

NIH Public Access

Author Manuscript

Org Lett. Author manuscript; available in PMC 2009 January 27

Published in final edited form as:

Org Lett. 2006 October 26; 8(22): 4985–4988. doi:10.1021/ol060946z.

Efficient microwave-assisted synthesis of amine substituted pentafluorophenylporphyrin

Diana Samaroo¹, Clifford E. Soll¹, Louis J. Todaro¹, and Charles M. Drain^{1,2,*}

1 Department of Chemistry & Biochemistry, Hunter College and the Graduate Center of the City University of New York, 695Park Ave, New York, NY 10021

2 The Rockefeller University, 1230 York Avenue, New York, NY 10021

Abstract



We report an efficient and rapid means for the synthesis of tetrapentafluorophenyl porphyrin (TPPF₂₀) derivatives by microwave irradiation in an environmentally acceptable solvent. The selective displacement of the para-fluorine groups in TPPF₂₀ by primary amines occurs in yields between 70–95%. This method demonstrates that TPPF₂₀ is an ideal platform for the rapid formation of porphyrin conjugates for therapeutic, catalytic and other applications.

Applications of porphyrin derivatives range from catalysts,¹ materials, devices² to photodynamic therapeutic agents (PDT)³ because of their rich photochemistry and redox chemistry. However, most applications require modification of the porphyrin macrocycle to allow attachment of additional substituents with various other functionalities.

Microwave-assisted reactions have become increasingly important in chemical synthesis in the last 20 years due to the advantages it provides over conventional heating methods.⁴ Significant reduction in reaction times, side reactions, increased yields, ease of purification and minimization of the amount of solvent used are only a few of these desirable qualities.⁵ Shorter reaction times (usually <15 min) allow rapid investigation into new methodologies and reaction optimization. Microwave-assisted reactions are believed to facilitate polarization of the substrates thereby promoting the reactions.⁶

We previously demonstrated a solvent-free⁷ porphyrin synthesis, and solid-supported reactions minimizes the amount of subsequent purification.⁸ In view of the significant need for rapid synthesis of porphyrin derivatives – especially for therapeutic applications – we present methods for the facile preparation of porphyrin derivatives around a core platform of 5,10,15,20-tetrakis(2,3,4,5,6-pentafluorophenyl)porphyrin (TPPF₂₀). As a demonstration of the method, a variety of amphipathic moieties are grafted onto the core for evaluation as photodynamic therapeutic (PDT) agents.

cdrain@hunter.cuny.edu.

Previously reported TPPF₂₀ derivatives were typically synthesized in refluxing DMF with a large excess of the amine or thiol reagents,⁹ but the yields were relatively poor and the substitution was sometimes incomplete. Better yields, ~90%, are obtained by using thiol reagents, and stirring for ~12 h at room temperature in DMF with a stoichiometric amount of dialkylamine.^{3a,c} To decrease the reaction time, improve yields, and employ a greener solvent, we developed a synthesis using microwave irradiation (MW) in N-methylpyrrolidone.

A diverse array of primary amines were chosen to demonstrate the method. Polyethylene glycols (PEG) are relevant to drug design and delivery as they enable the facile uptake of drugs into cells¹⁰; polylysine derivatives because these moieties are known to impart selectivity towards cancer cells; and a polyamine because these impart both cancer cell selectivity and cell permeability. The functional groups other than the intended amine are t-boc protected.

Reaction of 5–10 mg (5.1 or 10.2 μ mol) TPPF₂₀ with 10 equivalents of a primary amine in 1– 2 mL N-methylpyrrolidone (NMP) at 60–70 °C overnight leads to mixtures of six different TPPF₂₀ derivatives. However, the same amounts of porphyrin and amine in 100 μ L of NMP and 10 minutes MW irradiation yields only the tetra substituted products. The reaction time was dramatically reduced (see Table 1 and abstract). The nucleophilic aromatic substitution of the para-fluoro group by primary amines take 10–30 minutes in this procedure compared to 1– 2 days with conventional heating.¹¹ A significant added advantage for the MW reaction, is that it tolerates functional groups that can decompose under extended heating.

