Genetics of Sarcoidosis Candidate Genes and Genome Scans

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Human leukocyte antigen class II allele associations and T-cell receptor beta chain bias in sarcoidosis suggest a specific diseasetriggering antigen exposure in a genetically susceptible host. The cause of sarcoidosis has been elusive, but genetics provides one of the few promising avenues to further our understanding. We review the association studies and genome scans used to identify the genes involved in sarcoidosis.

Keywords: sarcoidosis; genetics; genomics

Racial differences in incidence rates and disease clustering in families (1–3) support the belief that genetics contribute to sarcoidosis etiology. In the United States, African Americans are three and a half times more commonly affected (2), and incidence rates vary widely across ethnic groups in Europe and Asia. Worldwide, familial sarcoidosis occurs in 3 to 14% of patients. In the United States, African Americans are three times more likely than whites to have an affected first- or second-degree relative (4). In African Americans, the sibling recurrence risk ratio, which compares disease risk among siblings with the disease prevalence in the general population, is about 2.2 (95% confidence interval [CI], 1.03–3.68) (3). The ACCESS study (A Case Control Etiologic Sarcoidosis Study) found that cases were five times more likely than control subjects to report an affected sibling or parent (1).

Investigators have relied mainly on candidate gene association studies to search for susceptibility genes in sarcoidosis. More recently, two genome scans, one in whites and the other in African Americans, have been reported. The ongoing refinement of genetic marker maps, genotyping technology, and statistical analyses makes genomic exploration for sarcoidosis genes appealing.

Because many sarcoidosis variants exist and because organ dysfunction may evolve over several years, it is often necessary to follow patients for several years to assign an accurate phenotype. Even patients who present with Löfgren's syndrome, which is a good prognostic sign, can develop chronic sarcoidosis many years later (5). A phenotyping scheme that may further aid linkage and association studies might be gained through evaluating clinical similarities among families with more than one member affected, but few studies exist. Valentonyte and colleagues, in a family analysis to evaluate the candidate gene CCR2, found that 25 of 75 informative sibling pairs exhibited discordance for acute or chronic sarcoidosis phenotypes, arguing against acute sarcoidosis as a distinct heritable phenotype (6). In a recent evaluation in

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340 affected African-American sib pairs, Judson and colleagues found only a modest sib concordance for liver and ocular involvement despite evaluating 15 different organ systems (7).

CANDIDATE GENE APPROACH

Investigators choose candidate genes that make sense based on the prevailing model of pathogenesis. Candidates have been chosen often based on their potential role in granuloma formation. Granuloma formation begins with the sarcoid antigen provoking mononuclear phagocyte and subsequent T-helper cell activation. Released chemokines and cytokines further regulate cell recruitment. Granuloma persistence and accumulation occur due to an inability to clear antigen. Perhaps sarcoidosis can be best thought of as "hereditary frustrated phagocytosis."

HUMAN LEUKOCYTE ANTIGENS

Given the importance of human leukocyte antigens (HLAs) in antigen presentation, it is not surprising that searching for HLA associations with sarcoidosis began over 30 yr ago. The first report revealed an association of acute sarcoidosis with the HLA class I antigen HLA-B8 (8). Other groups (9, 10) confirmed the HLA-B8 association and noted that HLA-B8/DR3 genes were inherited as a sarcoidosis risk haplotype (9), a haplotype also associated with autoimmune diseases in whites (11). These earlier studies of class I HLA antigens gave way to studies focused on HLA class II. A recent report by Grunewald and colleagues suggests that HLA class I and II genes work together in sarcoidosis pathophysiology (12). A summary of the most consistent HLA associations in sarcoidosis is shown in Table 1.

The HLA-DRB1 association predominates in the literature, with variation in the HLA-DRB1 gene affecting sarcoidosis susceptibility and prognosis (13, 14). Another consistent finding across populations has been the HLA-DQB1*0201 allele association with decreased risk and lack of disease progression (15). Other reports strongly support that several different HLA class II genes acting in concert or independently predispose to sarcoidosis (15–17). The linkage disequilibrium (LD) within the major histocompatibility complex (MHC) region limits the ability to precisely identify the involved HLA genes. LD exists when alleles at two distinctive loci occur in gametes more frequently than expected. For example, Grunewald and colleagues showed that the HLA-DRB1*03 associated with resolved disease and HLA-DRB1*15 with persistent disease were synonymous with HLA-DQB1*0201 with resolved disease and HLA-DQB1*0602 with persistent disease (12). Consequently, determining the effects of HLA-DQB1 on sarcoidosis risk apart from DRB1 or dissecting out other gene effects from closely linked haplotypes in the MHC region may be an intractable problem in whites. In African Americans, LD across the MHC region may not be strong (18).

A negative finding is the lack of association with HLA-DPB1*0201, the allele that carries glutamate in amino acid position 69 (Glu69) (16, 19). Glu69 has been associated with chronic beryllium disease, a disease similar to sarcoidosis histologically

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TABLE 1. SUMMARY OF HLA ASSOCIATION STUDIES OF SARCOIDOSIS

HLA	Risk Alleles	Finding	Reference
HLA-A	$A*1$	Susceptibility	126
HLA-B	$B*8$	Susceptibility in several populations	8, 9, 127
HLA-DPB1	$*0201$	Not associated with sarcoidosis	16,19
HLA-DOB1	$*0201$	Protection, Löfgren's syndrome, mild disease	
		in several populations	15, 128
	$*0602$	Susceptibility/disease progression in several groups	15, 129
HLA-DRB1	$*0301$	Acute onset/good prognosis in several groups	9, 127, 130
	$*04$	Protection in several populations	126
	$*1101$	Susceptibility in whites and African Americans.	
		Stage II/III chest X-ray	13
HLA-DRB3	$*1501$	Associated with Löfgren's syndrome	13, 129
	$*0101$	Susceptibility/disease progression in whites	131

and clinically. In 33 berylliosis cases and in 44 exposed individuals without berylliosis, Richeldi and colleagues found Glu69 in 97% of cases and in 30% of control subjects (20). This HLA-DPB1 Glu69 association in beryllium disease has been widely supported (21) but is not associated with sarcoidosis.

NON-HLA CANDIDATE GENES

Genes that influence antigen processing, antigen presentation, macrophage and T-cell activation, and cell recruitment and injury repair may be considered sarcoidosis candidate genes. Table 2 lists the non-HLA candidate genes reported to date.

Angiotensin-converting Enzyme

Epithelioid cells in granulomas produce angiotensin-converting enzyme (ACE), and serum angiotensin-converting enzyme (SACE) levels are believed to reflect granuloma burden. The ACE gene insertion (I)/deletion (D) polymorphism partially accounts for the SACE level variation, and investigators have proposed that this genotype should be used to adjust SACE reference values (22). Studies to support a role for ACE gene polymorphisms in susceptibility or severity have been inconsistent (23, 24).

CC-Chemokine Receptor 2

CC-chemokine receptor 2 (CCR2), a receptor for monocyte chemoattractant protein, plays an important role in monocyte/ macrophage/lymphocyte trafficking (25). CCR2 knockout mice die rapidly when challenged with mycobacteria (26) and display decreased IFN-γ production when challenged with *Leishmania donovani* or *Cryptococcus neoformans* (27, 28). Investigators have reported that a single nucleotide polymorphism (C190A, Val64Ile) shows protection in Japanese patients (29) and that an eight–single nucleotide polymorphism haplotype is associated with Löfgren's syndrome in Dutch patients (30). Underrepresentation of the Val64Ile variant was observed in 65 Czech patients and in 80 control subjects but did not achieve statistical significance (31). Despite using case control–based and family-based study designs and a sample much larger than the previous three studies, Valentonyte and colleagues could not replicate the CCR2 association (6).

CCR5

CCR5 serves as a receptor for CCL3 (macrophage inflammatory protein 1- α), CCL4 (macrophage inflammatory protein 1- β), CCL5 (RANTES [regulated upon activation, T-cell expressed and secreted]), and CCL8 (monocyte chemotactic protein 2) (32, 33). Despite CCR5 playing a dominant role in recruiting and activating lymphocytes and monocytes in sarcoidosis (34), no

firm association with sarcoidosis susceptibility has been found. Petrek and colleagues reported an association of the 32-bp deletion in CCR5 in Czech patients (31), whereas Spagnolo and colleagues, using haplotype analysis, found no association in evaluating 106 white British patients and 142 control subjects and 112 Dutch patients and 169 control subjects (35).

CCR10

Clara cell 10-kD protein. Clara cells serve as a major respiratory tract protector, acting as stem cells in bronchial epithelial repair, providing xenobiotic metabolism, and counterregulating inflammation (36). Clara cell 10-kD protein (CC10) has been shown to inhibit IFN- γ , tumor necrosis factor (TNF)- α , and interleukin (IL)-1 β . Murine and human CC10 gene promoter regions contain sites where inflammatory mediators, such as TNF- α and INF- α , - β , and - γ , alter transcriptional activity (37). Increased CC10 levels have been found in patients whose sarcoidosis had resolved compared with those whose sarcoidosis had progressed (38). The CC10 gene contains three short exons separated by a long first and short second intron. An adenine to guanine substitution at position 38 (A38G) downstream from the transcription initiation site within the noncoding region of exon 1 has been the most studied CC10 polymorphism. The A/A genotype results in decreased CC10 levels (39).

Ohchi and coworkers found that the CC10 A allele was associated with sarcoidosis (40). That patients with the A/A genotype had the lowest bronchoalveolar lavage fluid (BALF) levels of CC10 further strengthened this association. However, Janssen and colleagues found no association with the CC10 A38G SNP in a white Dutch population or in Japanese subjects (41).

CD80 and CD86

The B7 family of costimulatory molecules (CD80 and CD86) regulate T-cell activation. T-cell activation requires two signals: one mediated by T-cell receptor interaction with specific antigen in association with HLA molecules and an antigen-independent costimulatory signal provided by interaction between CD28 on T-cell surface and its ligands CD80 (B7-1) and CD86 (B7-2) on the antigen-presenting cells (42). Handa and colleagues investigated CD80 and CD86 SNPs for sarcoidosis susceptibility in 146 Japanese patients and found no significant difference compared with 157 control subjects (43).

Complement Receptor 1

Complement receptor 1 (CR1; CD35) is a widely distributed membrane glycoprotein present on polymorphonuclear leukocytes, macrophages, B lymphocytes, some T lymphocytes, dendritic cells, and erythrocytes (44). Immune complexes bound to CR1 are transferred to phagocytes as erythrocytes traverse the

Definition of abbreviations: IL = interleukin; LF = Löfgren's syndrome; TNF = tumor necrosis factor; VEGF = vascular endothelial growth factor.

* Type of association: $A =$ susceptibility; B = disease course; C = both.

 \dagger - = Association refuted; + = association replicated.

liver and spleen (45). Immune complex clearance rates correlate with CR1 density. Low expression of erythrocyte CR1 is associated with impaired immune complex clearance and deposition outside the reticuloendothelial system (46). These extrareticuloendothelial immune complex deposits incite local inflammatory responses and presumably granuloma formation. That immune complexes may be involved in sarcoidosis was suggested in the early 1970s. In a series involving 3,676 patients from 11 cities around the world, James and coworkers (47) reported elevated serum γ -globulin levels above 3.5 g/100 ml in 23 to 96% of patients, with IgG being the most consistently and persistently elevated (47). The different sensitivities of the techniques used explain in part the wide range in γ -globulin levels. It is generally accepted that immune complexes are always present in sarcoidosis depending on when and how they are detected.

Zorzetto and colleagues have been the only group to report a CR1 gene association with sarcoidosis (48). The GG genotype for the Pro1827Arg (C507G) polymorphism was associated with sarcoidosis versus healthy control subjects (odds ratio [OR], 3.13; 95% CI, 1.49–6.69) and versus control subjects with chronic obstructive pulmonary disease (OR, 2.82; 95% CI, 1.27–6.39). The GG genotype was most strongly associated with disease in

female patients (OR, 7.05; 95% CI, 3.10–1.61) versus healthy control subjects. No relationship with clinical variables was found.

Cystic Fibrosis Transmembrane Conductance Regulator

The R75Q mutation in the cystic fibrosis transmembrane conductance regulator (CFTR) occurs in high frequency in patients with atypical mild cystic fibrosis (49), bronchiectasis, and allergic bronchopulmonary aspergillosis (50). Bombieri and colleagues reported a R75Q association with sarcoidosis (51), but in followup using complete cystic fibrosis gene mutation screening they could not replicate their findings (52). Schurmann and colleagues could not demonstrate a CFTR association with sarcoidosis (53).

Heat Shock Protein A1L

Heat shock proteins (HSPs) comprise a conserved group of proteins with an average weight of 70 kD. Intracellular HSPs serve as molecular chaperones (54), whereas extracellular HSPs induce cellular immune responses (55). HSPs may also act as carrier molecules for the immunogenic peptides presented on antigen-presenting cells (56). Polymorphisms in the HSPA1L (alias HSP70-hom) have been associated with susceptibility to

rheumatoid arthritis (57). Antibodies to HSP70 in sarcoidosis have been reported (58, 59). To further evaluate the role of HSPs in sarcoidosis, the HSP70 $+2437$ C allele was evaluated and found to be associated with sarcoidosis and Löfgren's syndrome in Polish patients (60) but not in Japanese patients (61).

Inhibitor κΒ-α

Inhibitor κ B (I κ B) masks the nuclear factor (NF)- κ B nuclear localization sequence, thus retaining $NF - \kappa B$ in the cytoplasm and preventing DNA binding. On phosphorylation, IKB degrades, allowing $NF - \kappa B$'s nuclear localization and initiation of transcription (62). Terminating the NF- κ B response requires I κ B- α . I κ B- α knockout mice die 7 to 10 d after birth with increased levels of TNF- α mRNA in the skin and severe dermatitis (63). $NF-\kappa B$ –dependent signaling in alveolar macrophage makes $NF-\kappa B$ and thus I κB central to sarcoid pathophysiology (64). Abdallah and colleagues found the promoter $-297T$ allele associated with sarcoidosis (65). No other $I \kappa B$ studies in sarcoidosis have been reported.

IL-1

 $IL-1\beta$ produced mainly by macrophages maintains T-cell alveolitis and granuloma formation. Hunninghake and colleagues also demonstrated higher IL-1 β activity in the BALF of patients with sarcoidosis compared with normal subjects (66). Mikuniya and colleagues have suggested that the ratio of IL-1 receptor antagonist to IL-1 β in sarcoid alveolar macrophage culture supernatants could predict disease chronicity (67). The IL-1 α 5' flanking –889 C allele was found nearly two times more commonly among Czech patients with sarcoidosis compared with control subjects (68).

IL-4R

The inflammatory response in sarcoidosis is primarily Th1 mediated. IL-4 drives Th2 differentiation (69). To test whether variation in the IL-4R gene confers susceptibility to sarcoidosis, Bohnert and colleagues typed 241 members of 62 families with 136 affected siblings and 304 healthy control subjects for three functional SNPs within the IL-4R gene and found no evidence for linkage or association, thus excluding a significant role for IL-4R (70).

IL-18

IL-18 produced by monocytes/macrophages induces IFN- γ and drives the Th1 response. BALF and serum IL-18 levels are increased in sarcoidosis (71). An association between IL-18607 (A/C) polymorphism and sarcoidosis has been reported and refuted in Japanese (72, 73) and white subjects (74, 75).

IFN- α

The increasing number of reported cases of IFN- α –induced sarcoidosis supports that IFN- α is important in sarcoidosis (76). Akahoshi and colleagues found an IFN- α T551G (Ile184Arg) polymorphism associated with sarcoidosis susceptibility (OR, 3.27; 95% CI, 1.44–7.46; $p = 0.004$) (77). This allele is also associated with high IFN- α production and subsequent strong Th1 polarization.

