Population Changes in *Heterodera glycmes* **and Its Bacterial Parasite** *Pasteuria* **sp. in Naturally Infested Soil 1**

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Abstract: A two-year soil sampling study was conducted on four microplots naturally infested with *Heterodera glycines* and an undescribed species of *Pasteuria*. The objectives of the study were to investigate the population dynamics of both organisms and to assess the potential of *Pasteuria* sp. as a biological control agent of *H. glycines.* Seasonal fluctuations were observed in numbers of cysts, eggs per cyst, second-stage juveniles (I2) of *H. glycines*, number of *Pasteuria* endospores attached per I2, and percentages of endospore-encumbered J2. Percentages of endospore-encumbered J2, Y, increased with the mean numbers of endospores per J2, X, according to the equation $Y = 87.0(1 - e^{-0.53X})$. In contrast, numbers of J2 per 250 cm³ soil, Y, decreased with the numbers of endospores per J2, X, according to the exponential decay model $Y = 67.4 + 220.1e^{-1.2A}$. The equilibrium J2 density (67.4 \pm 3.3) derived from this function was consistent with the predictions of the Lotka-Volterra model of population dynamics based on the equation $0.0195\ln(y) - 0.000336y = 0.000049x - 0.00285\ln(x) + 0.06589$, where x and y represent the biweekly means of [2 densities and the percentages of endospore-encumbered [2, respectively. In all cases, predicted equilibrium densities of J2 were below the damage threshold reported from field studies. These results indicate that, given sufficient time following introduction into a field, *Paxteuria* may increase to levels that would be effective as one component in an integrated pest management program to control *H. glycines.*

Key words: biological control, *Glycine max, Heterodera glycines,* modeling, nematode, *Pasteuria,* population dynamics, soybean, soybean cyst nematode.

The soybean cyst nematode, *Heterodera glycines* Ichinohe, is the most economically damaging pathogen of soybean, *Glycine max* (L.) Merr., in the United States (Niblack, 1993). In the north-central and southern regions, the loss in average annual production is estimated at 1.3 million metric tons (\$279.5 million) and 0.6 million metric tons (\$127.7 million), respectively (Doupnik, 1993; Wrather et al., 1995). Planting resistant cultivars and cultural practices are the tactics used most often to manage the soybean cyst nematode. However, persistent

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crop losses indicate that additional management strategies are needed.

Species of *Pasteuria,* a gram-positive, mycelial, and endospore-forming bacterium of the order Actinomycetales (Starr and Sayre, 1988), are promising candidates for the biological control of several plant-parasitic nematodes (Brown et al., 1985; Chen et al., 1996, 1997; Nishizawa, 1987; Weibelzahl-Fulton et al., 1996). In spite of this potential, large-scale exploitation of *Pasteuria* spp. as biological control agents of nematodes has not been accomplished due to the lack of suitable procedures for in vitro mass cultivation (Bishop and Ellar, 1991; Williams et al., 1989) and to limited knowledge of the population dynamics of these parasites and the relationships with host populations in nature. Some studies have related low nematode densities to the suppressive action of *Pasteuria* (Bird and Brisbane, 1988; Oostendorp et al., 1991). However, the bacterium was associated with high nematode densities in one instance (Spaull, 1984), and in other instances no direct effect of *Pasteuria* was observed despite high percentages of infected nematodes (Ciancio et al., 1992; Giblin-Davis et al., 1990). Rates of parasitism were correlated with temporal fluctuations of the nematode population in only one of two

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kiwi orchards infested with both *Meloidogyne* spp. and *P. penetrans* (Thorne) Sayre & Starr (Verdejo-Lucas, 1992). In Italy, where parasitism of *Xiphinema diversicaudatum* (Micoletzky) Thorne by *P. penetrans* was investigated, nematode densities and rates of parasitism remained constant over time (Ciancio, 1995). Clearly, more data are needed to unveil the mechanisms that govern the *Pasteuria-nematode* interrelationships in nature. Understanding these mechanisms will assist in implementing biological control of nematodes as a component of integrated pest management.

An undescribed species of *Pasteu'ria* that infects *H. glycines* has been reported in North America (Noel and Stanger, 1994). Preliminary observations have shown that this isolate is not *P. nishizawae* Sayre, Wergin, Schmidt & Starr, the only species previously known to infect *H. glycines* (Sayre et al., 1991). The objectives of this study were to investigate the population dynamics of both *H. glycines and Pasteuria* sp. and to assess the potential of *Pasteuria* sp. as biological control agent of the soybean cyst nematode.

