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## Neuroendocrine Factors in the Regulation of Inflammation: Excessive Adiposity and Calorie Restriction

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### Abstract

Acute inflammation is usually a self-limited life preserving response, triggered by pathogens and/or traumatic injuries. This transient response normally leads to removal of harmful agents and to healing of the damaged tissues. In contrast, unchecked or chronic inflammation can lead to persistent tissue and organ damage by activated leukocytes, cytokines, or collagen deposition. Excessive energy intake and adiposity cause systemic inflammation, whereas calorie restriction without malnutrition exerts a potent anti-inflammatory effect. As individuals accumulate fat and their adipocytes enlarge, adipose tissue undergoes molecular and cellular alterations, macrophages accumulate, and inflammation ensues. Overweight/obese subjects have significantly higher plasma concentrations of C-reactive protein and several cytokines, including IL-6, IL-8, IL-18, and TNF-alpha. Experimental animals on a chronic CR regimen, instead, have low levels of circulating inflammatory cytokines, low blood lymphocyte levels, reduced production of inflammatory cytokines by the white blood cells in response to stimulation, and cortisol levels in the high normal range. Recent data demonstrate that CR exerts a powerful anti-inflammatory effect also in non-human primates and humans. Multiple metabolic and neuroendocrine mechanisms are responsible for the CR-mediated anti-inflammatory effects, including reduced adiposity and secretion of pro-inflammatory adipokines, enhanced glucocorticoid production, reduced plasma glucose and advanced glycation end-product concentrations, increased parasympathetic tone, and increased ghrelin production. Measuring tissue specific effects of CR using genomic, proteomic and metabolomic techniques in humans will foster the understanding of the complex biological processes involved in the anti-inflammatory and anti-aging effects of CR.

### Keywords

obesity; energy intake; adipocyte; adipokines; neuroendocrine

### Introduction

Inflammation is a complex set of responses to cellular/tissue injuries triggered by pathogenic invaders and/or trauma. This well-orchestrated and tightly controlled process normally leads to elimination of noxious agents and to healing of the damaged tissues (Nathan 2002).

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Therefore, acute inflammation usually is a self-limited life preserving response, as reflected by the augmented risk of life-threatening infections in individuals with genetic deficits in some components of the inflammatory cascade (Arnaout 1990).

In contrast, chronic inflammation triggered, for example, by some viral or intra-cellular pathogens (e.g. Hepatitis B and C, Mycobacterium tuberculosis, etc.) can lead to persistent tissue damage by activated leukocytes, cytokines, and collagen deposition, and in some cases may even promote neoplastic transformation (Schottenfeld and Beebe-Dimmer 2006). In addition, chronic inflammation plays a central role in the pathogenesis of several age-associated chronic diseases (e.g. atherosclerosis, cancer, arthritis, dementia and osteoporosis) and in the aging process itself (Akiyama et al. 2000; Coussens and Werb 2002; Libby 2002).

Elevated cellular and circulating IL-6 levels have emerged to be a common characteristic of senescence among several species, including rodents, monkeys and humans (Ershler et al. 1993; Harris et al. 1999). Recently, it has been shown that excessive energy intake and adiposity, in particular visceral adiposity, cause a state of systemic chronic inflammation (Rajala and Scherer 2003; Visser et al. 1999; Fontana et al. 2007a). In contrast, calorie restriction (CR) without malnutrition, the most robust intervention to slow down aging and prevent/delay the occurrence of many chronic diseases, exerts a potent anti-inflammatory effect (Fontana and Klein 2007).

## **Excessive adiposity, neuroendocrine alterations and inflammation**

Excessive caloric intake, particularly in sedentary individuals, results in a chronic positive energy balance, increased abdominal adiposity and several metabolic and neuroendocrine alterations, including: (1) insulin resistance (Xu et al. 2003); (2) chronic inflammation (Rajala and Scherer 2003; Visser et al. 1999; Fontana et al. 2007a); (3) sympathetic nervous system and angiotensin system activation (Seals and Bell 2004; Engeli et al. 2000); (4) accumulation of advanced glycation end-products (Soldatos et al. 2005), (5) oxidative stress-induced protein, lipids and DNA damage (Furukawa et al. 2004), and (6) alterations in neuroendocrine systems (Smith et al. 2001). All these metabolic abnormalities are implicated in the pathogenesis of several age-associated chronic diseases, such as type 2 diabetes, atherosclerosis, cancer and arthritis, which can cause accelerated dysfunction and damage of various tissues and organs (Must et al. 1999).

Chronic inflammation, in particular, is one of the key metabolic alterations associated with excessive calorie intake and adiposity (Rajala and Scherer 2003; Visser et al. 1999; Fontana et al. 2007a). Data from epidemiological studies have clearly shown that obesity is associated with increased non-infectious chronic inflammation, manifested by an up-regulation of cytokine and chemokine production by adipose tissue, presumably from both adipocytes and infiltrated macrophages (Rajala and Scherer 2003; Wellen and Hotamisligil 2003). Weight (fat) loss, induced by a negative energy balance, reduces inflammation by decreasing the production of inflammatory cytokines and chemokines by adipose tissue (Esposito et al. 2003). In contrast, removal of large amounts of subcutaneous abdominal fat by liposuction does not improve inflammation or other cardiometabolic risk factors for cardiovascular disease (Klein et al. 2004).

