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Gene expression in primate liver during viral hemorrhagic fever

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Abstract

Background: Rhesus macaques infected with lymphocytic choriomeningitis virus (LCMV) provide a model for human Lassa fever. Disease begins with flu-like symptoms and progresses rapidly with fatal consequences. Previously, we profiled the blood transcriptome of LCMV-infected monkeys (M. Djavani et al *J. Virol.* 2007) showing distinct pre-viremic and viremic stages that discriminated virulent from benign infections. In the present study, changes in liver gene expression from macaques infected with virulent LCMV-WE were compared to gene expression in uninfected monkeys as well as to monkeys that were infected but not diseased.

Results: Based on a functional pathway analysis of differentially expressed genes, virulent LCMV-WE had a broader effect on liver cell function than did infection with non-virulent LCMV-Armstrong. During the first few days after infection, LCMV altered expression of genes associated with energy production, including fatty acid and glucose metabolism. The transcriptome profile resembled that of an organism in starvation: mRNA for acetyl-CoA carboxylase, a key enzyme of fatty acid synthesis was reduced while genes for enzymes in gluconeogenesis were up-regulated. Expression was also altered for genes associated with complement and coagulation cascades, and with signaling pathways involving STAT1 and TGF- β .

Conclusion: Most of the 4500 differentially expressed transcripts represented a general response to both virulent and mild infections. However, approximately 250 of these transcripts had significantly different expression in virulent infections as compared to mild infections, with approximately 30 of these being differentially regulated during the pre-viremic stage of infection. The genes that are expressed early and differently in mild and virulent disease are potential biomarkers for prognosis and triage of acute viral disease.

Background

Arenaviruses are rodent-borne viruses that can be transmitted to primates, occasionally causing lethal hemor-

rhagic fever. Arenaviruses causing Lassa fever and South American hemorrhagic fevers have been classified as Category A bio-threats in the United States because of their

virulence. Human beings infected with a hemorrhagic fever virus initially exhibit flu-like symptoms, and disease progresses so rapidly that diagnosis and appropriate treatments are often too late. Laboratory studies using the arenavirus lymphocytic choriomeningitis virus (strain LCMV-WE) showed that rhesus macaques develop an acute viral disease similar to Lassa fever in human beings [1-8]. LCMV-associated hemorrhagic fever in macaques provided a practical model for disease in a well-controlled laboratory environment. Whereas LCMV-WE was highly pathogenic for primates and guinea pigs, animals infected with the Armstrong strain (LCMV-ARM) did not manifest disease or viremia and were protected from lethal challenge with LCMV-WE [8].

Our previous publications on the pathology of LCMV-WE infection described up-regulation of liver gene expression related to organ development, regeneration and inflammatory responses [2,3,5]. Blood profiles of LCMV-infected macaques revealed distinct pre-viremic and viremic stages of infection, with over 90 virulence-specific gene-expression changes detectable before the viremic stage [3]. The viremic or symptomatic stage of the virulent infection was characterized by high viral loads, high liver enzymes, thrombocytopenia, high plasma levels of IP-10, IFN- γ , MCP-1, IL-6, TNFR1 and TNFR2, as well as clinical signs of appetite loss, withdrawal, and fever [2-6,8]. Diseased liver tissue had disorganized parenchyma and mononuclear infiltrates (infiltrates were also seen in lung), whereas tissue from animals that were infected but not diseased had no infiltrates and appeared healthy [4-6]. Gene expression of PBMC was remarkable for its down-regulation of several signaling pathways, e.g. via IL-1 β receptor, epithelial growth factor receptor, and retinoic acid receptor [3] and this decrease was corroborated by studies in a guinea pig model for Lassa fever [9,10]. A dramatic and early drop in cyclo-oxygenase-2 gene (PTGS2) expression was observed in the primate model that could directly account for the drop in prostacyclin and platelet dysfunction described in Lassa fever [11-13].

Despite the complex clinical presentation of viral hemorrhagic fever, we chose to focus on liver gene expression because that organ had the highest virus titers. Liver tissue contains several cell types, and approximately 25% of the changes in transcriptome do not result in proteomic changes [14]; so with these *caveats* in mind, we examine the most prominent transcriptome changes in relation to published information about primate liver infections. Down-regulated genes involved in fatty acid synthesis and up-regulated genes involved in gluconeogenesis presented a profile that has been associated with starvation and also typifies LCMV infection of macaques. Although most of the gene expression changes controlling intermediary metabolism could be categorized as general homeostatic

responses to infection, some gene-expression changes, such as in transcripts related to amino-acid catabolism and protein phosphatase, were strongly associated with an early virulent profile and were more likely to contribute to fatal disease. Approximately 30 genes were identified as potential signature biomarkers for the onset of virulent LCMV-related liver disease. We discuss those gene expression changes that are similar to other viral diseases of liver and those changes that seem unique to an acute arenavirus infection.

Materials and methods

Experimental samples

Twenty healthy adult rhesus macaques, five to nine years of age, were used for a terminal study [3]. Liver tissue samples were obtained on the day of euthanasia, from uninfected controls and infected animals. Before euthanasia, blood was taken for clinical chemistry and hematology; tissues were processed for RNA extraction within 15 minutes after collection.

LCMV infection of rhesus macaques was described previously [3]. Briefly, animals were either uninfected or infected intravenously (i.v.) with LCMV-ARM or LCMV-WE using 10^3 plaque forming units (pfu) virus. LCMV-WE alone, at 10^3 pfu i.v. is uniformly lethal. Four animals infected with both LCMV-ARM and LCMV-WE (10^3 pfu each), did not develop symptoms and were classified as "infected-but-not-diseased". Infection in macaques was monitored by plaque assay of infectious particles in plasma, by infectious center assay of PBMC and liver tissues, and by RT-PCR to detect viral RNA in tissues [3].

RNA preparation and GeneChip hybridization

Total RNA was prepared from LCMV-infected or uninfected liver tissues following TRIzol (GIBCO-BRL) extraction and purification using an RNeasy system according to the manufacturer's instructions (QIAGEN, Valencia, CA). A QIAGEN RNase-free DNase supplement kit was used to ensure that the RNA had no DNA contamination. All RNA samples were checked for both quality and quantity as described previously [3]. RNA that passed this initial quality control screen was then labeled according to the standard target labeling protocols provided by Affymetrix and hybridized to the GeneChip human genome U133 Plus 2.0 array (Affymetrix, Santa Clara, CA) as described by the manufacturer <http://www.affymetrix.com>. The use of the Affymetrix human genome microarrays for monitoring transcriptome changes in rhesus macaque tissues has been validated by other studies [15-17].

Microarray data analysis

Microarray data analyses were performed using the Array Data Analysis and Management System (ADAMS), currently being developed at VBI <http://pathport.vbi.vt.edu>.

The system uses publicly available tools for analysis of the data. Briefly, raw probe intensities were normalized and summarized using a robust multichip average of G+C content algorithm (gcRMA algorithm) [18]. Detection calls (present, marginal, or absent) for each probe set were obtained using the mas5calls function in the Affy R package [19]. For paired comparisons, only genes with at least one present call among the compared samples were included.

A total of 20 samples were used to generate the microarray data <http://www.ncbi.nlm.nih.gov/geo>. