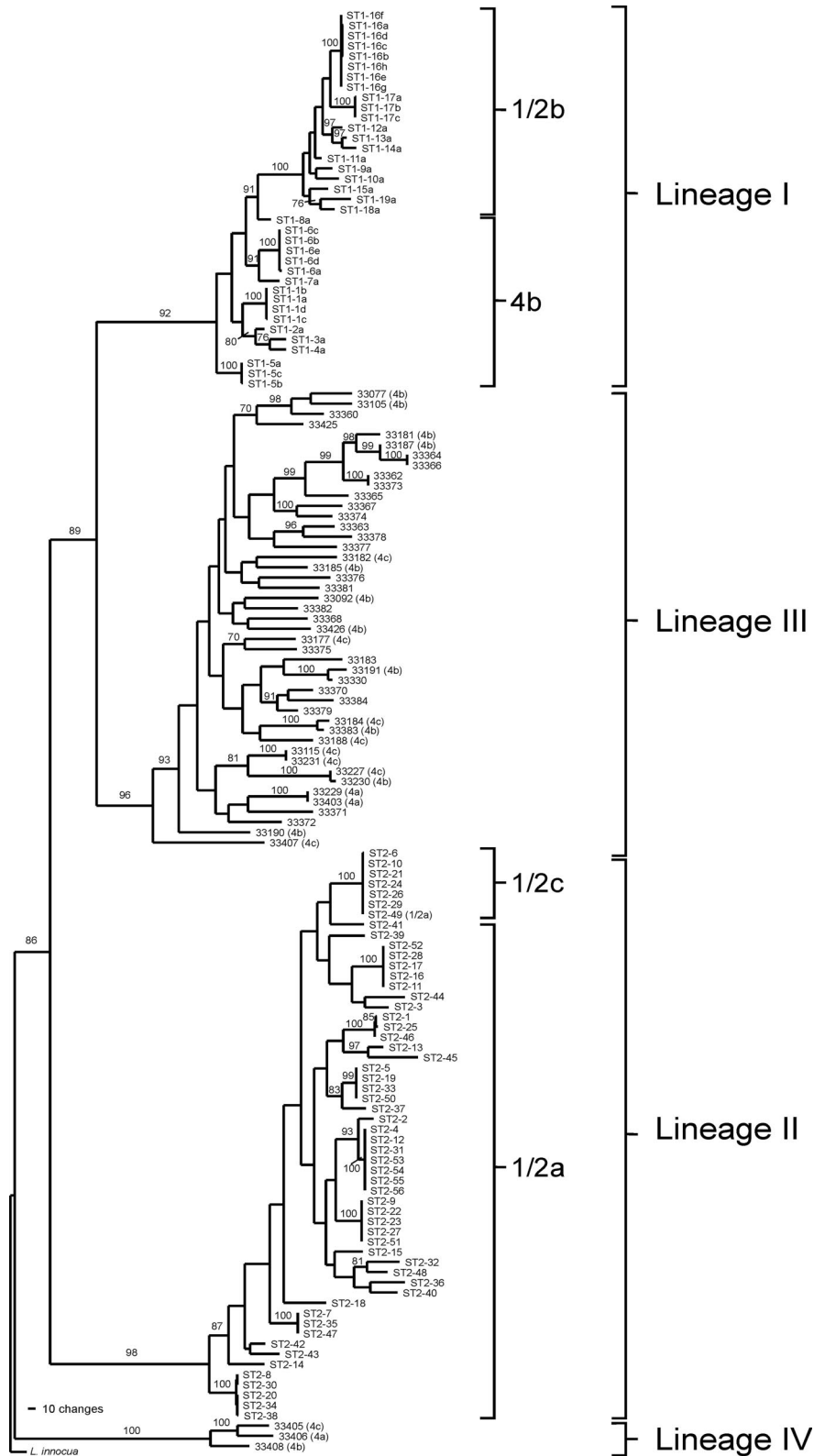


FIG. 2. One of four most parsimonious trees inferred from analysis of sequences from the LMO, VGC, and INL regions (Table 3). The analysis included sequences from all 48 lineage III and IV isolates examined (identified by NRRL-B numbers) and unique sequence types identified in the SNP discovery for lineages I (5) and II (Table 1). The LMO sequence from *Listeria innocua* strain Clip11262 (NC_003212) was used to root the tree. The frequency (percentage) with which a given branch was recovered in 2,000 bootstrap replications is shown above branches recovered in more than 70% of bootstrap replicates. Similar results were obtained with neighbor-joining analysis.

variation within these groups is shaped to a greater extent by selective constraints operating in food environments than is the case for strains from serogroup 4b of lineage I, or lineage III strains, both of which are relatively rare in food environments (5, 13, 41, 46). The relative rarity of serogroup 4b (lineage I) and lineage III strains in food environments could explain the absence of *InlA* truncation mutations in these groups, which in turn could contribute directly to their overrepresentation in human and animal listeriosis cases, respectively.