Until rather recently, adipocytes were simply considered to be fat storage depots. Instead, it is now clear that adipose tissue is an endocrine organ that secretes several adipokines, which act as hormones as well as pro-inflammatory cytokines (Rajala and Scherer 2003). These include leptin, adiponectin, resistin, tumor necrosis factor (TNF), IL-6, MCP-1, and other cytokines and chemokines (Rajala and Scherer 2003). As individuals accumulate fat and their adipocytes enlarge, adipose tissue undergoes molecular and cellular alterations, macrophages accumulate, and inflammation ensues (Cancello et al. 2005; Weisberg et al. 2003). Infiltrated and activated

macrophages in adipose tissue of obese individuals are responsible for most of the cytokine production, in particular IL-6 and TNF- $\alpha$  (Cancello et al. 2005; Weisberg et al. 2003).

Plasma IL-6 concentrations are much higher in the portal vein, which drains visceral fat, than in peripheral artery blood in obese subjects, demonstrating that visceral fat is an important source of IL-6 production in obese persons (Fontana et al. 2007a). These data are consistent with previous *ex vivo* findings that IL-6 secretion is higher in omental than in subcutaneous adipose tissue (Fain et al. 2004). Moreover, portal vein IL-6 concentrations correlate directly with arterial CRP concentrations in obese subjects (Fontana et al. 2007a). Recently, IL-6 was shown to cause systemic insulin resistance (Cai et al. 2005). Elevated IL-6 and CRP concentrations are predictors for the development of type 2 diabetes and myocardial infarction (Fontana et al. 2007a; Xu et al. 2003; Sattar et al. 2003).

## Calorie restriction and inflammation

Hundreds of studies have consistently shown that CR without malnutrition increases maximum lifespan and healthspan in different species, including yeast, flies, worms, fish, and rodents (Masoro 2000; Weindruch and Sohal 1997). Multiple overlapping mechanisms are responsible for the CR-mediated effects on aging-related diseases and aging itself in rodents, including decreased free radical-induced tissue damage, improved insulin sensitivity, decreased inflammation and several major neuroendocrine adaptive responses (Fontana and Klein 2007). In addition, CR enhances DNA repair processes, decreases protein glycation, and increases removal of damaged cellular proteins and oxidized lipids (Sell et al. 2003; Forster et al. 2000; Leeuwenburgh et al. 1997). Many of the effects of CR are likely mediated by regulation of gene expression, with 1) upregulation of genes involved in stress resistance, protection against oxidative damage, cellular repair and survival, and 2) downregulation of genes involved in mediating inflammation (Lee et al. 1999).

One of the most important mechanisms for the CR-mediated retardation of aging and prevention/delay of chronic diseases is the reduction in chronic systemic inflammation (Ershler et al. 1993). It is important to note that the CR-induced lower inflammatory state does not occur passively from an absence of inflammatory stimuli, but from positive actions of specific metabolic, hormonal and gene expression products that repress reactions to potential inflammatory stimuli that do not demand a full response (Higami et al. 2003; Lamas et al. 2003; Matsuzaki et al. 2001; Spaulding et al. 1997a).

Studies on animals have consistently shown that chronic CR results in low levels of circulating inflammatory cytokines, low blood lymphocyte levels, reduced production of inflammatory cytokines by the white blood cells in response to stimulation, and cortisol levels in the high normal range (Higami et al. 2003; Lamas et al. 2003; Matsuzaki et al. 2001; Spaulding et al. 1997a; Sabatino et al. 1991; Spaulding et al. 1997b; Tsuchiya et al. 2005). Furthermore, chronic CR enhances the ability to cope with intense inflammatory stressors. The inflammatory response to an injection of carageenan toxin into the footpad is attenuated in mice on a CR regimen (Klebanov et al. 1995). In addition, CR protects against aging-associated deterioration in immune function, including reduced immune response capacities, thymic involution, and shifts in leukocyte and lymphocyte subsets (Spaulding et al. 1997b; Poetschke et al. 2000; Weindruch et al. 1997; Messaoudi et al. 2006). CR also reduces the production of prostanoids (e.g. TXA<sub>2</sub>, PGI<sub>2</sub>, and PGE<sub>2</sub>) that occurs in sites of disease and inflammation (Kim et al. 2004).

Recent data show that CR also exerts a powerful anti-inflammatory effect in non-human primates and humans (Mascarucci et al. 2002). In monkeys CR attenuates the age-associated increase in IL-6 protein production by peripheral mononuclear cells (Mascarucci et al. 1999; Kim et al. 1997). Data from self-imposed CR, and from short-term CR intervention

studies have shown that CR in humans results in a reduction of markers of inflammation and pro-inflammatory cytokines. In non-obese, sedentary humans, a randomized clinical study demonstrated that a 20% reduction in calorie intake for 12 months decreases visceral fat mass, improves insulin sensitivity, increases plasma adiponectin concentration, reduces circulating inflammatory markers, decreases plasma T3 levels, and reverses some of the age-related deterioration in cardiac diastolic function (Fontana et al. 2007b; Racette et al. 2006; Weiss et al. 2006).

Information regarding the long-term effects of CR is coming from studies conducted in members of the Calorie Restriction Society, who practice self-imposed CR in the belief that CR will extend their lifespan (Fontana et al. 2004; Holloszy and Fontana 2007; Meyer et al. 2006). The CR Society members designed their diets to consist primarily of micronutrient-dense foods, supplying more than 100% of the Recommended Daily Intake (RDI) for all of the essential nutrients, while minimizing energy-dense foods and thus total energy content. These individuals have been eating ~ 30% fewer calories for an average of ~ 7 yr than age- and sex-matched, control subjects consuming a typical Western diet (1770±350 kcal/d and 2490±479 kcal/d, respectively).

