IMMUNOLOGY ORIGINAL ARTICLE

# B-cell-activating factor inhibits CD20-mediated and B-cell receptor-mediated apoptosis in human B cells

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### Summary

B-cell-activating factor (BAFF) is a survival and maturation factor for B cells belonging to the tumour necrosis factor superfamily. Among three identified functional receptors, the BAFF receptor (BAFF-R) is thought to be responsible for the effect of BAFF on B cells though details of how remain unclear. We determined that a hairy-cell leukaemia line, MLMA, expressed a relatively high level of BAFF-R and was susceptible to apoptosis mediated by either CD20 or B-cell antigen receptor (BCR). Using MLMA cells as an in vitro model of mature B cells, we found that treatment with BAFF could inhibit apoptosis mediated by both CD20 and BCR. We also observed, using immunoblot analysis and microarray analysis, that BAFF treatment induced activation of nuclear factor-KB2 following elevation of the expression level of Bcl-2, which may be involved in the molecular mechanism of BAFF-mediated inhibition of apoptosis. Interestingly, BAFF treatment was also found to induce the expression of a series of genes, such as that for CD40, related to cell survival, suggesting the involvement of a multiple mechanism in the BAFF-mediated antiapoptotic effect. MLMA cells should provide a model for investigating the molecular basis of the effect of BAFF on B cells in vitro and will help to elucidate how B cells survive in the immune system in which BAFFmediated signalling is involved.

Keywords: apoptosis; B-cell-activating factor; Bcl-2; B-cell receptor; CD20

### Introduction

The immune system comprises a variety of immune effector cells, including T and B lymphocytes and antigenpresenting cells, such as dendritic cells and others; it protects individuals from infections and cancer. To maintain these sophisticated mechanisms, a very subtle balance between the life and death of the immune effector cells must be maintained to eliminate, by apoptosis, potentially harmful self-reactive lymphocytes and only allow the survival, development and activation of safe and protective immune cells. For this purpose, a number of molecules are involved in this regulatory system.<sup>1</sup>

B-cell-activating factor (BAFF, also termed BlyS, TALL-1, THANK and zTNF4) produced by monocytes, dendritic cells and some T cells is a member of the tumour necrosis factor (TNF) superfamily and is a type 2 transmembrane-bound protein that can also be expressed as a soluble ligand.<sup>2</sup> BAFF was first described as a factor that stimulates cell proliferation and the secretion of immunoglobulin in B cells. $3-7$  Transgenic mice that overexpress BAFF in lymphoid tissues exhibited hyperplasia of the mature B-cell compartment. $8-10$  In contrast, mice deficient in BAFF showed a deficit in peripheral B lympho- $\text{cvtes}^{10,11}$  and an almost complete loss of follicular and marginal zone B lymphocytes in secondary lymphoid organs. This suggests an absolute requirement for BAFF in normal B-cell development.<sup>10</sup> In contrast, a later examination of immunized BAFF-null mice validated the BAFF-independent nature of germinal centre formation and that antibody responses, including high-affinity responses, were attenuated, indicating that BAFF is required for maintenance, but not initiation, of the germinal centre reaction.<sup>12</sup> Based on the above evidence, BAFF is considered to be a survival and maturation factor for B lymphocytes and has emerged as a crucial factor that modulates B-cell tolerance and homeostasis. $2,13$  However, the precise role of BAFF in B-cell development is

still controversial and it has been reported that the capacity of B lymphocytes to bind BAFF is correlated with their maturation state and that the effect of BAFF is dependent on the maturation stage of the B lymphocytes. $^{2,14}$ 

Recent studies have further shown that BAFF affects not only B lymphocytes but also T lymphocytes.<sup>15,16</sup> The three distinct receptors for BAFF, namely the BAFF receptor (BAFF-R, also termed BR3), the B-cell maturation antigen (BCMA), and the transmembrane activator and calcium modulator and cyclophilin ligand interactor (TACI), have been identified and BAFF binds with a similar high affinity to these receptors.<sup>7,17–23</sup> Among these receptors, however, BAFF-R is thought to be responsible for the survival and differentiation of B cells,<sup>24</sup> whereas the molecular basis of BAFF-mediated signalling remains unclear.