MATERIALS AND METHODS

During the 1994 and 1995 growing seasons, experiments were established in four 5-m x 5-m microplots on the USDA hematology farm at Urbana, Illinois. Microplots were infested for several years before initiation of the experiments with *H. glycines, two* with race 3 and two with race 4. *Pasteuria* sp. was present in all four microplots. The soil was a series Watseka (sandy, mixed, mesic Aquic Hapludolls), with the upper 20 cm containing 73.8% to 79.0% sand, 10.8% to 18.0% silt, 7.0% to 11.4% clay, 4% organic matter, and with pH ranging from 5.9 to 6.4. Microplots were planted with *the H. glycines*susceptible soybean cultivar Williams 82 on 15 May 1994 and 31 May 1995. Two soil samples, each consisting of 30 1.7-cm-diam. x 15 to 20-cm-deep cores, were collected randomly every 2 weeks from each microplot from 10 May to 4 October 1994 and from 5 April to 26 October 1995. White females and cysts of *H. glycines* (collectively referred to as cysts) were extracted from 250 cm³ soil with gravity sieving (Cobb, 1918) onto a 180-um-pore sieve. Second-stage juveniles (J2) and males of *H. glycines* were extracted from the same soil suspension with centrifugal flotation (Jenkins, 1964) and collected on a 38-µm-pore sieve. Cysts were crushed in a tissue grinder, and the numbers of eggs were determined with a Hawksley counting slide (Hawksley and Son, Lancing, England). All J2 and males of H. *glycines* were selected individually with the help of a stereomicroscope, mounted on temporary slides in 2.5% formalin, and examined under a compound microscope (x400) to determine the number *of Pasteuria* endospores attached to each nematode.

Poisson (PROC GENMOD, SAS Institute, Cary, NC) and negative binomial (LIMDEP, Econometric Software, Bellport, NY) models were used to estimate the frequency distribution of the numbers of endospores per J2. Both models involved three dummy variables (race, block, and year) and one continuous variable (date, expressed as number of weeks from the beginning of each sampling season) as covariates. The number of $J2$ from each 250 -cm³ soil sample was the exposure variable. Numbers of cysts, eggs per cyst, J2, endospores per J2, and percentages of endospore-encumbered J2 in each 250 -cm³ soil sample were analyzed with PROC GLM (SAS Institute, Cary, NC) for a split-split plot design with the race of *H. glycines* as the main plot, year as the split-plot, and sampling date as the split-split-plot. The biweekly means of each of the factors were interpolated with a cubic spline method (SigmaPlot, SPSS, Chicago, IL). A randomization test (Pollard and Lakhani, 1987) was used to detect density-dependent trends in the biweekly fluctuations of numbers of J2, endospores per J2, and percentages of endospore-encumbered J2. The test was based on Pearson's correlation coefficients, $r_{\rm xd}$, between $x_{ii} = \ln(X_{ii})$ and the corresponding differences, $d_{ii} = x_{ii+1} - x_{ii}$; where X_{ii} = the mean of variable X on date i and year j, $i =$ $1, \ldots, n_j - 1; j = 1, 2;$ and $n_j =$ the number of sampling dates in year j . Since the time series was discontinued from 4 October

1994 to 5 April 1995, the test was applied separately to the data from each year. However, when the date \times year interaction was not significant, the \overline{X}_{ii} were pooled over years. Conversely, when the race \times year interaction was significant, the test was carried out on the data from each race × year combination. Each run of the test consisted of 2,000 simulations (Marriot, 1979) performed with a random number generator and a personal computer. Regression analyses also were conducted with nonlinear curve-fitting procedures PROC NLIN (SAS Institute, Cary, NC) and TableCurve (SPSS, Chicago, IL). Two predator-prey models of population dynamics were estimated from the biweekly means of J2 densities and the percentages of endospore-encumbered J2. The two models included the Lotka-Volterra model (Lotka, 1925; Volterra, 1926) and a modification of the Nicholson-Bailey model (Nicholson and Bailey, 1935) proposed by Hassel and May (1973). The considerations for the use of the Lotka-Volterra model to describe the *Pasteuria-nematode* interrelationship, the estimation procedure, and the biological significance of each of the parameters involved were outlined by Ciancio (1995). Specifically, the differential equation,

$$
a\ln(y) - by = cx - d\ln(x) + k \qquad (1),
$$

was estimated from the system of equations,

$$
x_{t+1} = x_t + ax_t - bx_t y_t \tag{2}
$$

and

$$
y_{t+1} = y_t + cx_t y_t - dy_t \qquad (3),
$$

where x_t and y_t are the biweekly means of J2 densities and the percentages of endosporeencumbered $[2]$ at time t, respectively; whereas *a, b, c, d,* and k are the model parameters. On a per-capita basis, a represents the host growth rate in the absence of other limiting factors, b accounts for the host decrease due to parasitism, c is the parasite growth rate in relation to the host density, d indicates the parasite death rate, and k is an integration constant. For the Nicholson-Bailey model, the host-parasite interrelationship was represented by the system of equations,

$$
x_{t+1} = \lambda (1 - \gamma) x_t + \lambda \gamma x_t e^{-\alpha Y_t} \tag{4}
$$

and

$$
y_{\ell+1} = \gamma x_{\ell} (1 - e^{-\alpha Y_{\ell}})
$$
 (5),

under the assumption that only a proportion, γ , of $[2 \text{ may contact endospores at a}]$ given generation, t, with a probability α (Hassel, 1978; Hassel and May, 1973). The parameter λ represents the host rate of increase. The two models were fitted with Mathcad computer software (MathSoft, Cambridge, MA) by iteratively assigning values to the parameters and the initial values x_0 and y_0 so as to maximize the correlation between the observed and the interpolated values of y at each of the observed x values. Student's t-test was used to compare the observed and the predicted percentages of endospore-encumbered J2.

