

# Comparison of Two Steinernematid Species for Control of the Root Weevil *Diaprepes abbreviatus*

W. J. SCHROEDER<sup>1</sup>

**Abstract:** *Steinernema carpocapsae* Weiser All strain was compared to *Steinernema riobravisi* Cabanillas, Poinar, and Raulston for control of the root weevil, *Diaprepes abbreviatus* (L.), in the laboratory and in potted citrus. In the laboratory bioassay, *D. abbreviatus* larvae were exposed to 30, 60, and 120 nematodes/cm<sup>3</sup> in sand. Insect mortality 1 week after application was greater ( $P \leq 0.05$ ) for *S. riobravisi* than for *S. carpocapsae* in the laboratory bioassay. In the greenhouse bioassay, *D. abbreviatus* larvae were exposed to 3 and 9 nematodes per cm<sup>3</sup> of soil in potted citrus. Again, at each rate, mortality was greater ( $P \leq 0.05$ ) in pots treated with *S. riobravisi* than in pots treated with *S. carpocapsae*. The results of this study suggest that *S. riobravisi* is a better biological control agent against *D. abbreviatus* larvae in potted plants than *S. carpocapsae*.

**Key words:** biological control, citrus, *Diaprepes abbreviatus*, entomopathogenic nematode, nematode, *Steinernema carpocapsae*, *Steinernema riobravisi*, Steinernematidae.

The citrus root weevil complex consists of five species: the Fuller rose beetle, *Asynonychus godmani* (Crotch); the little leaf notcher, *Artipus floridanus* Horn; the citrus root weevils, *Pachnaeus litus* (Germar) and *P. opalus* (Oliver); and the sugarcane root-stalk borer weevil *Diaprepes abbreviatus* (L.). The life cycle of the five species is similar. Eggs are deposited in the canopy of the tree, and neonate larvae fall to the ground, enter the soil, and feed on roots. Major injury to citrus, sugarcane, ornamental plants, and vegetable crops in Florida and the Caribbean results from larval feeding damage to roots (5,11). *Diaprepes abbreviatus* is potentially the most destructive because it is the largest of the five species (11). Potted citrus is also a major concern to the industry and is considered one of the main methods for movement of *D. abbreviatus*.

Rhabditid nematodes of the family Steinernematidae are obligate parasites of insects that are characterized by a mutual-

istic relationship with *Xenorhabdus* spp. bacteria. They are lethal to a broad range of economically important insect pests (4,6). The nematode *Steinernema carpocapsae* (Weiser) has been evaluated for control of larvae of *D. abbreviatus* in Florida (9) and Puerto Rico (8). In one study, application of *S. carpocapsae* in the citrus grove reduced *D. abbreviatus*, *P. opalus*, and *P. litus* adult weevil emergence by 70% compared with the check (10). Subsequently, the commercial product BioVector®, containing the nematode *S. carpocapsae* All strain, was introduced as a biological agent for control of the citrus root weevil complex in Florida.

Recently, *Steinernema riobravisi* Cabanillas, Poinar, and Raulston (2) was isolated from the lower Rio Grande Valley in Texas. It is a parasite of the corn earworm, *Helicoverpa zea* (Boddie), and the fall armyworm, *Spodoptera frugiperda* (Smith) (7). This study compares *S. riobravisi* with *S. carpocapsae* as biological agents for control of the larvae of *D. abbreviatus* under controlled conditions.

## MATERIALS AND METHODS

**Nematodes:** *Steinernema carpocapsae* All strain and *S. riobravisi* were obtained from Biosys, Palo Alto, California. Additional nematode generations were produced from infected *D. abbreviatus* larvae using the method described by Dutky et al. (3).

**Weevil larvae:** *D. abbreviatus* were reared on diet (1). The average weight of the

Received for publication 2 August 1993.

<sup>1</sup>Research Entomologist, USDA ARS SAA, U.S. Horticultural Research Laboratory, 2120 Camden Road, Orlando, FL 32803.

The author thanks Manda Price for her study on the comparison of *Steinernema riobravisi* and *S. carpocapsae* as a biological control agent for the Apopka weevil, Science Fair, Oviedo High School, 1993.

