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Galanin: A Role in Mesolimbic Dopamine-Mediated Instrumental Behavior?

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Abstract

ROBINSON, J.K. and Brewer, A. Galanin: A Role in Mesolimbic-Dopamine Mediated Instrumental Behavior? NEUROSCI BIOBEHAV REV XX(X) XXX-XXX, 2008. The involvement of the neuropeptide galanin in the consumption of the primary "commodities" of food and water is well established. However, the present review describes anatomical and behavioral evidence that suggests that galanin may also modulate ascending mesolimbic dopamine function and thereby play an inhibitory role in the systems by which instrumental behavior is energized toward acquiring primary commodities. General anatomical frameworks for this interaction are presented and future studies that could evaluate it are discussed.

Keywords

drinking; galanin; feeding; progressive ratio; consumption; motivation

1. Introduction: Galanin as a multifunctional peptide

Galanin is a 29 amino acid (30 in humans) peptide that was isolated in 1983 (Tatemoto et al., 1983; Rökaeus et al., 1984). Though not as well known as many neuropeptides such as the opiates, it has been the focus of steady research attention for twenty-five years as evidenced by a recent symposium (Hökfelt, 2005) and over three thousand galanin-related publications in the literature. Galanin has been associated with a variety of important functions and dysfunctions, including learning and memory, anxiety, Alzheimer's disease, depression, pain sensation, and those described below (for reviews see Counts et al., 2003; Holmes, Mahony & Wynick, 2005; Lang, Gundlach & Kofler, 2007; Ögren et al, 2007; Robinson, 2004; Wiesenfeld-Hallin, Xu, Crawley & Hökfelt, 2005). However, new studies, as well as reassessment of older bodies of data, have prompted reevaluation of potential roles for galanin in psychological processes. The goal of this present theoretical review will be to consider one of these; in particular, a potential role for galanin in the modulation of mesolimbic dopamine

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systems underlying instrumental behavior. In doing so, we will first assess the distribution of galanin and known effects on the consumption of primary "commodities", food and water. We will then assess the anatomical evidence that puts galanin in the right place to modulate ascending dopamine function, and then review the physiological and behavioral evidence in favor of galanin-dopamine interactions. We will next discuss potential system-level models for this modulation and end by presenting a roadmap for future studies that could evaluate this assertion.

For the present discussion we will define *instrumental behavior* as behavior that reliably leads to an outcome, but not limit it to acquired operant responses or overt actions. In this way, the definition is broad and corresponds to "appetitive," in the sense used by of Craig (1917, 1918). This would allow it to include approach behaviors but also could include, for example, a reinforcer delivered contingently upon the instrumental response of remaining inactive for some set amount of time. The *outcome* is roughly analogous to Craig's consummatory act and Premack's (1962) and Timberlake & Allison's (1974) conception of a reinforcer as an opportunity to engage in a behavior that is contingent upon another.

2. Galanin: Anatomical Distribution

Galanin is expressed in many tissues throughout the body, though a major focus of work has been on its expression in the brain (Bartfai, Hökfelt, & Langel, 1993; Land, Gundlach & Kofler, 2007). Studies of the anatomical distribution of galanin, including immunohistochemical studies of galanin and galanin precursor protein mRNA and autoradiographic studies of binding to putative galanin receptors, showed expression in cortex, especially insular and cingulate cortices, hypothalamus, amygdala, central gray, dorsal raphe and locus coeruleus (Melander et al., 1988; Skofitsch & Jacobowitz, 1985; Skofitsch & Jacobowitz, 1986; Skofitsch, Sills, Jacobowitz, 1986; review by Merchenthaler, Lopez & Negro-Vilar, 1993). A vast majority of noradrenergic neurons express galanin and provide noradrenergic/galanergic input to the hypothalamus and ventral tegmentum (Holets et al., 1988; Xu, Shi, Hökfelt, 1998), though not the central amygdala (Barerra et al., 2006).

Studies in the late 1990s cloned three G-protein coupled receptor subtypes (GalR1, GalR2 and GalR3; reviewed in Branchek et al., 2000; Florén, Land, Langel, 2000) and mapped the distribution of mRNA for each in the rat. Of the three, GalR1 shows the broadest distribution patterns, with few areas of overlap with GalR2. The most prominent dissociations are that GalR1 is within discrete regions in hippocampus, amygdala, thalamus, and locus coeruleus, versus GalR2 predominance in dentate gyrus, substantia nigra/ventral tegmental area and dorsal raphe (O'Donnell et al., 1999). All three are found in low to moderate amounts in various hypothalamic nuclei, including arcuate, paraventricular, ventromedial and dorsomedial nuclei of the hypothalamus. GalR3 is also reported to have low levels of expression in motivationrelated brain areas including the bed nucleus of the stria terminalis and medial nucleus of the extended amygdala, the periaqueductal gray, the lateral parabrachial nucleus, the medial medullary reticular formation and in the subfornical organ (Mennicken et al., 2002). Selective radioligands or antibodies for all three subtypes are not yet available to make specific confirmation of these distribution patterns, though indirect inferences are generally consistent with these patterns of mRNA (Lu et al., 2005; Hawes & Picciotto, 2005). Thus, it appears that galanin and galanin receptors are well situated for involvement in multiple aspects of motivation.

Numerous biochemical and physiological studies have characterized galanin as a hyperpolarizing peptide with both presynaptic and postsynaptic actions. However, excitatory effects may occur in some systems mediated by the GalR2 subtype or by indirect actions (see Lang, Gundlach & Kofler, 2007 for review). For example, galanin, administered

intracerebroventricularly produces two particularly strong areas of metabolic activation of neurons in central amygdala and dorsomedial hypothalamus, as measured by cFos immunohistochemistry (Blackshear et al., 2007; Lawrence, Williams & Luckman, 2003; Parrado et al., 2007). Galanin may also exert broad tonic effects through a reduction in the firing rate of noradrenergic and serotonergic cells (Pieribone et al., 1995; Seutin et al., 1989; Sevcik, 1993; Xu, Zhang, Pieribone, Grillner & Hökfelt, 1998), and may have both inhibitory and excitatory influences on acetylcholine systems (Elvander et al., 2004; Elvander & Ögren, 2005; Jhamandas, et al., 2002).

3. Galanin and Feeding

Like many other CNS neuropeptides, galanin is implicated in diverse functions. However, a particular consistent and interesting role is in the regulation of feeding behavior. As detailed above, galanin mRNA expression is high in the rat hypothalamic nucleus, especially the supraoptic, paraventricular, arcuate, dorsomedial and ventromedial regions, all implicated in feeding regulation, as well as numerous other endocrine roles (Bonnefond et al., 1990). Galanin actions on circulating hormones related to appetite and body-weight is well documented, such as inhibition of insulin secretion and stimulation of growth hormone release (e.g. Dunning $\&$ Taborsky, 1988; Gundlach, Burazin, & Larm, 2001).