RESULTS

During the 2 years of experimentation, 9,606 J2 and a few males of *H. glycines* were examined for the attachment of *Pasteuria* endospores. On average, 64.4% of the J2 had endospores adhering to their cuticle, but none showed evidence of internal proliferation of the parasite. The highest J2 densities and the lowest rates of endospore attachment were observed in one of the microplots infested with race 3 of *H. glycines* (Table 1).

Temporal fluctuations of the H. glycines population: Over both years, population densities of *H. glycines* ranged from 0 to 41 cysts, 0 to 262 eggs/cyst, and 2 to 552 J2/250 cm^3 soil. Means and standard errors of the means were 7.9 ± 0.6 , 89.5 ± 4.4 , and 57.2 ± 7.1 , respectively. The effect of sampling dates was significant ($P \le 0.05$) for the numbers of cysts (Table 2). Specifically, mean numbers of cysts declined from 19.8 ± 4.2 on 10 May to 4.8 ± 0.6 on 1 September 1994 (Fig. 1). A similar trend was observed in 1995 when numbers of cysts declined from $12.0 \pm$ 3.9 on 25 April to 2.3 ± 0.6 on 21 September, although they subsequently increased to 8.6

TABLE 1. Mean numbers of *Heterodera glycines* cysts, eggs per cyst, second-stage juveniles (J2), *Pasteuria* endospores per $[2]$, and percentages of endospore-encumbered $[2]$ per 250 cm^3 of naturally infested microplot soil.²

^a Two soil samples were collected at 2-week intervals from each of four $5-m \times 5-m$ microplots, from 10 May to 4 October 1994 and from 5 April to 26 October 1995, resulting in 18 and 24 samples per microplot in 1994 and 1995, respectively.

 \pm 4.3 by the end of the season. For mean numbers of cysts, there were no significant differences between race 3 (7.9 \pm 0.8) and race 4 (7.9 \pm 0.9), or between 1994 (10.1 \pm 1.0) and 1995 (6.2 ± 0.7) .

Mean numbers of eggs per cyst decreased gradually from 133.1 ± 26.3 on 10 May to 69.9 ± 10.8 on 7 July 1994, then increased and peaked at 120.3 ± 15.4 on 19 September 1994 (Fig. 1). In 1995, the numbers of eggs per cyst oscillated with a decreasing amplirude and a regular periodicity of about 1 month from 5 April (153.6 ± 24.6) through mid-August (49.3 \pm 14.9). Thereafter, the numbers of eggs per cyst increased and peaked at 101.3 ± 15.5 on 6 October. The effect of sampling dates was not significant $(P > 0.05)$ for the numbers of eggs per cyst, nor were there significant differences between race 3 (99.4 \pm 6.6) and race 4 (79.2 \pm 5.7) or between $1994 (102.5 \pm 6.3)$ and 1995 $(79.2 \pm 6.0).$

 $a \ln(x + 1)$.

.

c Effect nested within those enclosed by brackets.

FIG. 1. Numbers of cysts and eggs per cyst of *Heterodera glycines* per 250 cm³ of naturally infested microplot soil. Each observation represents the mean of eight samples, and points are connected with a cubic spline curve.

Numbers of $]2$ peaked at 157.3 ± 73.3 on 25 July 1994, and at 130.4 ± 74.1 on 21 September 1995 (Fig. 2). Other less pronounced peaks were observed on 12 May (65.6 ± 11.1) and 5 July (68.8 ± 11.8) of the second year. However, the analysis of variance (Table 2) indicated that none of the effects of sampling dates, race (\overline{X} = 80.8 \pm 13.3 and 33.5 ± 3.0 for races 3 and 4, respectively), and year (\overline{X} = 50.6 \pm 9.8 and 62.1 \pm 9.9 for 1994 and 1995, respectively) was significant ($P > 0.05$) for numbers of J2. Also, no density-dependent trend was detected in the fluctuations of J2 numbers either in 1994 ($P = 0.417$) or in 1995 ($P = 0.156$), although the likelihood of density-dependence was significant ($P = 0.016$) when mean numbers of J2 were pooled over years.

