Alterations in Glucose Disposal in Sleep-disordered Breathing

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Rationale: It is well established that sleep-disordered breathing (SDB) is independently associated with insulin resistance, glucose intolerance, and type 2 diabetes mellitus. However, data on whether SDB alters *in vivo* kinetics of glucose and insulin are lacking.

Objectives: The primary goal of this study was to use the frequently sampled intravenous glucose tolerance test (FSIVGTT) in subjects with and without SDB to model the *in vivo* kinetics of glucose and insulin. Minimal model analysis of the FSIVGTT data was used to derive parameters of insulin sensitivity, glucose effectiveness (a measure of the ability of glucose to mediate its own disposal), and pancreatic β -cell function.

Results: A total of 118 nondiabetic subjects underwent polysomnography, the FSIVGTT, and body composition measurements including determination of percent body fat. Compared with normal subjects (apnea-hypopnea index < 5 events/h), those with mild, moderate, and severe SDB displayed a 26.7, 36.5 and 43.7% reduction in insulin sensitivity, respectively, independent of age, sex, race, and percent body fat. The disposition index, an integrated measure of pancreatic β -cell function, was also reduced in patients with moderate to severe SDB. The decrease in insulin sensitivity and the disposition index were correlated with the average degree of oxyhemoglobin desaturation. In contrast, glucose effectiveness was negatively correlated with the frequency of respiratory event–related arousals.

Conclusions: The results of this study suggest that, independent of adiposity, SDB is associated with impairments in insulin sensitivity, glucose effectiveness, and pancreatic β -cell function. Collectively, these defects may increase the risk of glucose intolerance and type 2 diabetes mellitus in SDB.

Keywords: sleep-disordered breathing; sleep apnea; glucose metabolism; insulin resistance; beta-cell function

Sleep-disordered breathing (SDB) is a relatively common condition that has increased dramatically in the general population in tandem with obesity. Characterized by recurrent collapse of the upper airway, SDB results in episodic drops in oxyhemoglobin saturation and transient arousals from sleep. Epidemiologic studies have shown that SDB is a risk factor for hypertension (1), cardiovascular disease (2), and all-cause mortality (3, 4). Independent of obesity, SDB has also been associated with insulin resistance, glucose intolerance, and type 2 diabetes mellitus (5, 6). Although much of the evidence on SDB and glucose metabolism has been relatively consistent,

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AT A GLANCE COMMENTARY

Scientific Knowledge on the Subject

Sleep-disordered breathing (SDB) is associated with insulin resistance and type 2 diabetes mellitus. Whether SDB is also associated with alterations in glucose disposal independent of insulin action and in pancreatic insulin secretion is not known.

What This Study Adds to the Field

Independent of obesity, patients with SDB exhibit impairments in insulin sensitivity, glucose disposal independent of insulin action, and pancreatic insulin secretion.

almost all of the published reports have used body mass index (BMI) to account for the confounding effects of adiposity. It is well established that BMI does not discriminate between muscle and adipose tissue and cannot provide an assessment of body fat distribution. Moreover, age, sex, and ethnicity modify the relation between BMI and body fat, indicating that BMI may not accurately reflect adiposity across different population subsets (7, 8). In contrast, techniques such as dual-energy X-ray absorptiometry (DEXA) provide a more precise estimate of body fat mass and thus would allow for a more rigorous assessment of whether SDB and metabolic dysfunction are independently associated.

In addition to the issue of residual confounding due to adiposity, it is important to recognize that a majority of available studies on SDB and metabolic function have used steady-state measurements to assess glucose metabolism. Simple surrogate metrics, such as the homeostasis model of insulin resistance (9), have been used to correlate the severity of SDB with the degree of insulin resistance. Although such studies have been highly informative and paved the way for additional research, a major drawback with the use of steady-state measures is their inability to characterize the dynamic relation between insulin sensitivity and insulin secretion. A decrease in peripheral insulin sensitivity is fed back to the pancreatic β -cell, which increases insulin output to maintain normal glucose tolerance (10, 11). A defect in this compensatory response in the face of insulin resistance is central to the pathogenesis of glucose intolerance and type 2 diabetes mellitus (12). Thus, although SDB may lead to deterioration in insulin sensitivity, information on SDB-related alterations in pancreatic β -cell function is lacking.