The most pronounced direct stimulatory effect on feeding is shown in the classic studies from Leibowitz and colleagues in which galanin microinjected into the paraventricular nucleus of the hypothalamus and central amygdala drastically increased food intake (Kyrkouli, Stanley, & Leibowitz, 1986; reviewed by Crawley, 1999). Subsequent experimental paradigms demonstrated this feeding stimulation effect in macronutrient diets that separate fat, protein and carbohydrates, as well as complex foods such as basic lab chow and cookie mash and showed that galanin preferentially increases the consumption of fat and carbohydrates (Tempel, Leibowitz & Leibowitz, 1988; Crawley et al., 1992; Corwin, Robinson & Crawley, 1993). Supporting this is a recent demonstration that galanin knockout mice consumed much less fat that wild-type controls under macronutrient choice conditions and when only fat was available (Adams, Clapham, Wynick & Speakman, 2008). It is important to note, however, that all of these feeding studies examined consumption in situations where food is made freely available to the animals.

Mounting evidence from several lines of research suggests galanin expression up-regulates rather than down-regulates in response to fat intake (Akabayashi, et al., 1994; Leibowitz, Akabayashi & Wang, 1998) and that galanin appears to be relatively insensitive to systemic signals of deprivation (Leibowitz, 2007). Thus, galanin seems to stimulate, in a nonhomeostatic manner, the consumption of foods that would promote weight gain rather than meeting short-term metabolic needs (Leibowitz, 2005). This view suggests that galanininduced feeding is more opportunistic (in response to availability) rather than drive-related (in response to deprivation conditions), and it follows from this that galanin induced consumption of foodstuffs may have the property of "elastic" rather than "inelastic" demand, where the increased "price" of the commodity results in its reduced consumption (Hursh, 1980).

4. Galanin in Fluid Intake Regulation

While the role for galanin in regulating food intake is well established, galanin regulation of consummatory behavior is not limited to feeding. Emerging evidence also shows that galanin regulates water intake and fluid balance. Brewer, Langel & Robinson (2005) demonstrated that relatively high doses galanin $(10-20 \mu g \text{ i.c.v.})$ moderately reduced the intake of water in twenty-three hour water restricted rats. This effect was seen regardless of whether the water was made available in a cup in a tub cage or whether delivered response-independently in discrete droplets on a Fixed-Time (FT 20 sec) schedule in an operant chamber. However, a

similar injection of galanin intracerebroventricularly into the nucleus accumbens or the lateral hypothalamus, produced no significant increase or decrease in low levels of water consumption in water-sated rats, though a slight trend toward increased consumption was apparent (Schneider et al., 2007). Kyrkouli et al. (1990) also reported that the proportion of time spent drinking relative to other activities, especially eating, did not increase from a low baseline level in response to galanin. Though the specific side-by-side experiment has not been performed, these studies together imply that fluid deprivation is a necessary precondition for a galanininduced decrease in fluid consumption to be apparent.

This may be explained by exploring the likely underlying mechanism. First, it is known that galanin co-exists with vasopressin in the hypothalamus (Skofitsch, Jacobowitz, Amann & Lembeck, 1989), though the two peptides are somewhat compartmentalized (Landry, Vila-Porcile, Hökfelt & Calas, 2003). Several studies have shown that galanin inhibits the release of vasopressin under various conditions of osmotic challenge, when vasopressin release is maximal (Ciosek & Cisowska, 2003; Ciosek, Cisowska & Dabrowski, 2003; Kondo et al., 1991; Landry, Roche & Callas, 1995), but had no substantial effects when the animals were not challenged (Balment & al Barazanji, 1992). Further, experimental manipulations producing hyperosmosis and hypovolemia upregulated galanin and GalR1 mRNA in magnocellular paraventricular nucleus of the hypothamaus (Burazin, Larm, Gundlach, 2003; Koenig, et al., 1989; Landry, Aman & Hökfelt., 1998; Meister et al., 1990; Rökaeus, Young, Mezey, 1988; Villar et al., 1990; Yagita, Okamura & Ibata, 1994;). A recent study has elegantly demonstrated galanin inhibition of angiotensin II-sensitive neurons in the subfornical organ, a brain structure thought to be critical to integrating osmotic and hypovolemic thirst messages and then activating behavior directed toward water seeking (Kai et al., 2006). Taken together, these studies point to a role for galanin in modulating homeostatic fluid regulation processes under conditions of deprivation. However an examination of the effects of galanin on habitual and cue-occasioned fluid ingestion has not been performed.

5. Instrumental behavior: Anatomy and Conceptualization

While it is possible to talk about feeding and drinking as distinct behaviors, in reality they are closely related consummatory processes in mammals. This is illustrated by the phenomena of adaptive postprandial drinking (de Castro, 1989) and maladaptive schedule-induced polydipsia (Falk, 1961). But more importantly, the consummatory acts of feeding and drinking are both inextricably linked with the instrumental, species-typical, appetitive behavior in the wild of foraging which is traditionally reduced in the psychological laboratory to traversing the length of a maze or pressing a bar in an operant chamber (e.g. Krebs & Kacelnik, 1983; Mathis, Johnson & Collier, 1995). These instrumental acts are presumed to converge on and modulate the same neural systems, regardless of whether the consummatory behavior involves feeding, drinking or otherwise.

While it is certainly a gross oversimplification to reduce a complex construct like instrumental behavior to a single anatomical component, a critical, if controversial, neural system identified with instrumental behavior is the ascending dopamine systems commonly called the mesolimbic dopamine pathway (see Björkland & Dunnett, 2007 for a discussion of terminology). This pathway is clearly part of a complex system involving multiple brain structures and numerous neurotransmitter systems within them. The ventral tegmental area is a major source of dopaminergic input to frontal cingulate and entorhinal cortices, central amygdala, hippocampus, hypothalamus, basal forebrain, periacquaductal gray, raphe and parabrachial nuclei, locus coeruleus and the nucleus accumbens (Beckstead, Domesick & Nauta, 1979; Simon, LeMoal & Calas, 1979;). The ventral tegmental area receives diffuse descending input from a frontal cortex, hypothalamus, amygdala, and many ventral midline structures (Phillipson, 1979; Geisler & Zahm, 2005). While the exact nature of mesolimbic

dopamine contributions to instrumental behavior remains controversial, there is good consensus that it is one critical component in the larger system that inputs information into executive and motor structures to direct or occasion behavior.

An influential theory by Wise (1982) suggested that mesolimbic dopamine underlies the subjective, pleasurable aspects of consumption (commonly called "the anhedonia hypothesis" to indicate that a blockade of dopamine would produce a loss of this pleasure from the act of consumption). However, a major focus of the current debate suggests that mesolimbic dopamine is maximally relevant to activating the behavior associated with the appetitive aspects rather than consuming (and the hedonic-perceptual) acts (e.g. Berridge, 2007; Berridge & Robinson, 2003; Baldo & Kelley, 2007; Barbano & Cador, 2006, 2007; Nicola, 2007; Phillips, Vacca & Ahn, in Press). To use the informal terms of Berridge & Robinson (1998), mesolimbic dopamine mediates the "wanting" rather than the "liking" of a commodity. One of the most clearly articulated theories is that of Salamone and colleagues (Salamone & Correa, 2002; Salamone et al., 2005) which suggests that mesolimbic dopamine "energizes" instrumental behavior, predicting that dopamine activity would be maximal during instrumental but not consummatory acts ("anhedonia" predicts the opposite), and that loss of dopamine though pharmacological blockade or lesioning would lessen or abolish this instrumental behavior.

