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Linkage Analysis of Glomerular Filtration Rate in American Indians: The Strong Heart Family Study

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Abstract

American Indians have a disproportionately high rate of kidney disease likely due to a combination of increased environmental and genetic risk factors. In an attempt to localize genes influencing kidney disease risk factors, we performed a genome wide scan of estimated glomerular filtration rate on participants of the Strong Heart Family Study. Over 3 600 men and women from 13 American Indian tribes were recruited from 3 centers (Arizona, North and South Dakota, Oklahoma). Using SOLAR 2.1.2, multipoint variance component linkage analysis was performed in each center as well as the entire cohort after controlling for center effects. Two modeling strategies were utilized: model 1 incorporated age, sex and interaction terms and model 2 additionally controlled for diabetic status, systolic and diastolic blood pressure, body mass index, low density lipoproteins, high density lipoproteins, triglycerides and smoking status. Significant evidence for linkage in Arizona lay on 12p12.2 at 39cM nearest marker D12S310 (LOD=3.5). Additional loci with suggestive evidence for linkage were detected at 1p36.31 (LOD=2.0–2.3), 2q33.3 (LOD=1.8) and 9q34.2 (LOD=2.4). No significant evidence for additive interaction with diabetes, hypertension or obesity was noted. In conclusion, we found evidence for linkage of a quantitative trait locus influencing estimated glomerular filtration rate to a region of chromosome 12p in a large cohort of American Indians.

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GFR in American Indians linked to 12p12.2

Disclosures

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Introduction

Chronic kidney disease (CKD) is an important public health problem affecting 8–11% of the U.S. population.(1,2) In addition to the rising burden of end stage renal disease (ESRD), CKD is also a strong risk factor for cardiovascular disease and death. Worsening glomerular filtration rate (GFR) can result in up to a 3-fold risk of cardiovascular events and a 6-fold risk of death. (3) In the U.S., diabetes and hypertension are the most common risk factors for CKD and end-stage renal disease.(4) Even in the absence of diabetes, metabolic syndrome is strongly associated with reduced GFR.(5,6) Other identified risk factors include glucose control(7,8), blood pressure control(9,10), body mass index(11), cholesterol levels(12,13) and smoking (14,15).

American Indians are known to have higher rates of ESRD than the general population, approximately twice that of Caucasians.(4,16,17) The vast majority of kidney disease in American Indians is due to diabetes, with rates of diabetic ESRD approximately four times higher than in Caucasians, and continuing to climb at a rapid pace in younger age groups.(4) The prevalence of earlier stages of CKD is also high. A cohort study of Navajo Indians found 3–6% of nondiabetics and 10–11% of diabetics had a creatinine clearance of less than 65ml/min.(18) Environmental risk factors for kidney disease are highly prevalent in this population. The prevalence of diabetes, hypertension, hypercholesterolemia and obesity in American Indians over 40 years of age has been reported to range from 40–60%, 25–35%, 30–40% and 20–40%, respectively.(19–21) Smoking is highly prevalent with 26–38% of American Indians being active smokers.(19)

Genetic predisposition to kidney disease is well accepted.(17,22–24) Heritable factors have long been considered a component of diabetic kidney disease,(25,26) with multiple genomic loci indicated in the development and progression of diabetic nephropathy.(23,24,27–31) Indeed, studies have implicated the same genes or genomic regions in the predisposition or progression of any kidney disease, irrespective of the initial insult.(32–36) Genetic factors may also play a role in the variability of creatinine and GFR in populations without kidney disease. (34,37–39) The aim of the present study was to identify genetic loci that are linked to the gene (s) that influence phenotypic variation of GFR in a large population of American Indian families.