In light of the above issues, the aim of the current study was to quantify the pathophysiologic alterations responsible for altered glucose metabolism in patients with SDB. To provide new insights into potential mechanisms, the frequently sampled intravenous glucose tolerance test (FSIVGTT) was used along with the minimal model analysis of the resulting data. The minimal model describes the *in vivo* kinetics of glucose and insulin and provides estimates of peripheral insulin sensitivity

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and pancreatic β -cell function. The minimal model also provides an assessment of glucose effectiveness, which represents the ability of glucose to facilitate its own disposal independent of an insulin response. DEXA-based body fat mass was determined and used in conjunction with BMI and waist circumference to account for the confounding effects of adiposity. It was hypothesized that, independent of adiposity, the frequency of apneas and hypopneas, the severity of nocturnal hypoxemia, and the degree of sleep fragmentation would be negatively correlated with insulin sensitivity, insulin secretion, and glucose effectiveness.

METHODS

Study Sample and Protocol

The study sample consisted of normal subjects and patients with newly diagnosed but untreated SDB. Normal subjects were recruited from the general community. Patients undergoing polysomnography and confirmed to have SDB were also recruited. Subjects with a fasting glucose greater than 125 mg/dl or those with a history of type 2 diabetes mellitus, angina, myocardial infarction, coronary revascularization, congestive heart failure, or stroke were excluded. In addition, history of a circadian rhythm disorder, chronic insufficient sleep (< 7 hours per night), obstructive lung disease, renal or hepatic dysfunction, upper airway surgery, cancer, or any chronic inflammatory condition also excluded participation. Finally, subjects using anti-inflammatory agents (e.g., steroids), supplemental oxygen, or positive airway pressure therapy were ineligible. After the initial screening visit, those enrolled were counseled on maintaining at least 7 hours of sleep per night and asked to consume at least 250 g of carbohydrates per day. Usual sleep habits were objectively assessed with an ambulatory wrist activity monitor that was worn for at least 5 days before completion of the metabolic tests. Informed consent was obtained from all subjects, and the protocol was approved by the local Institutional Review Board.

Polysomnography

The polysomnogram included recordings of C₃/M₂ and C₄/M₁ electroencephalograms (EEG), right and left electrooculograms, submental and bilateral anterior tibialis electromyograms, and body position. Respiration was monitored with a nasal pressure transducer, with thermocouples at the nose and mouth, and with thoracic and abdominal strain gauges. Continuous recording of SaO2 was obtained with an oximeter (Ohmeda 3700; Ohmeda, Englewood, CO). All physiologic signals were digitized for off-line analysis of sleep and breathing patterns (Embla recordings systems; Medcare, Buffalo, NY). Sleepstage scoring was performed on 30-second epochs according to the criteria of Rechtschaffen and Kales (13). Apneas were identified if airflow was absent in the themistor and nasal cannula channels for at least 10 seconds. Hypopneas were identified if there was at least 30% reduction in airflow lasting at least 10 seconds and associated with a 4% oyxhemoglobin desaturation or an EEG arousal. The apnea-hypopnea index (AHI) was defined as the number of apneas or hypopneas per hour of sleep. The degree of oxyhemoglobin desaturation (ΔSa_{O_2}) associated with each disordered breathing event was determined, and the overall average was used along with the percentage of total sleep time below an oxyhemoglobin saturation of 90% as measures of sleeprelated hypoxemia. Arousals were identified as abrupt shifts of at least 3 seconds duration in EEG frequency according to standard criteria (14).

Anthropometrics and Assessment of Body Fat

Weight was measured to the nearest 0.1 kg, and height was measured using a portable stadiometer to the nearest 0.5 cm. Waist circumference was measured midway between the lower rib margin and the iliac crest with the subject in the standing position at the end of a normal expiration. Fat mass and percent body fat were determined using a Hologic QDR-4500A (Hologic, Inc., Bedford, MA) DEXA scanner. Percent body fat was determined as the ratio of total fat mass to body weight. All body composition measurements were obtained on the same day as the intravenous glucose tolerance test.