To put this theoretical dissociation into practice, the clearest way to operationalize dopamine tone behaviorally would be in paradigms where some level of effort is applied to produce the **opportunity** to ingest, in the presence of a signal establishing the contingency between the effort and the consumption. This could be a Skinnerian three-term contingency, where a discriminative stimulus signals that responding on a reinforcement schedule will be reinforced, though could also be "foraging" on a radial arm maze, or an appetitive Pavlovian paradigm, where a CS signals the availability of a food UCS. Indeed, in many such tests, dopamine agonists are poor stimulators of direct consumption of food but in some cases will increase operant responding for food (for review see Baldo & Kelly, 2007) and lesions of the ventral tegmental area and dopamine antagonists reduce operant responding for food and other approach behavior for food, but not the consumption of the food itself: indeed, the effect of dopamine antagonism on operant responding is positively related to the amount of instrumental effort required to produce reinforcement (Salamone et al., 2007). These studies are all compelling because they show that the consumption of the commodity is itself not altered (or even enhanced) when made freely available, by a manipulation that reduces the effortful, appetitive behavior directed toward producing the opportunity to ingest. In anticipation of our later argument, one would predict that an endogenous inhibitor of mesolimbic dopamine, such as galanin perhaps, would also produce decreased instrumental behavior without decreasing or perhaps even stimulating consummatory behavior.

6. Galanin and Dopamine

Establishing some relationship between galanin and mesolimbic dopamine is a necessary, though not sufficient, condition for linking galanin to instrumental behavior. Indeed, some anatomical evidence associates galanin and mesolimbic dopamine. Classic autoradiographic receptor binding studies showed galanin-like binding in both the core and shell of the nucleus accumbens and ventral tegmental area (Skofitsch, Sills & Jacobowitz, 1986). The ventral tegmental area also expresses galanin mRNA and mRNA for the GalR2. One known noradrenergic input to ventral tegmental area is from the locus coeruleus, which is likely to also contain galanin, but other inputs could also arrive from known ventral tegmental area inputs regions that express galanin receptors and immunoreactivity, such as lateral hypothalamus, interpeduncular nucleus or central nucleus of the amygdala (Saper & Loewy, 1980; Akmaev, Kalimullina & Sharipova, 2004). However, no systematic studies using

colabelling and anterograde/retrograde tracers to look at galanin inputs to ventral tegmental area and nucleus accumbens have been conducted to confirm this association.

Physiological and biochemical studies also provide some, albeit largely indirect, evidence showing that galanin modulates mesocorticolimbic dopamine activity. Galanin reduced levels of tyrosine hydroxylase protein and attenuated dibutyryl cAMP-induced TH-immunoreactivity in ventral mesencephalon cell culture. This effect was correlated with a sizable increase in GalR1 mRNA expression (Counts et al., 2002). Another report showed reduced dopamine synthesis in ventral tegmental area following galanin treatment (Ericson & Arlenius, 1999). Tsuda et al. (1998), reported that galanin inhibited evoked DA release in striatal slice preparation. However, the only published study using *in vivo* microdialysis to measure galanin effects on dopamine release microinjected galanin into the paraventricular nucleus of the hypothalamus and observed an increase in nucleus accumbens dopamine levels (Rada, Mark & Hoebel, 1998). Together, these data suggest that a galanin modulatory influence may be complex and may involve several pathways, though the conclusive studies have not been conducted.

7. Galanin and Instrumental behavior Processes: Behavioral Evidence

The most straightforward evidence in favor of galanin modulation of instrumental behavior comes from behavioral experiments of the sort described above and used to demonstrate a selective role for dopamine in instrumental behavior, where appetitive and consummatory behavior are clearly separated procedurally and where dissociations between responsecontingent and free access consumption can be made.

The first such lines of evidence came from a reanalysis of some of the secondary measures from a line of research conducted in our lab examining the effects of galanin on various operant tasks (described in Brewer, Echevarria, Langel & Robinson, 2005). The primary focus of these experiments was to look at the effects of galanin on measures of working memory, executive and motor processes. Early studies had shown that galanin consistently impaired performance of delayed non-matching to sample and delayed matching to sample tests at even short memory delays. These multi-component operant tasks are usually described as measuring working memory, but also tap other psychological constructs such as perseveration, visual discrimination, and motor skills. This "delay-independent" pattern of impairment is usually thought to indicate disruption of task relevant processes other than working memory capacity, as performance is impaired even when little or no working memory appears to be needed. After replicating delay-independent delayed matching-to-position deficits produced by centrally administered galanin, follow up studies were able to rule out galanin-induced disruption of motor initiation or sustained responding (Brewer, Langel & Robinson, 2004) and simple and conditional discriminations (Echevarria et al., 2005; Saccone et al., unpublished) as explanations.

The common element of each of these tasks was that the rats were water restricted for twentythree hours prior to the test session and responding was reinforced by the delivery of 0.1 ml of water and the consistent effect of galanin across all of the tasks was to reduce the number of trials completed (and therefore, reinforcers received) in each session (shown in Table 1). For example, one study compared the effects of galanin on free access to water, both when presented in a cup in a tub cage or as free-droplets on a fixed time FT20 sec schedule, as well as responding on a progressive ratio schedule of reinforcement (Brewer, Langel & Robinson, 2005). A progressive ratio 1 (PR1) schedule increases the number of responses required per reinforcer on each subsequent reinforcer, and the "break point" measure, which is the point where the animal ceases to respond, is used commonly as a test of reinforcer strength. On the PR1 schedule, a substantially decreased break point was observed at a 5µg dose of galanin that

produced no reduction in consumption of the free water droplets on the FT schedule. These data clearly indicate reduced strength of the reinforcer, but only suggest that the effect of galanin on the instrumental response is added on top of the effect on reduced water consumption, rather than clearly dissociated. That is, when the experimental paradigm was made more complicated by including an instrumental response component, the effect of the galanin is observed at a lower dose. While this may simply reflect an additive effect of both an endocrine response (i.e., "thirst"), as discussed above, and an inhibition of mesolimbic dopamine appetitive activation, it does not provide a clear theoretical dissociation.

While these data are consistent with some sort of "effort-dependency" of the galanin effect, the clearest way to tease apart these two explanations is to perform the same experiments again with a food rather than a water reinforcer. Here, the nature of galanin provides the opportunity for a very compelling test. Since galanin stimulates eating, one could predict that galanin was affecting instrumental behavior through actions on the palatability or neurohormonal signals for the primary reinforcer, then here it would **enhance** instrumental behavior strength and the rats would persist **longer** on the progressive ratio schedule. In essence, if the commodity is valued more, they should "pay" more for it. Alternatively, a galanin-induced reduction in instrumental behavior should be independent of the properties of the food reinforcer, so one would predict less persistent responding on the progressive ratio schedule but a galanin **stimulation** of consumption of the food when made freely available. Indeed, it is the latter that was observed, regardless of whether standard lab chow pellets were used in both the free access conditions (Brewer & Robinson, In press) or if a high fat (15% fat) milk-cream product was used (shown in Figure 1). The addition of the PR1 response requirement abolished the galanininduction of feeding observed in the free-access condition, consistent with a selective inhibition of dopamine-mediated instrumental behavior.

