

Attenuation of changes in capillary fine structure and leukocyte adhesion improves muscle performance following chronic ischaemia in rats

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Acute ischaemia–reperfusion disrupts capillary fine structure and increases leukocyte adhesion in postcapillary venules. We determined whether chronic muscle ischaemia has similar consequences, and whether it is possible to ameliorate its effect on muscle performance. Following ischaemia (unilateral ligation, common iliac artery) rat hindlimb muscles were examined without other intervention or following treatment with an xanthine oxidase inhibitor (allopurinol), a Na⁺/H⁺ exchange blocker (amiloride), or an oxygen free radical scavenger (vitamin E). No significant leukocyte adhesion or rolling, nor changes in capillary fine structure were observed 3 days postsurgery, when limb use was limited. However, leukocyte rolling and adhesion almost trebled by 7 days ($P < 0.001$), when normal gait was largely restored. Capillary fine structure was disturbed over a similar time course, e.g. relative endothelial volume (control 46%, 7 days 61%; $P < 0.05$), that resolved by 5 weeks. Where activity was increased by mild electrical stimulation 3 days after ligation muscles showed enhanced capillary swelling (endothelial volume 66% versus 50%, $P < 0.005$), but improved fatigue index (52% versus 16%, $P < 0.001$) as a result of greater blood flow. Muscle fatigue after ligation was related to the extent of contraction-induced hyperaemia ($R^2 = 0.725$), but not capillary swelling. Amiloride, and to a lesser extent allopurinol but not vitamin E, significantly decreased leukocyte rolling and adhesion, as well as capillary endothelial swelling. We conclude that increased activity of ischaemic muscles on recovery is likely to accentuate acidosis accompanying changes in microcirculation and contribute to enhanced muscle fatigue, whereas formation of oxygen free radicals may be attenuated by endogenous protective mechanisms.

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Peripheral vascular diseases affect about 7% of people over 55 years of age (Leng *et al.* 1996). The incidence increases with age and is usually linked with impairment in coronary or cerebral circulation (Rothwell *et al.* 2005), thus representing serious health impairment in the ageing population. So far the most effective treatment of intermittent claudication, a symptom of peripheral vascular disease, has been exercise. However, intensive activity can also damage ischaemic muscles (Hudlická *et al.* 1994). One of the reasons for muscle damage may be an impaired muscle microcirculation, indicated by capillary endothelial swelling and increased leukocyte adhesion. Swollen endothelium has been demonstrated by electron microscopy in acutely ischaemic muscles (Strock

& Majno, 1969), especially when followed by reperfusion (Gidlöf *et al.* 1988), and increased permeability (Suval *et al.* 1987). Leukocyte adhesion to venular endothelium has been demonstrated in skeletal muscle (Nolte *et al.* 1992) and other tissue as a general feature of reperfusion injury (Ley, 1992). Either or both of these processes may contribute to impaired capillary perfusion due to microvascular narrowing or blockade. Changes in microcirculation similar to those following reperfusion were observed during haemorrhagic shock (Perry & Granger, 1992; Kretchmar & Engelhardt, 1994). Capillary endothelial swelling could be decreased by amiloride blockade of Na⁺/H⁺ exchange (Mazzoni *et al.* 1992), which is also activated during reperfusion (Masereel *et al.* 2003), while leukocyte adhesion in muscles exposed to ischaemia–reperfusion was prevented by oxygen free radical scavengers (Gute *et al.* 1998).

This paper has online supplemental material.

There are few data on capillary fine structure and leukocyte behaviour in chronically ischaemic muscles. Hou *et al.* (1995) described increased leukocyte adhesion in rat cremaster muscle 3 weeks after ligation of its feed artery. We have previously shown that capillary endothelial swelling and increased leukocyte adhesion occurred in chronically ischaemic muscles 2 weeks after unilateral ligation of the common iliac artery in the rat, and that some changes were also found in contralateral muscles (Hickey *et al.* 1992, 1993; Egginton *et al.* 1993). While it is impossible to assess the contribution of these two processes to impairment of capillary perfusion, clearly they are linked (this study; Anderson *et al.* 2006). Leukocyte adhesion represents an obstacle to capillary perfusion in acute reperfusion, and also results in increased capillary permeability, increased interstitial volume and hence tissue pressure, and consequently a drastic decrease in the number of perfused capillaries (Jerome *et al.* 1994). The highest percentage of capillaries with swollen endothelium (70% compared with 27% in control muscles) was found in extensor digitorum longus (EDL) 7 days after ligation (Hughes & Hudlická, 1992). Consistent with an impaired microcirculation, the time spent stationary by red blood cells was significantly increased, and surface P_{O_2} significantly decreased in comparison with control muscles (Dawson *et al.* 1990). Additionally, the number of adhering and rolling leukocytes was increased, suggesting activation of the venular endothelium (Anderson *et al.* 2006). In combination, these changes may explain the diminished muscle performance seen at this time (Hughes & Hudlická, 1992; Milkiewicz *et al.* 2006). Changes in leukocyte adhesion and the extent of capillary endothelial swelling may be related to muscle activity, since the animals start to use the ischaemic leg 3–7 days after ligation. In agreement with this, the proportion of rolling and adhering leukocytes was normal in EDL 3 days after ligation when the ischaemic leg is little used, but when the ligated muscles were subjected to a mild increase in activity by electrical stimulation for 2 out of the 3 days, the proportion of adhering leukocytes increased in comparison with muscles that were not stimulated (Anderson *et al.* 2006). We reasoned that similar changes would be observed in capillary fine structure.

The mechanism underlying the effect of muscle activity on capillary endothelial swelling is not clear. Ischaemic muscles have low pH (Challis *et al.* 1986) and muscle activity would certainly increase local acidosis. Acidosis was thought to be responsible for capillary swelling in shock (Mazzoni *et al.* 1992) which was diminished by amiloride, and an amiloride analogue also reduced capillary lumen narrowing resulting from systemic blood acidosis in combination with decreased blood flow (Mazzoni *et al.* 1994). Furthermore, it is possible that reperfusion which occurs after acute muscle ischaemia might also occur with a chronic insult: femoral

artery blood pressure dropped immediately after ligation but slightly recovered 7 days later, indicating partial reperfusion (Hudlická *et al.* 1994), and hence swelling as well as leukocyte adhesion could be triggered by generation of oxygen free radicals.

We hypothesised that increased muscle activity at an early stage after interruption of blood supply may accelerate metabolic changes induced by ischaemia (e.g. lowering of pH) and reperfusion that occurs gradually, and thus lead to capillary damage and microcirculatory dysfunction. Such changes may be accentuated by imposition of additional muscle activity, such as seen with therapeutic exercise. We tested this hypothesis by determining whether these responses could be attenuated by pharmacological intervention, or intensified by muscle stimulation.

Methods

All experiments were performed in accordance with the UK Animals (Scientific Procedures) Act 1986, and had local ethical committee approval.

Surgical procedures

Experiments were performed on male Sprague–Dawley rats (*Rattus norvegicus*) using animals of 390–450 g for blood flow and muscle performance estimations (5–7 animals were used in each of the treatment groups, Table 1), and ~290 g body mass for intravital microscopy observations (5 rats per group). The common iliac artery was ligated 3–5 mm below the bifurcation from the abdominal aorta under halothane anaesthesia (2% Fluothane, ICI, in oxygen at 2 l min⁻¹) and aseptic conditions. The animals were given systemic analgesic (2.5 ml kg⁻¹ buprenorphine, s.c., Temgesic, National Veterinary Services, Stoke-on-Trent, UK) and topical antibiotic (Duplocillin LA, National Veterinary Services, Stoke-on-Trent, UK) twice a day for the first 2 days. The animals recovered within 20–30 min and received either distilled water or 0.5 ml of the xanthine oxidase inhibitor allopurinol (50 mg kg⁻¹; Sigma-Aldrich Co., UK), the Na⁺/H⁺ exchanger blocker amiloride (5 mg kg⁻¹), or the oxygen free radical scavenger vitamin E (50 mg kg⁻¹ in castor oil and polyethyleneglycol) by gavage the afternoon after surgery, and twice daily for the following 6 days, with the last dose given ~16 h before being taken into the final experiment. To study the effect of additional activity on capillary fine structure, one group of animals had Teflon-coated stainless steel multistranded electrodes implanted close to the peroneal nerve at the time of the ligation of the iliac artery. In these animals the extensor digitorum longus and tibialis anterior muscles were stimulated for 2 days starting ~24 h after surgery. The stimulation was performed 7 times per day in periods of 15 min with a rest of 85 min