Attachment of Pasteuria endospores: The numbers of *Pasteuria__endospores* per J2 ranged from 0 to 133 ($X = 3.9 \pm 0.1$), and the percentages of endospore-encumbered J2 ranged from 0 to 100% (\bar{X} = 64.4% ± 2.3). The frequency distribution of the numbers of endospores per J2 was best described by a negative binomial distribution (expected mean $\mu = 2.1$ and clumping index $\theta =$ 0.2384) rather than a Poisson distribution $(X^2 = 403,037$ and 1,070,200, respectively, for the saturated models). Based on the negative binomial distribution, the probability of a J2 being encumbered with at least one *Pasteuria* endospore was $P = 0.42$ (Fig. 3). However, the numbers of endospores per J2 were significantly affected by the race of *H. glycines,* the year, the date, and the interactions among these factors (results not shown). Specifically, the marginal effect of race on the expected mean number of endospores per J2 was an increase of 9.6 for race 4 (μ = 10.8) compared to race 3 (μ = 1.2), and the probability of having at least one endospore per J2 was 25% higher for race 4 ($P = 0.60$) than for race 3 ($P = 0.35$).

FIG. 2. Numbers of second-stage juveniles (J2) of *Heterodera glycines* and of *Pasteuria* endospores per J2, and percentages of endospore-encumbered J2 per 250 cm³ of naturally infested microplot soil. Each observation represents the mean of eight samples, and points are connected with a cubic spline curve.

Similarly, the likelihood of having at least one endospore per J2 was higher in 1995 (P $= 0.59$) than in 1994 ($P = 0.45$), resulting in a greater expected mean number of endospores per J2 in 1995 (μ = 9.5) than in 1994 $(\mu = 2.6)$.

The analysis of variance (Table 2) also revealed significant $(P < 0.001)$ seasonal fluctuations in the numbers of endospores per J2, and the percentages of endospore-encumbered J2. Fluctuations in numbers of endospores per J2 were affected by year, whereas both year and race of *H. glycines* had a significant effect on the fluctuations of the percentages of endospore-encumbered J2. In 1994, the numbers of endospores per J2 declined progressively from 5.3 ± 0.5 on 27 May to 0.7 ± 0.1 on 25 July, and thereafter increased to 3.6 ± 1.0 on 1 September (Fig. 2). In contrast, the numbers of endospores per J2 increased markedly during the first half of the 1995 growing season, from $3.0 \pm$

0.4 on 5 April to 19.6 ± 1.1 on 5 July, and subsequently declined to 0.6 ± 0.1 by the end of the season. For sampling dates where observations were made in both years, mean numbers of endospores per J2 were higher in 1995 than in 1994, except for the last three sampling dates (Fig. 2). The fluctuations in mean numbers of endospores per J2 were density-independent in both years $(P =$ 0.368 and 0.898, respectively, for 1994 and 1995).

The percentages of endospore-encumbered J2 followed a trend similar to that of the numbers of endospores per J2. The mean percentages of endospore-encumbered J2 initially decreased from 62.9% \pm 10.2 on 10 May to $45.9\% \pm 10.9$ on 25 July, and then increased to $65.9\% \pm 11.0$ by the end of the 1994 growing season (Fig. 2). In 1995, the percentages of endospore-encumbered J2 increased rapidly at the beginning of the growing season from $63.3\% \pm 11.0$ on

FIG. 3. Observed and predicted (negative binomial and Poisson distributions) probabilities for the attachment of *Pasteuria* endospores to second-stage juveniles (]2) of *Heterodera glycines* in a naturally infested microplot soil.

5 April to $86.3\% \pm 7.8$ on 12 May. They leveled off at values $\geq 80.3\% \pm 8.3$ until 5 July, after which they declined to $43.4\% \pm 11.4$ on 26 October. Percentages of endospore-encumbered J2 were higher **in** 1995 than in 1994 throughout the time period from 10 May to mid-August (Fig. 2). Likewise percentages of endospore-encumbered J2 were significantly higher for race 4 than for race 3, except in July (results not shown), with ranges of variation of $58.3\% \pm 11.2$ to 90.6% \pm 2.3, and 28.5% \pm 18.4 to 75.2% \pm 11.4, respectively. No density-dependent trend was observed in the fluctuations of mean percentages of endospore-encumbered J2 for both races of *H. glycines* $(P = 0.128$ and 0.712 for race 3, $P = 0.737$ and 0.468 for race 4 in 1994 and 1995, respectively), **even** when the data were pooled over years ($P =$ 0.636 and 0.311, respecively, for race 3 and race 4).