Together, the findings from these varied operant and free consumption experiments paint a picture of galanin playing three regulatory roles: the simulation of food consumption, the reduction of deprivation-induced water consumption, and an inhibition of appetitive instrumental behavior, all by dissociable mechanisms.

8. Galanin & Instrumental Behavior: Theoretical Models

So let us consider the mechanism of this galanin-mesolimbic dopamine interaction. Two theories could be offered to explain these findings, each of which predicts fundamentally different connection networks. Specifically, they differ in the direction of the interaction.

A. Galanin inhibits dopamine

In this model, galanin, perhaps through the widespread and hyperpolarizing GalR1, inhibits dopamine-mediated appetitive activation at the ventral tegmental area or in nucleus accumbens. This might be accomplished directly by galanin from locus coeruleus, or indirectly by upstream inhibition by galanin of inputs to ventral tegmental area/nucleus accumbens from hippocampus, hypothalamus, central amygdala, or even medial prefrontal cortex. In this manner, galanin functions as a peptide neuromodulator of dopamine, adding nuance to the dopamine response.

B. Dopamine inhibits galanin

This model may not be as intuitive, but would fit the existing data as well. In this model, the addition of the high operant response requirement creates a condition in which mesolimbic dopamine activity is increased (e.g. increased release, burst firing). Here dopamine synapses on hypothalamic nuclei (e.g. paraventricular, lateral or arcuate), and blocks galanin induction of feeding and potentiates galanin reduction of drinking. In this model, the role of dopamine

in determining the elasticity of demand conditions is further articulated by linking it to galanin regulated consumptive functions.

C. Tests of the Theories

The critical evaluation of these models is dependent upon considerable amounts of information that does not yet exist in the literature, and a goal of this discussion is to stimulate new work to test them. While a decisive *experimentum crucius* is unlikely to emerge, data that could support or undermine both of these alternatives could be provided from several different approaches. While this is not an exhaustive list of possible experiments, they demonstrate the heuristic value of two theories in generating novel lines of work.

The most obvious first piece would be a substantial amount of anatomical work detailing the exact relationship between galanin and dopamine neurons. In particular, studies such as those of Barerra et al. (2004) that employed retrograde tracers and double labeling for galanin markers and tyrosine hydroxylase would help elucidate specific points of galanin–dopamine interaction. However, other pathways where galanin and dopamine may interact indirectly will be more complicated to establish.

Given the potentially numerous areas of interaction, and the difficulties of determining the specific relationships between neurotransmitter systems at the system, synaptic and intracellular levels, this anatomical work would be unlikely to serve as a strong test of either theory. However, anatomical evidence, in conjunction with site-specific microinjections would also help delineate the relevant systems and complement the anatomical studies. Therefore, the demonstration of direct effects of galanin and galanin antagonists on mesolimbic DA overflow as measured by *in vivo* microdialysis, voltametry and electrophysiology in conscious animals are essential. For example, if galanin is inhibitory of mesolimbic dopamine, then one might expect galanin microinjected into the ventral tegmental area or nucleus accumbens to reduce tonic dopamine firing rates, burst events, or dopamine overflow in target regions. Galanin antagonists might also have a facilitative action, which would support an endogenous role for galanin more strongly than the application of exogenous galanin. Inhibition of dopamine by intracerebroventricularly-administered galanin, in the absence of direct effects in VTA or accumbens, would point to indirect modulator pathways. A failure to observe direct effects of galanin on dopamine physiology and neurochemistry would bode poorly for a theory of galanin inhibition of dopamine. Overall, this work would expand upon and potentially reconcile the interesting but unclear direct and indirect measures of mesolimbic dopamine tone discussed previously (e.g. Counts et al., 2001; Ericsson & Arlenius, 1999; Rada et al., 1998; Tsuda et al., 1998).

Direct tests of the second theory, whose premise is that dopamine inhibits galanin, requires first that dopaminergic drugs be shown to affect galanin. Reliable measurement of galanin release *in vivo* is difficult but possible (Morilak et al., 2002), and would be the most direct way to elucidate the effects of dopamine agonists or antagonists on galanin overflow in specific brain regions. This influence of dopamine on galanin could also be examined less precisely by measuring gene expression of preprogalanin or galanin receptor subtype mRNA.

Showing anatomical or physiological interactions between galanin and dopamine enables but does not firmly establish the specific role in instrumental behavior that we have discussed previously. For this, various behavioral pharmacological and lesion experiments could also be conducted along the lines of dopamine devaluation experiments that showed that inhibition or loss of dopamine only reduced instrumental responding for food, but not the consumption of food made freely available (e.g. Salamone, Correa, Farrar & Mingote, 2007). For example, if galanin inhibits the dopamine that is maintaining instrumental responding for food, then galanin administration is like a pharmacological blockade of dopamine or a dopamine-specific

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lesion. Following from this argument, one could predict that dopamine agonists coadministered with galanin might offset the galanin-induced suppression of instrumental responding. Reciprocally, one could also predict that dopamine antagonism or 6-OHDA lesions would not impair galanin-induced feeding. If, according to the second theory of dopamine inhibiting galanin induced feeding, adding a response produces dopamine system activation that in turn inhibits galanin in hypothalamaus, then systemic or microinjected dopamine agonists would inhibit galanin-induced free-feeding by simulating response contingent feeding conditions.

A third line of research would establish the generality of the effects of galanin on instrumental behavior, and follows from the broader assumption that mesolimbic dopamine mediated appetitive activities should not be limited to food and water reinforced instrumental behaviors, but should generalize to other known operant reinforcers, such as sucrose and saccharine solutions, turning on a heat lamp in a cold environment, and avoiding shock and other aversive (negative) reinforcers. Other operants, such as wheel running for food or eating to gain access to wheel running in the style of the transposable response-instrumental behavior paradigms (reviewed in Dunham, 1976), also pose interesting challenges, such as whether galanin would stimulate versus inhibit eating to gain access to wheel running. Determining the effects of galanin on instrumental behavior for acquired, secondary reinforcers are also a fascinating potential experiments, given the importance of acquired reinforcers in motivating human activities.

A final and highly interesting set of tests also comes from the evaluation of drugs of abuse as reinforcers. A common theory is that the instrumentally energizing (reinforcing) effects of drugs of abuse are at least partially due to the activation of mesolimbic dopamine in a manner similar to that of "natural" rewards like food and sex (see Hyman, Malenka & Nestler, 2006, for a recent review). If examined in detail, this theory can generate some very complex discussions beyond the scope of the present review. However, it is not a point of debate that drugs of abuse can maintain operant behavior, and therefore one can make predictions about how galanin might affect instrumental responding for these substances. Some indirect evidence exists from transgenic mouse models that suggest that galanin chronically modulates the actions of several classes of drugs of abuse. For example, mice overexpressing galanin showed a diminished locomotor response to amphetamine challenge (Kuteeva, Hökfelt & Ögren, 2005) and galanin knockout mice showed increased activity and place preference to morphine (Hawes et al., 2007). Theses findings, when placed in the context of previous pharmacological tests with opiates that also showed that galanin inhibited the formation of opiate place preference and opiate withdrawal (Zachariou, Parikh & Picciotto, 1999; Zachariou et al., 2003), demonstrate that galanin may be an endogenous inhibitor of both opiate and dopamine systems. From this, one could suggest that it might also inhibit the self-administration of these (i.e. instrumental responding), especially at high response requirements, but not on a continuous reinforcement schedule (e.g. fixed-ratio 1). However, establishing the experimental conditions that would allow the dissociation between response contingent versus noncontingent consumption are not as straightforward as for food because of the difficulty of determining what free consumption levels of amphetamine or morphine would be and what "deprivation" means in this context.