The CR practitioners show many of the same alterations in metabolic and organ function that have been reported in CR rodents, including a low percent total body and truncal fat, low blood pressure, enhanced insulin sensitivity and lipid profile, and low serum T3 concentration (Fontana et al. 2004; Fontana et al. 2006). Also like mice and rats, middle-aged humans on CR have very low levels of chronic inflammation as reflected in remarkably low levels of C-reactive protein of ~0.3 mg/L compared to values in the 1.5 to 2mg/L range in healthy age-matched controls (Fontana et al. 2004; Holloszy and Fontana 2007; Meyer et al. 2006). Serum tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) levels are also low in humans on long-term CR, with values approximately 50% lower than those of age-matched controls (Fontana et al. 2006; Meyer et al. 2006).

In addition, left ventricular (LV) diastolic function (i.e. parameters of viscoelasticity and stiffness) in these CR subjects were similar to normal sex-matched subjects who were approximately 16 yr younger (Meyer et al. 2006). This finding suggests that CR might influence primary aging in humans, because aging results in a progressive increase in LV stiffness and impaired LV diastolic function (Kitzman et al. 1991), and is similar to the cardiac effects of CR observed in mice (Taffet et al. 1997). The mechanism for this beneficial effect on LV function is not known, but could represent decreased connective tissue accumulation and myocardial fibrosis caused by the effects of CR on inflammatory/fibrogenic mediators, such as TNF- $\alpha$  and transforming growth factor  $\beta$ 1 (TGF $\beta$ 1) (Sivasubramanian et al. 2001) (Border and Ruoslahti 1992).

## **Anti-inflammatory mechanisms/pathways triggered by chronic calorie restriction**

### **Reduced adiposity and circulating adipokines**

Excessive adiposity is associated with a state of low grade chronic inflammation as reflected in significant higher circulating levels of IL-6, IL-8, IL-18, TNF- $\alpha$ , MCP-1, fibrinogen and C-reactive protein (Rajala and Scherer 2003; Visser et al. 1999; Fontana et al. 2007a). Weight loss induced by CR or exercise reduces macrophage activation and infiltration in adipose tissue, and systemic inflammation (Clement et al. 2004; Esposito et al. 2003; Xydakis et al. 2004). The weight-loss mediated reduction in inflammatory markers is also related to improvements in insulin resistance, and serum adipokine concentrations. Weight loss reduces circulating levels of resistin, a proinflammatory molecule (Bokarewa et al. 2005), and increases adiponectin, an

insulin sensitizer and potent anti-inflammatory hormone produced by adipocytes (Engeli et al. 2003; Xydakis et al. 2004). In addition, weight loss reduces leptin, an adipokine with multiple biological effects in modulating inflammation and autoimmune reactivity. Leptin induces T cell proliferation, promotes Th1 cell differentiation and induces pro-inflammatory cytokine production (Fantuzzi and Faggioni 2000).

### Enhanced glucocorticoids production

Another mechanism by which CR may selectively exert its anti-inflammatory effects is via enhanced endogenous corticosteroid production (Sabatino et al. 1991). Chronic CR potentiates the diurnal elevation of plasma corticosterone. CR mice and rats have “moderately” but significantly higher daily mean plasma free corticosterone concentration than mice fed “ad libitum” throughout their lifespan. This moderate increase in corticosterone does not, however, reflect increased hypothalamic-pituitary activity, as plasma ACTH concentration is lower in CR than in “ad libitum” fed rodents (Han et al. 2001). The elevated free corticosterone is due to an enhanced response of the adrenal gland to ACTH (Han et al. 2001).

It is well known that the hypothalamic–pituitary–adrenal axis and glucocorticoids in particular are essential in limiting and resolving the inflammatory process (Sapolsky et al. 2000). Glucocorticoids have pleiotropic inhibitory effects on the immune system and inflammatory gene expression (Rhen and Cidlowski 2005). In addition, treatment with pharmacological doses of exogenous glucocorticoids has been used to block many inflammatory and autoimmune diseases, including rheumatoid arthritis, systemic lupus erythematosus, Graves’ disease, thyroiditis, glomerulonephritis, multiple sclerosis, and psoriasis.

### Reduced glycemia and advanced glycation end-products

Hyperglycemia and the accumulation of advanced glycation end products (AGE) have been shown to exert pro-inflammatory effects. Hyperglycemia, a major metabolic disorder associated with excessive adiposity and insulin resistance, has been shown to upregulate the expression of several genes involved in inflammation (e.g. activator protein-1, nuclear factor- $\kappa$ B, and early growth response), and the secretion of inflammatory cytokines in several cell types both *in vitro* and *ex vivo* (Nareika et al. 2006; Morohoshi et al. 1996).

Moreover, it has been shown that hyperglycemic spikes acutely increase plasma IL-6, TNF- $\alpha$ , and IL-18 concentrations, probably through an oxidative mechanism, in patients with impaired glucose tolerance as well as in healthy control subjects (Esposito et al. 2002). Finally, the interaction of the receptors for advanced glycation end products (RAGE) with AGE results in activation of signal transduction pathways that mediate diverse responses in a wide array of cell types, including induction of oxidative stress, increased extracellular matrix permeability, release of inflammatory cytokines and growth factors, and the increased expression of adhesion molecules and chemokines (Hofmann et al. 1999).