Comparative subtype analysis. The discriminatory power of the MLGT assay for lineage II was compared to that of PFGE, the six-gene multilocus sequence typing (MLST) approach described by Revazishvili et al. (37), and the multi-virulence locus sequence typing (MVLST) approach described by Zhang et al. (50). In an analysis of 20 sporadic listeriosis isolates provided by the CDC (Table 2), the lineage II MLGT provided the greatest discriminatory power (12 haplotypes; SDI, 0.93) among the DNA sequence-based approaches. SDI values are scored between 0 and 1, and SDI values above 0.9 are considered desirable for reliable subtyping (15). MVLST utilized 2,750 bp of DNA sequence from each of the 20 isolates and differentiated 10 sequence types (SDI, 0.88). MLST based on 4,322 bp of DNA sequence from each isolate identified 11 sequence types (SDI, 0.91) but utilized amplicons that were too large to be sequenced using the amplification primers, necessitating the design and use of several internal sequencing primers. In addition, the *pgm* primers used in the MLST produced small nontarget amplicons that prevented us from sequencing with the amplification primers, and the region of *actA* targeted by MLST included a 105-bp indel polymorphism that could make it impossible to assess positional homology.

As was the case in our previous analysis of lineage I isolates (5), PFGE provided greater strain-discriminating power (17 types; SDI, 0.98) than the DNA sequence-based approaches (Table 2). Combining data from MLGT, MLST, and MVLST provided a modest increase in discriminatory power (13 types; SDI, 0.94) above that provided by MLGT alone but still did not discriminate all of the strains that had unique PFGE patterns. Similarly, PFGE provided greater discriminatory power than a recently published method based on multiple-locus variable-number tandem repeat analysis (MLVA) (42). This suggests that there may be practical limitations to the development of locus-specific methods for DNA sequence-based subtyping of *L. monocytogenes* that achieve the level of discrimination provided by two-enzyme PFGE. However, DNA sequence-based methods offer solutions to many of the problems and limitations of PFGE described above, which make them attractive options for complementing or replacing PFGE as the primary subtyping tool for *L. monocytogenes* (5, 11, 16, 37, 43, 50). In addition, while PFGE has been reported to have the highest discriminatory power, it does not always discriminate between related but distinct strains (5, 40, 45). In our comparative analyses of typing methods, only the MLGT method differentiated strains with identical PFGE patterns. NRRL B-33575 and NRRL B-33578 shared identical PFGE, MLST, and MVLST patterns but were differentiated by MLGT because NRRL B-33578 harbors the T577 mutation in *inlA* that is associated with a weak attenuation of virulence in animal models (29, 30). This result demonstrates the potential for MLGT to complement PFGE data in strain discrimination and high-

lights the fact that strains with identical PFGE patterns can harbor differences in key virulence genes.

Combined with our previously developed MLGT assay for subtyping lineage I isolates (5), the MLGT assays described here provide a highly flexible and efficient system for DNA sequence-based subtyping of *L. monocytogenes*. These single-well assays provide high-throughput subtyping via direct interrogation of SNPs from multiple regions of the genome and produce data that are unambiguous, easily shared between laboratories, and amenable to phylogenetic analyses and risk assessment. Similar assays for detection and subtyping of food-borne pathogens have been described (6, 9) or are under development (16), indicating that SNP-based subtyping may offer a common approach for future subtyping of food-borne and other pathogens.

ACKNOWLEDGMENTS

We are indebted to Lewis Graves and the Centers for Disease Control and Prevention for providing clinical isolates and PFGE patterns and to Kitty Pupedis and the USDA Food Safety and Inspection Service for providing isolates. We thank Jennifer Steele for excellent technical assistance. We also thank Brent Page and Peter Evans for helpful advice and discussions.

The mention of firm names or trade products does not imply that they are endorsed or recommended by the U.S. Department of Agriculture over other firms or similar products not mentioned.