SLC11A1

Polymorphic variants of the natural resistance-associated macrophage protein 1 gene, now named SLC11A1, have been associated with tuberculosis and leprosy susceptibility in endemic areas of disease (78, 79). Immunolocalization studies demonstrate the presence of SLC11A1 in lysosomes of macrophages and polymorphonuclear leukocytes (80). SLC11A1 was found not to increase the risk of sarcoidosis among African Americans (81), although a more recent article has noted an association in Polish patients (OR, 1.68; 95% CI, 1.01–2.81) (82).

Transforming Growth Factor-

Polymorphisms for all three isoforms of transforming growth factor (TGF)- β (TGF- β 1, TGF- β 2, and TGF- β 3) have been associated with protein expression variation or functionality changes (83) . TGF- β 1 levels are increased in patients with sarcoidosis who have impaired pulmonary function (84). Kruit and colleagues reported that the TGF-β2 59941 G allele and the TGF-β3 4875 A and 17369 C alleles were associated with chest X-ray evidence of pulmonary fibrosis (85). The TFG-3 15101 G allele was lower in patients with fibrosis (85).

Toll-like Receptor 4

Toll-like receptor 4 (TLR4), the first and best described of the many TLRs, plays a crucial role in detecting infection and inducing inflammatory and adaptive immune responses (86). Pabst and colleagues examined 141 white German patients and control subjects for the TLR4 polymorphisms Asp299Gly and Thre399Ile and found no association with disease presence but did find a significant correlation with chronic disease (87).

TNF- α

TNF- α has a broad range of inflammatory and immunostimulatory actions, including playing a key role in granuloma formation. TNF- α stimulates cytokine production, enhances expression of adhesion molecules, and acts as a costimulator of T-cell activation. Alveolar macrophages from patients with active sarcoidosis secrete more TNF- α than those with inactive disease (88). TNF- α has been considered a target for therapy in sarcoidosis (89). Although it is unclear whether $TNF-\alpha$ promoter polymorphisms are functionally significant, studies suggest that a small but significant effect of the TNF- α promoter -307 A/G polymorphism may exist, with the A allele being associated with slightly greater levels of TNF- α transcription (90, 91). A higher frequency of TNF-307A allele has been found in patients presenting with Löfgren's syndrome and erythema nodosum (92–94). In evaluating five promoter polymorphisms, Grutters and colleagues found a significant increase in TNF $-857T$ allele in white British and Dutch patients and confirmed the TNF $-307A$ allele association with Löfgren's syndrome (95) . In these studies, it is not clear whether TNF-307A confers independent risk from HLA-DRB1 because TNF is in tight LD with HLA-DRBI (96). Using a family-based approach, $TNF-\alpha$ was not found to be significantly associated with sarcoidosis (23).

Vascular Endothelial Growth Factor

Dysregulated vascular endothelial growth factor (VEGF) expression has been implicated in several inflammatory diseases, such as rheumatoid arthritis and inflammatory bowel diseases (97, 98). VEGF modulates angiogenesis and has been reported to enhance monocyte migration, a key event in granuloma formation (99). Tolnay and colleagues reported increased VEGF transcription and protein production in activated alveolar macrophages in epithelioid cells and multinuclear giant cells of pulmonary sarcoid granulomas (100). Several polymorphisms have been associated with VEGF protein production (101, 102). Morohashi and colleagues found that the VEGF+813T allele was underrepresented in patients with sarcoidosis (associated with decreased risk). The $+813$ site is predicted to lie within a potential transcription factor binding site and could possibly reduce VEGF expression (102).

Vitamin D Receptors

The active form of vitamin D, 1,25-dihydroxy vitamin D3, modulates the immune response through control of cytokine expression, including IFN- γ and IL-2 (103). Increased expression of vitamin D receptors (VDRs) on sarcoid BAL T cells and sarcoid alveolar macrophage production of 1,25-dihydroxy vitamin D3 have been reported (104, 105). Niimi and colleagues reported a VDR Bsm1 restriction site polymorphism in intron 8 to be associated with sarcoidosis (106). Guleva and Seitzer examined a VDR Taq1 polymorphism in linkage disequilibrium with the BsmI polymorphism in 85 patients and 80 control subjects and could not confirm Niimi and colleagues' findings (107). Rybicki and colleagues also could not confirm VDRs as candidate genes in sarcoidosis (23).

Although the choice of each candidate gene (Table 2) makes sense based on their function, none have been consistently reproduced or confirmed in family studies. The variability in reported outcomes can be traced to methodologic differences between studies, such as the use of inappropriate control subjects (e.g., convenient samples, such as hospital personnel or patients with other diseases) or case heterogeneity. Furthermore, limitation to many of these studies likely resides in the case-control study design's susceptibility to a form of confounding known as population stratification. If the gene under study shows marked variation in allele frequency across population subgroups and if these subgroups differ in their baseline disease risk, then spurious associations may arise. Because infection and immune surveillance appear under strong negative population pressure, immune response genes seem to be particularly subject to confounding by population stratification.

The issue of population stratification can be overcome. One way is to use a family-based design that involves recruiting patients' siblings and parents if available. In this design, parental alleles not transmitted to affected offspring are used as the control alleles and thus control for genetic background. The transmission disequilibrium test, one of the statistical methods used, counts the number of parental gene variants transmitted to affected offspring. Deviation from expected transmission supports a predisposing effect of the more frequently transmitted allele. No candidate gene chosen based on its likely function in sarcoidosis pathophysiology has been confirmed using the family-based study design.

GENETIC ADVANCES IN OTHER GRANULOMATOUS DISEASE

Advances have been made in defining the genetics of inflammatory bowel disease and Blau syndrome, an autosomal dominant disease comprised of granulomatous uveitis, arthritis, and skin rash but without lung involvement or Kveim skin test positivity. CARD15 located on chromosome 16 is responsible for Blau's syndrome (108, 109) and is the gene identified in the inflammatory bowel disease 1 (IBD1) locus (110). NOD2, encoded by CARD15, recognizes peptidoglycan, a component of bacterial cell walls, and is expressed mainly by antigen-presenting cells and epithelial cells (111) . NOD2 activation leads to NF- κ B activation (111).

Using exclusion mapping among sarcoid-affected sib pairs, Rybicki and colleagues showed that the Blau syndrome/IBD1 locus did not confer risk for sarcoidosis (112). Further refuting CARD15 as a sarcoidosis susceptibility gene, Schurmann and colleagues, using case control–based and family-based samples, evaluated four coding CARD15 polymorphisms associated with increased risk of Crohn's disease and concluded that CARD15 mutations play no role in sarcoidosis susceptibility (113). Kanazawa and colleagues, however, analyzed 10 patients with early-onset sarcoidosis who had disease onset ranging from 6 mo to 4 yr of age (114). They reported that 9 of the 10 cases had heterozygous missense mutations in the CARD15 gene. Despite the negative studies reported by Rybicki and colleagues (112) and Schurmann and colleagues (113), the CARD15 mutations may play a role in the early-onset phenotype.

GENOME SCANNING: AFFECTED SIB PAIR LINKAGE ANALYSIS

Genomewide scans that use allele-sharing methods map disease genes by testing for marker similarity among affected relatives. If the affected individuals within a collection of pedigrees or sibships show an unusual propensity to share marker alleles identical by descent in a particular chromosomal region, then that region may harbor a disease gene. Genomewide scans offer the opportunity to detect candidate genes without regard to the current disease model but rather according to their chromosomal location detected by linkage signals.

First Sarcoidosis Genome Scan in Whites

Schurmann and colleagues, in the first sarcoidosis genome scan, used 225 microsatellite markers spanning the genome in 63 German families and found seven linkage peaks. The highest linkage signal was on chromosome 6p21 (D6S1666) (115). Despite the fact that this linkage signal represented only suggestive evidence of linkage with a nonparametric lod (logarithm of the odds) score of 2.99, the group used a three-stage SNP scan of the 16-MB region surrounding D6S1666 and identified a single SNP, rs2076530, in the BTNL2 gene associated with sarcoidosis (116). This SNP (G/A) was found at the 3' boundary of the exon 5 coding region. The A allele at this position introduces an alternate splice site at the transcript's exon $5'$ -3' intron that results in a premature truncation of the protein.