The improved yields of the tetra-substituted derivatives are concomitant with the reduced amounts of incompletely reacted products. Also, prolonged heating in DMF results in the fragmenting of the PEG moieties (data not shown). A 95% yield of the Por-PEG₄ product indicates that each reaction on the porphyrin proceeds with ~99% efficiency. In general, the protected acids are used because these are more soluble in the NMP and are found to result in greater yields. The deprotection of these groups is readily accomplished using literature procedures. For the EDA and the PEG the amide esters are cleaved using HCl in MeOH. For the Lys derivative, 6N HCl is used to cleave both the acetamide and methyl ester groups.

These conditions are specific for para fluoride substitution by primary amines, because under similar reaction conditions secondary amines exhibit no reactivity. Refluxing of TPPF₂₀ in DMF is known to yield dimethylamino substitution on the para position, thus previous methods require temperatures <60 °C and extended reaction times. When 5 mg TPPF₂₀ in DMF is heated in the microwave for 10 minutes, several dimethylamino derivatives are observed. Procedures in other solvents, such as toluene, DMSO and morpholine generally result in low reactivity or significantly diminished yields compared to NMP.

All porphyrins were characterized by electrospray ionization mass spectrometry (ESI-MS, see Supporting Information). For all porphyrins, the parent $[M+H^+]$ was seen and in some cases the sodium adduct. The electronic absorption spectra of the porphyrins were characterized at 1µM concentrations and were found to be typical for non-aggregated porphyrins. Substitution of the electron withdrawing fluorine with the electron donating amine results in a 7–9 nm red shift in the Soret. Table 2 summarizes the UV-Visible spectra of these porphyrins in methanol. All the derivatives show similar spectra, including the B band or Soret band at 417–419 nm, the Q-band at 508–509 nm together with three vibronic bands at 543–545 nm, 585–587 nm, and 643–649 nm. A singlet for the pyrrole H in the ¹H-NMR indicates the presence of the four para substituents, whereas incomplete substitution results in multiplets for this proton.

The X-ray crystal structure of Por-EDA₄ derivative was elucidated (Fig. 1). This derivative was crystallized from layered hexane over toluene at room temperature over several weeks. There are two different EDA conformations, 'bent' and extended; the bent conformation organized by an intramolecular H-bond. The most interesting aspect of the structure is that it

Org Lett. Author manuscript; available in PMC 2009 January 27.

forms a two-dimensional lattice (Fig. 2) organized by intermolecular N-H····O=C hydrogen bonds. The bent EDA moieties form a cyclic structure mediated intramolecular hydrogen bonds. The extended EDA moieties on opposite sides of the porphyrin H-bond to the bent EDA moieties on adjacent porphyrins. (See packing diagram Fig. 2). There is a surprising lack of porphyrin pi-stacking in the crystal structure, and the layers are organized by dispersion and packing forces. The intramolecular and intermolecular hydrogen-bonding motifs were not expected, and may explain the aggregation of these compounds at unexpectedly low concentrations.

One of the immediate goals for these porphyrin derivatives is to evaluate their uptake into cancer cells. Thus the solubility of these protected derivatives in an aqueous cell culture medium (Dulbecco's modified eagle medium without phenol red or phosphate buffered saline) must be considered. The absorption spectra of Por-EDA₄ in this medium show a red shift and substantial broadening of the Soret band, from 417 nm to 431 nm (supporting information), and similar spectral changes are observed for the other derivatives. These changes in the electronic spectra are indicative of aggregate formation.

The fluorescence emission of these derivatives in methanol and aqueous solvents was also recorded. For Por-EDA₄ in methanol, excitation at 417 nm in the Soret band, resulted in a strong fluorescence emission at 647–652 nm with a Stokes shift of 7–12 nm, which is typical of porphyrins. In the cell culture medium, the ground state spectra are red shifted by 6–10 nm, thus there is a similar red shift in the emission bands (Table 2).