Table 1. Body mass, EDL/TA muscle mass, peak tension development, activity-induced muscle blood flow, and fatigue index following iliac artery ligation

Days ligation (n)	Body mass (g)	Ischaemic muscle mass (g)	I/CL	Ipsilateral tension (g/g)	I/CL	Ischaemic MBF (ml min ⁻¹ (100 g) ⁻¹)	I/CL	Ipsilateral FI (%)	I/CL
0 (7)	391.9 ± 17.0	0.894 ± 0.054	0.966	280.0 ± 14.5	1.095	123.0 ± 18.1	0.936	66.0 ± 2.7	1.023
3 (5)	365.2 ± 17.3	0.814 ± 0.048	0.977	186.4 ± 17.3*	1.416	4.1 ± 0.5*	0.053	15.5 ± 0.9*	0.236
7 (7)	349.3 ± 5.2*	0.786 ± 0.036	0.892	188.3 ± 12.9*	1.360	4.3 ± 1.2*	0.069	20.0 ± 3.8*	0.309
35 (6)	408.3 ± 15.3	0.926 ± 0.051	0.894	145.3 ± 19.2*	0.824	29.1 ± 5.6*	0.498	52.2 ± 4.3*	0.841
3d SL (6)	349.2 ± 6.2*	0.816 ± 0.043	0.993	119.6 ± 29.7*	0.739	6.7 ± 0.8*	0.100	36.9 ± 7.0*	0.572

Means ± s.e.m. (number of animals). **P* < 0.05 versus control (ANOVA). Abbreviations: I, ischaemic muscle; CL, contralateral muscle; MBF, muscle blood flow; FI, fatigue index; 3d SL, 3 days ligation and 2 days stimulation.

at 10 Hz, pulse width 0.3 ms, and up to 5 V intensity using a Neurotech Multichannel Stimulator (Bio-Medical Research, West Galway, Ireland), as previously described (Hudlická *et al.* 1994). A group of animals with iliac artery ligation were taken into experiment 3 days later for studies of leukocyte adhesion (*cf.* Anderson *et al.* 2006) and capillary fine structure. All other animals were taken into experiment 7 days after unilateral ligation of the iliac artery. For acute experiments, animals were anaesthetised by intraperitoneal injection of sodium pentobarbitone (Sagatal; May & Baker, Dagenham, UK) 50 mg (kg body mass)⁻¹ diluted 1 : 1 with saline. At the end of observations, animals were killed by anaesthetic overdose.

Muscle performance and blood flow measurement

The right jugular vein was cannulated using vinyl tubing to supplement anaesthesia as necessary, and a tracheal cannula assisted spontaneous breathing. Both brachial arteries were cannulated, for the recording of blood pressure *via* a pressure transducer (Bell & Howell, Basingstoke, UK) and withdrawal of blood samples for measurement of blood flow. The left ventricle was cannulated *via* the right carotid artery, with the pressure pulse used to identify when the tip of the cannula was in the ventricle. For recording of muscle tension the thighs were fixed to a specially designed board where a U-shaped lead restraining bar was bent around the groin to prevent movement of the thigh and knee joint, thus providing a stable position of the calf from which muscle contractions could be measured. Feet were clamped into special holders, and the tendons of EDL and TA were exposed in both hindlimbs, cut and attached to strain gauges to record isometric muscle tension in response to indirect stimulation of the peroneal nerve at 4 Hz and a pulse width of 0.3 ms. The stimulation voltage and muscle length were adjusted to give maximum force of contraction. Tension output and blood pressure recordings were displayed on a six-channel recorder (Lectromed, Letchworth Garden City, UK). Muscle performance was assessed as peak tension and fatigue index (tension in g (g muscle)⁻¹ at the end of the period of contractions/peak tension × 100).

Radioactive microspheres (15 µm diameter and labelled with ⁴⁶Sc, ⁵⁷Co, or ¹¹³Sn; PerkinElmer (NEN), Groningen, the Netherlands) were used to measure muscle blood flow (MBF) at rest and at the end of 5 min isometric twitch contractions in the EDL and TA as previously described (Hudlická *et al.* 1994). Each animal received 1.5–1.8 × 10⁵ microspheres. The blood flow was measured in tibialis anterior and extensor digitorum longus muscles, where the number of microspheres was at least 400 (estimated on the basis of specific activity), which is sufficient to ensure reasonable accuracy (Buckberg *et al.* 1971). A reference blood sample was taken at a withdrawal rate of 0.5 ml min⁻¹ using a precision withdrawal pump (Braun, Melsungen, Germany) from the brachial artery simultaneously with injection of the microspheres into the left ventricle, to calculate MBF by scaling. The withdrawn blood volume was replaced with an intravenous infusion of 1% w/v bovine serum albumen in saline. At the end of the experiment radioactivity of EDL and TA muscles was determined (Packard Auto-Gamma Counter), along with samples from right and left kidney cortex (to confirm homogeneous distribution of microspheres) and from the lung (to assess the percentage of shunting). Any animal where the kidney samples had greater than a 10% discrepancy in blood flow, or where the lung sample showed greater than 5% shunting, was discounted.

Intravital microscopy

Another group of rats were surgically prepared with a jugular vein cannula for delivery of supplemental anaesthetic as required, a carotid artery cannula to record blood pressure, and a tracheal cannula to allow spontaneous breathing. Two coiled stainless steel multistranded Teflon-coated electrodes were implanted in the vicinity of the peroneal nerve and exteriorised on the back of the animal for later stimulation. The EDL was exposed for intravital observation, as described by Anderson *et al.* (1997). The muscle was superfused with warm (37°C) deoxygenated (95% N₂–5% CO₂) physiological salt solution (131.9 mM NaCl, 4.7 mM KCl, 2.0 mM CaCl₂, 1.2 mM MgSO₄ and 22 mM NaHCO₃,

pH 7.35–7.45) during dissection, and for the whole time of observation under the intravital microscope (ACM Zeiss, Oberkochen, Germany). The muscle was viewed using water immersion objective ($\times 25$, NA 0.6) and fibre optic epi-illumination (equipped with a green filter), and images of vessels were recorded as described in detail previously (Thomson *et al.* 1994; Anderson *et al.* 2006). The diameter of venules, together with the branching pattern, helped to classify them as collecting venules (V_4), V_3 draining them, or V_2 draining V_3 . The final magnification on the monitor was $\times 1000$, and monitor resolution was calculated as $0.45 \mu\text{m pixel}^{-1}$ from measurements using a graticule. Once the animal was transferred under the microscope, the hindlimb muscles were stimulated at 1 Hz (0.3 ms pulse width at 3–5 V; Grass S8 stimulator) for 30 min. This procedure increased leukocyte adhesion (Thomson *et al.* 1994) and thus enabled a better assessment of interventions. Five to 10 venules in different categories in each animal were observed for 1 min intervals to measure leukocyte rolling and adhesion offline, usually over a total period of 2 h. Leukocyte rolling or stationary adhesion was estimated as number of cells per $100 \mu\text{m}$ of each individual vessel adhering, or rolling for 10 s.

Capillary supply

Sections of EDL were mounted on cork discs using OCT medium and rapidly frozen in isopentane precooled in liquid nitrogen. Cryostat sections $8 \mu\text{m}$ thick were stained for alkaline phosphatase using the indoxyl-tetrazolium method, and capillary supply was estimated by counting number of fibres and capillaries in two areas of each muscle (each 0.25 mm^2) with the results expressed as capillary to fibre ratio (C:F).

Capillary fine structure

EDL from both legs were dissected free and thin strips fixed at resting length in 2.5% glutaraldehyde, 1.5% sucrose, 0.1 M phosphate buffer ($350 \text{ mosmol l}^{-1}$) pH 7.4 at room temperature for 1 h. We adopted immersion in isotonic fixative despite the use of high osmolarity and perfusion fixation in other studies, as the latter tends to obscure structural changes in the endothelium of ischaemic tissue (Egginton *et al.* 1993). The strips were then sliced into small blocks $\sim 2 \text{ mm}^3$ and placed in fresh fixative for 24–48 h at 4°C . Post-fixation was carried out using 1% OsO_4 in phosphate buffer for 1 h at room temperature. Samples were dehydrated in a graded series of alcohols and embedded in Agar 100/Epon substitute resin. Four blocks were prepared from each of four animals, and one per animal chosen at random for analysis. Semi-thin ($0.5 \mu\text{m}$) sections were used to ensure true transverse sections of muscle. Ultrathin sections were cut at 60–80 nm, stained with uranyl acetate for 7 min

and Reynolds lead citrate for 7 min, and viewed in a Jeol 100 CX II transmission electron microscope at 80 kV. A 35 mm camera was used to capture images of capillaries (approximately 40 per muscle) at an initial magnification of $\times 640$, using an unbiased sampling protocol. Negatives were then projected on a film reader ($\times 17.5$) and scanned into an Apple Macintosh computer, using NIH Image with purpose-written macros or Optilab software (Graftex, Meudon-la-Forêt, France) to quantify capillary structure by means of stereological measurements. As perfect circular profiles only exist with vessels that are fixed *in situ* whilst perfused (immersion fixation removes hydrostatic pressure and reveals unperfused vessels as collapsed profiles), and tortuosity of the capillary bed would also produce oblique profiles, both cross-sectional area and perimeter were determined. The former will be sensitive to section angle and degree of opening, but the latter will be less sensitive to such differences in *in vivo* status. The absolute and relative area of lumen and nucleus, area of endothelial cytoplasm and surface/volume ratio of lumen were used as indices of endothelial swelling.