A number of systems inducing apoptosis in B cells are present to eliminate inappropriate clones, such as selfacting B cells. For example, it is reported that stimulation via particular surface molecules, including B-cell receptor antigen (BCR) and CD20, induces apoptosis in cultured B cells.<sup>25,26</sup> The balance between apoptosis-inducing systems and survival systems, such as CD20 and BAFF-mediated signalling, would be important for the maintenance of appropriate B-cell development, though details are not known.

To elucidate the molecular basis of the interaction between apoptosis-inducing signals and BAFF-mediated cell survival signals in B cells, we have employed a B-cell line that expresses BAFF-R and is sensitive to CD20-mediated and BCR-mediated apoptosis. In this paper, we present evidence that BAFF-mediated stimulation inhibits the apoptosis induced by both CD20-mediated and BCRmediated signalling. The possible mechanisms involved in BAFF-mediated cell responses that regulate these apoptotic stimuli are discussed.

### Materials and methods

#### Cells and reagents

The human hairy cell leukaemia cell line MLMA was obtained from the Japanese Cancer Research Resources Bank (JCRB, Tokyo, Japan). Cells were cultured in RPMI-1640 medium supplemented with 10% fetal calf serum at 37 $\degree$  in a humidified 5% CO<sub>2</sub> atmosphere.

Recombinant human BAFF and a proliferation-inducing ligand (APRIL) were obtained from R&D Systems, Inc. (Minneapolis, MN), and used at a concentration of 400 ng/ml for cell stimulation unless otherwise described. The mouse monoclonal antibodies (mAbs) used for the immunofluorescence analysis were anti-CD10, anti-CD20, anti-CD21, anti-CD22, anti-CD24, anti-CD40, antihuman leucocyte antigen DR (HLA-DR; Beckman Coulter, Inc., Fullerton, CA); anti-CD19 (Becton Dickinson and Company, BD, Franklin Lakes, NJ); anti- $\kappa$ , anti- $\lambda$ ,

anti-u, anti- $\delta$ , anti- $\gamma$  (Dako, Denmark A/S); anti-BAFF-R (Santa Cruz Biotechnology, Santa Cruz, CA); and anti-CD45 (American Type Culture Collection, ATCC, Manassas, VA). The rat mAbs against BCMA (Vicky-1) and TACI (1A1) were purchased from Santa Cruz Biotechnology. The mouse mAbs used for the immunochemical analysis were anti-caspase-2, anti-caspase-3 and anti-glycogen synthase kinase-3b (GSK-3b; Becton Dickinson); anti-caspase-9 (Medical & Biological Laboratories Co., Ltd, Nagoya, Japan); anti-nuclear factor-KB (NF-KB) p52 (C-5), anti-Bcl-2 (100) from Santa Cruz; and anti- $\beta$ -actin (AC-15) from Sigma-Aldrich Co. (St Louis, MO). The rabbit polyclonal antibodies used were anti-cleaved poly ADP-ribose polymerase (PARP), anti-cleaved caspase-3, anti-phospho-GSK-3 $\beta$  (Ser9) and anti-phospho-GSK-3 $\alpha/\beta$ (Ser9, 21) from Cell Signaling Technology, Inc. (Danvers, MA). A goat anti-NF-KB p50 (C-19) from Santa Cruz was also used. Secondary antibodies, including fluorescein isothiocyanate- (FITC) and enzyme-conjugated antibodies, were purchased from either Jackson ImmunoResearch Laboratories, Inc. (West Grove, PA) or Dako. To crosslink BCR, purified anti-u rabbit polyclonal antibody (10 µg/ml) from Jackson ImmunoResearch Laboratories, Inc. was used. To cross-link CD20, a mouse anti-CD20 mAb from Beckman Coulter and a secondary anti-mouse immunoglobulin antibody from Jackson ImmunoResearch Laboratories, Inc. were used each at a concentration of  $5 \text{ µg/ml}.$ 