Heterodera glycines-Pasteuria interrelationship: The relationship between J₂ densities, *Y*,

and the numbers of endospores per J2, *X,* was described by an exponential decay model, $Y = 67.4 + 220.1e^{-1.2X}$ (Fig. 4). According to this model, J2 densities declined with increasing numbers of endospores per J2 from 287.5 ± 5.2 to 67.4 ± 3.3 at a relative rate of 1.2 ± 0.1 units J2 density per endospore per J2 per unit J2 density, which was an absolute decrease of 220.1 ± 4.0 units J2 density attributed to the attachment of endospores. No further reduction in J2 densities occurred after 39.9 endospores had attached to every individual J2. This model accounted for the totality of the explainable variation in J2 densities, after allowance for pure error (72% total variation). A regression curve, $Y = 87.0(1 - e^{-0.53X})$, similar to the "competition curve" (Nicholson, 1933), provided a good fit ($r^2 = 0.75$; $P < 0.0001$) to the relationship between the percentages of endospore-encumbered $[2, Y,$ and the mean numbers of endospores per $[2, X$ (Fig. 5). Based on this equation, the percentages of endospore-encumbered J2 increased with the mean numbers of endospores per J2 at a relative rate of 0.53 ± 0.04 percent units per unit increase in mean numbers of endospores per J2 per percent unit until $87.0\% \pm$ 2.0 of the J2 were each encumbered with an average of 36.1 endospores.

The parameters of the Lotka-Volterra model (eq. 1) were estimated as $a = 0.0195$, $b = 0.000336, c = 0.000049, d = 0.00285, and$ $k = 0.06589$ (Fig. 6). The model was fitted after 917 iterative solutions of the system of equations 2 and 3, starting from the **initial** values $x_0 = 11.250$ and $y_0 = 65.478$, respectively, for J2 densities and percentages of endospore-encumbered J2. The predicted equilibrium values for J2 densities (d/c) and the percentages of endospore-encumbered J2 *(a/b)* were 58.2 and 58.0, respectively. The corresponding ranges of variation were 10.5 to 175.3 and 33.1 to 93.6. A significant correlation ($r = 0.62$; $P < 0.01$) was obtained between the actual and predicted values of the percentages of endospore-encumbered J2, and the differences between these two sets of data were not significant ($t = 1.82; P$) 0.05). For the Nicholson-Bailey model, the

FIG. 4. Exponential decay of numbers of second-stage juveniles ([2) of *Heterodera glycines* with the numbers of Pasteuria endospores per J2 per 250 cm³ of naturally infested microplot soil.

parameter estimates were $\lambda = 1.955$, $\gamma =$ 0.7978, and $\alpha = 0.0789$ (Fig. 6). These values were obtained after 21 iterative solutions of the system of equations 4 and 5, beginning from $x_0 = 24.9$ and $y_0 = 68.7$. The steady values were calculated as 24.6 and 12.0, and the ranges of variation were 7.2 to 118.7 and 1.3 to 86.6, respectively, for J2 densities and percentages of endospore-encumbered J2. The actual and predicted values of the percentages of endospore-encumbered J2 were correlated $(r = 0.61; P \le 0.01)$, although they differed significantly according to Student's t-test ($t = 5.22$; $P < 0.01$). No correla-

tion was found between numbers of cysts and numbers of endospores per J2, or percentages of endospore-encumbered J2. In contrast, a significant but low correlation (r $= -0.24$; $P < 0.01$) was observed between the numbers of eggs per cyst and the numbers of endospores per J2.

DISCUSSION

The *Pasteuria* spp. that parasitize cyst nematodes are separated into two groups, depending on their life history. The first group includes those that infect the oat and

Fro. 5. Effect of the mean numbers of *Pasteuria* endospores per second-stage juvenile (]2) of *Heterodera glycines* on the percentages of endospore-encumbered $\tilde{J}2$ per 250 cm³ of naturally infested microplot soil.

pea cyst nematodes, *H. avenae* W011enweber and *H. goettingiana* Liebscher. In these species the bacterium develops and completes its life cycle in the J2 (Davies et al., 1990; Sturhan et al., 1994). In the second group invasion of the host root by the endosporeencumbered J2 occurs prior to the germination of the bacterium, e.g.P, *nishizawae* on *H. glycines* (Sayre et al., 1991).

The life history of the Illinois isolate of *Pasteuriawas* not known at the time the study was conducted. Therefore, its population dynamics and that of its host were investigated using attachment data of endospores to J2. Subsequent examinations of all life stages of *H. glycines* and cysts extracted from the rhizosphere of soybean plants revealed that the life history of this *Pasteuria* is similar to that of *P. nishizawae* (Atibalentja and Noel, 1997). Germ tubes develop from the endospores and penetrate the body of the nematode soon after the encumbered J2 invades the soybean root. The bacterium then proliferates and matures in the females, which