Statistical analysis

All results are presented as means \pm S.E.M. Statistical evaluation was carried out by ANOVA with Fisher's PLSD *post hoc* test used for intergroup comparisons. The distribution of data for both capillary endothelial swelling and leukocyte adherence was left skewed, and was normalised by a \log_{10} transformation before statistical processing. The distribution of capillary ultrastructural values among groups were also analysed using the Mann–Whitney *U* test. For all tests, $P < 0.05$ was taken as a significant difference.

Results

Body, muscle mass and blood pressure

The mean body and muscle mass (Table 1) and blood pressure (controls, $100.8 \pm 8.7 \text{ mmHg}$) of all animals used in this study were similar.

Muscle performance

Twitch, rather than tetanic tension was used to assess muscle performance since the use of tetanic contractions to elicit fatigue was considered to be too severe in ischaemic muscles. The right EDL and TA muscles in control animals produced a peak isometric twitch tension of $280.0 \pm 14.5 \text{ g g}^{-1}$, with significantly lower values in muscles 3, 7 and 35 days following ligation ($P < 0.05$; Table 1). Lower peak tension was also developed by the contralateral muscles at 3 days following ligation, and a reduction was apparent throughout the whole time

course following ligation (Table 1). Fatigue index (FI) was $66.0 \pm 2.7\%$ in the EDL and TA muscles of the right limb, and $64.5 \pm 4.1\%$ in the left limb in control animals. Fatiguability increased dramatically in animals 3 days following ligation, gradually reaching a value approaching that of control muscles after 35 days. The FI in contralateral muscles was not significantly different from control after ligation up to 35 days (see Table 1).

Muscle blood flow

Muscle blood flow (MBF) in EDL and TA at rest was significantly reduced at all time intervals following ligation, compared with muscles in control animals. Although some improvement was observed at 5 week after ligation ($5.03 \pm 1.14 \text{ ml min}^{-1} (100 \text{ g})^{-1}$), doubling that seen at 7 days ($2.60 \pm 0.41 \text{ ml min}^{-1} (100 \text{ g})^{-1}$), flow was still significantly lower than in control muscles ($7.27 \pm 1.13 \text{ ml min}^{-1} (100 \text{ g})^{-1}$ in control animals, $P < 0.05$). Resting blood flow in the contralateral EDL and TA showed no difference at any time point after ligation (data not shown; see also Milkiewicz *et al.* 2006).

MBF at the end of a 5 min period of muscle contractions at 4 Hz increased to $123.04 \pm 18.11 \text{ ml min}^{-1} (100 \text{ g})^{-1}$ in control animals. It was limited in all animals with iliac artery ligation, with the greatest reduction seen in muscles that had been ischaemic for 3 days ($4.06 \pm 0.50 \text{ ml min}^{-1} (100 \text{ g})^{-1}$ ($P < 0.01$ versus control; Table 1). The improvement was gradual, but MBF was still significantly lower 5 week after ligation ($29.10 \pm 5.62 \text{ ml min}^{-1} (100 \text{ g})^{-1}$, $P < 0.05$ versus control). Flow in the contralateral muscles (always on the left side) of animals 7 days after ligation was significantly lower than in the left EDL and TA muscles in control animals ($60.97 \pm 6.93 \text{ ml min}^{-1} (100 \text{ g})^{-1}$ versus $117.10 \pm 16.88 \text{ ml min}^{-1} (100 \text{ g})^{-1}$, $P < 0.05$) but there were no differences from controls in contralateral muscles at any other time point (Table 1; see also Milkiewicz *et al.* 2006). The fatigue resistance of ischaemic muscle fell in line with the reduced MBF (Figs 1 and 7).

Leukocyte rolling and adhesion

The behaviour of leukocytes was observed in postcapillary venules of rats with unilateral iliac artery ligation, and in those also treated with allopurinol, amiloride and vitamin E. Evaluation was performed in 45–50 collecting venules (V_4), 60–70 V_3 and 35–45 V_2 in the various groups of rats. Resting venular diameters were $V_4 = 7.0 \pm 0.3 \mu\text{m}$, $V_3 = 11.4 \pm 0.9 \mu\text{m}$ and $V_2 = 19.0 \pm 0.8 \mu\text{m}$ at 7 days, similar to those previously described (Anderson *et al.* 2006), and did not appear to have been affected by drug treatment. There was, however, a transient, though modest, dilatation at 3 days, possibly due to surgical

trauma, with venule diameters of 8.7 ± 0.5 , 12.1 ± 0.9 and $24.6 \pm 2.8 \mu\text{m}$, respectively (n.s.).

No rolling or adhering leukocytes were found in the smallest collecting venules (V_4) either in control or in muscles ischaemic for 7 days. Both rolling and adherent leukocytes were observed in V_3 with 12.8 ± 0.8 and $7.0 \pm 0.4 (100 \mu\text{m segment})^{-1} (60 \text{ s})^{-1}$, respectively ($4\times$ and $17\times$ increase from control values, $P < 0.05$). Values in V_2 were 22.3 ± 0.79 and $9.1 \pm 0.6 (100 \mu\text{m segment})^{-1} (60 \text{ s})^{-1}$, respectively ($2.4\times$ and $6.4\times$ increase from controls). The values in muscles ischaemic for 3 days only were similar to controls, but were increased by mild electrical stimulation, as seen previously (Anderson *et al.* 2006). Amiloride practically eliminated both rolling and

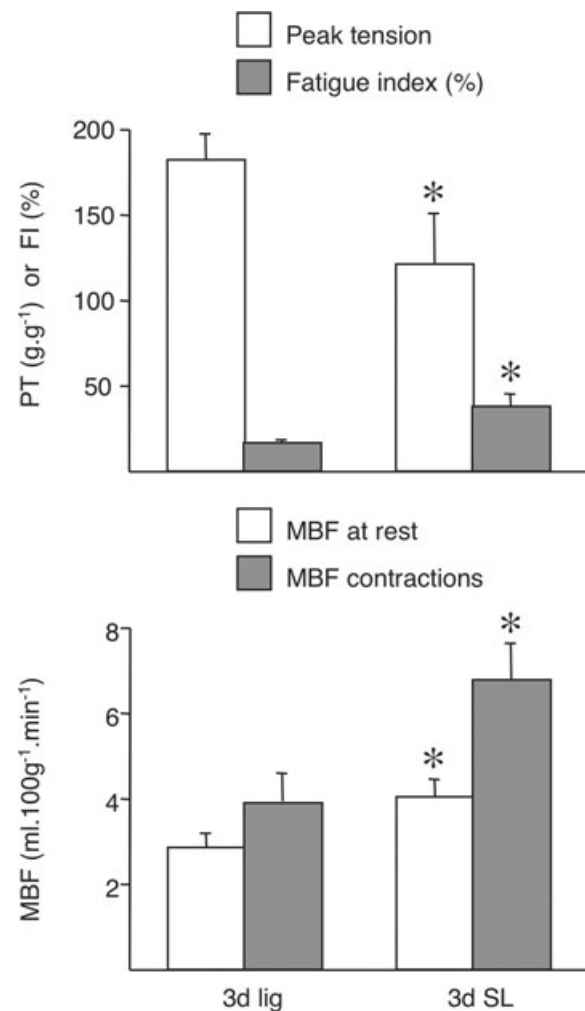


Figure 1. Muscle blood flow (MBF) and fatigue in combined EDL + TA muscles at rest and at the end of 5 min isometric contractions in 3 days ischaemic (3d lig) and 3 days ischaemic with 2 days stimulation (3d SL)

Fatigue index (FI) is the percentage of the peak tension (PT) at the end of the 5 min period of contractions. * denotes values significantly different from 3 days lig ($P < 0.05$). For control data, please refer to Table 1.

adherence in V_3 , and significantly decreased it in V_2 (to 8.7 ± 1.2 and 3.6 ± 0.6 ($100 \mu\text{m segment}^{-1}$) (60 s^{-1}), respectively, $P < 0.01$ versus ischaemia alone; Fig. 2). The effect of allopurinol was more modest (11.4 ± 0.8 and 7.2 ± 0.26 ($100 \mu\text{m segment}^{-1}$) (60 s^{-1}), respectively), with vitamin E having little or no effect (n.s., Fig. 2).