### Immunofluorescence analysis and detection of apoptosis

Cells were stained with FITC-labelled mAbs and analysed by flow cytometry (EPICS-XL, Beckman Coulter) as described previously.<sup>27</sup> To quantify the incidence of apoptosis, cells were incubated with FITC-labelled annexin V using a MEBCYTO-Apoptosis kit (Medical & Biological Laboratories Co., Ltd) and then analysed by flow cytometry according to the manufacturer's directions. Apoptotic cells were also detected by nuclear-staining with DAPI and examined by confocal microscopy as described previously.<sup>28</sup> The enzymatic activity of caspases -2, -3, -9 was assessed by using a colorimetric protease assay kit for each caspase (Medical & Biological Laboratories Co., Ltd) according to the manufacturer's protocol.

#### Immunoblotting

Immunoblotting was performed as described previously.<sup>29</sup> Briefly, cell lysates were prepared by solubilizing the cells in lysis buffer (containing  $20 \text{ mm}$   $\text{Na}_2\text{PO}_4$ , pH 7.4, 150 m<sup>M</sup> NaCl, 1% Triton X-100, 1% aprotinin, 1 m<sup>M</sup> phenylmethylsulphonylfluoride, 100 mm NaF, and 2 mm Na3VO4), and the total protein concentration was determined using a Bio-Rad protein assay kit (Bio-Rad, Hercules, CA). For each cell lysate, 20 µg was separated by sodium dodecyl sulphate–polyacrylamide gel electrophoresis and transferred to a nitrocellulose membrane using a semidry Transblot system (Bio-Rad). After blocking with 3% skimmed milk in phosphate-buffered saline, the membrane was incubated with the appropriate combination of primary and secondary antibodies as indicated, washed intensively, and examined using the enhanced chemiluminescence reagent system (ECL plus; GE Healthcare Bio-Sciences AB, Uppsala, Sweden).

#### DNA microarray analysis

The DNA microarray analysis was performed using GENE-CHIP (Affymetrix, Santa Clara, CA). Total RNA isolated from MLMA cells treated with and without BAFF for 12 hr was reverse transcribed and labelled using One-Cycle Target Labeling and Control Reagents as instructed by the manufacturer (Affymetrix). The labelled probes were hybridized to Human Genome U133 Plus 2.0 Arrays (Affymetrix). The arrays were analysed using GENECHIP OPERATING Software 1.2 (Affymetrix). Background subtraction and normalization were performed with GENESPRING GX 7.3 software (Agilent Technologies, Santa Clara, CA). Signal intensities were prenormalized based on the median of all measurements on that chip. To account for the difference in detection efficiency between the spots,

prenormalized signal intensities on each gene were normalized to the median of prenormalized measurements for that gene. The data were filtered with the following steps. (1) Genes that were scored as absent in both samples were eliminated. (2) Genes with a signal intensity lower than 90 in both samples were eliminated. (3) Performing cluster analysis using filtering genes, genes were selected that exhibited increased expression or decreased expression in BAFF-treated cells.

### Results

#### Immunophenotypic characterization of MLMA cells

While screening to identify human cell lines expressing BAFF-R, we found that MLMA cells expressed higher levels of BAFF-R than other human B-cell lines. Although the MLMA cell line is known to have been established from a patient with hairy-cell leukaemia, details were not reported. Therefore, we first examined the immunophenotypic characteristics of MLMA cells. Consistent with the JCRB records, flow cytometric analysis revealed that MLMA cells expressed high levels of  $\mu$  heavy chain and low levels of  $\delta$  heavy chain with expression of  $\kappa$  light chain (Fig. 1a). In addition to the CD19 and HLA-DR, MLMA cells were found to express mature B-cell



