Article Addendum

The Role of VIN3-LIKE Genes in Environmentally Induced Epigenetic Regulation of Flowering

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Addendum to:

A PHD Finger Protein Involved in Both the Vernalization and Photoperiod Pathways in Arabidopsis

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ABSTRACT

Given their sessile nature, it is critical for the survival of plants to adapt to their environment. Accordingly, plants have evolved the ability to sense seasonal changes to govern developmental fates such as the floral transition. Temperature and day length are among the seasonal cues that plants sense. We recently reported that VIN3-LIKE 1 (VIL1) is involved in mediating the flowering response to both cold and day length via regulation of two related genes, FLOWERING LOCUS C (FLC) and FLOWERING LOCUS M (FLM), respectively.

Vernalization renders plants competent to flower after exposure to the prolonged cold of winter.^{1,2} *Arabidopsis* exhibits facultative responses to both vernalization and photoperiod to initiate the floral transition. The facultative nature of the responses makes *Arabidopsis* a tractable genetic system to study these aspects of flowering time control.

In *Arabidopsis*, vernalization creates competence to flower via silencing of the potent floral repressor, *FLC*, in a mitotically stable manner.^{3,4} Thus, the vernalization response is an environmentally induced epigenetic switch in that exposure to cold permanently affects the plants' developmental program. This epigenetic switch is associated with increased levels of *FLC* chromatin methylation on Histone H3 Lys 9 and Lys 27.^{5,6} VERNALIZATION INSENSITIVE 3 (VIN3) plays an essential role in this switch since no modifications to *FLC* chromatin occur in *vin3* mutants.⁵ Furthermore, the levels of expression of *VIN3* mRNA are tightly correlated with the degree of the vernalization response.⁵ VIN3 encodes Plant HomeoDomain (PHD) finger-containing protein. PHD finger-containing proteins are often associated with protein complexes that are involved in chromatin remodeling.⁷

We performed a yeast two-hybrid screen to identify potential protein partners of VIN3. VIN3-LIKE 1 (VIL1) was identified by this screen.⁸ VIL1 encodes a PHD fingercontaining protein that is related to VIN3. As expected for proteins that are associated with VIN3, plants containing loss-of-function alleles of VIL1 do not respond to vernalization. Furthermore, no vernalization-mediated histone modifications occur at FLC in vill mutants similar to the situation in vin3 mutants. Thus, by yeast two hybrid and genetic analysis, VIL1 is a bona fide VIN3 partner that is required for vernalization-mediated histone modifications at FLC chromatin. Unlike VIN3, the expression of VIL1 does not change over the course of cold exposure. Rather, VIL1 mRNA levels are affected by photoperiod. VIL1 expression is significantly increased in non-inductive photoperiods (short days; SD). Consistent with this expression pattern, vil1 mutants in the Columbia accession exhibit a SD-specific late-flowering phenotype. Furthermore, VIL1 is required for attenuating expression of FLOWERING LOCUS M, a FLC-related gene, in a SD-specific manner. It is possible that the attenuation of FLM by VIL1 has a role in creating the facultative nature of photoperiod response in Arabidopsis since vil1 mutants tend towards an obligate photoperiod response (i.e., vil1 mutants often fail to flower in SD).

In *Arabidopsis*, there are four *VIN3*-related genes, which we named as *VIL1* ~ *VIL4*,⁸ and which have also been called *VRN5* and *VEL1* ~ *VEL3*.⁹ The C-terminal domain is highly conserved among these genes and was named the VIN3-Interacting Domain (VID) since it is required for protein-protein interaction between VIN3 and VIL1. The effect of cold on the expression patterns of *VIN3*-related genes varies. For example, VIL2 and VIL3 are induced specifically by vernalizing cold exposures whereas others such as VIL1 are, for the most part, constitutively expressed. It will be interesting to determine the functions of the remaining *VIL* genes.

FLC is the main target for vernalization in *Arabidopsis*. Interestingly, *FLC* orthologs have not been found in vernalization-responsive varieties of cereals. However, in wheat, VRN2 appears to have a role equivalent to that of *FLC* in *Arabidopsis*.¹⁰ *VRN2* encodes



Figure 1. Proposed relationship of *VIN3* family genes to the regulatory network controlling flowering time in response to environmental cues in *Arabidopsis* and diploid wheat (adapted from ref. 16).

a ZCCT type zinc-finger protein that does not have a homolog in the Arabidopsis genome. In diploid wheat, down regulation of VRN2 is correlated with the vernalization response.¹¹ Interestingly, wheat contains three VIN3-LIKE (VIL) genes, TmVIL1, TmVIL2 and TmVIL3.12 Furthermore, TmVIL1 is up-regulated by vernalization.¹² However, whether TmVIL1 has a direct role in the vernalization-mediated repression of VRN2 in wheat has not yet been addressed. Similar to VIL1, TmVIL3 shows elevated level of expression in SD. Furthermore, VRN2 is downregulated in SD;^{13,14} thus there is a correlation between the induction of *TmVIL* genes and the downregulation of the floral repressor VRN2 similar to the VIN3/ FLC and VIL1/FLM relationships (Fig. 1). Perhaps VIN3-related genes have similar roles both in Arabidopsis and in temperate wheat, but act on different target genes, possibly as a result of convergent evolution. Interestingly, the wheat gene TmVRN3 is homologous to FLOWERING LOCUS T (FT) of Arabidopsis, and TmVRN3 is repressed by TmVRN2 as FT is repressed by FLC,¹⁵ suggesting another similarity in the regulation of flowering time between Arabidopsis and temperate wheat (Fig. 1).

Although the PHD finger can be found in various eukaryotes, the VID is unique to plants. It is also noteworthy that VIN3-related genes can be found in various plant species, including rice, which does not have a vernalization response. It will be interesting to address whether the VIN3-related genes from various plant species are more broadly involved in relaying environmental signals to developmental programs.

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