BTNL2 (aliases "butyrophilinlike 2" and "BTL-2") is a butyrophilin gene that belongs to the B7 family (42, 117). Butyrophilin was initially cloned from cattle mammary epithelial cells (118) and is a member of a family of genes located in the MHC class II region (117). Members of the butyrophilin gene family share gene architecture that consists of a signal peptide, two immunoglobulin domains (the first a larger variable domain and the second a smaller constant domain), a transmembrane domain and cytoplasmic region, heptad repeats, and a PKA domain at the carboxy terminus.

In evaluating the BTNL2 gene as a sarcoidosis risk factor in whites and African Americans, Rybicki and colleagues characterized the variation in the BTNL2 exon/intron 5 region in three samples that consisted of 219 nuclear African American families (686 individuals) and two case-control samples (295 African American matched pairs and 366 white matched pairs) (119). BTNL2 moderately influences disease risk (ORs of 1.6 in heterozygotes and 2.8 in homozygotes) and seems to be somewhat less associated with sarcoidosis in African Americans compared with whites.

One question regarding BTNL2 as a sarcoidosis risk factor is whether it is independent of HLA-DRB1 risk alleles. HLADRB1 and BTNL2 are in linkage disequilibrium. HLA DRB lies about 180 kb centromeric to BTNL2. Based on regression models, BTNL2 seems to be an independent risk factor (116, 119). In the case of African Americans, where the BTNL2 conferred sarcoidosis risk is less significant than in whites, a negative interaction with HLA-DR seems to exist (119).

Sarcoidosis Genome Scan in African Americans

Eleven centers joined together in an NHLBI-sponsored effort (Sarcoidosis Genetic Analysis Consortium [SAGA]) to perform a genome scan in African American siblings. This group performed a 380-microsatellite genomewide scan across 22 autosomes in 519 African American sib pairs. The significant findings included 15 markers with p values less than 0.05 with the strongest linkage signal on chromosome 5 (120). Fine mapping studies indicated a sarcoidosis susceptibility gene on chromosome 5q11.2 and a gene protective effect for sarcoidosis on 5p15.2 (121). Although choosing to study African Americans to uncover sarcoidosis susceptibility genes makes sense because African Americans are more commonly and severely affected and have affected family members more often than whites, a disadvantage is that African Americans are admixed with white and other populations to varying degrees with possible admixture among our participating centers ranging from 12% in South Carolina to 20% in New York (122).

To address the possibility that admixed subpopulations existed in the SAGA sample and affected the power to detect linkage, the sample was stratified by genetically determined ancestry using the data from the 380 microsatellite markers genotyped in the genome scan. The African-American families were clustered into subpopulations based on ancestry similarity. Evidence of two genetically distinct groups was found: Stratified linkage results suggest that one subpopulation of families contributed to previously identified linkage signals at 1p22, 3p21- 14, 11p15, and 17q21 and that a second subpopulation of families contributed to those found at 5p15-13 and 20q13 (123). These findings support the presence of sarcoidosis susceptibility genes in regions previously identified but indicate that these genes are likely to be specific to ancestral groups that have combined to form modern-day African Americans. Additional studies are underway to evaluate candidate genes from these linked regions.

Markers on chromosome 16, close to the Blau syndrome/ IBD locus on chromosome 5 near the cytokine gene cluster and on chromosome 2 near the NRAMP locus, did not suggest linkage to sarcoidosis in either genome scan.

A genomewide association study in which high throughput genotyping methods are used to genotype a dense set of SNPs across the genome has yet to be performed in sarcoidosis. A significant advantage of this approach is that association studies are more powerful than affected sib pair methods of linkage analysis (124). Generally, large samples sizes (i.e., several hundred to a few thousand) will likely be required if several genes with modest affect are involved. Groups in Europe and in the United States are preparing to carry out this approach.

Outlook

Sarcoidosis likely develops from complex interactions between environmental agents and alleles of many genes. The environmental causes remain unknown, and over 80 yr has elapsed since Martenstein first reported that sarcoidosis can occur in siblings (125). Since then, HLA and other candidate genes have been associated with sarcoidosis susceptibility. Association studies have been motivated by the hopes that identifying alleles that affect risk and phenotype will help in understanding disease etiology. Unfortunately, many of the reported associations have not been replicated.

Emerging technologies and advances in genomics and proteomics are bringing us closer to answering three common questions patients ask: What causes sarcoidosis? Why did I get it? How will it affect my health? Methods such as multiplexed liquid chromatography and nano–high-performance liquid chromatography coupled to electrospray or matrix-assisted laser desorption/ionization mass spectrometry are expected to lead us to the inciting sarcoid antigens. Gene expression profiling in BALF and blood carried out at the time of presentation will likely allow us to better predict disease resolution or progression and

response to treatment. Functional analysis of the candidate genes identified by linkage analysis and whole genome association studies will provide insight into pathogenesis and disease risk.

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References

- 1. Rybicki BA, Iannuzzi MC, Frederick MM, Thompson BW, Rossman MD, Bresnitz EA, Terrin ML, Moller DR, Barnard J, Baughman RP, *et al.* Familial aggregation of sarcoidosis: a case-control etiologic study of sarcoidosis (ACCESS). *Am J Respir Crit Care Med* 2001; 164:2085–2091.
- 2. Rybicki BA, Major M, Popovich J Jr, Maliarik MJ, Iannuzzi MC. Racial differences in sarcoidosis incidence: a 5-year study in a health maintenance organization. *Am J Epidemiol* 1997;145:234–241.
- 3. Rybicki BA, Kirkey KL, Major M, Maliarik MJ, Popovich J Jr, Chase GA, Iannuzzi MC. Familial risk ratio of sarcoidosis in African-American sibs and parents. *Am J Epidemiol* 2001;153:188–193.
- 4. Harrington D, Major M, Rybicki B, Popovich J Jr, Maliarik M, Iannuzzi MC. Familial analysis of 91 families. *Sarcoidosis* 1994;11:240–243.
- 5. Mana J, Gomez-Vaquero C, Montero A, Salazar A, Marcoval J, Valverde J, Manresa F, Pujol R. Lofgren's syndrome revisited: a study of 186 patients. *Am J Med* 1999;107:240–245.
- 6. Valentonyte R, Hampe J, Croucher PJ, Muller-Quernheim J, Schwinger E, Schreiber S, Schurmann M. Study of C–C chemokine receptor 2 alleles in sarcoidosis, with emphasis on family-based analysis. *Am J Respir Crit Care Med* 2005;171:1136–1141.
- 7. Judson MA, Hirst K, Iyengar SK, Rybicki BA, El Ghormli L, Baughman RP, Donohue JF, Elston RC, Kavuru MS, Moller DR, *et al.* Comparison of sarcoidosis phenotypes among affected African-American siblings. *Chest* 2006;30:855–862.
- 8. Brewerton DA, Cockburn C, James DC, James DG, Neville E. HLA antigens in sarcoidosis. *Clin Exp Immunol* 1977;27:227–229.
- 9. Hedfors E, Lindstrom F. HLA-B8/DR3 in sarcoidosis: correlation to acute onset disease with arthritis. *Tissue Antigens* 1983;22:200–203.
- 10. Smith MJ, Turton CW, Mitchell DN, Turner-Warwick M, Morris LM, Lawler SD. Association of HLA B8 with spontaneous resolution in sarcoidosis. *Thorax* 1981;36:296–298.
- 11. Lio D, Candore G, Romano GC, D'Anna C, Gervasi F, Di Lorenzo G, Modica MA, Potestio M, Caruso C. Modification of cytokine patterns in subjects bearing the HLA-B8,DR3 phenotype: implications for autoimmunity. *Cytokines Cell Mol Ther* 1997;3:217–224.
- 12. Grunewald J, Eklund A, Olerup O. Human leukocyte antigen class I alleles and the disease course in sarcoidosis patients. *Am J Respir Crit Care Med* 2004;169:696–702.
- 13. Rossman MD, Thompson B, Frederick M, Maliarik M, Iannuzzi MC, Rybicki BA, Pandey JP, Newman LS, Magira E, Beznik-Cizman B, *et al.* HLA-DRB1*1101: a significant risk factor for sarcoidosis in blacks and whites. *Am J Hum Genet* 2003;73:720–735.
- 14. Ishihara M, Ohno S, Ishida T, Ando H, Naruse T, Nose Y, Inoko H. Molecular genetic studies of HLA class II alleles in sarcoidosis. *Tissue Antigens* 1994;43:238–241.
- 15. Iannuzzi MC, Maliarik MJ, Poisson LM, Rybicki BA. Sarcoidosis susceptibility and resistance HLA-DQB1 alleles in African Americans. *Am J Respir Crit Care Med* 2003;167:1225–1231.
- 16. Maliarik MJ, Chen KM, Major ML, Sheffer RG, Popovich J Jr, Rybicki BA, Iannuzzi MC. Analysis of HLA-DPB1 polymorphisms in African-Americans with sarcoidosis. *Am J Respir Crit Care Med* 1998; 158:111–114.
- 17. Rybicki BA, Maliarik MJ, Poisson LM, Sheffer R, Chen KM, Major M, Chase GA, Iannuzzi MC. The major histocompatibility complex gene region and sarcoidosis susceptibility in African Americans. *Am J Respir Crit Care Med* 2003;167:444–449.
- 18. Zachary AA, Bias WB, Johnson A, Rose SM, Leffell MS. Antigen, allele, and haplotype frequencies report of the ASHI minority antigens workshops: part 1, African-Americans. *Hum Immunol* 2001;62: 1127–1136.
- 19. Schurmann M, Bein G, Kirsten D, Schlaak M, Muller-Quernheim J, Schwinger E. HLA-DQB1 and HLA-DPB1 genotypes in familial sarcoidosis. *Respir Med* 1998;92:649–652.
- 20. Richeldi L, Sorrentino R, Saltini C. HLA-DPB1 glutamate 69: a genetic marker of beryllium disease. *Science* 1993;262:242–244.
- 21. Amicosante M, Sanarico N, Berretta F, Arroyo J, Lombardi G, Lechler R, Colizzi V, Saltini C. Beryllium binding to HLA-DP molecule