Based on the above results for primary amines we hypothesized that combinatorial libraries of porphyrins can be created by this method as long as the individual reagents react with the $TPPF_{20}$ with similar rates and efficiencies. As a proof-of-principle experiment, a 1:1:1 mixture of the three amine reagents were used in a solution phase combinatorial reaction. The total amount of the three primary amines were present in the reaction at the same 10:1 ratio with the porphyrin as used above. Statistically this should yield 21 compounds, but ESI-MS analysis indicated incomplete reactivity as a significant number of compounds with one or more unsubstituted fluorines were observed. Sequential addition of the less reactive amine followed by the more reactive amines also yields inconsistent results.

In summary, protected amine conjugates of tetrapentafluorophenyl porphyrin with polyethylene glycol, ethylenediamine, and lysine groups were synthesized by microwave irradiation. The protocol described allows for significant reduction in reaction times yet results in high yields and limits by-product formation compared to earlier reports.¹¹ The procedure uses commercially availabe reagents at ca. 10-fold greater concentrations in an environmentally friendly solvent. In general the MW method works for primary nucleophiles significantly better than secondary, and better for amines than for sulfides. ¹²

In future studies, the prepared protected and deprotected porphyrin conjugates will be assessed for their photodynamic efficiency using MDA-MB-231 cancer cells. The crystal structure indicates these types of porphyrins may self-organize into photonic materials.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgements

This work was supported by NIH SCORE (GM60654), NSF CHE-0554703 and IGERT DGE-9972892. DS acknowledges NSF LS-AMP and MAGNET graduate fellowships. Hunter College Chemistry infrastructure is supported by the NSF, NIH, including the RCMI program (G12-RR-03037), and the City University of New York.

Org Lett. Author manuscript; available in PMC 2009 January 27.

We thank Dr. Mikki Vinodu of Hunter College of the City University of New York for careful reading of the manuscript.

References

- 1. (a) Benaglia M, Danelli T, Fabris F, Sperandio D, Pozzi G. Org Lett 2002;4:4229. [PubMed: 12443065]
 (b) Yang J, Weinberg R, Breslow R. Chem Commun 2000:531.
- 2. Drain CM, Hupp JT, Suslick KS, Wasielewski MR, Chen X. J Porphyrins Phthalocyanines 2002;6:243.
- (a) Chen X, Hui L, Foster DA, Drain CM. Biochemistry 2004;43:10918. [PubMed: 15323552] (b) Sylvain I, Zerrouki R, Granet R, Huang YM, Lagorce JF, Guilloton M, Blais JC, Krausz P. Bioorg Med Chem 2002;10:57. [PubMed: 11738607] (c) Pasetto P, Chen X, Drain CM, Franck RW. Chem Commun 2001:82.
- 4. Sauer DR, Kalvin D, Phelan KM. Org Lett 2003;5:4721. [PubMed: 14627424]
- Yoon DS, Han Y, Stark TM, Haber JC, Gregg BT, Stankovich SB. Org Lett 2004;6:4775. [PubMed: 15575683]
- 6. (a) Liu MO, Tai CH, Wang WY, Chen JR, Hu AT, Wei TH. J Organomet Chem 2004;689:1078. (b) Liu MO, Hu AT. J Organomet Chem 2004;689:2450. (c) Boufatah N, Gellis A, Maldonado J, Vanelle P. Tetrahedon 2004;60:9131.
- 7. Drain CM, Gong X. Chem Commun 1997:2117.
- (a) Chaouchi M, Loupy A, Marque S, Petit A. Eur J Org Chem 2002:1278. (b) Mojtahedi MM, Saidi MR, Shirzi JS, Bolourtchian M. Syn Commun 2002;32:851.
- (a) Shaw SJ, Elgie KJ, Edwards C, Boyle RW. Tetrahedron Lett 1999;40:1595. (b) Suzuki M, Shimizu S, Shin JY, Osuka A. Tetrahedron Lett 2003;44:4597. (c) Battioni P, Brigaud O, Desvaux H, Mansuy D, Traylor TG. Tetrahedron Lett 1991;32:2893.
- (a) Molineux G. Pharmacotherapy 2003;23:3S. [PubMed: 12921216] (b) Hamblin MR, Miller JL, Rizvi I, Ortel B, Maytin EV, Hasan T. Cancer Res 2001;61:7155. [PubMed: 11585749]
- 11. Battioni P, Brigaud O, Desvaux H, Mansuy D, Traylor TG. Tetrahedron Lett 1991;32:2893.
- 12. General Procedure for the Preparation of Amine Derivatives of TPPF_{20} by Microwave Irradiation. Into a 3.4 mL vial, 5 mg of 5,10,15,20-tetrakispentafluorophenylporphyrin (5.1 µmol) and 10 equivalents of mono-N-t-boc EDA (Quanta BioDesign) in NMP (0.1 mL) was added. The closed vial was irradiated using a domestic microwave oven (1100 W, Samsung MW4250W) at 2 min intervals until no starting material was visualized by TLC (10 min). After the vial was cooled, the solvent (NMP) was removed in vacuo. The reaction mixture was purified on a silica gel prep TLC plate (hexane : ethyl acetate : methanol, 2 : 1.5 : 0.5). The porphyrin was redissolved in methanol and evaporated under reduced pressure to afford >90 % yield.