Chronic CR improves insulin sensitivity and reduces serum glucose and AGEs concentrations (Masoro et al. 1989; Cefalu et al. 1995). Moreover, CR significantly reduces AGE accumulation in several tissues of experimental animals, including kidney, aorta and skin (Cefalu et al. 1995; Sell et al. 2003; Teillet et al. 2000).

### Sympatholitic activity and increased parasympathetic tone

The two principal branches of the autonomic nervous system (parasympathetic and sympathetic pathways) control heart rate, blood pressure, and respiratory rate, gastrointestinal motility, and other essential life functions such as the innate immune system (Tracey 2007). Rodents maintained on a CR diet have lower resting heart rate and higher heart and blood pressure



variability than rodents on ad libitum diet, suggesting that CR decreases sympathetic activity and augments parasympathetic or vagal tone (Mager et al. 2006).

Interestingly, vagus nerve stimulation has been shown to effectively suppress cytokine-mediated inflammation and damage in several pathological conditions, including endotoxemia, ischemia reperfusion injury, sepsis, and arthritis (Borovikova et al. 2000). In contrast, vagotomy aggravates cytokine responses to inflammatory stimuli and sensitizes animals to the lethal effects of endotoxin (Borovikova et al. 2000). The neurotransmitter acetylcholine is released by firing efferent vagus nerve endings, binds to nicotinic- $\alpha 7$  receptors on resident macrophages, and thus prevents the release of TNF, IL-1, and IL-18 (Wang et al. 2003). Data from experimental animal studies indicate that the threshold of vagus nerve activity that stimulates the vagal anti-inflammatory pathway is significantly lower than that required to change heart rate variability.

### Increased ghrelin production

Ghrelin is a circulating orexigenic peptide secreted predominantly from stomach cells (Lee et al. 2002). Ghrelin modulates growth hormone release, energy homeostasis and central feeding regulation. Ghrelin levels increase following fasting and chronic food deprivation (Lee et al. 2002). Chronic CR significantly increases plasma ghrelin concentration as well as total ghrelin production in the stomach of rodents (Yang et al. 2007).

Chronic ghrelin administration potently stimulates food intake, and increases body weight and adiposity in rodents (Tschop et al. 2000). Intravenous ghrelin infusion enhances appetite and increases food intake in humans, too (Wren et al. 2001). In addition, ghrelin exerts potent anti-inflammatory effects (Li et al. 2004). It has been shown that ghrelin, via the functional cell surface growth hormone secretagogue receptor (GHS-R), inhibits proinflammatory cytokine production, cell binding, and nuclear factor-kappaB activation in a wide variety of cells in vitro (Dixit et al. 2004). Further, ghrelin attenuates endotoxin-induced cytokine production in a murine cytokine-induced anorexia model. Interestingly, ghrelin and leptin exert mutually antagonistic effects on food intake and immune function (Dixit et al. 2004).

## CONCLUSIONS

The neuroendocrine system and inflammation are deeply involved in the pathogenesis of many age-related diseases and in the biology of the aging process itself. Calorie intake and adiposity, and their associated metabolic and neuroendocrine alterations, are important determinants of inflammation in experimental animals and humans.

Excessive calorie intake and adiposity up-regulate the expression of several genes and proteins involved in mediating inflammation in many tissues and organs. In contrast, CR with adequate nutrition, a robust life extending intervention, prevents or attenuates local and systemic inflammation via multiple metabolic and neuroendocrine mechanisms. Recent data indicate that CR exerts a powerful anti-inflammatory effect also in non-human primates and humans. Measuring tissue specific effects of CR using genomic, proteomic and metabolomic techniques in humans will promote an understanding of the complex biological processes involved in the anti-inflammatory and anti-aging effects of CR.