REFERENCES

- Borucki, M. K., S. H. Kim, D. R. Call, S. C. Smole, and F. Pagotto. 2004. Selective discrimination of *Listeria monocytogenes* epidemic strains by a mixed-genome DNA microarray compared to discrimination by pulsed-field gel electrophoresis, ribotyping, and multilocus sequence typing. *J. Clin. Microbiol.* **42**:5270–5276.
- Doumith, M., C. Buchrieser, P. Glaser, C. Jacquet, and P. Martin. 2004. Differentiation of the major *Listeria monocytogenes* serovars by multiplex PCR. *J. Clin. Microbiol.* **42**:3819–3822.
- Doumith, M., C. Cazalet, N. Simoes, L. Frangeul, C. Jacquet, F. Kunst, P. Martin, P. Cossart, P. Glaser, and C. Buchrieser. 2004. New aspects regarding evolution and virulence of *Listeria monocytogenes* revealed by comparative genomics and DNA arrays. *Infect. Immun.* **72**:1072–1083.
- Doumith, M., C. Jacquet, V. Goulet, C. Oggioni, F. Van Loock, C. Buchrieser, and P. Martin. 2006. Use of DNA arrays for the analysis of outbreak-related strains of *Listeria monocytogenes*. *Int. J. Med. Microbiol.* **296**:559–562.
- Ducey, T. F., B. Page, T. Usgaard, M. K. Borucki, K. Pupedis, and T. J. Ward. 2007. A single-nucleotide-polymorphism-based multilocus genotyping assay for subtyping lineage I isolates of *Listeria monocytogenes*. *Appl. Environ. Microbiol.* **73**:133–147.
- Dunbar, S. A., and J. W. Jacobson. 2007. Quantitative, multiplexed detection of *Salmonella* and other pathogens by Luminex xMAP suspension array. *Methods Mol. Biol.* **394**:1–19.
- Farber, J. M., and P. I. Peterkin. 1991. *Listeria monocytogenes*, a food-borne pathogen. *Microbiol. Rev.* **55**:476–511.
- Felsenstein, J. 1985. Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* **39**:783–791.
- Fitzgerald, C., M. Collins, S. van Duynne, M. Mikoleit, T. Brown, and P. Fields. 2007. Multiplex, bead-based suspension array for molecular determination of common *Salmonella* serogroups. *J. Clin. Microbiol.* **45**:3323–3334.
- Fugett, E., E. Fortes, C. Noka, and M. Wiedmann. 2006. International Life Sciences Institute North America *Listeria monocytogenes* strain collection: development of standard *Listeria monocytogenes* strain sets for research and validation studies. *J. Food Prot.* **69**:2929–2938.
- Gerner-Smidt, P., K. Hise, J. Kincaid, S. Hunter, S. Rolando, E. Hyttia-Trees, E. M. Ribot, and B. Swaminathan. 2006. PulseNet USA: a five-year update. *Foodborne Pathog. Dis.* **3**:9–19.
- Gravesen, A., T. Jacobsen, P. L. Moller, F. Hansen, A. G. Larsen, and S. Knochel. 2000. Genotyping of *Listeria monocytogenes*: comparison of RAPD, ITS, and PFGE. *Int. J. Food Microbiol.* **57**:43–51.
- Gray, M. J., R. N. Zadoks, E. D. Fortes, B. Dogan, S. Cai, Y. Chen, V. N. Scott, D. E. Gombas, K. J. Boor, and M. Wiedmann. 2004. *Listeria monocytogenes* isolates from foods and humans form distinct but overlapping populations. *Appl. Environ. Microbiol.* **70**:5833–5841.
- Hamon, M., H. Bierne, and P. Cossart. 2006. *Listeria monocytogenes*: a multifaceted model. *Nat. Rev. Microbiol.* **4**:423–434.