The effects of galanin on the free-consumption of another drug of abuse, ethanol, is known. Galanin microinjected into the paraventricular nucleus of the hypothalamus or third ventricle has been shown to stimulate consumption of a 4% ethanol in ethanol preferring rats (Rada et al., 2004; Schneider, et al., 2007), and voluntary ethanol consumption or ethanol loading also stimulate galanin expression in the hypothalamus (Leibowitz et al., 2003). These authors suggest that this stimulation of ethanol consumption by galanin is related to galanin stimulation of fat consumption. So, one could predict that making ethanol consumption contingent upon

a response requirement, would, as with food, show galanin reducing rather than increasing ethanol consumption.

9. Conclusions

The goal of this review was to point to an underappreciated relationship between galanin, dopamine and instrumental behavior. Clearly, the current state of this proposed role for galanin in instrumental behavior awaits much future work. However, what makes this role especially interesting as a target of investigation is that galanin may be involved in both the appetitive/ instrumental and the consummatory aspects of motivation. Assessment of galanin involvement in the "hedonic", non-homeostatic aspects of food intake, perhaps through interactions with opiate systems in the nucleus accumbens and hypothalamus (e.g. Kelly et al., 2005; Barton, York, & Bray, 1996; Dube et al., 1994), should also be further explored, since galanin may play a unique integrative function in these systems as well. These data serve to further illustrate the unique position of galanin in multiple aspects of motivation, all questions of central importance to neuroscience.

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References

- Adams AC, Clapham JC, Wynick D, Speakman JR. Feeding behaviour in galanin knockout mice supports a role of galanin in fat intake and preference. J. Neuroendocrinol 2008;20:199–206. [PubMed: 18088361]
- Akabayashi A, Koenig JI, Watanabe Y, Alexander JT, Leibowitz SF. Galanin-containing neurons in the paraventricular nucleus: a neurochemical marker for fat ingestion and body weight gain. Proc. Natl. Acad. Sci. U S A 1994;91:10375–10379. [PubMed: 7524093]
- Akmaev IG, Kalimullina LB, Sharipova LA. The central nucleus of the amygdaloid body of the brain: cytoarchitectonics, neuronal organization, connections. Neurosci. Behav. Physiol 2004;34:603–610. [PubMed: 15368908]
- Baldo A, Kelley AE. Discrete neurochemical coding of distinguishable motivational processes: insights from nucleus accumbens control of feeding. Psychopharmacol. (Berl) 2007;191:439–459.
- Balment RJ, al Barazanji K. Renal, cardiovascular and endocrine effects administered galanin in the anaesthetised rat. Regul. Pept 1992;38:71–77. [PubMed: 1374192]
- Barbano MF, Cador M. Differential regulation of the consummatory, motivational and anticipatory aspects of feeding behavior by dopaminergic and opioidergic drugs. Neuropsychopharmacol 2006;31:1371–1381.
- Barbano MF, Cador M. Opioids for hedonic experience and dopamine to get ready for it. Psychopharmacol. (Berl) 2007;191:497–506.
- Bartfai T, Hökfelt T, Langel Ü. Galanin--a neuroendocrine peptide. Crit. Rev. Neurobiol 1993;7:229– 274. [PubMed: 7693357]
- Barton C, York DA, Bray GA. Opioid receptor subtype control of galanin-induced feeding. Peptides 1996;17(2):237–240. [PubMed: 8801527]
- Barrera G, Hernandez A, Poulin JF, Laforest S, Drolet G, Morilak DA. Galanin-mediated anxiolytic effect in rat central amygdala is not a result of corelease from noradrenergic terminals. Synapse 2006;59:27–40. [PubMed: 16237681]
- Beckstead RM, Domesick VB, Nauta WJ. Efferent connections of the substantia nigra and ventral tegmental area in the rat. Brain Res 1979;175:191–217. [PubMed: 314832]

- Berridge KC. The debate over dopamine's role in instrumental behavior: the case for incentive salience. Psychopharmacol (Berl) 2007;191:391–431.
- Berridge KC, Robinson TE. What is the role of dopamine in instrumental behavior: hedonic impact, instrumental behavior learning, or incentive salience? Brain Res Brain Res Rev 1998;28:309–369. [PubMed: 9858756]
- Berridge KC, Robinson TE. Parsing reward. Trends Neurosci 2003;26:507–513. [PubMed: 12948663]
- Björklund A, Dunnett SB. Dopamine neuron systems in the brain: an update. Trends Neurosci 2007;30:194–202. [PubMed: 17408759]
- Blackshear A, Yamamoto M, Anderson BJ, Holmes PV, Lundström L, Langel Ü, Robinson JK. Intracerebroventricular administration of galanin or galanin receptor subtype 1 agonist M617 induces c-Fos activation in central amygdala and dorsomedial hypothalamus. Peptides 2007;28:1120–1124. [PubMed: 17337094]
- Bonnefond C, Palacios JM, Probst A, Mengod G. Distribution of Galanin mRNA Containing Cells and Galanin Receptor Binding Sites in Human and Rat Hypothalamus. Eur. J. Neurosci 1990;2:629–637. [PubMed: 12106297]
- Branchek TA, Smith KE, Gerald C, Walker MW. Galanin receptor subtypes. Trends Pharmacol. Sci 2000;21:109–117. [PubMed: 10689365]
- Brewer A, Echevarria DJ, Langel Ü, Robinson JK. Assessment of new functional roles for galanin in the CNS. Neuropeptides 2005;39:323–326. [PubMed: 15944029]
- Brewer A, Langel Ü, Robinson JK. Intracerebroventricular administration of galanin decreases free water intake and operant water reinforcer efficacy in water-restricted rats. Neuropeptides 2005;39:117– 124. [PubMed: 15752545]
- Brewer A, Langel Ü, Robinson JK. Intracerebroventricularly administered galanin does not alter operant reaction time or differentially reinforced high rate schedule operant responding in rats. Neurosci Lett 2004;369:245–249. [PubMed: 15464273]
- Brewer A, Robinson JK. Galanin stimulation of nonhomeostatic feeding is blocked by the addition of a response requirement. Behavioral Neuroscience. In Press
- Burazin TC, Larm JA, Gundlach AL. Regulation by osmotic stimuli of galanin-R1 receptor expression in magnocellular neurones of the paraventricular and supraoptic nuclei of the rat. J Neuroendocrinol 2001;13:358–370. [PubMed: 11264724]
- Ciosek J, Cisowska A. Centrally administered galanin modifies vasopressin and oxytocin release from the hypothalamo-neurohypophysial system of euhydrated and dehydrated rats. J. Physiol Pharmacol 2003;54:625–641. [PubMed: 14726616]
- Ciosek J, Cisowska A, Dabrowski R. Galanin affects vasopressin and oxytocin release from the hypothalamo-neurohypophysial system in haemorrhaged rats. J. Physiol. Pharmacol 2003;54:233– 246. [PubMed: 12832724]2003
- Corwin RL, Robinson JK, Crawley JN. Galanin antagonists block galanin-induced feeding in the hypothalamus and amygdala of the rat. Eur. J. Neurosci 1993;5:1528–1533. [PubMed: 7506975]
- Counts SE, McGuire SO, Sortwell CE, Crawley JN, Collier TJ, Mufson EJ. Galanin inhibits tyrosine hydroxylase expression in midbrain dopaminergic neurons. J. Neurochem 2002;83:442–451. [PubMed: 12423254]
- Counts SE, Perez SE, Ginsberg SD, De Lacalle S, Mufson EJ. Galanin in Alzheimers disease. Mol Interv 2003;23:137–156. [PubMed: 14993421]
- Craig W. Appetites and aversions as constituents of instincts. P.N.A.S 1917;3:685–688. [PubMed: 16586767]
- Craig W. Appetites and aversions as constituents of instincts. Biol. Bull 1918;34:91–107.
- Crawley JN. The role of galanin in feeding behavior. Neuropeptides 1999;33:369–375. [PubMed: 10657514]
- Crawley JN, Robinson JK, Langel Ü, Bartfai T. Galanin receptor antagonists M40 and C7 block galanininduced feeding. Brain Res 1993;600:268–272. [PubMed: 7679604]
- de Castro JM. The interactions of fluid and food intake in the spontaneous feeding and drinking patterns of rats. Physiol Behav 1989;45:861–870. [PubMed: 2780871]