Results

A total of 3 665 individuals were available for analysis from the Strong Heart Family Study (SHFS) (Arizona = 1 235; Dakotas = 1 220; Oklahoma = 1 210). Descriptive characteristics of all SHFS participants are summarized in table 1. The average age \pm standard deviation of participants in each center was approximately 39 years \pm 15; 41 years \pm 17 and 44 years \pm 17 in Arizona, North and South Dakota and Oklahoma, respectively. Diabetes, hypertension and obesity were highly prevalent, especially in the Arizona center. Kidney disease was common, with albuminuria (urine albumin:creatinine ratio[ACR] >30 $\mu\text{g}/\text{mg}$) present in approximately 19% and depressed GFR ($<60\text{ml}/\text{min}/1.73\text{m}^2$) present in approximately 7% of individuals in all centers (center-specific data not shown). There was wide variation in albuminuria and estimated GFR with means of $106\mu\text{g}/\text{mg} \pm 598$ and $99\text{ml}/\text{min}/1.73\text{m}^2 \pm 27$ in all centers, respectively. The distribution of CKD as defined by the Kidney Disease Quality of Outcomes Initiative (KDOQI), amongst all centers was: 11% stage 1, 28% stage 2, 4% stage 3, 0.5% stage 4 and 1% stage 5 CKD. Twenty percent (n=722) of all observations had an eGFR $> 120\text{ml}/\text{min}/1.73\text{m}^2$. Five percent (n=182) of all observations had an eGFR $> 120\text{ml}/\text{min}/1.73\text{m}^2$ and no proteinuria. For the analyses of those not on antihypertensive medications, there were 738 participants deleted from analysis and 486 of these had either diabetes and/or proteinuria.

Genetic data were available for > 60 000 relative pairs in the full dataset and > 18 000, 22 000 and 18 000 relative pairs in Arizona, the Dakotas and Oklahoma, respectively. The number of participants with available genetic and covariate data for the model 1 and model 2 analyses were: 3 605 and 3 536 in all centers; 1 215 and 1 177 in Arizona; 1 186 and 1 174 in the Dakotas; and 1 204 and 1 185 in Oklahoma. Using the fully adjusted models, the heritability estimates (standard errors [SE]) of eGFR were consistent across centers: approximately 0.33 (SE=0.03), 0.33 (SE=0.06), 0.34 (SE=0.05), 0.33 (SE=0.06) in the full sample, Arizona, the Dakotas and Oklahoma, respectively.

Analyses yielding logarithm of odds (LOD) scores ≥ 1.8 are displayed in table 2. There was evidence for a quantitative trait locus (QTL) for GFR in the Arizona center using model 1 (LOD = 3.5) on chromosome 12 at 39cM (nearest marker D12S31), with a 16 cM 1-LOD unit support interval spanning the regions 12p11.23 – 12p13.1.(40) Loci with suggestive evidence for linkage included 1p36.31 in all centers using both models (LOD=2.3, 2.0, respectively), 2q33.3 in the Dakotas using model 1 (LOD=1.8) and 9q34.2 using model 1 in the Dakotas (LOD=2.4).(40) Data were very similar with the use of ranked inverse serum creatinine as the outcome measure (results not shown).

When we excluded individuals on antihypertensive medications from analysis the LOD score on chromosome 12 increased to 4.6 in model 1 and 2.8 in model 2. LOD scores on chromosome 1 remained similar in all centers, however analyses within Arizona now also yielded suggestive LOD scores using both models (LOD= 2.5, 2.2, for models 1 and 2, respectively). Additionally, loci on chromosomes 2 and 9 were no longer significant and new loci were evident on 3q21.2 in Arizona using model 2 (LOD=1.8) and 7p21.3 in all centers and the Dakotas, using both models (LODs =1.9–2.3). Tests of additive interaction for gene-by-diabetes, -hypertension and -obesity were nonsignificant in the full sample and every center at the 0.006 p-value threshold level which accounts for multiple testing.