FIG. 6. Phase space diagram showing the relationship between densities of second-stage juveniles (J2) of *Heterodera glycines* and percentages of J2 encumbered with *Pasteuria* endospores in a naturally infested microplot soil. Each observation represents the mean of eight samples. The predictions for the Lotka-Volterra model were based on the system of equations

$$
x_{t+1} = x_t + 0.0195x_t - 0.000336x_t y_t
$$
 and

$$
y_{t+1} = y_t + 0.000049x_t y_t - 0.00285y_t
$$

where x_t and y_t are the numbers of J2 per 250 cm³ soil and the percentages of endospore-encumbered J2 at time t, respectively; $t = 0, \ldots, 916$; $x_0 = 11.250$, and $y_0 =$ 65.478. For the Nicholson-Bailey model, calculated values were obtained from the system of equations

$$
x_{t+1} = 0.3953x_t + 1.55970x_t e^{-0.0789y_t}
$$
 and

$$
y_{t+1} = 0.7978x_t(1 - e^{-0.0789y_t});
$$

where x and y_t are the numbers of J2 per 250 cm³ soil and the percentages of endospore-encumbered J2 at generation t, respectively; $t = 0, \ldots, 20$; $x_0 = 24.9$, and $y_0 = 68.7.$

ultimately become filled with an average of 3 \times 10⁵ endospores. Parasitized females normally do not produce eggs but, occasionally, a female may produce very few eggs. The life cycle is completed when endospores are liberated into the soil upon disintegration of the diseased female.

Seasonal fluctuations of irregular ampli-

tude and periodicity were observed in both the *Pasteuria* and the *H. glycines* populations. Typically, the greatest peaks in J2 densities coincided with the decline in the numbers of endospores per J2 and the percentages of endospore-encumbered J2. Such an observation suggests a regulation of the *H. glycines* population by *Pasteuria* sp., a hypothesis that has been substantiated by the fit of an exponential decay model to the relationship between J2 densities and numbers of endospores per J2. This model demonstrated a reduction of J2 densities with increasing numbers of endospores per J2, and a stable equilibrium density consistent with the obligately parasitic nature of *Pasteuria* sp. A similar conclusion was obtained from the analysis of variance, which showed that densities of J2 remained stable about their mean value. The steady density of J2 predicted by the Lotka-Volterra model also was consistent with that derived from the exponential decay model. In addition, the Lotka-Volterra model allowed the estimation of such biologically significant parameters as the host and parasite growth and death rates upon which the equilibrium of the host-parasite interrelationship hinges. The knowledge of these parameters may provide important clues about how *the H. glycines-Pasteuria* pathosystem could be manipulated to achieve effective control of the nematode by the bacterium.

The choice of the modified Nicholson-Bailey model (Hassel and May, 1973) was indicated by the form of the equation describing the relationship between the percentages of endospore-encumbered J2 and the mean numbers of endospores per J2. The similarity between this equation and the "random contact" equation (]affee et al., 1992; Nicholson, 1933; Perry, 1978) is apparent if the mean number of endospores per J2 is assumed to reflect the density, *S,* of endospores in the soil. In such a case, our equation gives the percentage of endosporeencumbered J2 as the product of two quantities: γ = the probability of a J2 being susceptible to infection, equal to 0.87 ± 0.02 , and $P_s = 1 - e^{-0.535}$ = the probability of having at least one endospore within the critical

distance of a J2 (Perry, 1978). The critical distance, equal to 0.53, is the fraction of the soil pore volume covered per J2 per unit time, and is equivalent in concept to the transmission efficiency parameter of pathogen propagules (Anderson and May, 1981; Jaffee et al., 1992). These two terms are preferred to the term "searching efficiency" (Nicholson, 1933) since endospores are nonmotile and do not seek out their prey. However, unlike the spores of the nematophagous fungus *Hirsutella rhossiliensis* Minter & Brady (Jaffee et al., 1992; Perry, 1978), *Pasteu74a* endospores are not transmitted directly through contact between parasitized and healthy nematodes (Ciancio, 1995). Lack of motility and of direct endospore transmission may explain why, despite the stabilizing effect of the refuge (Hassel and May, 1973) in which 11% to 20% of the J2 escape infection at each generation, the Nicholson-Bailey model did not fit the data as well as did the Lotka-Volterra model. In addition, the Nicholson-Bailey model and its variants (Hassel, 1978; Hassel and May, 1973;Jaffee et al., 1992; Perry, 1978) assume that encounters between the host and the parasite follow a Poisson distribution. We have shown that, for *the H. glycines-Pasteuria* pathosystem, the negative binomial distribution is more appropriate than the Poisson distribution.

This study indicated that the race of H. *glycines,* alone or in conjunction with other factors such as the date and the year, could have a significant effect on the numbers of endospores per J2 and the percentages of endospore-encumbered J2. Such results would imply that either the two races of H. *glycines* exhibit differential susceptibility to this *Pasteuria* or two distinct strains of the bacterium exist. Microscopic examinations of endospores from both races did not provide evidence to support the second of these hypotheses (Noel and Stanger, 1994). On the other hand, the results from individual microplots showed that the race effect could have originated from spatial and(or) temporal variations of *Pasteuria* inoculum densities in the soil. Further, these data suggest that the *Pasteuria* infestation of one of the race 3 microplots may have occurred later than that of the other three microplots.