Robinson and Brewer Page 12

- Dube MG, Horvath TL, Leranth C, Kalra PS, Kalra SP. Naloxone reduces the feeding evoked by intracerebroventricular galanin injection. Physiol. Behav 1994;56:811–813. [PubMed: 7528433]
- Dunham, P. The nature of reinforcing stimuli. In: Honig, WK.; Staddon, JER., editors. Handbook of operant behavior. Englewood Cliffs, NJ: Prentice-Hall; 1977. p. 98-124.
- Dunning BE, Taborsky GJ Jr. Galanin-sympathetic neurotransmitter in endocrine pancreas? Diabetes 1988;37:1157–1162. [PubMed: 2457528]
- Echevarria DJ, Brewer A, Bushell G, Manuzon H, Langel Ü, Robinson JK. Galanin and perseveration. Brain Res 2005;1041:143–148. [PubMed: 15829223]
- Elvander E, Ogren SO. Medial septal galanin and acetylcholine: influence on hippocampal acetylcholine and spatial learning. Neuropeptides 2005;39:245–248. [PubMed: 15944017]
- Elvander E, Schött PA, Sandin J, Bjelke B, Kehr J, Yoshitake T, Ogren SO. Intraseptal muscarinic ligands and galanin: influence on hippocampal acetylcholine and cognition. Neurosci 2004;126:541–557.
- Ericson E, Ahlenius S. Suggestive evidence for inhibitory effects of galanin on mesolimbic dopaminergic neurotransmission. Brain Res 1999;822:200–209. [PubMed: 10082897]
- Falk JL. Production of polydipsia in normal rats by an intermittent food schedule. Science 1961;133:195– 196. [PubMed: 13698026]
- Florén A, Land T, Langel Ü. Galanin receptor subtypes and ligand binding. Neuropeptides 2000;34:331– 337. [PubMed: 11162289]
- Geisler S, Zahm DS. Afferents of the ventral tegmental area in the rat-anatomical substratum for integrative functions. J. Comp. Neurol 2005;490:270–294. [PubMed: 16082674]
- Gundlach AL, Burazin TC, Larm JA. Distribution, regulation and role of hypothalamic galanin systems: renewed interest in a pleiotropic peptide family. Clin. Exp. Pharmacol. Physiol 2001;28:100–105. [PubMed: 11153523]
- Hawes JJ, Brunzell DH, Narasimhaiah R, Langel Ü, Wynick D, Picciotto MR. Galanin Protects Against Behavioral and Neurochemical Correlates of Opiate Reward. Neuropsychopharmacology. 2007Oct 24 epub ahead of print
- Hawes JJ, Picciotto MR. Characterization of GalR1, GalR2, and GalR3 immunoreactivity in catecholaminergic nuclei of the mouse brain: Erratum. J Comp Neurol 2005;490:98–100.
- Holets VR, Hökfelt T, Rökaeus A, Terenius L, Goldstein M. Locus coeruleus neurons in the rat containing neuropeptide Y, tyrosine hydroxylase or galanin and their efferent projections to the spinal cord, cerebral cortex and hypothalamus. Neurosci 1988;24:893–906.
- Holmes FE, Mahoney SA, Wynick D. Use of genetically engineered transgenic mice to investigate the role of galanin in the peripheral nervous system after injury. Neuropeptides 2005;39:191–199. [PubMed: 15944011]
- Hökfelt T. Galanin and its receptors: introduction to the Third International Symposium, San Diego, California, USA, 21–22 October 2004. Neuropeptides 2005;39:125–142.
- Hursh SR. Economic concepts for the analysis of behavior. J. Exp. Anal. Behav 1980;34:219–238. [PubMed: 16812188]
- Hyman SE, Malenka RC, Nestler EJ. Neural Mechanisms of Addiction: The Role of Reward-Related Learning and Memory. Annu. Rev. Neurosci 2006;29:565–598. [PubMed: 16776597]
- Jhamandas JH, Harris KH, MacTavish D, Jassar BS. Novel excitatory actions of galanin on rat cholinergic basal forebrain neurons: implications for its role in Alzheimer's disease. J. Neurophysiol 2002;87:696–704. [PubMed: 11826038]
- Kai A, Ono K, Kawano H, Honda E, Nakanishi O, Inenaga K. Galanin inhibits neural activity in the subfornical organ in rat slice preparation. Neuroscience 2006;143:769–777. [PubMed: 17027169]
- Kelley AE, Baldo BA, Pratt WE, Will MJ. Corticostriatal-hypothalamic circuitry and food motivation: integration of energy, action and instrumental behavior. Physiol. Behav 2005;86:773–795. [PubMed: 16289609]
- Koenig JI, Hooi S, Gabriel SM, Martin JB. Potential involvement of galanin in the regulation of fluid homeostasis in the rat. Regul Pept 1989;24:81-86. [PubMed: 2472648]
- Kondo K, Murase T, Otake K, Ito M, Oiso Y. Centrally administered galanin inhibits osmotically stimulated arginine vasopressin release in conscious rats. Neurosci. Lett 1991;22:245–248. [PubMed: 1719452]