Capillary supply

Capillary to fibre ratio (C : F) in the EDL muscles of control animals was 1.32 ± 0.07 . No significant differences were observed in the ischaemic muscles following ligation at 3 or 7 days but increased to 1.70 ± 0.1 ($P < 0.05$ versus control) 35 days after ligation. All contralateral muscles had C : F values not significantly different from those seen in control animals.

Capillary fine structure

Stereological analysis of the total capillary and luminal cross sectional area or surface/volume ratio (S/V) revealed

differential changes in capillary fine structure according to length of ischaemia and drug treatment. Muscle ischaemia *per se* did not significantly alter the cross sectional area of capillaries, but resulted in swelling of the capillary endothelium. The cytoplasm was more translucent in capillaries that showed swelling, and many capillaries had protrusions into the lumen (Fig. 3). The values for cytoplasmic area (i.e. volume of endothelium – volume of nucleus), volume density and S/V of the capillary lumen – which are indications of capillary endothelium swelling – are given in Fig. 4 for ischaemic muscles, with other parameters measured included in Table 2. Capillaries in control muscles had an endothelial cell (EC) area of $7.52 \pm 0.87 \mu\text{m}^2$ and the lumen occupied $54.3 \pm 3.1\%$ of the total capillary area (cross sectional area of capillary profiles, i.e. lumen + endothelium). A significant increase in relative EC area was observed at 7 days following ligation ($P < 0.05$, Mann–Whitney U test), but values were not significantly different from controls after 5 week (Fig. 4). Surprisingly, there was no evidence of endothelial swelling in muscles 3 days after ligation. The increased percentage of EC cytoplasm resulted in a smaller luminal area

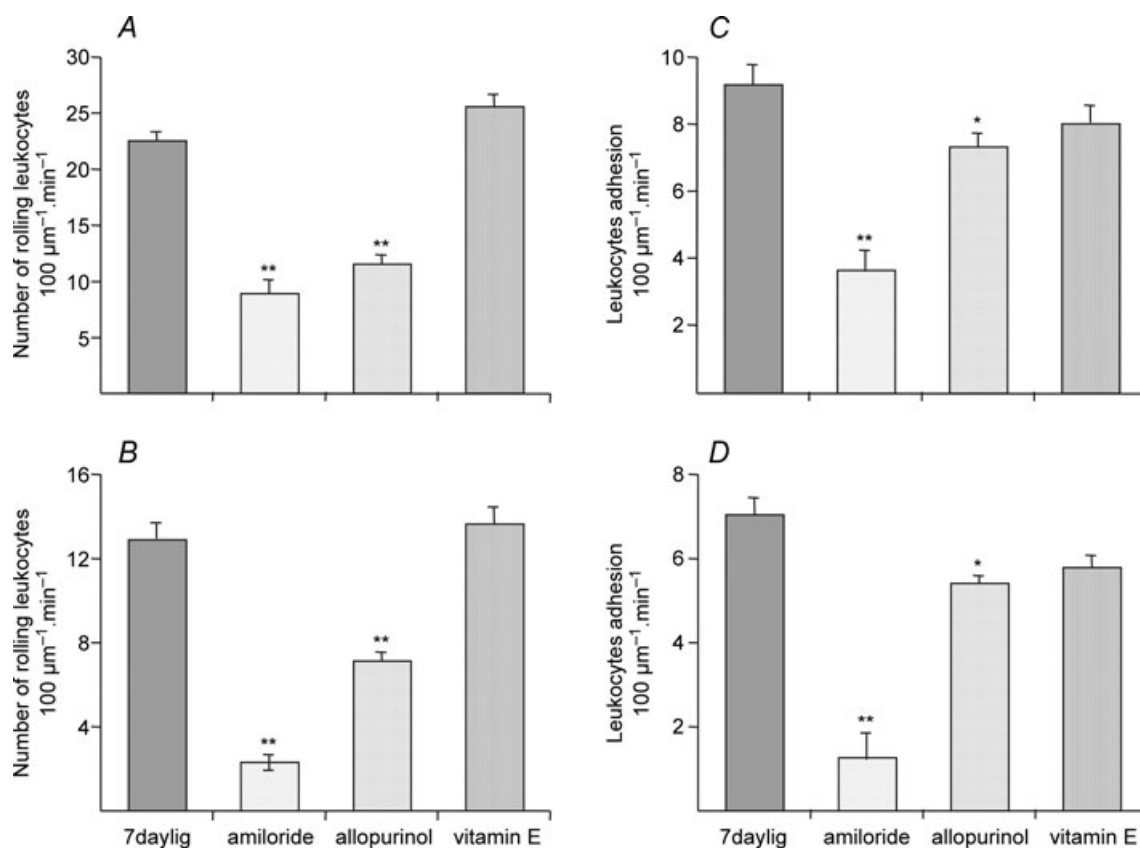


Figure 2. Leukocyte rolling and adhesion in untreated 7 days ischaemic muscles, and ischaemic muscles in animals treated with amiloride, allopurinol or vitamin E

* $P < 0.05$, ** $P < 0.01$ versus ischaemia alone. For comparison, in untreated control muscles the corresponding values are: V_3 rolling 3.0 ± 1.7 , stationary 0.4 ± 0.4 ; V_2 rolling 9.3 ± 3.0 , stationary $1.4 \pm 0.6 \text{ min}^{-1}$ (Anderson *et al.* 2006).

and, reflecting this, luminal S/V was increased from $3.2 \pm 0.2 \mu\text{m}^{-1}$ in control muscles up to $4.2 \pm 0.3 \mu\text{m}^{-1}$ 7 days after ligation. The values for cytoplasmic area and lumen S/V were significantly greater in muscles that had been ischaemic for 7 days or in those ischaemic for 3 days with increased activity (Fig. 4).

As there were minimal changes in the morphological parameters described above in contralateral muscles (see online supplemental material, Supplementary Table 1), the effect of drugs was evaluated as a proportion of change related to the contralateral side. Neither drug altered the capillary profile area. Allopurinol and amiloride, but not vitamin E decreased the cytoplasmic area, indicating their beneficial effect on reducing capillary endothelial swelling (Fig. 5 and Table 3). There was a significant correlation between resting MBF and the proportion of the capillary occupied by endothelium ($R^2 = 0.603$, $P < 0.05$).

Changes in capillary fine structure were also observed in papillary muscle (Fig. 6 and Table 4). Unlike in skeletal muscles, capillary area in the papillary muscle was increased as a result of ischaemia, and although the changes in the cytoplasmic area was smaller the surface to volume ratio was decreased in all muscles from ischaemic animals, even 35 days after ligation of the common iliac artery. Other parameters (capillary and luminal cross-sectional area, proportion of lumen, and lumen radius) showed a gradual change with time after ligation. Lumen S/V gradually decreased, indicating that the increase in lumen volume was greater than the increase in the luminal perimeter, possibly as a result of fewer luminal protrusions (Table 4).

Effect of activity on capillary fine structure, blood flow and performance in ischaemic muscles

Morphometric data based on quantitative electron microscopy showed that marked capillary EC swelling was absent at 3 days but appeared 7 days after ligation, i.e. at a time when the animals were observed first using the ischaemic limb, suggesting that structural changes could be due to a combination of ischaemia with muscle activity. To test this hypothesis, we increased muscle activity by electrical stimulation for 2 days starting 24 h after ligation of the iliac artery. This resulted in EC swelling at 3 days that was greater than with ligation alone ($P < 0.05$) and similar to that found in ischaemic muscles at 7 days (n.s., Fig. 4), and induced modest changes in the contralateral muscles (Supplementary Table 1, some data not shown). Although the total capillary area was greater than in controls ($17.4 \pm 1.4 \mu\text{m}^2$ versus $14.4 \pm 1.0 \mu\text{m}^2$, $P < 0.05$) and in 3 days ischaemic muscles ($13.4 \pm 0.58 \mu\text{m}^2$, $P < 0.05$), the luminal area did not change and thus the relative area of lumen (0.39 ± 0.02 versus 0.34 ± 0.03) as well as the luminal radius ($1.38 \pm 0.09 \mu\text{m}$ versus $1.56 \pm 0.62 \mu\text{m}$

in 3 day ischaemic muscles) were significantly smaller ($P < 0.05$; Table 2, Fig. 4).