The author gratefully acknowledges critical review of the manuscript by W. R. Martin, W. R. Nickle, and R. G. Bullock.

Mention of a trademark, warranty, proprietary product, or vendor does not constitute a guarantee by the U.S. Department of Agriculture and does not imply its approval to the exclusion of other products or vendors that may also be suitable.

3-month-old larvae was 0.516 g (range 0.310–0.901 g).

**Laboratory bioassay:** Each 3.5-cm-d bioassay cup contained 16 cm<sup>3</sup> (25 g) of sterile dry Astatula fine sand (hyperthermic, uncoated typic quartzsammments). The moisture of the sand was adjusted to 10% wt/wt with deionized water. A single *D. abbreviatus* larvae was placed in each cup. Nematodes were added at 30, 60, and 120 per cm<sup>3</sup> of sand, and a slice of carrot was placed on the sand as food for the larvae. The cups were maintained at a temperature of 26 C for 1 week. After 1 week, all weevil larvae were dissected to confirm nematode infection. There were 10 cups per replication and eight replications per treatment. A bioassay was conducted simultaneously without food to determine if this was a factor in nematode infection. Also, comparison of the efficacy of F1 and F2 generations of *S. carpocapsae* and *S. riobravivis* was done to eliminate variables in shipping, formulation, and storage that might have affected each nematode species differently.

**Potted plant bioassay:** Sour orange *Citrus aurantium* (L.) seedlings were established in 15-cm-d pots with 2 liters of soil. The potting soil media was three parts Florida peat and one part coarse builder's sand (v/v). Ten *D. abbreviatus* larvae were placed 5 cm below the soil in each pot. After 2 weeks, nematodes were added to the pots at the rate of three and nine nematodes/cm<sup>3</sup> of soil. The study was conducted from October through March with ambient weather conditions (5–28 C), and plants were watered once a week. The soil was removed from the pot after 2–4 weeks and the number of live larvae determined. There were 20 plants per treatment for a total of 100 plants.

Arcsine-transformed data were subjected to an analysis of variance (ANOVA) and means were separated by a Student-Newman-Keuls multiple-range test.

## RESULTS AND DISCUSSION

**Bioassay:** The results of the laboratory study comparing *S. riobravivis* with *S. carpo-*

TABLE 1. Mortality of *Diaprepes abbreviatus* caused by *Steinernema riobravivis* or *S. carpocapsae* with and without food in a laboratory bioassay.

Nematode species and treatment	% mortality (nematodes per cm <sup>3</sup> )				
	0	30	60	120	Total
<i>S. riobravivis</i>					
No food	02	60	57	75	63 a
Food	03	57	63	60	60 a
<i>S. carpocapsae</i>					
No food	03	13	22	35	26 b
Food	02	32	48	27	36 b

Means within the same column followed by the same letter are not significantly different ( $P \geq 0.05$ ; Student-Newman-Keuls multiple-range test).

*capsae* at 30, 60, and 120 nematodes/cm<sup>3</sup> are shown in Table 1. More *D. abbreviatus* larvae were killed by *S. riobravivis* than by *S. carpocapsae* at each of the three rates tested ( $P \leq 0.05$ ). In the bioassay that was conducted simultaneously with or without food, it was determined that food was not a factor in infection of weevil larvae. Apparently, nematodes that were consumed with the carrot did not affect ( $P \leq 0.05$ ) the mortality of the larvae. Therefore, food was not included in the laboratory bioassay when nematode generations were compared.

When the parent, F1, and F2 generations of *S. riobravivis* and *S. carpocapsae* were compared, mortality by *S. riobravivis* was different ( $P \leq 0.05$ ) from *S. carpocapsae* (Table 2). This difference in activity indicates

TABLE 2. Mortality of *Diaprepes abbreviatus* caused by *Steinernema riobravivis* or *S. carpocapsae* in a laboratory bioassay.

Nematode species and generation	% mortality (nematodes per cm <sup>3</sup> )				
	0	30	60	120	Total
<i>S. riobravivis</i>					
Parent	0	43	50	50	48 a
F1	0	78	68	80	75 b
F2	7	55	63	80	66 b
<i>S. carpocapsae</i>					
Parent	5	20	23	25	23 c
F1	0	20	35	48	34 c
F2	0	30	48	20	33 c

Means within the same column followed by the same letter are not significantly different ( $P \geq 0.05$ ; Student-Newman-Keuls multiple-range test).