Discussion

This study is only the third genome-wide scan of eGFR in a general population not targeted for diabetes, hypertension or ESRD (34,41) and is one of the first to examine gene-by-environment interactions in a linkage study of estimated GFR. We found evidence of linkage to eGFR on 12p12.2 and suggestive evidence for linkage on 1p36.31, 2q33.3 and 9q34.2 (Figure 1). The locus on 12p12.2 replicates findings from a previous genome scan of diabetic nephropathy in African Americans with a LOD score as high as 2.9 in a subset of patients with late onset diabetes.(27) Additionally, our finding of suggestive evidence for linkage to 2q33.3 replicates a study of GFR using cystatin-C in diabetics (LOD=4.1) and another of creatinine clearance in a general population of Mexican Americans.(41)

The lack of gene-by-diabetes, -hypertension and -obesity interaction on GFR in our study population was somewhat surprising. It is logical that there are genetic polymorphisms that predispose to kidney disease only in the setting of permissive environmental factors. Given that diabetes is the most common cause of kidney disease in American Indians, this seems a likely candidate to influence genetic susceptibility. Interaction-specific linkage analysis is underpowered and may alone explain the difference in findings for diabetes interactions. Placha et al. (2006) found strong evidence of gene-by-diabetes interaction on GFR using cystatin C in multiple extended families enriched for diabetes.(41) Cystatin C was used to calculate GFR which may be more accurate than creatinine-based methods for detecting mild CKD specifically in American Indians and/or diabetics.(42, 43) The population used in the study by Placha et al. was roughly ten years older than the SHFS and also had a lower mean BMI. It is possible that misclassification and the presence of glucose intolerance/metabolic syndrome without overt diabetes may have contributed to our finding of no interaction.

Our analysis isolated a QTL for GFR with a maximum LOD score of 3.5 at 12p12.2. In our analysis, restricting the dataset to those not on antihypertensive medications resulted in an increase of the LOD score to 4.6. Placha et al. (2006) also isolated significantly higher LOD scores after inclusion of antihypertensive treatment as a covariate.(41) The purpose of this subgroup analysis was two-fold. First, it is plausible that treatment with renin-angiotensin system (RAS) antagonists diminish the genetic propensity towards reduced kidney function. We would have liked to have stratified by treatment with RAS antagonists, however, this specific information was not available. Hence, we used treatment with antihypertensives as a surrogate for RAS antagonism. Second, since severe hypertension may also act as either a confounder or effect modifier on the propensity towards reduced kidney function, we used treatment with antihypertensives as a marker of the more severe hypertensive phenotype. The higher LOD score on 12p obtained in the subanalysis of those not on antihypertensive medications provides support for these hypotheses.

Within the one-LOD drop support interval of our peak on 12p12.2 lies the gene protein-tyrosine phosphatase receptor type-O (*PTPRO*). The protein, termed glomerular epithelial protein 1 (GLEPP1), is primarily located on the apical surface of glomerular podocytes and is present from early-on in development.(44,45) Abnormalities in distribution are present in multiple human primary glomerulopathies, including minimal change disease and focal segmental glomerulosclerosis.(46) *PTPRO* knockout mice are viable, but podocytes have blunted, widened foot processes and decreased amounts of nephrin.(47) Knockout mice have no difference in urine albumin excretion or GFR as long as renal reserve is intact. Uninephrectomized knockout mice, however, have a 25–50% lower GFR than their wild-type counterparts, suggesting an increase in susceptibility to decreased GFR in the presence of states of hyperfiltration.(47) This is highly applicable to the Strong Heart population, given the high prevalence of obesity, diabetes and hypertension which also result in glomerular hyperfiltration.

There are several caveats to our reported findings. The MDRD equation is less accurate in those with an estimated GFR $> 60\text{ml}/\text{min}/1.73\text{m}^2$ and the vast majority of the SHFS falls into this category. Additionally, the MDRD equation was not developed in American Indians, the estimates of GFR may have been biased. However, the MDRD equation has been validated in two studies of Pima Indians using iothalamate as the gold standard measurement.(42) Although estimates of GFR may be more accurate using cystatin C than creatinine, cystatin C measures were not available. Analyses were not repeated using the Cockcroft-Gault equation since the high prevalence of obesity in our study population would have made it difficult to interpret results of creatinine clearance based on weight.