In spite of the changes in capillary fine structure that would limit capillary perfusion, animals which received muscle stimulation showed a modest improvement in resting MBF when compared with those ischaemic for 3 days ($3.83 \pm 0.07 \text{ ml min}^{-1} (100 \text{ g})^{-1}$ in stimulated animals compared with $2.81 \pm 0.32 \text{ ml min}^{-1} (100 \text{ g})^{-1}$ with ligation alone, $P < 0.05$), as well as during muscle

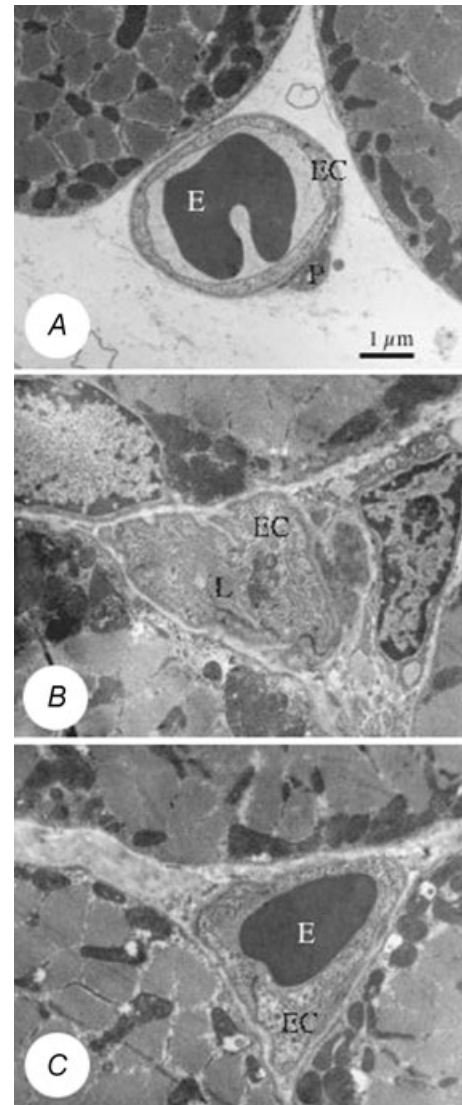


Figure 3. Changes in capillary fine structure with ischaemia A, electron micrograph illustrating the normal capillary phenotype from a control muscle showing the thin endothelium and smooth luminal and abluminal surfaces. B, a capillary with a swollen endothelium from a muscle 7 days after ligation of the iliac artery, where endothelial protrusions obscure the lumen. C, a capillary with a swollen endothelium from a muscle after 7 days ligation treated with amiloride, where the endothelial swelling is reduced sufficiently to allow likely passage of an erythrocyte. Abbreviations: E, erythrocyte; EC, endothelial cell; L, lumen; P, pericyte. Scale bar = $1.0 \mu\text{m}$.

Table 2. Morphometric analysis of the capillary response to ischaemia and imposed activity in rat EDL

	Control	3d lig	7d lig	35d lig	3d stim	3d SL
Cap perim (μm)	16.33 \pm 0.27	16.89 \pm 0.20	17.37 \pm 0.70	17.82 \pm 1.04	16.34 \pm 0.50	18.70 \pm 0.54*†
S/V (capillary) (μm^{-1})	2.59 \pm 0.32	2.57 \pm 0.17	2.02 \pm 0.08*	2.36 \pm 0.15	2.19 \pm 0.08	2.12 \pm 0.07
Endo area (μm^2)	7.52 \pm 0.87	7.82 \pm 0.31	9.90 \pm 0.72*	8.58 \pm 1.91	8.63 \pm 0.40	10.45 \pm 0.72*
Lum area (μm^2)	6.92 \pm 0.23	8.57 \pm 0.69*	6.31 \pm 0.28	8.56 \pm 1.48	4.73 \pm 0.36*	6.93 \pm 0.69†
Vv(endo,cap)	0.457 \pm 0.021	0.496 \pm 0.024	0.611 \pm 0.015**	0.479 \pm 0.072	0.660 \pm 0.024**	0.612 \pm 0.017**
Lum perim (μm)	13.96 \pm 0.20	15.37 \pm 0.36	14.03 \pm 0.51	15.31 \pm 0.81	13.60 \pm 0.54	15.68 \pm 0.40*†
Sv(lum,cap) (μm^{-1})	1.068 \pm 0.077	1.079 \pm 0.065	0.953 \pm 0.059	1.025 \pm 0.087	1.097 \pm 0.029	0.988 \pm 0.071

Abbreviations: Cap, capillary; perim, perimeter; S/V, surface to volume ratio; Endo, endothelium; Lum, lumen; Vv, volume density; Sv, surface density. * $P < 0.05$ ** $P < 0.001$ versus control; † $P < 0.05$ versus 3 days stimulation.

contractions ($6.73 \pm 0.84 \text{ ml min}^{-1} (100 \text{ g})^{-1}$ versus $4.06 \pm 0.50 \text{ ml min}^{-1} (100 \text{ g})^{-1}$, respectively, $P < 0.05$). There was no difference in MBF in contralateral limbs at rest, with only a slight improvement in contraction-induced hyperaemia (data not shown). The peak tension in the stimulated ischaemic muscles was lower than with ligation alone, but muscle stimulation resulted in a significant improvement in the FI when compared to the 3 days ligated animals without stimulation ($P < 0.05$), albeit with a lower peak tension (Fig. 1). The contra-

lateral muscles of animals receiving muscle stimulation in conjunction with 3 days of ligation showed no significant differences in either peak tension or in fatigability (data not shown).

Discussion

This study shows that increased activity during recovery of chronically ischaemic skeletal muscles parallels changes in microvasculature, demonstrated as swelling of capillary

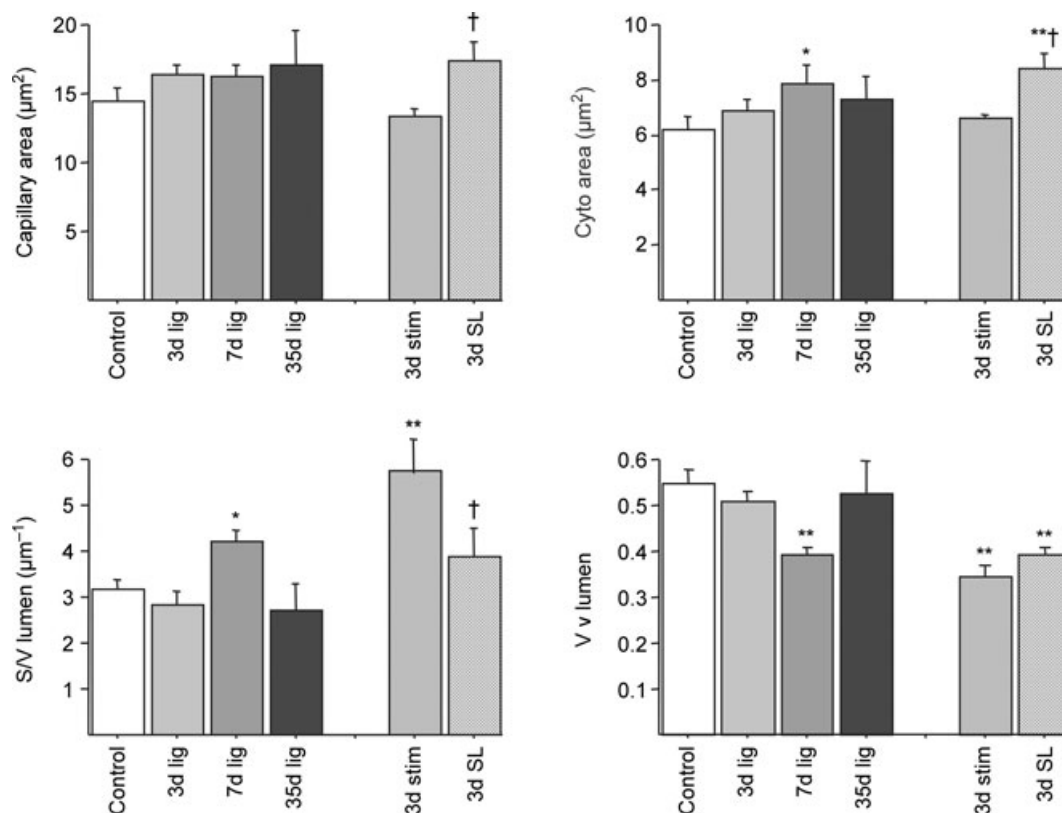


Figure 4. Capillary indices used to evaluate endothelial swelling on the basis of quantitative electron microscopy in control and ischaemic rat EDL

Abbreviations: cyto, endothelial cytoplasm; S/V, surface to volume ratio; Vv, volume density. * $P < 0.05$, ** $P < 0.01$ versus control; † $P < 0.05$ versus 3d stimulation alone.