TABLE 3. Mortality of *Diaprepes abbreviatus* larvae caused by *Steinernema riobravisi* or *S. carpocapsae* in potted citrus.

Nematode species	Nematodes per cm <sup>3</sup>	Larvae per plant		% mortality
		Mean	Range	
<i>S. riobravisi</i>	3	1.4	0-5	86 a
	9	2.3	0-7	77 a
<i>S. carpocapsae</i>	3	6.8	2-10	32 bc
	9	5.8	2-10	42 b
Check	0	7.8	5-10	23 c

Means within the same column followed by the same letter are not significantly different ( $P \geq 0.05$ ; Student-Newman-Keuls multiple-range test).

that *S. riobravisi* is a more virulent biocontrol agent compared with *S. carpocapsae*.

**Potted plant bioassay:** This study evaluated treatment effects under field conditions for plants with soil attached to the roots. Mortality of *D. abbreviatus* larvae in plants treated with *S. riobravisi* was greater ( $P \leq 0.05$ ) than for plants treated with *S. carpocapsae* (Table 3). Mortality of larvae in the check plants was apparently due to cannibalism by other larvae.

The results of this study suggest that the entomopathogenic nematode, *S. riobravisi*, is a more effective biological control agent against *D. abbreviatus* larvae than *S. carpocapsae*. This was evident in the laboratory bioassay and in potted citrus. Citrus was the larval host used in this study; however, the data should apply to other potted plant species that are infested with *D. abbreviatus*.

#### LITERATURE CITED

1. Beavers, J. B. 1982. Biology of *Diaprepes abbreviatus* (Coleoptera: Curculionidae) reared on artificial diet. *Florida Entomologist* 65:263-269.
2. Cabanillas, H. E., G. O. Poinar, Jr., and J. R. Raulston. 1994. *Steinernema riobravisi* n. sp. (Rhabditiidae:Steinernematidae) from Texas. *Fundamental and Applied Nematology* 17:123-131.
3. Dutky, S. R., J. V. Thompson, and G. E. Cantwell. 1964. A technique for the mass propagation of the DD-136 nematode. *Journal of Insect Pathology* 6:471-472.
4. Gaugler, R., and H. K. Kaya, eds. 1990. Entomopathogenic nematodes in biological control. Boca Raton, FL: CRC Press.
5. Martorel, L. F. 1976. Annotated food plant catalog of the insects of Puerto Rico. Puerto Rico: Agricultural Experiment Station, University of Puerto Rico.
6. Poinar, G. O., Jr. 1971. Use of nematodes for control of insects. Pp. 181-203 in H. D. Burges and N. W. Hussey, eds. *Microbial control of insects and mites*. London: Academic Press.
7. Raulston, J. R., S. D. Pair, J. Loera, and H. E. Cabanillas. 1992. Prepupal and pupal parasitism of *Helicoverpa zea* and *Spodoptera frugiperda* (Lepidoptera: Noctuidae) by *Steinernema* sp. in cornfields in the lower Rio Grande Valley. *Journal of Economic Entomology* 85:1666-1670.
8. Roman, J., and W. Figueroa. 1985. Control of the sugarcane rootstalk borer weevil *Diaprepes abbreviatus* (L.) with entomogenous nematode *Neoplectana carpocapsae* Weiser. *Journal of Agriculture, University of Puerto Rico* 69:153-158.
9. Schroeder, W. J. 1987. Laboratory bioassays and field trials of entomogenous nematodes for control of *Diaprepes abbreviatus* (Coleoptera: Curculionidae) in citrus. *Environmental Entomology* 16:987-989.
10. Schroeder, W. J. 1992. Entomopathogenic nematodes for control of root weevils of citrus. *Florida Entomologist* 75:563-567.
11. Woodruff, R. E. 1964. A Puerto Rican weevil new to the United States (Coleoptera: Curculionidae). *Florida Department of Agriculture, Division of Plant Industry, Entomology Circular* 30:1-2.