The intra-individual variability of serum creatinine tends to be high thus an average of repeated measures of serum creatinine, had they been available, would have provided a more optimal measurement of serum creatinine and estimated GFR. Other factors which may affect levels of measured serum creatinine in populations include hydration status, concomitant medications and random testing errors.

Estimates of GFR may also have been affected by obesity, hypertension and/or diabetes as a consequence of glomerular hyperfiltration. The presence of a supranormal GFR is the first stage of kidney disease for these conditions and hence may misrepresent the true pathologic status when placed within a continuous outcome variable paradigm. According to this logic, 20% of our subjects fell into this category of having a supranormal GFR, and excluding these participants would not have been appropriate. Further complicating this issue is that persons with a GFR $90\text{--}120\text{ml}/\text{min}/1.73\text{m}^2$ may, in fact, possess more severe renal pathology than those with a supranormal GFR, as they may have surpassed this state and have declining renal function even though their GFR is in the ‘normal range’. This is an issue inherent to renal

pathology which cannot be overcome or corrected for in the analysis. From a physiologic standpoint, it would have been interesting to have an ordinal outcome variable using the KDOQI staging system for CKD, however, such an approach would be difficult to implement, as linkage analysis requires either a two category discrete or continuous outcome variable.

There was a significant decrease in the LOD score for the locus on 12p12.2 in the fully adjusted model, however, this may be explained by a loss of power alone. Analysis of the fully adjusted model in Arizona incorporated 1177 individuals whereas the minimally adjusted model incorporated 1215 individuals. Although this difference would be miniscule in a general epidemiologic study, it is possible in a linkage study that crucial individuals were dropped from the analysis. The alternative possibility is that covariates introduced in the maximum model are confounders in the minimum model. We addressed this by running the analysis using the covariates diabetes, hypertension, hypertension medication, obesity, BMI, systolic and diastolic blood pressures as the outcome variable using the minimum model over the 20 centimorgan region on 12p. None of these analyses were remotely suggestive of linkage. To address whether albuminuria had confounded our analysis, we also ran the analysis with UACR as the outcome variable and found no evidence of linkage. In addition, there was no evidence of genetic correlation between GFR and UACR. Since we did not incorporate hypertension, hypertensive medication or UACR into the model, we also ran the analysis incorporating these variables into the model and obtained the exact same results.

The validity of our findings is strengthened by two indicators of robustness. First, the addition of hypertension, hypertensive medication and UACR did not alter the LOD score isolated on 12p12.2. Second, this locus is a replication of a previous genome scan of kidney disease and we additionally replicated effect modification by antihypertensive treatment. Lastly, within our 1-LOD drop support interval is the candidate gene for GLEPP-1 which could logically influence eGFR in a population prone to states of hyperfiltration. Since very few genome wide linkage scans of eGFR have been published, and this region of 12p has been implicated in other populations, we should direct future research into the refined mapping of the 12p12.2 region.

Materials and Methods

Study Population

The Strong Heart Study (SHS) was begun in 1988 to investigate cardiovascular disease (CVD) and its risk factors in a geographically diverse group of resident American Indian tribal members at three study centers in Arizona, Oklahoma, and North and South Dakota. The SHFS, a component of the SHS, was initiated in 1996 with the goal of localizing genes that influence CVD and its risk factors. The data utilized for the current analysis were gathered during phase IV of the SHFS, which occurred between 2001 and 2003. More than 3600 men and women aged 14 to 93 were recruited from over 90 extended families originating from 13 separate American Indian tribes. The Arizona center is primarily composed of Pima Indians, but there are also representatives of the closely related Maricopa and Tohono O'odham Indian tribes. The vast majority of participants in the Dakotas center are Dakota/Lakota/Nakota. The Oklahoma Center is represented by members of the Kiowa, Comanche, Delaware, Apache, Caddo, Fort Sill Apache, and Wichita tribes. All protocols were approved by the Indian Health Service Institutional Review Board, by the institutional review boards of the participating institutions and by the 13 American Indian tribes participating in these studies.