Table 3. Relative capillary response in rat EDL to 7 days ligation, as a ratio of ischaemic to contralateral limb

	Untreated	Allopurinol	Amiloride	Vitamin E
SV(capillary)	0.91 ± 0.04	0.92 ± 0.03	0.90 ± 0.02	0.92 ± 0.06
Endo area	1.08 ± 0.11	0.83 ± 0.05†	0.90 ± 0.03†	1.34 ± 0.16
Lum area	1.25 ± 0.05	1.70 ± 0.26†	1.62 ± 0.19†	1.12 ± 0.12
Cyto:Lum	0.92 ± 0.09	0.54 ± 0.10*†	0.56 ± 0.06*†	1.14 ± 0.14

* $P < 0.05$ versus untreated; † $P < 0.05$ versus vitamin E. Cyto, endothelial cytoplasm

endothelium and increased rolling and adhesion of leukocytes in venules, which are attenuated by use of the xanthine oxidase inhibitor allopurinol or the Na^+/H^+ exchange inhibitor amiloride. The addition of muscle activity by electrical stimulation accentuated the structural changes (this study), and also increased leukocyte adhesion (Anderson *et al.* 2006). An important question is how these two responses may be linked. Various agents liberated from ischaemic tissues could explain increased capillary permeability and leukocyte adhesion (Ley, 1992), the latter induced primarily by changes in venules followed by modification of the capillary

endothelium (Mayrovitz *et al.* 1987), as described during inflammatory reactions in the rat cremaster (Joris *et al.* 1992). Subsequently, increased leukocyte rolling and adhesion would increase the resistance to flow in capillaries (Harris & Skalak, 1993) and accentuate ischaemia-induced hypoxia, leading to capillary endothelial swelling.

Muscle performance and blood flow

There was a striking reduction in peak tension (i.e. tension within 1–20 s after the beginning of muscle contractions) in ischaemic muscle, which we reason is due

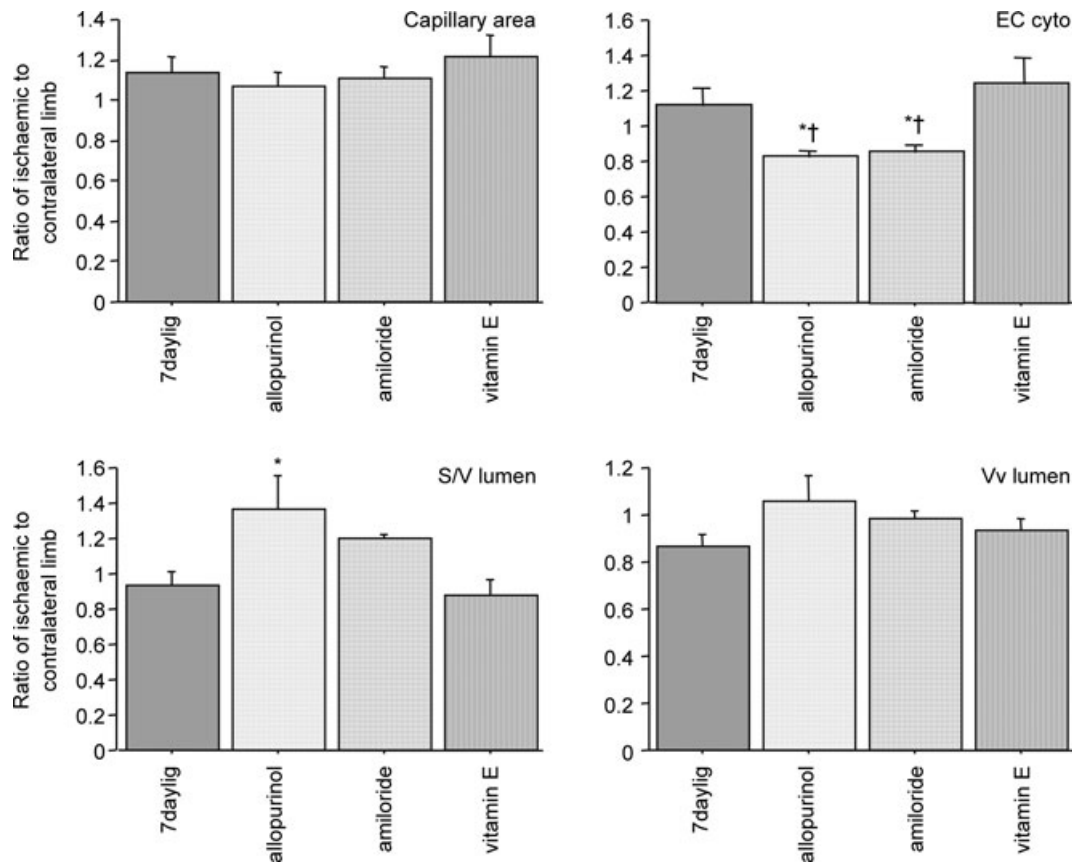


Figure 5. Several parameters used for evaluation of capillary endothelial swelling expressed as a ratio between the values for contralateral and ischaemic muscles following drug treatment

* $P < 0.05$ versus control; † $P < 0.05$ versus vitamin E.

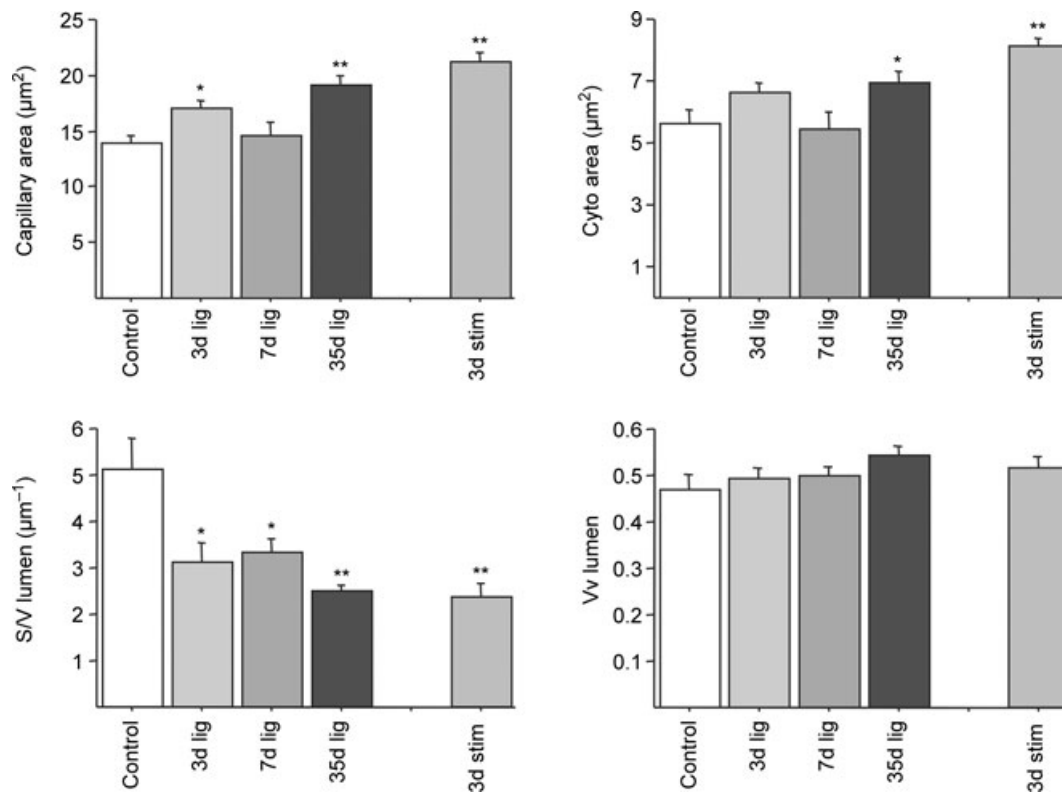
Table 4. Morphometric analysis of the capillary response to ischaemia in rat papillary muscle