Frequently Sampled Intravenous Glucose Tolerance Test

Subjects were admitted to a clinical research unit (~ 9 AM) after an overnight fast for the insulin-modified FSIVGTT (15, 16). An intravenous line was inserted in the antecubital vein for glucose and insulin administration with a second line in the contralateral antecubital vein for blood sampling. Baseline blood samples were obtained at -15, -10, -5, and -1 minute before the glucose injection. At time 0 minutes, a weight-adjusted dose of glucose (50% dextrose, 0.3 g/kg) was administered intravenously over 1 minute followed by a continuous infusion of normal saline. Twenty minutes after the glucose injection, a weight-adjusted dose of regular insulin (0.03 U/kg) was administered. Blood samples were collected after the glucose injection at 2, 3, 4, 5, 6, 8, 10, 12, 14, 16, 19, 22, 24, 25, 27, 30, 40, 50, 60, 70, 80, 90, 100, 120, 140, 160, and 180 minutes. Glucose and insulin concentrations for each sample were determined in duplicate. The resulting values were subjected to the minimal model analysis using the MINMOD software (15–17).

The minimal model provides a mathematical representation of glucose disposal during the FSIVGTT using two coupled differential equations (see online supplement). The first equation represents the physiologic factors that facilitate the return of glucose to basal levels after the glucose injection. These include the effects of insulin, which normalize plasma glucose through the glucose transporter system, and the effects of glucose, which can normalize its concentration independent of an insulin response. The second equation describes the movement of insulin from plasma into the interstitial space where it exerts its physiologic actions. By using the measured insulin profile during the FSIVGTT as the input to the model, the glucose concentration profile is determined by nonlinear least-squares fitting, and parameters of insulin-sensitivity (S_I) and glucose effectiveness (S_G) are numerically estimated. S_I quantifies the effect of insulin in enhancing glucose disposal. S_G quantifies the effects of glucose on its own disposal independent of any insulin response. SG is further divided into two components: the contribution of hyperglycemia per se to increase glucose disposal and the effect of basal insulin levels. The basal component of S_G is referred to as the basal insulin effect and is the product of basal insulin levels (Ib) and SI. The contribution of noninsulin-dependent glucose uptake (glucose effectiveness at zero insulin [GEZI]) to glucose disposal is the difference between total S_{G} and the basal insulin effect: $GEZI = S_G - (Ib \times S_I)$. In addition to estimating S_I , S_G, and GEZI, the acute insulin response to glucose (AIRg), an index of pancreatic β -cell response, was determined as the area under the insulin curve between 0 and 10 minutes. Finally, the disposition index (DI), which is an integrated measure of pancreatic β-cell function, was calculated as the crosss-product of $S_{\rm I}$ and AIRg. Additional details regarding the minimal model are provided in the online supplement.

Statistical Analysis

The dependent variables derived from the FSIVGTT were S_I, S_G, GEZI, AIRg, and DI. Each was examined for normality and logtransformed if necessary. Results are reported as arithmetic means along with the standard errors. Bivariate analyses were conducted to describe the association between each FSIVGTT parameter and SDB status. To account for differences in adiposity and other factors across groups of SDB severity, multivariable linear regression models were used. Covariates considered in these multivariable models included age, gender, race, BMI, waist circumference, and percent body fat. To minimize the influence of outlying values, SI and DI were log-transformed before modeling. For variables that remained skewed despite transformation (S_G and GEZI), median regression analyses (18) were used. Continuous and categorical forms of each independent variable (e.g., AHI, ΔSa_{Ω_2}) were considered. Results from categorical analyses are presented for ease of interpretation. All statistical analyses were conducted using the SAS 9.0 or Stata 7.0 statistical software packages.