Phenotypes

Detailed descriptions of the SHS and SHFS study design and laboratory protocols have been published previously.(48,49) Blood pressure and body morphometrics were measured during the physical exam. After five minutes of rest, upper arm seated blood pressure was measured

three times by a trained technician using a mercury column sphygmomanometer (WA Baum Co) and size-adjusted cuffs. The first and fifth Korotkoff sounds were recorded. The average of the last two measures was used for all analyses. Hypertension was defined by a systolic blood pressure (SBP) ≥ 140 mm Hg or diastolic blood pressure (DBP) ≥ 90 mm Hg or use of antihypertensive drugs.(10) Body mass index was calculated as weight (kg)/height (m)², and obesity was defined as a BMI ≥ 30 kg/m². Type II diabetes was determined according to the American Diabetes Association criteria.(50)

Fasting blood samples were assayed at MedStar Research Institute, Washington, D.C., using standard laboratory methods as previously described.(49) Triglycerides (TG), total cholesterol and high density lipoprotein cholesterol (HDL-C) were measured using enzymatic reagents and the Hitachi 717. Low density lipoprotein cholesterol (LDL-C) was derived using the Friedewald equation; it was directly measured in subjects with triglyceride values of >400 mg/dl.(51) Serum creatinine was measured by the picric acid method.(52) GFR was estimated by the abbreviated MDRD equation: $GFR (\text{ml}/\text{min}/1.73\text{m}^2) = 186.3 \times (\text{serum creatinine})^{-1.154} \times (\text{Age})^{-0.203} \times (0.742 \text{ if female}) \times (1.210 \text{ if African American})$.(53) This prediction equation has been previously validated in an American Indian population.(42) GFR was calculated using the formula for Caucasians. Information regarding smoking status was obtained during a personal interview and was defined as having ever had at least 100 cigarettes.

Genotypes

The SHFS genotyping procedures have been previously described.(54) In brief, DNA was isolated from fasting blood samples using organic solvents, and then amplified in separate polymerase chain reactions (PCR) with primers specific for short tandem repeat markers using the ABI PRISM Linkage Mapping Set-MD10 Version 2.5 (Applied Biosystems, Foster City, CA). PCR products were loaded into an ABI PRISM 377 DNA sequencer for laser-based automated genotyping. Analyses and assignment of the marker alleles were done using computerized algorithms (Applied Biosystems).

Genetic distances were obtained using sex-averaged chromosomal maps from the Marshfield Center for Medical Genetics (<http://research.marshfieldclinic.org/genetics>) and are reported in Haldane centiMorgans (cM). Pedigree relationships were verified using the PREST (pedigree relationship statistical tests) package, which employs likelihood-based inference statistics for genome-wide identity-by-descent (IBD) allele sharing.(55) Mendelian inconsistencies and spurious double recombinants were detected using the SimWalk2 package.(56) The overall blanking rate for both types of errors was less than 1% of the total number of genotypes for Arizona, Dakotas and Oklahoma. The cytogenetic locations of markers were determined using the web resources of the University of California Santa Cruz (UCSC) (<http://genome.ucsc.edu>) and the Marshfield Linkage maps (<http://research.marshfieldclinic.org/genetics/MarkerSearch/buildMap.asp>).

Quantitative Genetic Analyses

GFR was rank transformed to normalize the distribution and attain a kurtosis of less than 1.0. SAS, version 9.1, was used to calculate the residual variability after covariate adjustment. To maximize the power to detect genetic effects, we considered two different models of covariate adjustment in each center and the full sample. In model 1, adjustments were made for age, sex, age², as well as age-by-sex interactions. Model 2 incorporated additional covariates supported by current literature as potential confounders including: BMI, SBP, DBP, TG, HDL-C, LDL-C, diabetic status (yes versus no) and smoking status (current and former versus never). TG levels were log transformed and outliers from the distribution of systolic blood pressure were set to 200 mmHg (Winsorization) to maintain normal distributions. The presence of ACEI/ARB medications as well as hypertension severity could potentially confound or modify the

genetic influence over GFR. Therefore, we additionally ran models 1 and 2 excluding individuals on any antihypertensive medication (specific medication use was unknown). All models were stratified by center.