carrying the marker of susceptibility to berylliosis glutamate beta 69. *Hum Immunol* 2001;62:686–693.

- 22. Sharma P, Smith I, Maguire G, Stewart S, Shneerson J, Brown MJ. Clinical value of ACE genotyping in diagnosis of sarcoidosis. *Lancet* 1997;349:1602–1603.
- 23. Rybicki BA, Maliarik MJ, Poisson LM, Iannuzzi MC. Sarcoidosis and granuloma genes: a family-based study in African-Americans. *Eur Respir J* 2004;24:251–257.
- 24. Maliarik MJ, Rybicki BA, Malvitz E, Sheffer RG, Major M, Popovich J Jr, Iannuzzi MC. Angiotensin-converting enzyme gene polymorphism and risk of sarcoidosis. *Am J Respir Crit Care Med* 1998;158:1566– 1570.
- 25. Boring L, Gosling J, Chensue SW, Kunkel SL, Farese RV Jr, Broxmeyer HE, Charo IF. Impaired monocyte migration and reduced type 1 (Th1) cytokine responses in C–C chemokine receptor 2 knockout mice. *J Clin Invest* 1997;100:2552–2561.
- 26. Peters W, Scott HM, Chambers HF, Flynn JL, Charo IF, Ernst JD. Chemokine receptor 2 serves an early and essential role in resistance to Mycobacterium tuberculosis. *Proc Natl Acad Sci USA* 2001;98: 7958–7963.
- 27. Traynor TR, Kuziel WA, Toews GB, Huffnagle GB. CCR2 expression determines T1 versus T2 polarization during pulmonary Cryptococcus neoformans infection. *J Immunol* 2000;164:2021–2027.
- 28. Sato N, Kuziel WA, Melby PC, Reddick RL, Kostecki V, Zhao W, Maeda N, Ahuja SK, Ahuja SS. Defects in the generation of IFNgamma are overcome to control infection with Leishmania donovani in CC chemokine receptor (CCR) 5-, macrophage inflammatory protein-1 alpha-, or CCR2-deficient mice. *J Immunol* 1999;163:5519– 5525.
- 29. Hizawa N, Yamaguchi E, Furuya K, Jinushi E, Ito A, Kawakami Y. The role of the C–C chemokine receptor 2 gene polymorphism V64I (CCR2- 64I) in sarcoidosis in a Japanese population. *Am J Respir Crit Care Med* 1999;159:2021–2023.
- 30. Spagnolo P, Renzoni EA, Wells AU, Sato H, Grutters JC, Sestini P, Abdallah A, Gramiccioni E, Ruven HJ, du Bois RM, *et al.* C–C chemokine receptor 2 and sarcoidosis: association with Lofgren's syndrome. *Am J Respir Crit Care Med* 2003;168:1162–1166.
- 31. Petrek M, Drabek J, Kolek V, Zlamal J, Welsh KI, Bunce M, Weigl E, Du Bois R. CC chemokine receptor gene polymorphisms in Czech patients with pulmonary sarcoidosis. *Am J Respir Crit Care Med* 2000; 162:1000–1003.
- 32. Blanpain C, Migeotte I, Lee B, Vakili J, Doranz BJ, Govaerts C, Vassart G, Doms RW, Parmentier M. CCR5 binds multiple CC-chemokines: MCP-3 acts as a natural antagonist. *Blood* 1999;94:1899–1905.
- 33. Combadiere C, Ahuja SK, Tiffany HL, Murphy PM. Cloning and functional expression of CC CKR5, a human monocyte CC chemokine receptor selective for MIP-1(alpha), MIP-1(beta), and RANTES. *J Leukoc Biol* 1996;60:147–152.
- 34. Standiford TJ, Rolfe MW, Kunkel SL, Lynch JP III, Burdick MD, Gilbert AR, Orringer MB, Whyte RI, Strieter RM. Macrophage inflammatory protein-1 alpha expression in interstitial lung disease. *J Immunol* 1993;151:2852–2863.
- 35. Spagnolo P, Renzoni EA, Wells AU, Copley SJ, Desai SR, Sato H, Grutters JC, Abdallah A, Taegtmeyer A, du Bois RM, *et al.* C–C chemokine receptor 5 gene variants in relation to lung disease in sarcoidosis. *Am J Respir Crit Care Med* 2005;172:721–728.
- 36. Singh G, Katyal SL. Clara cells and Clara cell 10 kD protein (CC10). *Am J Respir Cell Mol Biol* 1997;17:141–143.
- 37. Cowan MJ, Huang X, Yao XL, Shelhamer JH. Tumor necrosis factor alpha stimulation of human Clara cell secretory protein production by human airway epithelial cells. *Ann N Y Acad Sci* 2000;923:193–201.
- 38. Shijubo N, Itoh Y, Shigehara K, Yamaguchi T, Itoh K, Shibuya Y, Takahashi R, Ohchi T, Ohmichi M, Hiraga Y, *et al.* Association of Clara cell 10-kDa protein, spontaneous regression and sarcoidosis. *Eur Respir J* 2000;16:414–419.
- 39. Laing IA, Hermans C, Bernard A, Burton PR, Goldblatt J, Le Souef PN. Association between plasma CC16 levels, the A38G polymorphism, and asthma. *Am J Respir Crit Care Med* 2000;161:124–127.
- 40. Ohchi T, Shijubo N, Kawabata I, Ichimiya S, Inomata S, Yamaguchi A, Umemori Y, Itoh Y, Abe S, Hiraga Y, *et al.* Polymorphism of Clara cell 10-kD protein gene of sarcoidosis. *Am J Respir Crit Care Med* 2004;169:180–186.
- 41. Janssen R, Sato H, Grutters JC, Ruven HJ, du Bois RM, Matsuura R, Yamazaki M, Kunimaru S, Izumi T, Welsh KI, *et al.* The Clara cell10 adenine38guanine polymorphism and sarcoidosis susceptibility in Dutch and Japanese subjects. *Am J Respir Crit Care Med* 2004;170: 1185–1187.
- 42. Sharpe AH, Freeman GJ. The B7–CD28 superfamily. *Nat Rev Immunol* 2002;2:116–126.
- 43. Handa T, Nagai S, Ito I, Tabuena R, Shigematsu M, Hamada K, Kitaichi M, Izumi T, Aoyama T, Toguchida J, *et al.* Polymorphisms of B7 (CD80 and CD86) genes do not affect disease susceptibility to sarcoidosis. *Respiration (Herrlisheim)* 2005;72:243–248.
- 44. Loegering DJ, Blumenstock FA. Depressing hepatic macrophage complement receptor function causes increased susceptibility to endotoxemia and infection. *Infect Immun* 1985;47:659–664.
- 45. Cornacoff JB, Hebert LA, Smead WL, VanAman ME, Birmingham DJ, Waxman FJ. Primate erythrocyte-immune complex-clearing mechanism. *J Clin Invest* 1983;71:236–247.
- 46. Schifferli JA, Ng YC, Estreicher J, Walport MJ. The clearance of tetanus toxoid/anti-tetanus toxoid immune complexes from the circulation of humans: complement- and erythrocyte complement receptor 1 dependent mechanisms. *J Immunol* 1988;140:899–904.
- 47. James DG, Neville E, Walker A. Immunology of sarcoidosis. *Am J Med* 1975;59:388–394.
- 48. Zorzetto M, Bombieri C, Ferrarotti I, Medaglia S, Agostini C, Tinelli C, Malerba G, Carrabino N, Beretta A, Casali L, *et al.* Complement receptor 1 gene polymorphisms in sarcoidosis. *Am J Respir Cell Mol Biol* 2002;27:17–23.
- 49. Hughes D, Dork T, Stuhrmann M, Graham C. Mutation and haplotype analysis of the CFTR gene in atypically mild cystic fibrosis patients from Northern Ireland. *J Med Genet* 2001;38:136–139.
- 50. Luisetti M, Pignatti PF. Genetics of idiopathic disseminated bronchiectasis. *Semin Respir Crit Care Med* 2003;24:179–184.
- 51. Bombieri C, Luisetti M, Belpinati F, Zuliani E, Beretta A, Baccheschi J, Casali L, Pignatti PF. Increased frequency of CFTR gene mutations in sarcoidosis: a case/control association study. *Eur J Hum Genet* 2000;8:717–720.
- 52. Bombieri C, Belpinati F, Pignatti PF, Luisetti M. Comment on 'CFTR gene mutations in sarcoidosis'. *Eur J Hum Genet* 2003;11:553–554.
- 53. Schurmann M, Albrecht M, Schwinger E, Stuhrmann M. CFTR gene mutations in sarcoidosis. *Eur J Hum Genet* 2002;10:729–732.
- 54. Matouschek A. Protein unfolding: an important process in vivo? *Curr Opin Struct Biol* 2003;13:98–109.