- Krebs JR, Kacelnik A. Time horizons of foraging animals. Ann. N. Y. Acad. Sci 1984;423:278–291. [PubMed: 6588792]
- Kuteeva E, Hökfelt T, Ögren SO. Behavioural characterisation of young adult transgenic mice overexpressing galanin under the PDGF-B promoter. Regul. Pept 2005;125:67–78. [PubMed: 15582716]
- Kyrkouli SE, Stanley BG, Leibowitz SF. Galanin: stimulation of feeding induced by medial hypothalamic injection of this novel peptide. Eur. J. Pharmacol 1986;122:159–160. [PubMed: 2420618]
- Kyrkouli SE, Stanley BG, Seirafi RD, Leibowitz SF. Stimulation of feeding by galanin: anatomical localization and behavioral specificity of this peptide's effects in the brain. Peptides 1990;11:995– 1001. [PubMed: 1704616]
- Landry M, Aman K, Hökfelt T. Galanin-R1 receptor in anterior and mid-hypothalamus: distribution and regulation. J. Comp. Neurol 1998;399:321–340. [PubMed: 9733081]
- Landry M, Roche D, Calas A. Short-term effects of centrally administered galanin on the hyperosmotically stimulated expression of vasopressin in the rat hypothalamus. An in situ hybridization and immunohistochemistry study. Neuroendocrinology 1995;61:393–404. [PubMed: 7540263]
- Landry M, Vila-Porcile E, Hökfelt T, Calas A. Differential routing of coexisting neuropeptides in vasopressin neurons. Eur. J. Neurosci 2003;17:579–589.
- Lang R, Gundlach AL, Kofler B. The galanin peptide family: receptor pharmacology, pleiotropic biological actions, and implications in health and disease. Pharmacol. Ther 2007;115:177–207. [PubMed: 17604107]
- Lawrence CB, Williams T, Luckman SM. Intracerebroventricular galanin-like peptide induces different brain activation compared with galanin. Endocrinol 2003;144:3977-39784.
- Leibowitz SF. Regulation and effects of hypothalamic galanin: relation to dietary fat, alcohol ingestion, circulating lipids and energy homeostasis. Neuropeptides 2005;39:327–332. [PubMed: 15944030]
- Leibowitz SF. Overconsumption of dietary fat and alcohol: mechanisms involving lipids and hypothalamic peptides. Physiol. Behav 2007;91:513–521. [PubMed: 17481672]
- Leibowitz SF, Akabayashi A, Wang J. Obesity on a high-fat diet: role of hypothalamic galanin in neurons of the anterior paraventricular nucleus projecting to the median eminence. J. Neurosci 1998;18:2709– 2719. [PubMed: 9502828]
- Leibowitz SF, Avena NM, Chang GQ, Karatayev O, Chau DT, Hoebel BG. Ethanol intake increases galanin mRNA in the hypothalamus and withdrawal decreases it. Physiol. Behav 2003;79:103–111. [PubMed: 12818715]
- Lu X, Mazarati A, Sanna P, Shinmei S, Bartfai T. Distribution and differential regulation of galanin receptor subtypes in rat brain: effects of seizure activity. Neuropeptides 2005;39:147–152. [PubMed: 15944003]
- Mathis CE, Johnson DF, Collier GH. Procurement time as a determinant of meal frequency and meal duration. J. Exp. Anal. Behav 1995;63:295–311. [PubMed: 7751834]
- Meister B, Cortes R, Villar MJ, Schalling M, Hökfelt T. Peptides and transmitter enzymes in hypothalamic magnocellular neurons after administration of hyperosmotic stimuli: comparison between messenger RNA and peptide/protein levels. Cell Tissue Res 1990;260:279–297. [PubMed: 1694105]
- Melander T, Kohler C, Nilsson S, Hökfelt T, Brodin E, Theodorsson E, Bartfai T. Autoradiographic quantitation and anatomical mapping of $125I$ -galanin binding sites in the rat central nervous system. J. Chem. Neuroanat 1988;1:213–233. [PubMed: 2477035]
- Mennicken F, Hoffert C, Pelletier M, Ahmad S, O'Donnell D. Restricted distribution of galanin receptor 3 (GalR3) mRNA in the adult rat central nervous system. J. Chem. Neuroanat 2002;24:257–268. [PubMed: 12406501]
- Merchenthaler I, Lopez FJ, Negro-Vilar A. Anatomy and physiology of central galanin-containing pathways. Prog. Neurobiol 1993;40:711–769. [PubMed: 7683433]
- Morilak DA, Cecchi M, Khoshbouei H. Interactions of norepinephrine and galanin in the central amygdala and lateral bed nucleus of the stria terminalis modulate the behavioral response to acute stress. Life Sci 2003;73:715–726. [PubMed: 12801593]
- Nicola SM. The nucleus accumbens as part of a basal ganglia action selection circuit. Psychopharmacol. (Berl) 2007;191:521–550.

- O'Donnell D, Ahmad S, Wahlestedt C, Walker P. Expression of the novel galanin receptor subtype GALR2 in the adult rat CNS: distinct distribution from GALR1. J. Neurol 1999;409:469–481.
- Ögren SO, Razani H, Elvander-Tottie E, Kehr J. The neuropeptide galanin as an in vivo modulator of brain 5-HT1A receptors; possible relevance for affective disorders. Physiol. Behav 2007;92:172– 179. [PubMed: 17585970]
- Parrado C, Díaz-Cabiale Z, García-Coronel M, Agnati LF, Coveñas R, Fuxe K, Narváez JA. Region specific galanin receptor/neuropeptide Y Y1 receptor interactions in the tel- and diencephalon of the rat. Relevance for food consumption. Neuropharmacol 2007;52:684–692.
- Phillipson OT. Afferent projections to the ventral tegmental area of Tsai and interfascicular nucleus: A horseradish peroxidase study in the rat. J. Comp. Neurol 1979;187:117–143. [PubMed: 489776]
- Phillips AG, Vacca G, Ahn S. A top-down perspective on dopamine, motivation and memory. Pharmacol. Biochem. Behav. 2007in press
- Pieribone VA, Xu ZQ, Zhang X, Grillner S, Bartfai T, Hökfelt T. Galanin induces a hyperpolarization of norepinephrine-containing locus coeruleus neurons in the brainstem slice. Neurosci 1995;64:861– 874.
- Premack D. Reversibility of the reinforcement relation. Science 1962;136:255–257. [PubMed: 14488597]
- Rada P, Avena NM, Leibowitz SF, Hoebel BG. Ethanol intake is increased by injection of galanin in the paraventricular nucleus and reduced by a galanin antagonist. Alcohol 2004;33:91–97. [PubMed: 15528006]
- Rada P, Mark GP, Hoebel BG. Galanin in the hypothalamus raises dopamine and lowers acetylcholine release in the nucleus accumbens: a possible mechanism for hypothalamic initiation of feeding behavior. Brain Res 1998;798:1–6. [PubMed: 9666056]
- Robinson JK. Galanin and cognition. Behav. Cogn. Neurosci. Rev 2004;3:222–242. [PubMed: 15812108]
- Robinson JK, Crawley JN. Intraventricular galanin impairs delayed nonmatching-to-sample performance in rats. Behav. Neurosci 1993;107:458–467. [PubMed: 7687133]
- Rökaeus Å, Melander T, Hökfelt T, Lundberg JM, Tatemoto K, Carlquist M, Mutt VA. A galanin-like peptide in the central nervous system and intestine of the rat. Neurosci. Lett 1984;47:161–166. [PubMed: 6205331]
- Rökaeus Å, Young WS III, Mezey É. Galanin coexists with vasopressin in the normal ra hypothalamus and galanin's synthesis is increased in the Brattleboro (diabetes insipidus) rat. Neurosci. Lett 1988;19:45–50.
- Salamone JD, Correa M. Motivational views of reinforcement: implications for understanding the behavioral functions of nucleus accumbens dopamine. Behav. Brain Res 2002;137:3–25. [PubMed: 12445713]
- Salamone JD, Correa M, Mingote SM, Weber SM. Beyond the instrumental behavior hypothesis: alternative functions of nucleus accumbens dopamine. Curr. Opin. Pharmacol 2005;5:34–41. [PubMed: 15661623]
- Salamone JD, Correa M, Farrar A, Mingote SM. Effort-related functions of nucleus accumbens dopamine and associated forebrain circuits. Psychopharmacology (Berl) 2007;191:461–482. [PubMed: 17225164]
- Saper CB, Loewy AD. Efferent connections of the parabrachial nucleus in the rat. Brain Res 1980;197:291–317. [PubMed: 7407557]
- Schneider ER, Rada P, Darby RD, Leibowitz SF, Hoebel BG. Orexigenic peptides and alcohol intake: differential effects of orexin, galanin, and ghrelin. Alcohol Clin Exp Res 2007;31:1858–1865. [PubMed: 17850217]
- Seutin V, Verbanck P, Massotte L, Dresse A. Galanin decreases the activity of locus coeruleus neurons in vitro. Eur. J. Pharmacol 1989;164:373–376. [PubMed: 2474450]
- Sevcik J, Finta EP, Illes P. Galanin receptors inhibit the spontaneous firing of locus coeruleus neurones and interact with mu-opioid receptors. Eur. J. Pharmacol 1993;230:223–230. [PubMed: 7678551]
- Simon H, Le Moal M, Calas A. Efferents and afferents of the ventral tegmental-A10 region studied after local injection of [3H]leucine and horseradish peroxidase. Brain Res 1979;178:17–40. [PubMed: 91413]