Figure 1. Immunophenotypic characterization of MLMA cells. (a) MLMA cells were stained with specific fluorescein isothiocyanate (FITC) labelled monoclonal antibodies (mAbs) against B-cell differentiation antigens and analysed by flow cytometry. The x-axis represents fluorescence intensity and the  $\gamma$ -axis the relative cell number; control was isotype-matched mouse immunoglobulin. (b) The expression of B-cell-activating factor receptor (BAFF-R), transmembrane activator and calcium modulator and cyclophilin ligand interactor (TACI), and B-cell maturation antigen (BCMA) on MLMA cells was also examined as in (a).

antigens, including CD20, CD21, CD22 and CD40, but not CD24. Notably, MLMA cells showed the expression of CD10. When the expression of three types of receptors for BAFF was similarly examined, MLMA cells exhibited apparent expression of BAFF-R, while the levels of BCMA and TACI were found to be quite low (Fig. 1b). The data indicate that MLMA cells exhibit immunophenotypic characteristics of mature B cells expressing BAFF-R.

### Cross-linking of BCR and CD20 induces apoptosis in MLMA cells

It has been well documented that cross-linking of BCR using anti- $\mu$  heavy chain antibodies induces apoptosis in some B cells in vitro.<sup>25</sup> Recent studies including our own have also shown that CD20 cross-linking mediates apoptosis in human B-cell lines in a manner involving raftmediated signalling.<sup>26,30</sup> Therefore, we next examined whether cross-linking of either BCR or CD20 mediated apoptosis in MLMA cells. As shown in Fig. 2(a), when anti-µ antibodies were added to the culture, a time-dependent increase in the number of cells bound to annexin V was observed, suggesting the occurrence of

apoptosis in MLMA cells after BCR cross-linking. The apoptosis was confirmed by the morphological appearance of nuclear fragmentation, a typical feature of apoptosis, detected by either Giemsa-staining or nuclearstaining with DAPI (Fig. 2b). Immunoblotting revealed the cleavage of caspases -9, -3 and -2 and of PARP after treatment with anti- $\mu$  antibodies (Fig. 2c), indicating that caspase activation was involved in the apoptosis. In the case of caspase-3, we also detected a 17 000 molecular weight cleaved fragment by using a specific antibody (Fig. 2c). In addition, elevation of the enzymatic activity of each caspase after cross-linking of BCR was detected by a colorimetric protease assay (Fig. 2d). We also examined the effect of anti-CD20 antibodies and found that CD20 cross-linking signalling induced apoptosis in MLMA cells (Fig. 2).

### BAFF inhibits CD20-mediated and BCR-mediated apoptosis in MLMA cells

Next, we examined whether BAFF was able to inhibit apoptosis mediated by cross-linking of CD20 and BCR. As shown in Fig. 3(a), when BAFF was added to the



Figure 2. Induction of apoptosis in MLMA cells mediated by CD20 and B-cell antigen receptor. (a) MLMA cells were treated with either rabbit anti-µ heavy-chain polyclonal antibody (xµ, 10 µg/ml) or a combination of anti-CD20 monoclonal antibody (mAb; xCD20, 5 µg/ml) and secondary rabbit anti-mouse immunoglobulin antibody (RaM, 5 µg/ml) for 48 hr and binding with fluorescein iosthiocyanate (FITC)-conjugated annexin V was examined by flow cytometry. Each experiment was performed in triplicate and the means + SD are indicated. (b) The same sample preparations as in (a) were cytocentrifuged and morphological appearance was examined by Giemsa-staining and nuclear staining with DAPI, using light microscopy and confocal microscopy, respectively. (c) Cell lysates were obtained from the same sample preparation as in (a) and the proforms of each caspase, cleaved caspase-3 and cleaved PARP were detected by immunoblotting.