- 55. Asea A, Kraeft SK, Kurt-Jones EA, Stevenson MA, Chen LB, Finberg RW, Koo GC, Calderwood SK. HSP70 stimulates cytokine production through a CD14-dependant pathway, demonstrating its dual role as a chaperone and cytokine. *Nat Med* 2000;6:435–442.
- 56. Srivastava PK, Udono H, Blachere NE, Li Z. Heat shock proteins transfer peptides during antigen processing and CTL priming. *Immunogenetics* 1994;39:93–98.
- 57. Jenkins SC, March RE, Campbell RD, Milner CM. A novel variant of the MHC-linked hsp70, hsp70-hom, is associated with rheumatoid arthritis. *Tissue Antigens* 2000;56:38–44.
- 58. de Smet MD, Ramadan A. Circulating antibodies to inducible heat shock protein 70 in patients with uveitis. *Ocul Immunol Inflamm* 2001;9:85–92.
- 59. Hrycaj P, Wurm K, Mennet P, Muller W. Antibodies to heat shock proteins in patients with pulmonary sarcoidosis. *Sarcoidosis* 1995;12: 124–130.
- 60. Bogunia-Kubik K, Koscinska K, Suchnicki K, Lange A. HSP70-hom gene single nucleotide $(+2763 \text{ G/A}$ and $+2437 \text{ C/T})$ polymorphisms in sarcoidosis. *Int J Immunogenet* 2006;33:135–140.
- 61. Ishihara M, Ohno S, Ishida T, Mizuki N, Ando H, Naruse T, Ishihara H, Inoko H. Genetic polymorphisms of the TNFB and HSP70 genes located in the human major histocompatibility complex in sarcoidosis. *Tissue Antigens* 1995;46:59–62.
- 62. Perkins ND. The Rel/NF-kappa B family: friend and foe. *Trends Biochem Sci* 2000;25:434–440.
- 63. Klement JF, Rice NR, Car BD, Abbondanzo SJ, Powers GD, Bhatt PH, Chen CH, Rosen CA, Stewart CL. IkappaBalpha deficiency results in a sustained NF-kappaB response and severe widespread dermatitis in mice. *Mol Cell Biol* 1996;16:2341–2349.
- 64. Conron M, Bondeson J, Pantelidis P, Beynon HL, Feldmann M, duBois RM, Foxwell BM. Alveolar macrophages and T cells from sarcoid, but not normal lung, are permissive to adenovirus infection and allow analysis of NF-kappa b-dependent signaling pathways. *Am J Respir Cell Mol Biol* 2001;25:141–149.
- 65. Abdallah A, Sato H, Grutters JC, Veeraraghavan S, Lympany PA, Ruven HJ, van den Bosch JM, Wells AU, du Bois RM, Welsh KI. Inhibitor kappa B-alpha (IkappaB-alpha) promoter polymorphisms in UK and Dutch sarcoidosis. *Genes Immun* 2003;4:450–454.
- 66. Hunninghake GW. Release of interleukin-1 by alveolar macrophages of patients with active pulmonary sarcoidosis. *Am Rev Respir Dis* 1984;129:569–572.
- 67. Mikuniya T, Nagai S, Takeuchi M, Mio T, Hoshino Y, Miki H, Shigematsu M, Hamada K, Izumi T. Significance of the interleukin-1 receptor antagonist/interleukin-1 beta ratio as a prognostic factor in patients with pulmonary sarcoidosis. *Respiration (Herrlisheim)* 2000; 67:389–396.
- 68. Hutyrova B, Pantelidis P, Drabek J, Zurkova M, Kolek V, Lenhart K, Welsh KI, Du Bois RM, Petrek M. Interleukin-1 gene cluster polymorphisms in sarcoidosis and idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med* 2002;165:148–151.
- 69. Rengarajan J, Szabo SJ, Glimcher LH. Transcriptional regulation of Th1/Th2 polarization. *Immunol Today* 2000;21:479–483.
- 70. Bohnert A, Schurmann M, Hartung A, Hackstein H, Muller-Quernheim J, Bein G. No linkage of the interleukin-4 receptor locus on chromosome 16p11.2–12.1 with sarcoidosis in German multiplex families. *Eur J Immunogenet* 2002;29:269–272.
- 71. Shigehara K, Shijubo N, Ohmichi M, Yamada G, Takahashi R, Okamura H, Kurimoto M, Hiraga Y, Tatsuno T, Abe S, *et al.* Increased levels of interleukin-18 in patients with pulmonary sarcoidosis. *Am J Respir Crit Care Med* 2000;162:1979–1982.
- 72. Zhou Y, Yamaguchi E, Hizawa N, Nishimura M. Roles of functional polymorphisms in the interleukin-18 gene promoter in sarcoidosis. *Sarcoidosis Vasc Diffuse Lung Dis* 2005;22:105–113.
- 73. Takada T, Suzuki E, Morohashi K, Gejyo F. Association of single nucleotide polymorphisms in the IL-18 gene with sarcoidosis in a Japanese population. *Tissue Antigens* 2002;60:36–42.
- 74. Kelly DM, Greene CM, Meachery G, O'Mahony M, Gallagher PM, Taggart CC, O'Neill SJ, McElvaney NG. Endotoxin up-regulates interleukin-18: potential role for gram-negative colonization in sarcoidosis. *Am J Respir Crit Care Med* 2005;172:1299–1307.
- 75. Janssen R, Grutters JC, Ruven HJ, Zanen P, Sato H, Welsh KI, du Bois RM, van den Bosch JM. No association between interleukin-18 gene polymorphisms and haplotypes in Dutch sarcoidosis patients. *Tissue Antigens* 2004;63:578–583.
- 76. Goldberg HJ, Fiedler D, Webb A, Jagirdar J, Hoyumpa AM, Peters J. Sarcoidosis after treatment with interferon-alpha: A case series and review of the literature. *Respir Med* 2006;100:2063–2068.
- 77. Akahoshi M, Ishihara M, Remus N, Uno K, Miyake K, Hirota T, Nakashima K, Matsuda A, Kanda M, Enomoto T, *et al.* Association between IFNA genotype and the risk of sarcoidosis. *Hum Genet* 2004; 114:503–509.
- 78. Bellamy R. Identifying genetic susceptibility factors for tuberculosis in Africans: a combined approach using a candidate gene study and a genome-wide screen. *Clin Sci (Lond)* 2000;98:245–250.
- 79. Abel L, Sanchez FO, Oberti J, Thuc NV, Hoa LV, Lap VD, Skamene E, Lagrange PH, Schurr E. Susceptibility to leprosy is linked to the human NRAMP1 gene. *J Infect Dis* 1998;177:133–145.
- 80. Gruenheid S, Pinner E, Desjardins M, Gros P. Natural resistance to infection with intracellular pathogens: the Nramp1 protein is recruited to the membrane of the phagosome. *J Exp Med* 1997;185:717–730.
- 81. Maliarik MJ, Chen KM, Sheffer RG, Rybicki BA, Major ML, Popovich J Jr, Iannuzzi MC. The natural resistance-associated macrophage protein gene in African Americans with sarcoidosis. *Am J Respir Cell Mol Biol* 2000;22:672–675.
- 82. Dubaniewicz A, Jamieson SE, Dubaniewicz-Wybieralska M, Fakiola M, Nancy Miller E, Blackwell JM. Association between SLC11A1 (formerly NRAMP1) and the risk of sarcoidosis in Poland. *Eur J Hum Genet* 2005;13:829–834.
- 83. Awad MR, El-Gamel A, Hasleton P, Turner DM, Sinnott PJ, Hutchinson IV. Genotypic variation in the transforming growth factor-beta1 gene: association with transforming growth factor-beta1 production, fibrotic lung disease, and graft fibrosis after lung transplantation. *Transplantation* 1998;66:1014–1020.
- 84. Salez F, Gosset P, Copin MC, Lamblin Degros C, Tonnel AB, Wallaert B. Transforming growth factor-beta1 in sarcoidosis. *Eur Respir J* 1998;12:913–919.
- 85. Kruit A, Grutters JC, Ruven HJ, van Moorsel CH, Weiskirchen R, Mengsteab S, van den Bosch JM. Transforming growth factor-beta gene polymorphisms in sarcoidosis patients with and without fibrosis. *Chest* 2006;129:1584–1591.
- 86. Akira S, Takeda K, Kaisho T. Toll-like receptors: critical proteins linking innate and acquired immunity. *Nat Immunol* 2001;2:675–680.
- 87. Pabst S, Baumgarten G, Stremmel A, Lennarz M, Knufermann P, Gillissen A, Vetter H, Grohe C. Toll-like receptor (TLR) 4 polymorphisms