RESULTS

A total of 118 subjects met the enrollment criteria and completed overnight polysomnography and the FSIVGTT. Eleven subjects had missing actigraphy data on habitual sleep patterns. Sensitivity analysis that excluded these subjects showed no major differences in the overall inferences regarding the association between SDB and glucose disposal during the FSIVGTT. Thus, the analysis included all 118 subjects that were grouped as follows: 39 subjects had no SDB (AHI < 5 events/h), 34 had mild SDB (AHI, 5.0-14.9 events/h), 22 had moderate SDB (AHI, 15.0–29.9 events/h), and 23 had severe SDB (AHI \ge 30.0 events/h). Table 1 summarizes the characteristics of the study sample by SDB status and shows that older age, male sex, and increasing adiposity were associated with greater severity of SDB. Actigraphy monitoring of usual sleep habits showed little difference in the average time spent in bed across the four groups. Estimated habitual sleep duration was similar and was at least 7 hours except in subjects with severe SDB. Finally, compared with normal subjects, those with moderate to severe SDB displayed typical alterations in sleep architecture, including frequent arousals, more stage 1 sleep, and less slow-wave and REM sleep. During the FSIVGTT, patients with SDB had a slower rate of decline in glucose levels after the glucose injection than normal subjects (Figure 1, left). In contrast, the first-phase insulin response or the area under the insulin curve during the first 10 minutes was comparable across the four groups (Figure 1, right). Table 2 shows the unadjusted parameters derived from the minimal model analysis of the glucose and insulin profiles during the FSIVGTT. Insulin sensitivity (S_I), glucose effectiveness (S_G), and glucose effectiveness at zero insulin (GEZI) progressively decreased with increasing severity of SDB. The acute insulin response to glucose (AIRg) did not vary despite a progressive decline in S_I. The disposition index (DI = $S_I \times AIRg$) was also lower in patients with moderate to severe SDB.

To characterize the independent associations between measures of SDB severity and the FSIVGTT-derived parameters, multivariable regression methods were used. Models were initially constructed with all three measures of adiposity (i.e., BMI, percent fat, and waist circumference) as covariates. However, given the modest to strong correlation between percent body fat and BMI (r = 0.63; P < 0.0001) and between BMI and waist circumference (r = 0.78; P < 0.0001), sensitivity analyses were conducted by including each measure individually and in all possible pairwise combinations. Irrespective of the variables used in the multivariable models, parameter estimates remained essentially unchanged after the inclusion of percent body fat. Thus, to minimize collinearity, percent body fat was used as the single adiposity measure along with age, sex, and race. Despite multivariable adjustments, SDB severity (i.e., AHI) remained inversely associated with S_I, DI, S_G, and GEZI (Figure 1). Compared with normal subjects, the adjusted mean S_I was 26.7, 36.5, and 43.7% lower in patients with mild, moderate, and severe SDB, respectively (Figure 2A; P < 0.007 for linear trend). Although DI was comparable in normal subjects and in subjects with mild SDB, its adjusted mean was also 28.0 and 44.8% lower in patients with moderate and severe SDB, respectively (P <0.035 for linear trend). SG and GEZI also declined with increasing severity of SDB (Figures 2C and 2D). Although a linear association between AHI and GEZI was not observed, GEZI was lower in patients with moderate and severe SDB than in normal subjects or those with mild SDB (Figure 1D).

Analyses were undertaken to determine whether measures of nocturnal hypoxemia and sleep fragmentation were correlated with the results of the FSIVGTT. S_I was most notably associated with the average degree of oxyhemoglobin desaturation (ΔSa_{O_2}) in analyses that adjusted for age, sex, race, and percent body fat (Figure 3). DI showed a similar inverse as-