Figure 3. Effect of B-cell-activating factor (BAFF) on B-cell receptor (BCR)-induced and CD20-induced apoptosis in MLMA cells. (a) MLMA cells were treated with either rabbit anti-u heavy-chain polyclonal antibody (mAb) ( $\alpha\mu$ , 10  $\mu$ g/ml, left panel) or a combination of anti-CD20 mAb ( $\alpha$ CD20, 5 µg/ml) and secondary rabbit anti-mouse immunoglobulin antibody ( $R\alpha M$ , 5  $\mu g/ml$ ) (right panel) for 48 hr in the presence or absence of different concentrations of BAFF as indicated and binding with fluorescein isothiocyanate (FITC) conjugated annexin V was examined as in Fig. 2(a). (b) MLMA cells preincubated with or without 400 ng/ml of BAFF for the indicated periods were treated with either  $\alpha\mu$  (left panel) or a combination of aCD20 and RaM (right panel) and examined as in (a). (c) The effect of APRIL on apoptosis induction was also examined as in (a). (d) MLMA cells were treated as in (a) and apoptosis was induced. The inhibitory effect of simultaneous addition of BAFF (200 ng/ml) against apoptosis was examined at different time-points as in (a).

culture, the incidence of apoptosis induced by both BCRmediated and CD20-mediated stimuli was reduced as assessed by annexin V-binding. Although inhibition tended to be more effective with a higher dose of BAFF, the effect was not significant. We also examined the effect of pretreatment with BAFF on the inhibition of apoptosis but found none (Fig. 3b). In contrast, APRIL, another ligand for BCMA and TACI, did not affect apoptosis induced by the BCR-mediated and CD20-mediated stimuli, indicating the specificity of BAFF's effect (Fig. 3c). Therefore, we concluded that BAFF-mediated stimuli are able to inhibit apoptosis mediated by the cross-linking of either CD20 or BCR and simultaneous treatment with apoptosis-inducing stimuli is almost sufficient to achieve maximum BAFF-mediated inhibition of apoptosis, at least in these cases. However, the inhibitory effect of BAFF against apoptosis mediated by the cross-linking of either CD20 or BCR was only partial and it was more obvious when the inhibition of apoptosis was examined at several different time-points.

### Cellular effect of BAFF involved in the inhibition of apoptosis in MLMA cells

We further examined the molecular basis of the BAFFmediated inhibition of apoptosis in MLMA cells. First, we tested the effect of BAFF on the growth of MLMA cells. As shown in Fig. 4, when BAFF was added to the culture, the cell proliferation was slightly enhanced, as assessed by cell counting, suggesting that BAFF promotes the growth of MLMA cells.

Next, we examined the intracellular signalling induced in MLMA cells by BAFF treatment. As shown in Fig. 5(a), immunoblot analysis revealed cleavage of p100, the precursor of NF- $\kappa$ B2, and a increase in p52, the active form of NF-KB2 after BAFF treatment, suggesting that the



Figure 4. Effect of B-cell-activating factor (BAFF) on MLMA cell proliferation. Starting from a cell concentration at  $5 \times 10^5$ /ml, MLMA cells were cultured in the presence (solid line) and absence (dotted line) of 400 ng/ml of BAFF and cell numbers were counted at the time-points indicated. Each experiment was performed in triplicate and the means + SD are indicated.



Figure 5. Intracellular signalling events and induction of Bcl-2 protein by B-cell-activating factor (BAFF). Cell lysates were prepared from MLMA cells treated with 400 ng/ml of BAFF for the periods indicated and an immunoblot analysis was performed using the antibodies indicated.

activation of  $NF$ - $\kappa$ B2 occurred after the treatment. We also observed the cleavage of the precursor of  $NF-\kappa B1$ after BAFF treatment (Fig. 5a). We further examined the activation of other molecules after treatment with BAFF and found that GSK-3b was transiently phosphorylated (Fig. 5b). In addition, we observed an elevation in the level of Bcl-2, an anti-apoptotic protein, after BAFF treatment.

To investigate the early responses to BAFF in MLMA cells, global screening of candidate genes whose expression is regulated by BAFF was performed by employing a microarray system. First, we selected up-regulated genes that are expressed in MLMA cells treated with BAFF for 12 hr at a level at least 1-5-fold higher than in untreated cells. Under these conditions, 178 probes were selected as up-regulated genes (Table 1). Consistent with the results of the immunoblot analysis presented in Fig. 5(a), the gene expression of Bcl-2 was found to be up-regulated by BAFF treatment (Table 1). Interestingly, the gene expression of CD40, a member of the TNF-receptor family involved in B-cell survival, was also increased after treatment with BAFF. The genes that are known to be involved in anti-apoptotic effect, including Myb, Epstein– Barr virus (EBV)-induced gene 3 (EBI3), and caspase 8 and FADD-like apoptosis regulator (CFLAR), were also up-regulated by BAFF treatment.