15. Hunter, P. R., and M. A. Gaston. 1988. Numerical index of the discriminatory ability of typing systems: an application of Simpson's index of diversity. *J. Clin. Microbiol.* **26**:2465–2466.
16. Hyttiä-Trees, E. K., K. Cooper, E. M. Ribot, and P. Gerner-Smidt. 2007. Recent developments and future prospects in subtyping of foodborne bacterial pathogens. *Future Microbiol.* **2**:175–185.
17. Jacquet, C., M. Doumith, J. I. Gordon, P. M. Martin, P. Cossart, and M. Lecuit. 2004. A molecular marker for evaluating the pathogenic potential of foodborne *Listeria monocytogenes*. *J. Infect. Dis.* **189**:2094–2100.
18. Jeffers, G. T., J. L. Bruce, P. L. McDonough, J. Scarlett, K. J. Boor, and M. Wiedmann. 2001. Comparative genetic characterization of *Listeria monocytogenes* isolates from human and animal listeriosis cases. *Microbiology* **147**:1095–1104.
19. Jonquieres, R., H. Bierre, J. Mengaud, and P. Cossart. 1998. The *inlA* gene of *Listeria monocytogenes* LO28 harbors a nonsense mutation resulting in release of internalin. *Infect. Immun.* **66**:3420–3422.
20. Kathariou, S. 2003. Foodborne outbreaks of listeriosis and epidemic-associated lineages of *Listeria monocytogenes*. Iowa State University Press, Ames.
21. Lecuit, M., S. Vandormael-Pournin, J. Lefort, M. Huerre, P. Gounon, C. Dupuy, C. Babinet, and P. Cossart. 2001. A transgenic model for listeriosis: role of internalin in crossing the intestinal barrier. *Science* **292**:1722–1725.
22. Liu, D., M. L. Lawrence, M. Wiedmann, L. Gorski, R. E. Mandrell, A. J. Ainsworth, and F. W. Austin. 2006. *Listeria monocytogenes* subgroups IIIA, IIIB, and IIIC delineate genetically distinct populations with varied pathogenic potential. *J. Clin. Microbiol.* **44**:4229–4233.
23. McClure, P. J., T. M. Kelly, and T. A. Roberts. 1991. The effects of temperature, pH, sodium chloride and sodium nitrite on the growth of *Listeria monocytogenes*. *Int. J. Food Microbiol.* **14**:77–91.
24. Mead, P. S., L. Slutsker, V. Dietz, L. F. McCaig, J. S. Bresee, C. Shapiro, P. M. Griffin, and R. V. Tauxe. 1999. Food-related illness and death in the United States. *Emerg. Infect. Dis.* **5**:607–625.
25. Nightingale, K. K., R. A. Ivy, A. J. Ho, E. D. Fortes, B. L. Njaa, R. M. Peters, and M. Wiedmann. 2008. *inlA* premature stop codons are common among *Listeria monocytogenes* isolated from foods and yield virulence-attenuated strains that confer protection against infection by fully virulent *L. monocytogenes* strains. *Appl. Environ. Microbiol.* **74**:6570–6583.
26. Nightingale, K. K., K. Windham, K. E. Martin, M. Yeung, and M. Wiedmann. 2005. Select *Listeria monocytogenes* subtypes commonly found in foods carry distinct nonsense mutations in *inlA*, leading to expression of truncated and secreted internalin A, and are associated with a reduced invasion phenotype for human intestinal epithelial cells. *Appl. Environ. Microbiol.* **71**:8764–8772.
27. Nightingale, K. K., K. Windham, and M. Wiedmann. 2005. Evolution and molecular phylogeny of *Listeria monocytogenes* isolated from human and animal listeriosis cases and foods. *J. Bacteriol.* **187**:5537–5551.
28. Olier, M., D. Garmyn, S. Rousseaux, J. P. Lemaitre, P. Piveteau, and J. Guzzo. 2005. Truncated internalin A and asymptomatic *Listeria monocytogenes* carriage: in vivo investigation by allelic exchange. *Infect. Immun.* **73**:644–648.
29. Olier, M., F. Pierre, J. P. Lemaitre, C. Divies, A. Rousset, and J. Guzzo. 2002. Assessment of the pathogenic potential of two *Listeria monocytogenes* human faecal carriage isolates. *Microbiology* **148**:1855–1862.
30. Olier, M., F. Pierre, S. Rousseaux, J. P. Lemaitre, A. Rousset, P. Piveteau, and J. Guzzo. 2003. Expression of truncated internalin A is involved in impaired internalization of some *Listeria monocytogenes* isolates carried asymptotically by humans. *Infect. Immun.* **71**:1217–1224.
31. Olsen, S. J., M. Patrick, S. B. Hunter, V. Reddy, L. Kornstein, W. R. MacKenzie, K. Lane, S. Bidol, G. A. Stoltman, D. M. Frye, I. Lee, S. Hurd, T. F. Jones, T. N. LaPorte, W. Dewitt, L. Graves, M. Wiedmann, D. J. Schoonmaker-Bopp, A. J. Huang, C. Vincent, A. Bugenhagen, J. Corby, E. R. Carloni, M. E. Holcomb, R. F. Woron, S. M. Zansky, G. Dowdle, F. Smith, S. Ahrabi-Fard, A. R. Ong, N. Tucker, N. A. Hynes, and P. Mead. 2005. Multistate outbreak of *Listeria monocytogenes* infection linked to delicatessen turkey meat. *Clin. Infect. Dis.* **40**:962–967.