	AHI < 5.0 events/h (<i>n</i> = 39)	AHI 5.0–14.9 events/h (n = 34)	AHI 15.0–29.9 events/h (n = 22)	$AHI \ge 30.0$ events/h (n = 23)
Demographics				
Male sex, %	43.6*	61.8	63.6	82.6‡
White race, %	92.3	79.4	95.5	78.3
Age, yr	37.7 (1.9)	46.5 (2.0)	51.6 (2.0)	52.4 (2.0) [§]
Body composition				
BMI, kg/m ²	26.5 (0.8)	29.6 (1.1)	30.7 (0.8)	32.8 (1.3) [§]
Waist circumference, cm	84.1 (1.8)	91.8 (2.3)	96.1 (2.3)	106.4 (3.2)
Percent body fat, %	30.3 (1.7)	31.9 (1.8)	32.6 (1.6)	34.9 (1.6) [†]
Actigraphy monitoring		. ,		
Usual time in bed, h	7.8 (0.2)	7.6 (0.2)	7.8 (0.1)	7.5 (0.2)
Sleep duration, h	7.2 (0.2)	7.0 (0.2)	7.3 (0.1)	6.5 (0.2)
Polysomnography				
Total sleep time, h	6.8 (0.2)	6.9 (0.2)	6.7 (0.2)	6.4 (0.2)
Stage 1 sleep, %	8.1 (0.8)	10.4 (0.9)	11.1 (1.5)	15.8 (2.0)
Stage 2 sleep, %	55.7 (1.3)	60.0 (1.4)	62.7 (1.6)	62.4 (2.5) [§]
Slow wave sleep, %	13.7 (1.2)	9.8 (1.0)	7.6 (1.1)	5.0 (1.4) [§]
REM sleep, %	22.5 (0.8)	19.8 (1.1)	18.6 (1.6)	16.7 (1.2)§
Arousal index, n/h	8.1 (0.4)	13.3 (1.0)	19.7 (1.3)	38.3 (4.3) [§]
ΔSa_{O_2} %	3.0 (0.2)	3.8 (0.2)	3.8 (0.2)	7.1 (0.9)§
Fasting serum measurements				
Glucose, mg/dl	94.7 (1.2)	97.9 (1.4)	100.9 (1.9)	102.5 (2.2) [§]
Insulin, μU/ml	9.0 (0.7)	12.9 (1.0)	11.9 (1.2)	16.4 (1.7) [§]

TABLE 1. CHARACTERISTIC OF THE STUDY SAMPLE BY SLEEP-DISORDERED BREATHING SEVERITY

Definition of abbreviations: AHI = apnea-hypopnea index; BMI = body mass index; REM = rapid eye movement; ΔSa_{O_2} = average oxyhemoglobin desaturation.

* Values reported are either percentages or means (SE).

[†] P < 0.05 for comparisons across AHI groups (*P* values determined by χ^2 for categorical variables or Cuzick's nonparametric test for trend for continuous variables).

^{*} P < 0.02 for comparisons across AHI groups (*P* values determined by χ^2 for categorical variables or Cuzick's nonparametric test for trend for continuous variables).

[§] P < 0.001 for comparisons across AHI groups (*P* values determined by χ^2 for categorical variables or Cuzick's nonparametric test for trend for continuous variables).



Figure 1. Glucose and insulin profiles from the frequently sampled intravenous glucose tolerance test in subjects with and without sleep-disordered breathing. Values are average concentrations of glucose (*left*) and insulin (*right*) versus time.

sociation with average ΔSa_{O_2} . For every 1% increase in the average ΔSa_{O_2} , the adjusted average DI decreased by 5.7% (P < 0.065). S_G and GEZI were not associated with the degree of nocturnal hypoxemia (i.e., ΔSa_{O_2}). However, S_G was independently associated with the arousal frequency even after accounting for ΔSa_{O_2} . For every five-unit increase in the respiratory event–related arousal frequency (events/h), the median adjusted value of S_G decreased by 4.3×10^{-4} /min. Analyses correlating sleep stage amounts (e.g., slow-wave sleep, stage 1 sleep) to metabolic parameters showed no significant associations after adjusting for percent body fat.

DISCUSSION

The importance of SDB as an independent risk factor for type 2 diabetes mellitus has been a topic of significant research. The current study adds to the body of existing literature by providing strong evidence that, independent of adiposity, SDB is associated with several metabolic defects, including a decrease in insulin sensitivity, glucose effectiveness, and pancreatic β -cell

function. The major implication of these findings is that SDB is characterized by multiple physiologic deficits that are known to increase the predisposition for type 2 diabetes mellitus.