We further confirmed the increased CD40 protein expression by flow cytometry (Fig. 6a). Similarly, downregulated genes that were expressed in BAFF-treated cells at a level at least 0-75-fold lower than in untreated cells were selected. As shown in Table 2, 517 probes were selected as down-regulated genes. The above results of global gene expression profiling suggest that the expression of various types of genes was influenced by BAFF stimulation in MLMA cells.

Figure 6. Effect of B-cell-activating factor (BAFF) on CD40 expression in MLMA cells. (a) MLMA cells cultured with or without BAFF for 3 days were stained with fluorescein isothiocyanate (FITC)-labelled monoclonal antibody (mAb) against CD40 and analysed by flow cytometry as in Fig. 1. (b) The inhibitory effect of CD40 stimulation on apoptosis induction was examined. MLMA cells were treated with 500 ng/ml of CD40-ligand in the presence of 2-5 ng/ml of interleukin-4 to stimulate CD40. The effects of either stimulation of CD40 alone or simultaneous stimulation of CD40 and BAFF receptor on apoptosis similarly induced as in Fig. 2 were examined.



Table 1. Up-regulated genes after BAFF stimulation

Affy ID	Gene name	Symbol	Fold-change
204798_at	V-myb myeloblastosis viral oncogene homolog	<b>MYB</b>	3.6934717
207861_at	Chemokine (C-C motif) ligand 22	CCL <sub>22</sub>	3.2195807
201669_s_at	Myristoylated alanine-rich protein kinase C substrate	<b>MARCKS</b>	3.0888734
213138_at	AT rich interactive domain 5A	ARID5A	2.9730885
203927_at	IkBe	<b>NFKBIE</b>	2.6444874
239412_at	Interferon regulatory factor 5	IRF5	2.6300144
205173_x_at	CD58 antigen	CD58	2.5103657
230543_at	Similar to Chloride intracellular channel protein 4	USP9X	2.4929807
201932_at	Leucine rich repeat containing 41	MUF1	2.3778253
203835_at	Leucine rich repeat containing 32	GARP	2.3439856
221912 <sub>_8_at</sub>	Human DNA sequence from clone RP4-622L5	<b>MGC1203</b>	2.296463
205599_at	TNF receptor-associated factor 1	TRAF1	2.283286
218470_at	Tyrosyl-tRNA synthetase 2	$CGI-04$	2.2782216
203685_at	B-cell CLL/lymphoma 2	BCL <sub>2</sub>	2.2648098
202644_s_at	Tumor necrosis factor, alpha-induced protein 3	TNFAIP3	2.2458956
204897_at	Prostaglandin E receptor 4 (subtype EP4)	PTGER4	2.2327275
217728_at	S100 calcium binding protein A6	S100A6	2.188896
234339 <sub>_8_at</sub>	Glioma tumor suppressor candidate region gene 2	GLTSCR2	2.1786015
226354_at	Lactamase, beta	LACTB	2.1238286
209680_s_at	Kinesin family member C1	KIFC1	2.1163168
206508_at	Tumor necrosis factor (ligand) superfamily, member 7	TNFSF7	2.0985012
223319_at	Gephyrin	GPHN	2.0934505
242312_x_at	AV736963 CB		2.0774355
207608_x_at	Cytochrome P450, family 1, subfamily A, polypeptide 2	CYP1A2	2.019507
229437_at	BIC transcript	BIC	1.9920377
224468_s_at	Multidrug resistance-related protein	MGC13170	1.9605879
$214101$ <sub>-S</sub> _at	Aminopeptidase puromycin sensitive	<b>NPEPPS</b>	1.949555
208624_s_at	Eukaryotic translation initiation factor 4 gamma, 1	EIF4G1	1.9305304
218819_at	DEAD/H (Asp-Glu-Ala-Asp/His) box polypeptide 26	DDX26	1.925604
200648_s_at	Glutamate-ammonia ligase	GLUL	1.9081395
210686_x_at	Solute carrier family 25, member 16	GDA; GDC; ML7; hML7; HGT.1; D10S105E; MGC39851	1.903342
48659_at	Invasion inhibitory protein 45	FLJ12438	1.8990294
204283_at	Phenylalanine-tRNA synthetase 2	FARS2	1.8852582
1563796_s_at	KIAA1970 protein	KIAA1970	1.8749646
219424_at	Epstein-Barr virus induced gene 3	EBI3	1.8620349
213747_at	Antizyme inhibitor 1	OAZIN	1.8602847
205419_at	Epstein-Barr virus induced gene 2	EBI2	1.8580037
210978_s_at	Transgelin 2	TAGLN2	1.8418014
$201502\_s\_at$	IkBa	NFKBIA	1.8220907
207688_s_at	Inhibin, beta C	<b>INHBC</b>	1.8162365
208949_s_at	Lectin, galactoside-binding, soluble, 3	LGALS3	1.8159895
$216252_x$ _x_at	Fas	FAS	1.7921783
203422_at	Polymerase (DNA directed), delta 1	POLD1	1.7856169
227299_at	Cyclin I	<b>CCNI</b>	1.7854006