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## Reference List

1. Akiyama H, Barger S, Barnum S, Bradt B, Bauer J, Cole GM, Cooper NR, Eikelenboom P, Emmerling M, Fiebich BL, Finch CE, Frautschy S, Griffin WST, Hampel H, Hull M, Landreth G, Lue LF, Mraz R, Mackenzie IR, McGeer PL, O'Banion MK, Pachter J, Pasinetti G, Plata-Salaman C, Rogers J, Rydel R, Shen Y, Streit W, Strohmeyer R, Tooyoma I, Van Muiswinkel FL, Veerhuis R, Walker D, Webster S, Wegrzyniak B, Wenk G, Wyss-Coray T. Inflammation and Alzheimer's disease. *Neurobiol Aging* 2000;21:383–421. [PubMed: 10858586]
2. Arnaout MA. Leukocyte Adhesion Molecules Deficiency - Its Structural Basis, Pathophysiology and Implications for Modulating the Inflammatory Response. *Immunol Rev* 1990;114:145–180. [PubMed: 1973407]
3. Bokarewa M, Nagaev I, Dahlberg L, Smith U, Tarkowski A. Resistin, an adipokine with potent proinflammatory properties. *J Immunol* 2005;174:5789–5795. [PubMed: 15843582]
4. Border WA, Ruoslahti E. Transforming Growth-Factor-Beta in Disease - the Dark Side of Tissue-Repair. *J Clin Invest* 1992;90:1–7. [PubMed: 1634602]
5. Borovikova LV, Ivanova S, Zhang MH, Yang H, Botchkina GI, Watkins LR, Wang HC, Abumrad N, Eaton JW, Tracey KJ. Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxin. *Nature* 2000;405:458–462. [PubMed: 10839541]
6. Cai DS, Yuan MS, Frantz DF, Melendez PA, Hansen L, Lee J, Shoelson SE. Local and systemic insulin resistance resulting from hepatic activation of IKK-beta and NF-kappa B. *Nat Med* 2005;11:183–190. [PubMed: 15685173]
7. Canello R, Henegar C, Viguier N, Taleb S, Poitou C, Rouault C, Coupaye M, Pelloux V, Hugol D, Bouillot JL, Bouloumie A, Barbatelli G, Cinti S, Svensson PA, Barsh GS, Zucker JD, Basdevant A, Langin D, Clement K. Reduction of macrophage infiltration and chemoattractant gene expression changes in white adipose tissue of morbidly obese subjects after surgery induced weight loss. *Diabetes* 2005;54:2277–2286. [PubMed: 16046292]
8. Cefalu WT, Bellfarrow AD, Wang ZQ, Sonntag WE, Fu MX, Baynes JW, Thorpe SR. Caloric Restriction Decreases Age-Dependent Accumulation of the Glycoxidation Products, N-Epsilon-(Carboxymethyl)Lysine and Pentosidine, in Rat Skin Collagen. *Journals of Gerontology Series A-Biological Sciences and Medical Sciences* 1995;50:B337–B341.
9. Clement K, Viguier N, Poitou C, Carette C, Pelloux V, Curat CA, Sicard A, Rome S, Benis A, Zucker JD, Vidal H, Laville M, Barsh GS, Basdevant A, Stich V, Canello R, Langin D. Weight loss regulates inflammation-related genes in white adipose tissue of obese subjects. *FASEB J* 2004;18:1657–1669. [PubMed: 15522911]
10. Coussens LM, Werb Z. Inflammation and cancer. *Nature* 2002;420:860–867. [PubMed: 12490959]
11. Dixit VD, Schaffer EM, Pyle RS, Collins GD, Sakthivel SK, Palaniappan R, Lillard JW, Taub DD. Ghrelin inhibits leptin- and activation-induced proinflammatory cytokine expression by human monocytes and T cells. *J Clin Invest* 2004;114:57–66. [PubMed: 15232612]
12. Engeli S, Feldpausch M, Gorzelniak K, Hartwig F, Heintze U, Janke J, Mohlig M, Pfeiffer AFH, Luft FC, Sharma AM. Association between adiponectin and mediators of inflammation in obese women. *Diabetes* 2003;52:942–947. [PubMed: 12663465]
13. Engeli S, Negrel R, Sharma AM. Physiology and pathophysiology of the adipose tissue renin-angiotensin system. *Hypertension* 2000;35:1270–1277. [PubMed: 10856276]
14. Ershler WB, Sun WH, Binkley N, Gravenstein S, Volk MJ, Kamoske G, Klopp RG, Roecker EB, Daynes RA, Weindruch R. Interleukin-6 and Aging - Blood-Levels and Mononuclear Cell Production Increase with Advancing Age and In-Vitro Production Is Modifiable by Dietary Restriction. *Lymphokine Cytokine Res* 1993;12:225–230. [PubMed: 8218595]
15. Esposito K, Nappo F, Marfella R, Giugliano G, Giugliano F, Ciotola M, Quagliari L, Ceriello A, Giugliano D. Inflammatory cytokine concentrations are acutely increased by hyperglycemia in humans - Role of oxidative stress. *Circulation* 2002;106:2067–2072. [PubMed: 12379575]