The obligate nature and host specificity of *Pasteuria* spp. suggest density-dependent relationships with their hosts (Ciancio, 1995). The failure of the Pollard's test to detect any density-dependent trend in either the *H. glycines* or the *Pasteuria* populations does not rule out the existence of such relationships in these populations. In fact, the negative results obtained with the Pollard's test could be attributed to the small size of our time series, since the chance of detecting density dependence increases with the number of observations in the time series (Bulmer, 1975).

The research presented herein demonstrates that *Pasteuria* sp. is capable of maintaining *H. glycines* populations at equilibrium densities that are below the damage threshold reported from field studies (Noel, 1984). Whether fungi (Chen et al., 1996; McLean and Lawrence, 1995; Meyer and Meyer, 1996) contributed to this phenomenon is not known. Stiles et al. (1993) evaluated the pathogenicity of several fungi isolated from *H. glycines* in Illinois. Those fungi did not reduce either the numbers of cysts or numbers of eggs and J2. The soil used in that study originated from the same field that provided the soil for the microplots utilized in the current investigation. If other factors did affect *the H. glycines* population, their action may then explain the inflation of the pure error (variation in J2 densities at fixed levels of endospores per J2) observed and accounted for during the fitting of the exponential decay model. It is not known whether the equilibrium densities observed in this study can provide adequate control of *H. glycines* in commercial soybean fields. Nevertheless, these results are encouraging in that, given sufficient time following introduction into a field, this *Pasteuria* sp. may increase to levels that would be effective as one component in an integrated pest management program to control *H. glycines.*

LITERATURE CITED

Anderson, R. M., and R. M. May. 1981. The population dynamics of microparasites and their invertebrate hosts. Philosophical Transactions of the Royal Society of London B 291:451-524.

Atibalentja, N., and G. R. Noel. 1997. Life cycle and host specificity of *Pasteuria* sp. parasitizing *Heterodera glycines.* Journal of Nematology 29:568 (Abstr.)

Bird, A. F., and P. G. Brisbane. 1988. The influence of *Pasteuria penetrans* in field soils on the reproduction of root-knot nematodes. Revue de Nématologie 11:75-81.

Bishop, A. H., and D. J. Ellar. 1991. Attempts to culture *Pasteuria penetrans* in vitro. Biocontrol Science and Technology 1:101-114.

Brown, S. M., J. L. Kepner, and G. C. Smart, Jr. 1985. Increased crop yields following application of *Bacillus penetrans* to field plots infested with *Meloidogyne incognita.* Soil Biology and Biochemistry 17:483-486.

Bulmer, M. G. 1975. The statistical analysis of density dependence. Biometrics 31:901-911.

Chen, S.Y., D. W. Dickson, and D.J. Mitchell. 1996. Pathogenicity of fungi to eggs of *Heterodera glycines.* Journal of Nematology 28:148-158.

Chen, Z.X., D.W. Dickson, R. McSorley, D.J. Mitchell, and T.E. Hewlett. 1996. Suppression of *Meloidogyne arenaria* race 1 by soil application of endospores of *Pasteuria penetrans.* Journal of Nematology 28: 159-168.

Chen, Z. X., D. W. Dickson, D.J. Mitchell, R. McSorley, and T. E. Hewlett. 1997. Suppression mechanisms of *Meloidogyne arenaria* race 1 by *Pasteuria penetrans.* Journal of Nematology 29:1-8.

Ciancio, A. 1995. Density-dependent parasitism of *Xiphinema diversicaudatum* by *Pasteuria penetrans* in a naturally infested soil. Phytopathology 85:144-149.

Ciancio, A., R. Mankau, and M. Mundo-Ocampo. 1992. Parasitism of *Helicotylenchus lobus* by *Pasteuria pet~ etrans* in naturally infested soil. Journal of Nematology 24:29-35.

Cobb, N.A. 1918. Estimating the nema populations of soil. U.S. Department of Agriculture Technology Circular 1. Washington, DC: U.S. Government Printing Office.

Davies, K. G., C. A. Flynn, V. Laird, and B. R. Kerry. 1990. The life cycle, population dynamics, and host specificity of a parasite of *Heterodera avenae,* similar to Pasteuria penetrans. Revue de Nématologie 13:303-309.

Doupnik, B., Jr. 1993. Soybean production and disease loss estimates for north-central United States from 1989 to 1991. Plant Disease 77:1170-1171.

Giblin-Davis, R.M., L.L. McDaniel, and F. G. Bilz. 1990. Isolates of the *Pasteuria penetrans* group from phytoparasitic nematodes in bermudagrass turf. Supplement to the Journal of Nematology 22:750-762.

Hassel, M.P. 1978. The dynamics of arthropod predator-prey systems. Princeton: Princeton University Press.

Hassel, M. P., and R. M. May. 1973. Stability in insect host-parasite models. Journal of Animal Ecology 42: 693-736.

Jaffee, B., R. Phillips, A. Muldoon, and M. Mangel. 1992. Density-dependent host-pathogen dynamics in soil microcosms. Ecology 73:495-506.

Jenkins, W.R. 1964. A rapid centrifugal-flotation technique for separating nematodes from soil. Plant Disease Reporter 48:692.