### Table 1. (Continued)



### Table 2. Down-regulated genes after BAFF stimulation















Table 2. (Continued)



















Next we examined the effect of CD40 stimulation on apoptosis induction in MLMA cells. As shown in Fig. 6(b), when CD40 was stimulated by the addition of CD40-ligand (CD40L) in the presence of interleukin-4, induction of apoptosis in MLMA cells was mediated by

both BCR- and CD20-cross-linking. Furthermore, when both CD40 and BAFF were simultaneously stimulated, better inhibition of BCR-induced and CD20-induced apoptosis was observed than those mediated by each of them alone.

### **Discussion**

In the present study, we have clearly shown that BAFF can inhibit apoptosis mediated by BCR or CD20 in MLMA cells that exhibit a mature B-cell phenotype. The BCR is thought to play a crucial role in clonal selection and clonal expansion in the process of B-cell development to expand high-affinity clones against exogenous antigens and eliminate self-acting or low-affinity clones.<sup>31</sup> Although its precise function is yet to be clarified, CD20 is thought to play a role in B-cell development by mediating lipid raft-related signalling.<sup>32</sup> Our findings indicate that BAFF contributes to the regulation of B-cell development by modulating apoptotic elimination of B cells mediated by BCR and CD20. Furthermore, our results also indicate that BAFF can enhance cell proliferation in MLMA cells.

A number of studies have attempted to elucidate the molecular basis of the function of BAFF.<sup>24,33-39</sup> A major focus of recent investigations has been the pro-survival signalling of BAFF-R. Activation of the alternative  $NF-\kappa B$ pathway (processing of  $NF$ - $\kappa$ B2 and the nuclear translocation of p52/RelB heterodimers) is a major outcome of BAFF-R-stimulation,  $2,24,36$  whereas BAFF-R also weakly activates the classical  $NF-\kappa B$  pathway mediated by NF-KB1 and low-level nuclear translocation of p50/RelA DNA-binding activity is induced. Coincident with previous reports, we also observed that BAFF induced cleavage of both NF- $\kappa$ B2 and NF- $\kappa$ B1 in MLMA cells (Fig. 4). APRIL only activates NF-KB1 via either BCMA or TACI and did not inhibit CD20- and BCR-mediated apoptosis in MLMA cells, so the anti-apoptotic effect of BAFF is thought to be mediated mainly by NF- $\kappa$ B2 activation.