are associated with a chronic course of sarcoidosis. *Clin Exp Immunol* 2006;143:420–426.

- 88. Zheng L, Teschler H, Guzman J, Hubner K, Striz I, Costabel U. Alveolar macrophage TNF-alpha release and BAL cell phenotypes in sarcoidosis. *Am J Respir Crit Care Med* 1995;152:1061–1066.
- 89. Baughman RP, Iannuzzi M. Tumour necrosis factor in sarcoidosis and its potential for targeted therapy. *BioDrugs* 2003;17:425–431.
- 90. Allen RD. Polymorphism of the human TNF-alpha promoter: random variation or functional diversity? *Mol Immunol* 1999;36:1017–1027.
- 91. Wilson AG, di Giovine FS, Blakemore AI, Duff GW. Single base polymorphism in the human tumour necrosis factor alpha (TNF alpha) gene detectable by NcoI restriction of PCR product. *Hum Mol Genet* 1992;1:353.
- 92. Somoskovi A, Zissel G, Seitzer U, Gerdes J, Schlaak M, Muller Quernheim J. Polymorphisms at position -308 in the promoter region of the TNF-alpha and in the first intron of the TNF-beta genes and spontaneous and lipopolysaccharide-induced TNF-alpha release in sarcoidosis. *Cytokine* 1999;11:882–887.
- 93. Seitzer U, Swider C, Stuber F, Suchnicki K, Lange A, Richter E, Zabel P, Muller-Quernheim J, Flad HD, Gerdes J. Tumour necrosis factor alpha promoter gene polymorphism in sarcoidosis. *Cytokine* 1997;9: 787–790.
- 94. Labunski S, Posern G, Ludwig S, Kundt G, Brocker EB, Kunz M. Tumour necrosis factor-alpha promoter polymorphism in erythema nodosum. *Acta Derm Venereol* 2001;81:18–21.
- 95. Grutters JC, Sato H, Pantelidis P, Lagan AL, McGrath DS, Lammers JW, van den Bosch JM, Wells AU, du Bois RM, Welsh KI. Increased frequency of the uncommon tumor necrosis factor -857T allele in British and Dutch patients with sarcoidosis. *Am J Respir Crit Care Med* 2002;165:1119–1124.
- 96. Wilson AG, de Vries N, Pociot F, di Giovine FS, van der Putte LB, Duff GW. An allelic polymorphism within the human tumor necrosis factor alpha promoter region is strongly associated with HLA A1, B8, and DR3 alleles. *J Exp Med* 1993;177:557–560.
- 97. Kanazawa S, Tsunoda T, Onuma E, Majima T, Kagiyama M, Kikuchi K. VEGF, basic-FGF, and TGF-beta in Crohn's disease and ulcerative colitis: a novel mechanism of chronic intestinal inflammation. *Am J Gastroenterol* 2001;96:822–828.
- 98. Kasama T, Shiozawa F, Kobayashi K, Yajima N, Hanyuda M, Takeuchi HT, Mori Y, Negishi M, Ide H, Adachi M. Vascular endothelial growth factor expression by activated synovial leukocytes in rheumatoid arthritis: critical involvement of the interaction with synovial fibroblasts. *Arthritis Rheum* 2001;44:2512–2524.
- 99. Flamme I, Frolich T, Risau W. Molecular mechanisms of vasculogenesis and embryonic angiogenesis. *J Cell Physiol* 1997;173:206–210.
- 100. Tolnay E, Kuhnen C, Voss B, Wiethege T, Muller KM. Expression and localization of vascular endothelial growth factor and its receptor flt in pulmonary sarcoidosis. *Virchows Arch* 1998;432:61–65.
- 101. Watson CJ, Webb NJ, Bottomley MJ, Brenchley PE. Identification of polymorphisms within the vascular endothelial growth factor (VEGF) gene: correlation with variation in VEGF protein production. *Cytokine* 2000;12:1232–1235.
- 102. Renner W, Kotschan S, Hoffmann C, Obermayer-Pietsch B, Pilger E. A common 936 C/T mutation in the gene for vascular endothelial growth factor is associated with vascular endothelial growth factor plasma levels. *J Vasc Res* 2000;37:443–448.
- 103. Hewison M. Vitamin D and the immune system. *J Endocrinol* 1992;132: 173–175.
- 104. Biyoudi-Vouenze R, Cadranel J, Valeyre D, Milleron B, Hance AJ, Soler P. Expression of 1,25(OH)2D3 receptors on alveolar lymphocytes from patients with pulmonary granulomatous diseases. *Am Rev Respir Dis* 1991;143:1376–1380.
- 105. Adams JS, Singer FR, Gacad MA, Sharma OP, Hayes MJ, Vouros P, Holick MF. Isolation and structural identification of 1,25-dihydroxyvitamin D3 produced by cultured alveolar macrophages in sarcoidosis. *J Clin Endocrinol Metab* 1985;60:960–966.
- 106. Niimi T, Tomita H, Sato S, Kawaguchi H, Akita K, Maeda H, Sugiura Y, Ueda R. Vitamin D receptor gene polymorphism in patients with sarcoidosis. *Am J Respir Crit Care Med* 1999;160:1107–1109.
- 107. Guleva I, Seitzer U. Vitamin D receptor gene polymorphism in patients with sarcoidosis. *Am J Respir Crit Care Med* 2000;162:760–761.
- 108. Blau EB. Familial granulomatous arthritis, iritis, and rash. *J Pediatr* 1985;107:689–693.
- 109. Miceli-Richard C, Lesage S, Rybojad M, Prieur AM, Manouvrier-Hanu S, Hafner R, Chamaillard M, Zouali H, Thomas G, Hugot JP. CARD15 mutations in Blau syndrome. *Nat Genet* 2001;29:19–20.
- 110. Hugot JP, Chamaillard M, Zouali H, Lesage S, Cezard JP, Belaiche J, Almer S, Tysk C, O'Morain CA, Gassull M, *et al.* Association of NOD2 leucine-rich repeat variants with susceptibility to Crohn's disease. *Nature* 2001;411:599–603.
- 111. Strober W, Murray PJ, Kitani A, Watanabe T. Signalling pathways and molecular interactions of NOD1 and NOD2. *Nat Rev Immunol* 2006;6:9–20.