SOLAR, version 2.1.2, was used to perform multipoint variance component linkage analysis of the residuals. Details of this model have been described previously.(57,58) The use of the variance component approach requires an estimate of the IBD matrix. We used the Loki package which employs a Markov chain Monte Carlo stochastic procedure to compute the IBD allele sharing at points throughout the genome conditional on the genotype information available at neighboring points.(59)

Interaction Analyses

Genotype-by-diabetes, -hypertension and -obesity interactions on GFR were explored using a three step strategy. We initially tested for evidence of additive interaction by accounting for the genetic covariance differences according to diabetic, hypertension or obesity status in relative pairs. In these analyses, the likelihood of a model including genotype-by-diabetes, -hypertension or -obesity interaction is compared to the likelihood of restricted models in which such interactions are excluded. We tested for differential additive genetic effects among diabetic versus non-diabetic, hypertensive versus normotensive or obese versus non-obese participants (genetic correlation (ρ_g) $\neq 1$); for differences in the magnitude of the genetic effects among diabetic versus non-diabetic, hypertensive versus normotensive and obese versus non-obese participants (genetic variance (σ_g) \neq among two groups); and for differences in residual environmental interaction with diabetes, hypertension or obesity status (environmental variance (σ_e) \neq among two groups).

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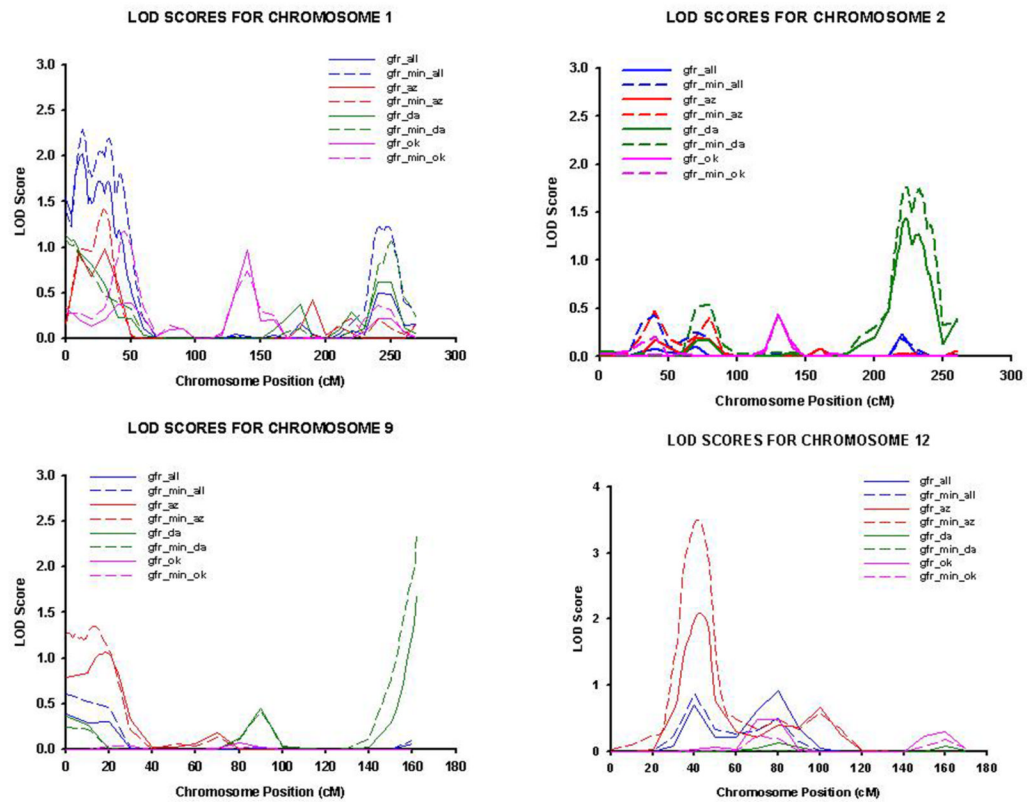