	Control	3d lig	7d lig	35d lig	3d stim
Cap perim (μm)	21.30 \pm 1.69	18.94 \pm 0.65	17.96 \pm 0.81*	21.56 \pm 0.68	21.33 \pm 0.55
S/V(capillary) (μm^{-1})	3.357 \pm 0.061	2.637 \pm 0.040*	2.890 \pm 0.102*	2.867 \pm 0.192*	2.527 \pm 0.090**
Endo area (μm^2)	7.18 \pm 0.59	8.50 \pm 0.34	7.10 \pm 0.63	8.70 \pm 0.78	10.26 \pm 0.48**
Lum area (μm^2)	6.72 \pm 0.57	8.46 \pm 0.22	7.43 \pm 0.88	10.35 \pm 0.46*	10.85 \pm 1.05**
Vv(endo,cap)	0.541 \pm 0.037	0.529 \pm 0.022	0.512 \pm 0.027	0.474 \pm 0.009	0.495 \pm 0.024
Vv(cyto,cap)	0.423 \pm 0.029	0.413 \pm 0.017	0.393 \pm 0.021	0.378 \pm 0.007	0.391 \pm 0.019
Lum perim (μm)	20.97 \pm 0.86	17.49 \pm 0.74*	16.75 \pm 1.34*	22.65 \pm 1.20	19.36 \pm 0.52
Sv(lum,cap) (μm^{-1})	1.624 \pm 0.108	1.148 \pm 0.049**	1.258 \pm 0.092*	1.240 \pm 0.019*	0.980 \pm 0.054**

Abbreviations as per Table 2. * $P < 0.05$ ** $P < 0.001$ versus control; † $P < 0.05$ versus 3 days stimulation.

to a lower content of glycogen, and possibly ATP or phosphocreatine. Chronic stimulation leads to depletion of glycogen with very slow repletion (Maier & Pette, 1987), and this repletion is less complete in ischaemic muscles (Hudlická *et al.* 1994). The content of ATP in muscles at rest that were chronically stimulated was lower than in controls (Hudlická *et al.* 1986), and ATP content in resting, stimulated ischaemic muscles was lower than in untreated ischaemic muscles (Elander *et al.* 1985). Interestingly, the systemic effect of muscle ischaemia was seen in a reduction in tension developed in the contralateral limbs, although the cause of this is unknown.

We have shown previously (Milkiewicz *et al.* 2006) that blood flow was reduced to 60% of control values ($7.5 \pm 0.9 \text{ ml } (100 \text{ g})^{-1} \text{ min}^{-1}$) in muscles ischaemic for 7 days, with a gradual return towards control values after 35 days. A similar reduction of blood flow to that seen after 7 days was observed after 3 days. Mild chronic stimulation of these muscles increased blood flow at rest and during contractions relative to ischaemia alone, and improved fatigue resistance without recovery of peak muscle tension (Fig. 1). It is unclear whether there is any direct relationship between peak tension and muscle blood flow, although both decline after ligation, but

**Figure 6. Capillary indices used to evaluate endothelial swelling in papillary muscle**

* $P < 0.05$, ** $P < 0.01$ versus control.

muscle perfusion is more likely to dictate the relative endurance, indicated by the fatigue index. This includes 3 days ischaemia, when there is low MBF but capillary swelling has not yet occurred (see below). Indeed, the resistance to fatigue was related to MBF during contractions (Fig. 7). Indirect electrical stimulation is likely to have improved fatigue resistance because it improved blood flow, and it has been shown to restore vasoreactivity of arterioles which is lost in ischaemic muscle (Hudlická *et al.* 1994). It also decreased the proportion of capillaries with intermittent flow, but increased the time red cells spent stationary in individual capillaries, thus enabling greater extraction of oxygen (Anderson *et al.* 1997).

Improved MBF in this model is likely to involve some formation of collateral vessels, as demonstrated by measurements of perfusion pressure below the site of ligation in the femoral artery at its branch to the saphenous artery, i.e. well below the site of ligation. This dropped to 20% of the original value shortly after ligation, recovering by only 10% after 7 days but recovering to 50% of the original value 35 days after ligation (Hudlická *et al.* 1994).

Leukocyte adhesion

The changes in capillary fine structure were linked with increased leukocyte rolling and adhesion, although it is not clear to what extent events affecting capillaries would also alter properties of the venular endothelium (Mayrovitz *et al.* 1987). One possible link might be that low shear stress favours expression of some adhesion molecules in tissue cultures (Ando *et al.* 1994; Tsou *et al.* 2008). In addition, low shear stress in venules may induce aggregation of red blood cells, thus forcing leukocytes towards the vessel wall (Nazziola & House, 1992) favouring increased rolling and adhesion. However, Nazziola & House (1992) consider the most important factor enabling leukocyte adhesion in venules (as opposed to arterioles) to be the different quality of venular endothelium. This may be modified by chronic ischaemia and explain greater adhesion of leukocytes in patients with peripheral vascular diseases, which was accentuated by treadmill exercise (Ciuffetti *et al.* 1994). Indeed, chronic ischaemia in conjunction with increased activity led to increased expression of ICAM-1 in small venules (Anderson *et al.* 2006). However, after 7 days stimulation alone there was an increase in leukocyte flux from 12.3 ± 4.6 to $34.9 \pm 6.3 \text{ min}^{-1}$, but without significant change in adhesion (JM. Dawson & O. Hudlická, unpublished observation). Similar data may be found in contralateral muscles (Anderson *et al.* 2006). The pH in chronically ischaemic muscles is reduced (Angersbach *et al.* 1988), while chronic stimulation increased the lactate content 3-fold (Elander *et al.* 1985). While such changes are unlikely to be responsible for the responses observed in contralateral muscles, they may modify leukocytes leaving

ischaemic muscles in the venular drainage (Anderson *et al.* 1997), providing a mechanism by which signals are transferred to the endothelium in non-ischaemic muscles.

Capillary fine structure

Capillary endothelial cell swelling occurs in acutely ischaemic muscles (Strock & Majno, 1969), in conjunction with reperfusion (Gidlöf *et al.* 1988) and during haemorrhage (in skeletal muscle, Mazzoni *et al.* 1992; in the mesentery Kretschmar & Engelhardt, 1994). It has been linked with increased permeability (Suval *et al.* 1987), which could be prevented by calcium entry blockers (Paul *et al.* 1990), suggesting that swelling may be due to increased Ca^{2+} influx, possibly mediated by accumulation of adenosine in ischaemic muscles. Muscle ischaemia results, of course, in acidosis that is increased during muscle activity, and could induce increased sodium entry into the cells as suggested by Mazzoni *et al.* (1992, 1994). The fact that amiloride decreased swelling in ischaemic muscles favours the second explanation. Moreover, inhibition of Na^+/H^+ exchanger decreased injury in the heart caused by reperfusion (Masereel *et al.* 2003). Amiloride and its analogues also decreased the swelling of cultured endothelial cells exposed to low pH (Hamada *et al.* 2000). Similar changes in capillary swelling and fatigue resistance may imply an additional resistance to tissue perfusion, due to vessel stenosis or blockade, and this may contribute to heterogeneity within the muscle at a finer scale than would be the case with alteration of individual arteriolar reactivity previously reported in ischaemic muscle. In particular, the swollen endothelial cells were shown to impair capillary perfusion: red blood cells spent more time stationary than flowing in individual capillaries 1 week after ligation, and the

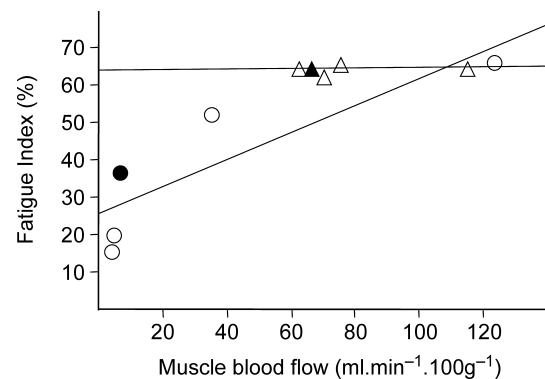


Figure 7. The relationship between muscle fatigue resistance and muscle blood flow during contractions

Mean values are given for groups of animals at different times after ligation. Circles, ipsilateral muscle; triangles, contralateral muscle. Filled symbols represent 3d SL. Linear regression of group means give $\text{FI} = 31.482 + 0.301 \times \text{MBF}$, $R^2 = 0.725$ (ipsilateral), and $\text{FI} = 63.645 + 0.008 \times \text{MBF}$, $R^2 = 0.017$ (contralateral).

proportion of capillaries with intermittent flow increased from 60% in control muscle to 85% 3 days after ligation, whereas stimulation of ischaemic muscles reduced this proportion to 70% (Anderson *et al.* 1997). In the absence of compensatory changes in autonomic tone, the shear stress-mediated NO response may be dominant. Although we know of no evidence for changes in ascending dilatation, modulation of autonomic tone and/or humoral regulators of vascular smooth muscle cells may influence the relative importance of our findings.