16. Esposito K, Pontillo A, Di Palo C, Giugliano G, Masella M, Marfella R, Giugliano D. Effect of weight loss and lifestyle changes on vascular inflammatory markers in obese women - A randomized trial. *Jama-Journal of the American Medical Association* 2003;289:1799–1804.
17. Fain JN, Madan AK, Hiler ML, Cheema P, Bahouth SW. Comparison of the release of adipokines by adipose tissue, adipose tissue matrix, and Adipocytes from visceral and subcutaneous abdominal adipose tissues of obese humans. *Endocrinology* 2004;145:2273–2282. [PubMed: 14726444]
18. Fantuzzi G, Faggioni R. Leptin in the regulation of immunity, inflammation, and hematopoiesis. *J Leukoc Biol* 2000;68:437–446. [PubMed: 11037963]
19. Fontana L, Eagon JC, Trujillo ME, Scherer PE, Klein S. Visceral fat adipokine secretion is associated with systemic inflammation in obese humans. *Diabetes* 2007a;56:1010–1013. [PubMed: 17287468]
20. Fontana L, Klein S. Aging, adiposity, and calorie restriction. *Jama-Journal of the American Medical Association* 2007;297:986–994.
21. Fontana L, Klein S, Holloszy JO, Premachandra BN. Effect of long-term calorie restriction with adequate protein and micronutrients on thyroid hormones. *J Clin Endocrinol Metab* 2006;91:3232–3235. [PubMed: 16720655]
22. Fontana L, Meyer TE, Klein S, Holloszy JO. Long-term calorie restriction is highly effective in reducing the risk for atherosclerosis in humans. *Proc Natl Acad Sci U S A* 2004;101:6659–6663. [PubMed: 15096581]
23. Fontana L, Villareal DT, Weiss EP, Racette SB, Steger-May K, Klein S, Holloszy JO. Calorie restriction or exercise: effects on coronary heart disease risk factors. A randomized, controlled trial. *American Journal of Physiology-Endocrinology and Metabolism* 2007b;293:E197–E202. [PubMed: 17389710]
24. Forster MJ, Sohal BH, Sohal RS. Reversible effects of long-term caloric restriction on protein oxidative damage. *Journals of Gerontology Series A-Biological Sciences and Medical Sciences* 2000;55:B522–B529.
25. Furukawa S, Fujita T, Shimabukuro M, Iwaki M, Yamada Y, Nakajima Y, Nakayama O, Makishima M, Matsuda M, Shimomura I. Increased oxidative stress in obesity and its impact on metabolic syndrome. *J Clin Invest* 2004;114:1752–1761. [PubMed: 15599400]
26. Han ES, Evans TR, Shu JH, Lee S, Nelson JF. Food restriction enhances endogenous and corticotropin-induced plasma elevations of free but not total corticosterone throughout life in rats. *Journals of Gerontology Series A-Biological Sciences and Medical Sciences* 2001;56:B391–B397.
27. Harris TB, Ferrucci L, Tracy RP, Corti MC, Wacholder S, Ettinger WH, Heimovitz H, Cohen HJ, Wallace R. Associations of elevated interleukin-6 and C-reactive protein levels with mortality in the elderly. *Am J Med* 1999;106:506–512. [PubMed: 10335721]
28. Higami Y, Pugh TD, Page GP, Allison DB, Prolla TA, Weindruch R. Adipose tissue energy metabolism: altered gene expression profile of mice subjected to long-term caloric restriction. *FASEB J* 2003;17:415. [PubMed: 14688200]
29. Hofmann MA, Drury S, Fu CF, Qu W, Taguchi A, Lu Y, Avila C, Kambham N, Bierhaus A, Nawroth P, Neurath MF, Slattery T, Beach D, McClary J, Nagashima M, Morser J, Stern D, Schmidt AM. RAGE mediates a novel proinflammatory axis: A central cell surface receptor for S100/calgranulin polypeptides. *Cell* 1999;97:889–901. [PubMed: 10399917]
30. Holloszy JO, Fontana L. Caloric restriction in humans. *Exp Gerontol* 2007;42:709–712. [PubMed: 17482403]
31. Kim JW, Zou Y, Yoon S, Lee JH, Kim YK, Yu BP, Chung HY. Vascular aging: Molecular modulation of the prostanoid cascade by calorie restriction. *Journals of Gerontology Series A-Biological Sciences and Medical Sciences* 2004;59:876–885.
32. Kim MJ, Aiken JM, Havighurst T, Hollander J, Ripple MO, Weindruch R. Adult-onset energy restriction of rhesus monkeys attenuates oxidative stress-induced cytokine expression by peripheral blood mononuclear cells. *J Nutr* 1997;127:2293–2301. [PubMed: 9405577]
33. Kitzman DW, Sheikh KH, Beere PA, Philips JL, Higginbotham MB. Age-Related Alterations of Doppler Left-Ventricular Filling Indexes in Normal Subjects Are Independent of Left-Ventricular Mass, Heart-Rate, Contractility and Loading Conditions. *J Am Coll Cardiol* 1991;18:1243–1250. [PubMed: 1918701]