Figure 1. Cumulative multipoint Logarithm of Odds (LOD) scores for ranked estimated glomerular filtration rate using the simplified Modification of Diet in Renal Disease (MDRD) equation on chromosomes 1, 2, 9 and 12 in phase IV participants of the Strong Heart Family Study (2001–2003). gfr_all: model 2 in all centers; gfr_min_all: model 1 in all centers; gfr_az: model 2 in Arizona; gfr_min_az: model 1 in Arizona; gfr_da: model 2 in Dakotas; gfr_min_da: model 1 in Dakotas; gfr_ok: model 2 in Oklahoma; gfr_min_ok: model 1 in Oklahoma

Table 1
 Descriptive statistics* of Phase IV Strong Heart Family Study participants (2001–2003).

Characteristics	All Centers N=3665	Arizona N=1235	Dakotas N=1220	Oklahoma N=1210
Age, years	40 (17)	37 (16)	39 (17)	44 (17)
Female gender, N (%)	2 197 (60)	769 (62)	717 (59)	711 (59)
Urine ACR, µg/mg	97 (574)	152 (728)	76 (508)	64 (439)
EstimatedGFR, † ml/min/1.73m ²	100 (29)	112 (33)	96 (25)	93 (24)
Body mass index, kg/m ²	32 (8)	35 (9)	30 (7)	31 (7)
Systolic BP, mmHg	123 (17)	121 (17)	120 (16)	127 (17)
Diastolic BP, mmHg	76 (11)	77 (12)	75 (11)	77 (11)
LDL-C, mmol/L	2.5 (0.8)	2.4 (0.7)	2.6 (0.8)	2.6 (0.8)
HDL-C, mmol/L	1.3 (0.4)	1.3 (0.4)	1.3 (0.4)	1.4 (0.4)
Triglycerides, mmol/L	4.3 (4.4)	4.4 (3.5)	4.2 (5.2)	4.5 (4.4)
Hemoglobin A1C, %	8.4 (2.1)	8.7 (2.2)	7.8 (1.9)	8.2 (2.0)
Hypertension, N (%)	1 153 (31)	420 (34)	285 (23)	448 (37)
Antihypertensive treatment, N (%)	738 (20)	276 (22)	186 (15)	276 (23)
Diabetes, N (%)	830 (23)	410 (33)	172 (14)	248 (21)
	Current	311 (25)	518 (42)	401 (33)
	Former	303 (25)	284 (23)	298 (25)
	Never	606 (49)	416 (34)	510 (42)

* Means (± standard deviations) unless otherwise specified.

† Calculated using simplified MDRD.

ACR= Albumin to Creatinine Ratio; BP=blood pressure; LDL-C=Low Density Lipoprotein Cholesterol; HDL-C= High Density Lipoprotein Cholesterol

Logarithm of Odds (LOD) scores suggestive of linkage* (LOD \geq 1.8) using two modeling strategies for multipoint quantitative trait linkage analyses of ranked estimated glomerular filtration rate using the simplified Modification of Diet in Renal Disease (MDRD) equation in phase IV participants of the Strong Heart Family Study (2001–2003).

Table 2

Center	Chromosome	Location (cM)	Chromosomal Region	Nearest Marker	Empirical LOD score (model 1, model 2) [†]
All Centers	1	12	1p36.31	DIS214	2.3, 2.0
Dakotas	2	205	2q33.3	D2S325	1.8, 0.5
Dakotas	9	150	9q34.2	D9S164	2.4, 1.7
Arizona	12	39	12p12.2	D12S310	3.5, 2.1

* Linkage significance criteria were as suggested by Rao and Gu (2001).

[†] Model 1 was adjusted for age, sex, age squared and age-sex interactions.

Model 2 was additionally adjusted for diabetic status, body mass index, systolic blood pressure, diastolic blood pressure, high density lipoprotein cholesterol, low density lipoprotein cholesterol, triglycerides and smoking status.