Interestingly, capillary endothelial swelling was also observed in muscle remote from the ischaemic ones, and it is unlikely that muscles such as spinotrapezius (Hickey *et al.* 1993) or the papillary muscle (this study; Egginton *et al.* 1993) would have lower pH as a result of ischaemia produced in the hindlimb. Although the degree of swelling was considerably smaller than in ischaemic muscles, it persisted for longer with the area of cytoplasm still elevated even 35 days after ligation, at which time capillaries in the ischaemic muscles were not different from controls. As white blood cells leaving the ischaemic limbs are morphologically modified (Anderson *et al.* 1997) they might conduct some so far unknown signals to capillaries in other parts of the body. Changes in papillary muscle may thus be an indicator of generalized changes resulting from muscle ischaemia, and patients suffering from limb ischaemia frequently experience altered coronary vasculature (Rothwell *et al.* 2005).

Amelioration of systemic injury

Some systemic effects of muscle ischaemia are observed (e.g. papillary muscle capillaries), but as we used systemic drug treatment the limitation of using contralateral muscles as reference may be similar to that using a separate set of animals, given the inherent biological variability and its effect on the sensitivity of cross-sectional studies. The issue of a reduced maximum tension in contralateral muscles may be related to spinal reflexes, as discussed elsewhere (Hudlická *et al.* 2003), but as the fatigue index did not change from that of control muscle (e.g. 65% at 7 days *versus* 66% in controls, Shiner, 1994), and capillary structure was largely unaffected, we consider the responses as described to be valid as relative, if not absolute, comparisons.

Reperfusion after acute ischaemia initiates a number of events that act as pro-inflammatory stimuli, including release of leukotriene B4 (Gute *et al.* 1998). A stable prostacyclin analogue (iloprost) which antagonises the effect of leukotriene B4 reduced leukocyte rolling and adhesion under these conditions (Thomson *et al.* 1994). However, these stimuli are unlikely to play a major role in the observed changes in microcirculation, as there were no overt signs of inflammation in our model of muscle ischaemia (Anderson *et al.* 2006).

Breakdown of high-energy phosphates in ischaemic muscles, particularly while contracting, results in the formation of hypoxanthine, which xanthine oxidase (XO) converts to xanthine and oxygen free radicals in the presence of oxygen during reperfusion (Carden & Granger, 2000). As XO is present in capillary endothelium in skeletal muscles (Hellstenwesting, 1993), this process may be responsible for the capillary endothelium swelling, as well as leukocyte adhesion. Allopurinol, an inhibitor of xanthine oxidase, attenuated swelling in acute reperfusion (Ferrari *et al.* 1996) and was as effective in our experiments as amiloride. However, its effect on leukocyte adhesion was less than that of amiloride. The improvement of microcirculation by allopurinol might explain the drugs beneficial effect on muscle function (McCutchan *et al.* 1990) and blood flow (Hardy *et al.* 1992) during acute reperfusion.

Local ischaemia may result in the release of endothelin 1 from hypoxic endothelium as described in cultured rat aortic endothelial cell (Bodin *et al.* 1992). Although there are no data on the release of endothelin in either peripheral vascular diseases, or in animal models of muscle ischaemia, increased plasma levels were found in coronary ischaemia resulting in myocardial infarction (Miyachi *et al.* 1989). Endothelin 1 is a potent activator of the Na^+/H^+ antiport (Khandoudi *et al.* 1994) and this could explain capillary endothelial cell swelling observed in the ischaemic muscles. Inhibitors of Na^+/H^+ antiport also decreased swelling of cultured endothelial cell exposed to low pH (Huck *et al.* 2007), and in endothelium in femoral arteries after ischaemia–reperfusion (Hamada *et al.* 2000).

The effect of allopurinol in this study is in agreement with those obtained by others in acute ischaemia and reperfusion. In contrast, data regarding efficacy of vitamin E under these conditions is varied and controversial, and in common with the literature we are unable to explain the lack of effect in our experiments. Vitamin E may prevent the vascular damage after acute reperfusion, as it inhibited superoxide production from rat macrophages (Hiramatsu *et al.* 1991) and in skeletal muscle (Novelli *et al.* 1997; Formigli *et al.* 1997), and improved walking distance in patients with peripheral vascular disease (Teoh *et al.* 1992). In contrast, we saw no attenuation of either endothelial swelling or leukocyte behaviour, in agreement with another report showing vitamin E had no effect on leukocyte adhesion (Lehr *et al.* 1995). Whereas we used scavengers to reduce the effect of ROS, in a review of complimentary ischaemia studies Gute *et al.* (1998) demonstrate leukocyte adhesion and endothelial permeability that was ameliorated by reduced ROS generation following xanthine oxidase inhibition. Similarly, Suzuki *et al.* (1995) used dimethylurea as a scavenger, but we have been unable to uncover any data that would allow a comparison of its influence on muscle performance with the scavengers used in this

study. Interestingly, damage was predominantly restricted to glycolytic fibres (Suzuki *et al.* 1995), consistent with their earlier glycogen depletion compared to oxidative fibres, and increased depletion in contracting ischaemic muscles (Hudlická *et al.* 1994).

Effect of activity

The period of recovery after ligation includes gradual restoration of blood flow and muscle metabolism, as well as recuperation of muscle activity. However, from extant data it is unclear whether the imposition of additional muscle activity may aid the recovery process. In spite of the changes in microcirculation caused by electrical stimulation in muscles 3 days after ligation of the iliac artery, stimulation improved muscle blood flow and performance assessed as fatigue resistance (though not peak twitch tension). This apparent discrepancy can be explained by reference to the regulation of muscle blood flow. Arterioles determine the amount of blood entering muscle and their dilatation is under the influence of metabolites released from skeletal muscle (e.g. lactic acid and adenosine). Interestingly, arterioles in ischaemic muscles do not dilate (Kelsall *et al.* 2004), but their dilatation was restored by chronic electrical stimulation (Hudlická *et al.* 1994). Muscle contractions result in increased shear stress and generation of NO which could help dilatation of arterioles but little, if anything, is known about its effect on the ultrastructure of microvessels, although it is known that it is essential in the prevention of leukocyte adhesion in cremaster muscle (Akimitsu *et al.* 1995). Although the NO response may be dominant, modulation of autonomic tonus and/or humoral regulators of vascular smooth muscle may influence the relative importance of these findings. Stimulation of non-ischaemic muscle did not cause such changes to the same extent, as the fatigue index was the same as controls at 3 days and 7 days in contralateral muscles, although it did increase in ipsilateral muscles despite the presence of capillary swelling (Egginton & Hudlická, 1999). Hence, while increased muscle activity imposes an additional insult on capillary structure to that elicited by ischaemia alone, this was outweighed by the beneficial effect on blood flow (Fig. 7). It also decreased the proportion of capillaries with intermittent flow, but increased the time red cells spent stationary in individual capillaries, thus enabling greater extraction of oxygen (Anderson *et al.* 1997).

Conclusions

This paper demonstrates that ischaemic muscle exposed to mild, ambulatory activity shortly after the ischaemic insult has impaired microcirculation function and structure (increased number of rolling and adhering leukocytes

in venules, and swollen capillary endothelium). These symptoms were alleviated by inhibition of the formation of oxygen free radicals with allopurinol or by the Na⁺/H⁺ channel blocker amiloride. However, an oxygen free radical scavenger, vitamin E, was not effective. Although the microcirculation showed signs of deterioration, additional activity imposed by electrical stimulation improved muscle blood flow and fatigue resistance, probably because it restores the dilatation of arterioles otherwise lost in ischaemic muscles. As activation of capillary endothelium may be one cause of leukocyte adhesion, the implication of this work is that combination treatment may attenuate impaired function of the microcirculation in working ischaemic muscles.

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Supplemental material

Online supplemental material for this paper can be accessed at: <http://jp.physoc.org/cgi/content/full/jphysiol.2008.158055/DC1>