Robinson and Brewer Page 15

- Skofitsch G, Jacobowitz DM. Immunohistochemical mapping of galanin-like neurons in the rat central nervous system. Peptides 1985;6:509–546. [PubMed: 2415952]
- Skofitsch G, Jacobowitz DM. Quantitative distribution of galanin-like immunoreactivity in the rat central nervous system. Peptides 1986;7:609–613. [PubMed: 2429289]
- Skofitsch G, Jacobowitz DM, Amann R, Lembeck F. Galanin and vasopressin coexist in the rat hypothalamo-neurohypophyseal system. Neuroendocrinol 1989;49:419–427.
- Skofitsch G, Sills MA, Jacobowitz DM. Autoradiographic distribution of ^{125}I -galanin binding sites in the rat central nervous system. Peptides 1986;7:1029–1042. [PubMed: 2436195]1986
- Tatemoto K, Rokaeus A, Jornvall H, McDonald TJ, Mutt V. Galanin a novel biologically active peptide from porcine intestine. FEBS Lett 1983;164:124–128. [PubMed: 6197320]
- Tempel DL, Leibowitz KJ, Leibowitz SF. Effects of PVN galanin on macronutrient selection. Peptides 1988;9:309–314. [PubMed: 2453854]
- Timberlake W, Allison J. Response deprivation: An empirical approach to instrumental performance. Psychological Review 1974;81:146–164.
- Tsuda K, Tsudab S, Nishioa I, Masuyamac Y, Goldstein M. Effects of galanin on dopamine release in the central nervous system of normotensive and spontaneously hypertensive rats. American Journal of Hypertension 1998;11:1475–1479. [PubMed: 9880130]
- Villar MJ, Meister B, Cortes R, Schalling M, Morris M, Hökfelt T. Neuropeptide gene expression in hypothalamic magnocellular neurons of normal and hypophysectomized rats: a combined immunohistochemical and in situ hybridization study. Neurosci 1990;36:181–199.
- Wiesenfeld-Hallin Z, Xu XJ, Crawley JN, Hökfelt T. Galanin and spinal nociceptive mechanisms: recent results from transgenic and knock-out models. Neuropeptides 2005;39:207–210. [PubMed: 15944013]
- Wise RA. Neuroleptics and operant behavior: the anhedonia hypothesis. Behav Brain Sci 1982;5:39–87.
- Xu ZQ, Shi TJ, Hökfelt TJ. Galanin/GMAP- and NPY-like immunoreactivities in locus coeruleus and noradrenergic nerve terminals in the hippocampal formation and cortex with notes on the galanin-R1 and -R2 receptors. J. Comp. Neurol 1998;392:227–251. [PubMed: 9512271]
- Xu ZQ, Zhang X, Pieribone VA, Grillner S, Hökfelt T. Galanin-5-hydroxytryptamine interactions: electrophysiological, immunohistochemical and in situ hybridization studies on rat dorsal raphe neurons with a note on galanin R1 and R2 receptors. Neurosci 1998;87:79–94.
- Yagita K, Okamura H, Ibata Y. Rehydration process from salt-loading: recovery of vasopressin and its coexisting galanin, dynorphin and tyrosine hydroxylase immunoreactivities in the supraoptic and paraventricular nuclei. Brain Res 1994;667:13–23. [PubMed: 7534608]
- Zachariou V, Brunzell DH, Hawes J, Stedman DR, Bartfai T, Steiner RA, Wynick D, Langel Ü, Piccioto MR. The neuropeptide galanin modulates behavioral and neurochemical signs of opiate withdrawal. Proc. Natl. Acad. Sci. USA 2003;100:9028–9033. [PubMed: 12853567]
- Zachariou V, Parikh K, Picciotto MR. Centrally administered galanin blocks morphine place preference in the mouse. Brain Res 1999;831:33–42. [PubMed: 10411981]

Progressive Ratio Schedule

Figure 1.

Galanin (10µg i.c.v.) stimulated consumption of a high-fat milk solution when it was made freely available in a dish in a tub cage (top panel, $** = p \lt 0.001$) but not when consumption was based upon the completion of a progressive ratio reinforcement schedule (bottom panel).

Table 1

Effects of galanin (i.c.v) on measures of water consumption. Across a variety of instrumental and response-independent test, galanin consistently reduced the persistence of responding and/or amount of water consumed. In the operant tasks, no instance of a separable effect on the main performance measures (i.e. discrimination, working memory) were observed, consistent with an interpretation that the primary effects of galanin across all of the tasks was on the incentive value of the reinforcer and not secondary to the impairment of discrimination, non-matching, working memory and such. All studies were conducted in twenty-three hour water restricted rats. $nd = not$ determined.

1 Brewer, Langel & Robinson, 2004

2 Echevarria, Brewer, Manuzon, Langel & Robinson, 2005

3 Robinson & Crawley, 1993a

4 Brewer, Langel & Robinson, 2005

5 Saccone, Brewer, Echevarria & Robinson, Unpublished.