Figure 1. X-ray Crystal Structure of Por-EDA₄ H white; C grey; N blue; O red; F green.

Org Lett. Author manuscript; available in PMC 2009 January 27.



Figure 2. X-ray Crystal Packing Structure of Por-EDA₄ Illustrating the Inter- and Intra- molecular H-bonding H white; C grey; N blue; O red; F green

NIH-PA Author Manuscript	
IIH-PA Author Manuscript	7
IH-PA Author Manuscript	~
H-PA Author Manuscript	=
I-PA Author Manuscript	
PA Author Manuscript	T
PA Author Manuscript	÷.
A Author Manuscript	Ū
Author Manuscript	$\mathbf{\Sigma}$
Author Manuscript	-
Author Manuscript	~
uthor Manuscript	1
thor Manuscript	
hor Manuscript	=
or Manuscript	-
or Manuscript	5
r Manuscript	0
Manuscript	_
Manuscript	-
lanuscript	\leq
anuscript	5
nuscript	<u><u></u></u>
uscript	⊐
script	–
script	5
cript	0
ript	0
ਰੂ	Ξ.
¥	
-	9

 Table 1

 Comparison of Conventional versus Microwave Heating in the Synthesis of Protected Porphyrin Derivatives^a

	solvent	Conventional heating ^b	yield	Microwave irradiation	yield
R=-HN~~o~o~o~n~o~o~	NMP	2 days	~50%	12 min	94 ± 19
R=-HN_NLOX Por-EDA4	NMP	1 day	~50%	10 min	$94 \pm 2^{\circ}$
R= _HN ~~~ HC _NH Por-Lys 4 COCH 3	NMP	20 h	72%	30 min	77 ± 5'

Samaroo et al.

residue was purified by silica gel column chromatography.

 $b_{\rm In}$ an oil bath maintained at 60–70 °C.

Table 2
ESI-MS, UV-Vis and Emission data of Protected Porphyrin Derivatives

derivative	mass ^a	UV-vis ^b	emission ^b
TPPF ₂₀	975	406, 502 (536, sh), 580 (634, sh)	640, 707
Por-PEG ₄	2176	418, 509, 545, 586, 643	652, 714
$Por-EDA_4$	1535	417, 508, 543, 585, 644	647, 713
Por-Lys ₄	1703	419, 509, 545 587, 649	652, 711

^a[M+H]⁺

^bAbsorption and emission in and methanol.