- 112. Rybicki BA, Maliarik MJ, Bock CH, Elston RC, Baughman RP, Kimani AP, Sheffer RG, Chen KM, Major M, Popovich J Jr, *et al.* The Blau syndrome gene is not a major risk factor for sarcoidosis. *Sarcoidosis Vasc Diffuse Lung Dis* 1999;16:203–208.
- 113. Schurmann M, Valentonyte R, Hampe J, Muller-Quernheim J, Schwinger E, Schreiber S. CARD15 gene mutations in sarcoidosis. *Eur Respir J* 2003;22:748–754.
- 114. Kanazawa N, Okafuji I, Kambe N, Nishikomori R, Nakata-Hizume M, Nagai S, Fuji A, Yuasa T, Manki A, Sakurai Y, *et al.* Early-onset sarcoidosis and CARD15 mutations with constitutive nuclear factorkappaB activation: common genetic etiology with Blau syndrome. *Blood* 2005;105:1195–1197.
- 115. Schurmann M, Reichel P, Muller-Myhsok B, Schlaak M, Muller-Quernheim J, Schwinger E. Results from a genome-wide search for predisposing genes in sarcoidosis. *Am J Respir Crit Care Med* 2001; 164:840–846.
- 116. Valentonyte R, Hampe J, Huse K, Rosenstiel P, Albrecht M, Stenzel A, Nagy M, Gaede KI, Franke A, Haesler R, *et al.* Sarcoidosis is associated with a truncating splice site mutation in BTNL2. *Nat Genet* 2005;37:357–364.
- 117. Rhodes DA, Stammers M, Malcherek G, Beck S, Trowsdale J. The cluster of BTN genes in the extended major histocompatibility complex. *Genomics* 2001;71:351–362.
- 118. Jack LJ, Mather IH. Cloning and analysis of cDNA encoding bovine butyrophilin, an apical glycoprotein expressed in mammary tissue and secreted in association with the milk-fat globule membrane during lactation. *J Biol Chem* 1990;265:14481–14486.
- 119. Rybicki BA, Walewski JL, Maliarik MJ, Kian H, Iannuzzi MC. The BTNL2 gene and sarcoidosis susceptibility in African Americans and Whites. *Am J Hum Genet* 2005;77:491–499.
- 120. Iannuzzi MC, Iyengar SK, Gray-McGuire C, Elston RC, Baughman RP, Donohue JF, Hirst K, Judson MA, Kavuru MS, Maliarik MJ, *et al.* Genome-wide search for sarcoidosis susceptibility genes in African Americans. *Genes Immun* 2005;6:509–518.
- 121. Gray-McGuire C, Sinha R, Iyengar SK, Millard C, Rybicki BA, Elston RC, Iannuzzi MC. Genetic characterization and fine mapping of susceptibiltiy genes for sarcoidosis in African Americans on chromsome 5. *Hum Genet* 2006;120:420–430.
- 122. Parra EJ, Marcini A, Akey J, Martinson J, Batzer MA, Cooper R, Forrester T, Allison DB, Deka R, Ferrell RE, *et al.* Estimating African American admixture proportions by use of population-specific alleles. *Am J Hum Genet* 1998;63:1839–1851.
- 123. Thompson CL, Rybicki BA, Iannuzzi MC, Elston RC, Iyengar SK, Gray-McGuire C. Stratified linkage analysis based on population substructure in a population of African-American sarcoidosis families. *Am J Hum Genet* 2006;79:603–613.
- 124. Risch N, Merikangas K. The future of genetic studies of complex human diseases. *Science* 1996;273:1516–1517.
- 125. Martenstein H. Knochveranderungen bei lupus pernio. *Zentralbl Haut Geschlechtskr Grenzeb* 1923;7:308.
- 126. Martinetti M, Tinelli C, Kolek V, Cuccia M, Salvaneschi L, Pasturenzi L, Semenzato G, Cipriani A, Bartova A, Luisetti M. "The sarcoidosis map": a joint survey of clinical and immunogenetic findings in two European countries. *Am J Respir Crit Care Med* 1995;152:557–564.
- 127. Gardner J, Kennedy HG, Hamblin A, Jones E. HLA associations in sarcoidosis: a study of two ethnic groups. *Thorax* 1984;39:19–22.
- 128. Sato H, Grutters JC, Pantelidis P, Mizzon AN, Ahmad T, Van Houte AJ, Lammers JW, Van Den Bosch JM, Welsh KI, Du Bois RM. HLA-DQB1*0201: a marker for good prognosis in British and Dutch patients with sarcoidosis. *Am J Respir Cell Mol Biol* 2002;27:406–412.
- 129. Voorter CE, Drent M, van den Berg-Loonen EM. Severe pulmonary sarcoidosis is strongly associated with the haplotype HLA-DQB1*0602– DRB1*150101. *Hum Immunol* 2005;66:826–835.
- 130. Berlin M, Fogdell-Hahn A, Olerup O, Eklund A, Grunewald J. HLA-DR predicts the prognosis in Scandinavian patients with pulmonary sarcoidosis. *Am J Respir Crit Care Med* 1997;156:1601–1605.
- 131. Bogunia-Kubik K, Tomeczko J, Suchnicki K, Lange A. HLA-DRB1*03, DRB1*11 or DRB1*12 and their respective DRB3 specificities in clinical variants of sarcoidosis. *Tissue Antigens* 2001;57:87–90.
- 132. McGrath DS, Foley PJ, Petrek M, Izakovicova-Holla L, Kolek V, Veeraraghavan S, Lympany PA, Pantelidis P, Vasku A, Wells AU, *et al.* Ace gene I/D polymorphism and sarcoidosis pulmonary disease severity. *Am J Respir Crit Care Med* 2001;164:197–201.
- 133. Furuya K, Yamaguchi E, Itoh A, Hizawa N, Ohnuma N, Kojima J, Kodama N, Kawakami Y. Deletion polymorphism in the angiotensin I converting enzyme (ACE) gene as a genetic risk factor for sarcoidosis. *Thorax* 1996;51:777–780.
- 134. Arbustini E, Grasso M, Leo G, Tinelli C, Fasani R, Diegoli M, Banchieri N, Cipriani A, Gorrini M, Semenzato G, *et al.* Polymorphism of angiotensin-converting enzyme gene in sarcoidosis. *Am J Respir Crit Care Med* 1996;153:851–854.
- 135. Morohashi K, Takada T, Omori K, Suzuki E, Gejyo F. Vascular endothelial growth factor gene polymorphisms in Japanese patients with sarcoidosis. *Chest* 2003;123:1520–1526.