32. Orsi, R. H., D. R. Ripoll, M. Yeung, K. K. Nightingale, and M. Wiedmann. 2007. Recombination and positive selection contribute to evolution of *Listeria monocytogenes* *inlA*. *Microbiology* **153**:2666–2678.
33. Orsi, R. H., Q. Sun, and M. Wiedmann. 2008. Genome-wide analyses reveal lineage specific contributions of positive selection and recombination to the evolution of *Listeria monocytogenes*. *BMC Evol. Biol.* **8**:233.
34. Palumbo, J. D., M. K. Borucki, R. E. Mandrell, and L. Gorski. 2003. Serotyping of *Listeria monocytogenes* by enzyme-linked immunosorbent assay and identification of mixed-serotype cultures by colony immunoblotting. *J. Clin. Microbiol.* **41**:564–571.
35. Penny, D., and M. D. Hendy. 1985. The use of tree comparison metrics. *Syst. Zool.* **34**:75–82.
36. Rasmussen, O. F., P. Skouboe, L. Dons, L. Rossen, and J. E. Olsen. 1995. *Listeria monocytogenes* exists in at least three evolutionary lines: evidence from flagellin, invasive associated protein and listeriolysin O genes. *Microbiology* **141**:2053–2061.
37. Revazishvili, T., M. Kotetishvili, O. C. Stine, A. S. Kreger, J. G. Morris, Jr., and A. Sulakvelidze. 2004. Comparative analysis of multilocus sequence typing and pulsed-field gel electrophoresis for characterizing *Listeria monocytogenes* strains isolated from environmental and clinical sources. *J. Clin. Microbiol.* **42**:276–285.
38. Roberts, A., K. Nightingale, G. Jeffers, E. Fortes, J. M. Kongo, and M. Wiedmann. 2006. Genetic and phenotypic characterization of *Listeria monocytogenes* lineage III. *Microbiology* **152**:685–693.
39. Rousseaux, S., M. Olier, J. P. Lemaitre, P. Piveteau, and J. Guzzo. 2004. Use of PCR-restriction fragment length polymorphism of *inlA* for rapid screening of *Listeria monocytogenes* strains deficient in the ability to invade Caco-2 cells. *Appl. Environ. Microbiol.* **70**:2180–2185.
40. Sauters, B. D., E. D. Fortes, D. L. Morse, N. Dumas, J. A. Kiehlbauch, Y. Schukken, J. R. Hibbs, and M. Wiedmann. 2003. Molecular subtyping to detect human listeriosis clusters. *Emerg. Infect. Dis.* **9**:672–680.
41. Shen, Y., Y. Liu, Y. Zhang, J. Cripe, W. Conway, J. Meng, G. Hall, and A. A. Bhagwat. 2006. Isolation and characterization of *Listeria monocytogenes* isolates from ready-to-eat foods in Florida. *Appl. Environ. Microbiol.* **72**:5073–5076.
42. Sperry, K. E., S. Kathariou, J. S. Edwards, and L. A. Wolf. 2008. Multiple-locus variable number tandem repeat analysis as a subtyping tool for *Listeria monocytogenes*. *J. Clin. Microbiol.* **46**:1435–1450.
43. Swaminathan, B., T. J. Barrett, S. B. Hunter, and R. V. Tauxe. 2001. PulseNet: the molecular subtyping network for foodborne bacterial disease surveillance, United States. *Emerg. Infect. Dis.* **7**:382–389.
44. Swaminathan, B., and P. Gerner-Smidt. 2007. The epidemiology of human listeriosis. *Microbes Infect.* **9**:1236–1243.
45. Tenover, F. C., R. D. Arbeit, R. V. Goering, P. A. Mickelsen, B. E. Murray, D. H. Persing, and B. Swaminathan. 1995. Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing. *J. Clin. Microbiol.* **33**:2233–2239.
46. Ward, T. J., L. Gorski, M. K. Borucki, R. E. Mandrell, J. Hutchins, and K. Papedis. 2004. Intraspecific phylogeny and lineage group identification based on the *prfA* virulence gene cluster of *Listeria monocytogenes*. *J. Bacteriol.* **186**:4994–5002.
47. Wiedmann, M., J. L. Bruce, C. Keating, A. E. Johnson, P. L. McDonough, and C. A. Batt. 1997. Ribotypes and virulence gene polymorphisms suggest three distinct *Listeria monocytogenes* lineages with differences in pathogenic potential. *Infect. Immun.* **65**:2707–2716.
48. Wong, A. C. 1998. Biofilms in food processing environments. *J. Dairy Sci.* **81**:2765–2770.
49. Xia, X., and Z. Xie. 2001. DAMBE: software package for data analysis in molecular biology and evolution. *J. Hered.* **92**:371–373.
50. Zhang, W., B. M. Jayarao, and S. J. Knabel. 2004. Multivirulence-locus sequence typing of *Listeria monocytogenes*. *Appl. Environ. Microbiol.* **70**:913–920.