34. Klebanov S, Diai S, Stavinoha WB, Suh YM, Nelson JF. Hyperadrenocorticism, Attenuated Inflammation, and the Life-Prolonging Action of Food Restriction in Mice. *Journals of Gerontology Series A-Biological Sciences and Medical Sciences* 1995;50:B78–B82.
35. Klein S, Fontana L, Young VL, Coggan AR, Kilo C, Patterson BW, Mohammed BS. Absence of an effect of liposuction on insulin action and risk factors for coronary heart disease. *N Engl J Med* 2004;350:2549–2557. [PubMed: 15201411]
36. Lamas O, Moreno-Aliaga MJ, Martinez JA, Marti A. NF-kappa B-binding activity in an animal diet-induced overweightness model and the impact of subsequent energy restriction. *Biochem Biophys Res Commun* 2003;311:533–539. [PubMed: 14592449]
37. Lee CK, Klopp RG, Weindruch R, Prolla TA. Gene expression profile of aging and its retardation by caloric restriction. *Science* 1999;285:1390–1393. [PubMed: 10464095]
38. Lee HM, Wang GY, Englander EW, Kojima M, Greeley GH. Ghrelin, a new gastrointestinal endocrine peptide that stimulates insulin secretion: Enteric distribution, ontogeny, influence of endocrine, and dietary manipulations. *Endocrinology* 2002;143:185–190. [PubMed: 11751608]
39. Leeuwenburgh C, Wagner P, Holloszy JO, Sohal RS, Heinecke JW. Caloric restriction attenuates dityrosine cross-linking of cardiac and skeletal muscle proteins in aging mice. *Arch Biochem Biophys* 1997;346:74–80. [PubMed: 9328286]
40. Li WG, Gavrilu D, Liu XB, Wang LX, Gunnlaugsson S, Stoll LL, McCormick ML, Sigmund CD, Tang CS, Weintraub NL. Ghrelin inhibits proinflammatory responses and nuclear factor-kappa B activation in human endothelial cells. *Circulation* 2004;109:2221–2226. [PubMed: 15117840]
41. Libby P. Inflammation in atherosclerosis. *Nature* 2002;420:868–874. [PubMed: 12490960]
42. Mager DE, Wan RQ, Brown M, Cheng AW, Wareski P, Abernethy DR, Mattson MP. Caloric restriction and intermittent fasting alter spectral measures of heart rate and blood pressure variability in rats. *FASEB J* 2006;20:631–637. [PubMed: 16581971]
43. Mascarucci P, Taub D, Handy A, De Angelis H, Weininger C, Roth G, Lane M, Ingram DK. Endotoxin-induced cytokine responses in rhesus monkeys: Effects of age and calorie restriction. *Age* 1999;22:130.
44. Mascarucci P, Taub D, Saccani S, Paloma MA, Dawson H, Roth GS, Lane MA, Ingram DK. Cytokine responses in young and old rhesus monkeys: Effect of caloric restriction. *J Interferon Cytokine Res* 2002;22:565–571. [PubMed: 12060495]
45. Masoro EJ. Caloric restriction and aging: an update. *Exp Gerontol* 2000;35:299–305. [PubMed: 10832051]
46. Masoro EJ, Katz MS, McMahan CA. Evidence for the Glycation Hypothesis of Aging from the Food-Restricted Rodent Model. *Journals of Gerontology* 1989;44:B20–B22. [PubMed: 2910985]
47. Matsuzaki J, Kuwamura M, Yamaji R, Inui H, Nakano Y. Inflammatory responses to lipopolysaccharide are suppressed in 40% energy-restricted mice. *J Nutr* 2001;131:2139–2144. [PubMed: 11481408]
48. Messaoudi I, Warner J, Fischer M, Park B, Hill B, Mattison J, Lane MA, Roth GS, Ingram DK, Pickler LJ, Douek DC, Mori M, Nikolich-Zugich J. Delay of T cell senescence by caloric restriction in aged long-lived nonhuman primates. *Proc Natl Acad Sci U S A* 2006;103:19448–19453. [PubMed: 17159149]
49. Meyer TE, Kovacs SJ, Ehsani AA, Klein S, Holloszy JO, Fontana L. Long-term caloric restriction ameliorates the decline in diastolic function in humans. *J Am Coll Cardiol* 2006;47:398–402. [PubMed: 16412867]
50. Morohoshi M, Fujisawa K, Uchimura I, Numano F. Glucose-dependent interleukin 6 and tumor necrosis factor production by human peripheral blood monocytes in vitro. *Diabetes* 1996;45:954–959. [PubMed: 8666148]
51. Must A, Spadano J, Coakley EH, Field AE, Colditz G, Dietz WH. The disease burden associated with overweight and obesity. *Jama-Journal of the American Medical Association* 1999;282:1523–1529.
52. Nareika A, Im YB, Game BA, Slate EH, Sanders JJ, London SD, Lopes-Virella MF, Huang Y. High glucose markedly enhances lipopolysaccharide-induced CD14 expression in U937 histiocytes. *Diabetes* 2006;55:A171.
53. Nathan C. Points of control in inflammation. *Nature* 2002;420:846–852. [PubMed: 12490957]

54. Poetschke HL, Klug DB, Perkins SN, Wang TTY, Richie ER, Hursting SD. Effects of caloric restriction on thymocyte growth, death and maturation. *Carcinogenesis* 2000;21:1959–1964. [PubMed: 11062154]
55. Racette SB, Weiss EP, Villareal DT, Arif H, Steger-May K, Schechtman KB, Fontana L, Klein S, Holloszy JO. One year of caloric restriction in humans: Feasibility and effects on body composition and abdominal adipose tissue. *Journals of Gerontology Series A-Biological Sciences and Medical Sciences* 2006;61:943–950.
56. Rajala MW, Scherer PE. Minireview: The adipocyte - At the crossroads of energy homeostasis, inflammation, and atherosclerosis. *Endocrinology* 2003;144:3765–3773. [PubMed: 12933646]
57. Rhen T, Cidlowski JA. Antiinflammatory action of glucocorticoids - New mechanisms for old drugs. *N Engl J Med* 2005;353:1711–1723. [PubMed: 16236742]
58. Sabatino F, Masoro EJ, McMahan CA, Kuhn RW. Assessment of the Role of the Glucocorticoid System in Aging Processes and in the Action of Food Restriction. *Journals of Gerontology* 1991;46:B171–B179. [PubMed: 1890278]
59. Sapolsky RM, Romero LM, Munck AU. How do glucocorticoids influence stress responses? Integrating permissive, suppressive, stimulatory, and preparative actions. *Endocr Rev* 2000;21:55–89. [PubMed: 10696570]
60. Sattar N, Gaw A, Scherbakova O, Ford I, O'Reilly DS, Haffner SM, Isles C, Macfarlane PW, Packard CJ, Cobbe SM, Shepherd J. Metabolic syndrome with and without C-reactive protein as a predictor of coronary heart disease and diabetes in the West of Scotland Coronary Prevention Study. *Circulation* 2003;108:414–419. [PubMed: 12860911]
61. Schottenfeld D, Beebe-Dimmer J. Chronic inflammation: A common and important factor in the pathogenesis of neoplasia. *Ca-A Cancer Journal for Clinicians* 2006;56:69–83. [PubMed: 16514135]
62. Seals DR, Bell C. Chronic sympathetic activation - Consequence and cause of age-associated obesity? *Diabetes* 2004;53:276–284. [PubMed: 14747276]
63. Sell DR, Lane MA, Obrenovich ME, Mattison JA, Handy A, Ingram DK, Cutler RG, Roth GS, Monnier VM. The effect of caloric restriction on glycation and glycoxidation in skin collagen of nonhuman primates. *Journals of Gerontology Series A-Biological Sciences and Medical Sciences* 2003;58:508–516.
64. Sivasubramanian N, Coker ML, Kurrelmeyer KM, MacLellan WR, Demayo FJ, Spinale FG, Mann DL. Left ventricular remodeling in transgenic mice with cardiac restricted overexpression of tumor necrosis factor. *Circulation* 2001;104:826–831. [PubMed: 11502710]
65. Smith SR, Lovejoy JC, Greenway F, Ryan D, deJonge L, de la Bretonne J, Volafova J, Bray GA. Contributions of total body fat, abdominal subcutaneous adipose tissue compartments, and visceral adipose tissue to the metabolic complications of obesity. *Metabolism-Clinical and Experimental* 2001;50:425–435. [PubMed: 11288037]
66. Soldatos G, Cooper ME, Jandeleit-Dahm KAM. Advanced-glycation end products in insulin-resistant states. *Current Hypertension Reports* 2005;7:96–102. [PubMed: 15748532]
67. Spaulding CC, Walford RL, Effros RB. Caloric restriction inhibits the age-related dysregulation of the cytokines TNF-alpha and IL-6 in C3B10RF1 mice. *Mech Ageing Dev* 1997a;93:87–94. [PubMed: 9089573]
68. Spaulding CC, Walford RL, Effros RB. The accumulation of non-replicative, non-functional, senescent T cells with age is avoided in calorically restricted mice by an enhancement of T cell apoptosis. *Mech Ageing Dev* 1997b;93:25–33. [PubMed: 9089568]
69. Taffet GE, Pham TT, Hartley CJ. The age-associated alterations in late diastolic function in mice are improved by caloric restriction. *Journals of Gerontology Series A-Biological Sciences and Medical Sciences* 1997;52:B285–B290.
70. Teillet L, Verbeke P, Gouraud S, Bakala H, Borot-Laloi C, Heudes D, Bruneval P, Corman B. Food restriction prevents advanced glycation end product accumulation and retards kidney aging in lean rats. *J Am Soc Nephrol* 2000;11:1488–1497. [PubMed: 10906162]
71. Tracey KJ. Physiology and immunology of the cholinergic antiinflammatory pathway. *J Clin Invest* 2007;117:289–296. [PubMed: 17273548]
72. Tschop M, Smiley DL, Heiman ML. Ghrelin induces adiposity in rodents. *Nature* 2000;407:908–913. [PubMed: 11057670]