Lotka, A.J. 1925. Elements of physical biology. Baltimore: Williams and Wilkins.

Marriot, F. H. C. 1979. Banard's Monte Carlo tests: How many simulations? Applied Statistics 28:75-77.

McLean, K. S., and G. W. Lawrence. 1995. Development of *Heterodera glycines as* affected by *Fusarium solani,* the causal agent of sudden death syndrome of soybean. Journal of Nematology 27:70-77.

Meyer, S. L. F., and R.J. Meyer. 1996. Greenhouse studies comparing strains of the fungus *Verticillium lecanii* for activity against the nematode *Heterodera glycines.* Fundamental and Applied Nematology 19:305-308.

Niblack, T. L., ed. 1993. Protect your soybean profits: Manage soybean cyst nematodes. St. Louis, MO: United Soybean Board and American Soybean Association.

Nicholson, A.J. 1933. The balance of animal populations. Journal of Animal Ecology 2:132-178.

Nicholson, A.J., and V. A. Bailey. 1935. The balance of animal populations. Part I. Proceedings of the Zoological Society of London 3:551-598.

Nishizawa, T. 1987. A decline phenomenon in a population of the upland rice cyst nematode, *Heterodera elachista,* caused by a bacterial parasite, *Pasteuria pen*etrans. Journal of Nematology 19:546 (Abstr.).

Noel, G. R. 1984. Relating numbers of soybean cyst nematode to crop damage. Pp. 17-19 *in* D. P. Schmitt, ed. Proceedings of the Fifth Cyst Nematode Workshop. Raleigh, NC: North Carolina State University.

Noel, G. R., and B. A. Stanger. 1994. First report of *Pasteuria* sp. attacking *Heterodera glycines* in North America. Supplement to the Journal of Nematology 26: 612-615.

Oostendorp, M., D.W. Dickson, and D.J. Mitchell. 1991. Population development of *Pasteuria penetrans* on *Meloidogyne arenaria.* Journal of Nematology 23:58-64.

Perry, J. N. 1978. A population model for the effect of parasitic fungi on numbers of the cereal cyst-nematode, *Heterodera avenae.* Journal of Applied Ecology 15:781- 787.

Pollard, E., and K. H. Lakhani. 1987. The detection of density-dependence from a series of annual censuses. Ecology 68:2046-2055.

Sayre, R. M., W. P. Wergin, J. M. Schmidt, and M. P.

Starr. 1991. *Pasteuria nishizawae* sp. nov., a mycelial and endospore-forming bacterium parasitic on cyst nematodes of genera *Heterodera and Globodera.* Research in Microbiology 142:551-564.

Spaull, V. W. 1984. Observations on *Bacillus penetrans* infecting *Meloidogyne* in sugarcane fields in South Africa. Revue de Nématologie 7:277-282.

Starr, M. P., and R. M. Sayre. 1988. Pasteuria thornei sp. nov. and *Pasteuria penetrans* sensu stricto emend., mycelial and endospore-forming bacteria parasitic, respectively, on plant-parasitic nematodes of the genera *Pratylenchus* and *Meloidogyne.* Annales de l'Institut Pastenr/Microbiologie 139:11-31.

Stiles, C. M., D. A. Glawe, G. R. Noel, and J. K. Pataky. 1993. Reproduction of *Heterodera glycines* on soybean in nonsterile soil infested with cyst-colonizing fungi. Nematropica 23:81-89.

Sturhan, D., R. Winkelheide, R. M. Sayre, and W. P. Wergin. 1994. Light and electron microscopical studies of the life cycle and developmental stages of a *Pasteuria* isolate parasitizing the pea cyst nematode, *Heterodera goettingiana.* Fundamental and Applied Nematology 17: 29-42.

Verdejo-Lucas, S. 1992. Seasonal fluctuations of *Meloidogyne* spp. and the *Pasteuria penetrans* group in kiwi orchards. Plant Disease 76:1275-1279.

Voherra, V. 1926. Variations and fluctuations of the number of individuals in animal species living together. Pp. 409-448 *in* R.N. Chapman, ed. Animal ecology. New York: McGraw-Hill.

Weibelzahl-Fulton, E., D.W. Dickson, and E. B. Whitty. 1996. Suppression of *Meloidogyne incognita and M. javanica* by *Pasteuria penetrans* in field soil. Journal of Nematology 28:43-49.

Williams, A. B., G. R. Stirling, A. C. Hayward, and J. Perry. 1989. Properties and attempted culture of Pasteuria penetrans, a bacterial parasite of root-knot nematode *(Meloidogynejavanica).Journal* of Applied Bacteriology 67:145-156.

Wrather, J.A., A.Y. Chambers, J.A. Fox, W. F. Moore, and G. L. Sciumbato. 1995. Soybean disease loss estimates for the southern United States, 1974 to 1994. Plant Disease 79:1076-1079.