Early investigations of metabolic disturbances in SDB correlated sleep-related symptoms (e.g., snoring) or polysomnographic metrics (e.g., AHI) to a wide array of measures of insulin sensitivity and glycemic status (19, 20). Although pioneering, many of these initial studies had several methodologic limitations, including small sample sizes, the use of selected clinic-based cohorts, and the lack of an appropriate control group. Thus, it is not surprising that these studies offered mixed conclusions, with some arguing for and others against an independent association. A number of published reports have addressed these concerns by including an adequate number of subjects along with objective sleep-related assessments. However, two major limitations remain. First, because of the relatively low burden, investigators have frequently relied on using a static single blood draw measure of fasting insulin levels as a proxy for insulin sensitivity. Although the fasting condition represents a steady state in which glucose is homeostatically



Figure 2. Adjusted values of insulin sensitivity (A: average S_1 and 95% confidence intervals), the disposition index (B: average DI and 95% confidence intervals), glucose effectiveness (C: median S_G and 95% confidence intervals), and glucose effectiveness at zero insulin (D: median GEZI and 95% confidence interval). Values adjusted for age, sex, race, and percent body fat.

TABLE 2. MINIMAL MODEL ANALYSIS OF THE FREQUENTLY SAMPLED INTRAVENOUS GLUCOSE TOLERANCE TEST DATA

FSIVGTT parameter	AHI < 5.0 events/h (n = 39)	AHI 5.0–14.9 events/h $(n = 34)$	AHI 15.0–29.9 events/h (n = 22)	$AHI \ge 30.0 \text{ events/h}$ $(n = 23)$
S _I , (mU/L) ⁻¹ min ⁻¹	3.7 (0.3)*	2.5 (0.2)	2.5 (0.4)	1.8 (0.3) [†]
AIRg, (mU/L) min^{-1}	510.9 (80.1)	601.0 (69.9)	518.7 (80.7)	527.8 (85.6)
DI, dimensionless	1,592.0 (215.3)	1,332.1 (176.4)	1,054.5 (219.8)	845.0 (148.2)†
$S_{\rm G} \times 10^{-2}$, min ⁻¹	2.1 (0.1)	1.7 (0.1)	1.6 (0.3)	1.5 (0.1)†
GEZI \times 10 ⁻³ , min ⁻¹	1.8 (0.1)	1.4 (0.1)	1.4 (0.1)	1.3 (0.1)†

Definition of abbreviations: AIRg = acute insulin response to glucose; DI = disposition index (SI \times AIRg); FSIVGTT = frequently sampled intravenous glucose tolerance test; GEZI = glucose effectiveness at zero insulin; S_G = glucose effectiveness; S_I = insulin sensitivity.

* Values reported as means (standard error).

[†] P < 0.001 for comparisons across AHI groups (P values determined by Cuzick's nonparametric test for trend).

maintained in the normal range, fasting insulin levels do not reflect pancreatic insulin secretion in response to nutrient intake. Even after incorporating fasting glucose values, as in the homeostasis model of insulin resistance index (9), it is difficult, if not impossible, to quantify the dynamic relation between insulin sensitivity and insulin secretion. Second, to account for the confounding effects of adiposity, most studies have used BMI as an index for body fat. Although BMI is relatively easy to measure, it does not discriminate lean from fat mass and thus is an imperfect measure of adiposity (7, 8). Because fat mass is a major determinant of whole-body insulin resistance (21), greater precision in measurement of body fat is needed to adequately assess the independent effects of SDB on glucose metabolism. Hence, the most unique aspects of the current study are that total body fat was quantified and that a more informative approach was used that included simultaneous assessments of insulin sensitivity and insulin secretion. Minimal model analysis of the FSIVGTT data showed that insulin sensitivity, glucose effectiveness, and the disposition index were lower in patients with SDB. It is also important to note that as insulin sensitivity decreases, the acute insulin response to glucose (or AIRg) should increase if the pancreatic β-cell can respond adequately. The fact that AIRg was comparable across groups of SDB severity indicates that the pancreatic β -cell does not compensate for the decrease in insulin sensitivity observed in patients with moderate to severe SDB. The primary assertion of the foregoing is that the SDB is associated with multiple deficits that are critical in the pathogenesis of type 2 diabetes mellitus.