73. Tsuchiya T, Higami Y, Komatsu T, Tanaka K, Honda S, Yamaza H, Chiba T, Ayabe H, Shimokawa I. Acute stress response in calorie-restricted rats to lipopolysaccharide-induced inflammation. *Mech Ageing Dev* 2005;126:568–579. [PubMed: 15811426]
74. Visser M, Bouter LM, McQuillan GM, Wener MH, Harris TB. Elevated C-reactive protein levels in overweight and obese adults. *Jama-Journal of the American Medical Association* 1999;282:2131–2135.
75. Wang H, Yu M, Ochani M, Amella CA, Tanovic M, Susarla S, Li JH, Wang HC, Yang H, Ulloa L, Al-Abed Y, Czura CJ, Tracey KJ. Nicotinic acetylcholine receptor alpha 7 subunit is an essential regulator of inflammation. *Nature* 2003;421:384–388. [PubMed: 12508119]
76. Weindruch R, Lane MA, Ingram DK, Ershler WB, Roth GS. Dietary restriction in Rhesus monkeys: Lymphopenia and reduced mitogen-induced proliferation in peripheral blood mononuclear cells. *Aging-Clinical and Experimental Research* 1997;9:304–308.
77. Weindruch R, Sohal RS. Caloric intake and aging. *N Engl J Med* 1997;337:986–994. [PubMed: 9309105]
78. Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW. Obesity is associated with macrophage accumulation in adipose tissue. *J Clin Invest* 2003;112:1796–1808. [PubMed: 14679176]
79. Weiss EP, Racette SB, Villareal DT, Fontana L, Steger-May K, Schechtman KB, Klein S, Holloszy JO. Improvements in glucose tolerance and insulin action induced by increasing energy expenditure or decreasing energy intake: a randomized controlled trial. *Am J Clin Nutr* 2006;84:1033–1042. [PubMed: 17093155]
80. Wellen KE, Hotamisligil GS. Obesity-induced inflammatory changes in adipose tissue. *J Clin Invest* 2003;112:1785–1788. [PubMed: 14679172]
81. Wren AM, Seal LJ, Cohen MA, Brynes AE, Frost GS, Murphy KG, Dhillon WS, Ghatei MA, Bloom SR. Ghrelin enhances appetite and increases food intake in humans. *J Clin Endocrinol Metab* 2001;86:5992–5995. [PubMed: 11739476]
82. Xu HY, Barnes GT, Yang Q, Tan Q, Yang DS, Chou CJ, Sole J, Nichols A, Ross JS, Tartaglia LA, Chen H. Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *J Clin Invest* 2003;112:1821–1830. [PubMed: 14679177]
83. Xydakis AM, Case CC, Jones PH, Hoogveen RC, Liu MY, Smith EO, Nelson KW, Ballantyne CM. Adiponectin, inflammation, and the expression of the metabolic syndrome in obese individuals: The impact of rapid weight loss through caloric restriction. *J Clin Endocrinol Metab* 2004;89:2697–2703. [PubMed: 15181044]
84. Yang H, Youm YH, Nakata C, Dixit VD. Chronic caloric restriction induces forestomach hypertrophy with enhanced ghrelin levels during aging. *Peptides* 2007;28:1931–1936. [PubMed: 17875344]