Recent studies have shown that NF-KB directly binds to the promoter region of the Bcl-2 gene and induces transcriptional activation.<sup>39</sup> Since Bcl-2 has an anti-apoptotic function, the elevated level of Bcl-2 protein is thought to be important for BAFF-mediated B-cell survival. In addition, BAFF is reported to temporarily inactivate GSK-3b via AKT-mediated phosphorylation.<sup>34</sup> Since GSK-3 $\beta$  has been found to cause apoptosis by inducing the degradation of Mcl-1 (an anti-apoptotic Bcl-2 family member) and compromising mitochondrial membrane integrity,<sup>40</sup> the BAFF-mediated phosphorylation of  $GSK-3\beta$  is thought to also participate in the anti-apoptotic effect of BAFF. Consistent with previous reports, we observed that BAFF treatment also induced both an increase in Bcl-2 expression and the transient phosphorylation of GSK-3b in MLMA cells. Therefore, it is likely that the inhibitory effect of BAFF against apoptosis mediated by CD20 or BCR is also mainly the result of NF-KB2-mediated Bcl-2 expression and the transient inactivation of GSK-3b.

The pro-apoptotic BH3-only Bcl-2 family member Bim was shown to sequestrate Bcl-2 and play an essential role for BCR cross-linking-induced apoptosis. $41$  Moreover, Mcl-1 was shown to inhibit Bim selectively and to be essential both early in lymphoid development and later on in the maintenance of mature B lymphocytes. $42$  Therefore, future investigation of the involvement of Bim and MCL-1 in the BAFF-induced inhibition of CD20-mediated and BCR-mediated apoptosis in MLMA cells should be interesting.

Interestingly, our findings indicated that BAFF treatment also induces the expression of a series of genes related to cell survival. For example, both the microarray analysis and the flow cytometric analysis revealed increased expression of CD40 after the treatment. Since CD40 is known to mediate pro-survival signalling upon interaction with CD40L expressed on activated T cells,  $43$ it is suggested that BAFF-mediated up-regulation of CD40 expression also contributes to B-cell survival in vivo. Indeed, we observed in this study that simultaneous stimulation with CD40 and BAFF resulted in better inhibition of BCR-induced and CD20-induced apoptosis than stimulation with by each of them alone. Therefore, it may be possible that BAFF-mediated up-regulation of CD40 inhibits apoptosis induction synergistically with the effect of BAFF in vivo.

The microarray analysis also revealed up-regulation of several genes involved in either the inhibition of apoptosis or the proliferation of B cells (Table 1). For example, Myb has been demonstrated to directly up-regulate Bcl-2 and suppresses apoptosis.<sup>44,45</sup> EBI3 is a subunit of interleukin-27 that increases proliferation of B cells.<sup>46</sup> CFLAR is known to inhibit the activation of caspase  $8.^{47}$  Therefore, our data may indicate that mechanisms other than  $NF$ - $\kappa$ B2-mediated Bcl-2 up-regulation are involved in the anti-apoptotic effect of BAFF in MLMA cells. Furthermore, because direct cross-talk between the BCR-signalling and BAFF-signalling systems has been reported, $48$ BAFF-mediated signals may also be able to directly influence the BCR-mediated apoptotic signalling system.

It is well documented that BAFF-mediated signalling is involved in the survival of malignant B cells. $2,49$  For example, BAFF is reported to be an autocrine pro-survival and proliferation factor for B-cell chronic lymphocytic leukaemia and multiple myeloma.2,49–51 BAFF is also thought to promote cell survival and proliferation in Hodgkin and non-Hodgkin lymphoma. Therefore, BAFF and BAFF-R might be potential molecular targets in the treatment of B-cell malignancies.<sup>52-54</sup> Using a combination of blocking of BAFF-signalling and activation of apoptosis induction, such as CD20 cross-linking, a novel therapeutic approach would be developed.

In conclusion, BAFF-mediated signalling inhibited CD20-mediated or BCR-mediated apoptosis in MLMA cells. Although more detailed experiments are clearly needed, MLMA cells should provide a model for investigating the molecular basis of BAFF's effect on B cells in vitro and will help to elucidate how B cells survive in an immune system in which BAFF-mediated signalling is involved.

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