Figure 3. Adjusted average values of insulin sensitivity (S_1) as a function of average oxyhemoglobin desaturation (ΔSa_{O_2}) during sleep. Displayed are measured S_1 values (*squares*) and the fitted regression values (*dark line* with 95% confidence bands) adjusted for age, sex, race, and percent body fat.

The underlying mechanisms for impaired glucose metabolism in SDB are largely unknown. The genesis of metabolic dysfunction in SDB likely involves several distinct but synergistic processes, including activation of the sympathetic nervous system, increase in oxidative stress, dysregulation of the hypothalamic-pituitary-adrenal (HPA) axis, and low-grade systemic inflammation. It is well established that SDB has profound effects on the sympathetic nervous system (22) and is characterized by higher levels of circulating catecholamines (23, 24). Elevations in norepinephrine and epinephrine levels can induce insulin resistance, inhibit pancreatic insulin secretion, decrease glucose effectiveness, and stimulate hepatic glucose release (25-28). Repetitive cycles of hypoxemia with reoxygenation in SDB can also initiate a cascade of biochemical reactions that increase in oxidative stress in a manner similar to that seen with ischemiareperfusion injury. Indeed, SDB has been associated with excess production of reactive oxygen species (29-32), which can be harmful to the pancreatic β -cell given its relatively low levels of antioxidant enzymes (33). An increase in oxidative stress suppresses insulin secretion and diminishes insulin-medicated peripheral glucose uptake.

Glucose metabolism could be further disturbed in SDB through activation of the HPA axis by sleep fragmentation (34, 35) or intermittent hypoxemia (36, 37). Glucocorticoids inhibit insulin secretion and induce insulin resistance by increasing free fatty acid flux and modifying multiple aspects of the insulin-mediated glucose transport system (38, 39). Finally, low-grade systemic inflammation may be another mechanism relating SDB to metabolic dysfunction. Despite strong biologic plausibility for a causal role, the majority of interventional studies indicate that continuous positive pressure therapy does not mitigate SDB-related metabolic dysfunction. It is possible that chronic exposure to intermittent hypoxemia and sleep fragmentation lead to irreversible metabolic changes. Alternatively, the small number of subjects in most interventional studies coupled with the overwhelming effects of obesity, a strong correlate of SDB, could explain why treatment has not been found to have favorable effects.

Several caveats should be considered in interpreting our results. First, given the cross-sectional nature of our study, inferences on the cause-and-effect sequence are not possible. Second, despite the numerous insights gained by modeling the kinetics of glucose and insulin, the minimal model is not without its drawbacks (40). Because it is uses a single compartment to model glucose disposal, the minimal model oversimplifies the physiology of glucose homeostasis. Finally, visceral adiposity, which is an important contributor to metabolic risk, was not directly assessed with an abdominal CT or MRI scan. Despite these limitations, the current study has several strenghts. These include a relatively large sample size with full-montage polysomnography, exclusion of prevalent medical conditions, assessment of adiposity, and use of a dynamic test to quantify insulin sensitivity, glucose effectiveness, and pancreatic β -cell function.

In conclusion, our results demonstrate that, independent of adiposity, SDB is associated with impairments in insulin sensitivity, glucose effectiveness, and pancreatic β -cell function. These findings are especially relevant given that it is the concurrence of such defects that increases the propensity for type 2 diabetes mellitus. A critical area that must be addressed in future studies is the relative paucity of information on the specific mechanisms that could explain the metabolic alterations associated with SDB. Perhaps most urgent, however, is the need for well-controlled clinical trials to address whether positive pressure therapy can improve metabolic function and potentially avert some of the risk for type 2 diabetes mellitus and its innumerable sequelae.

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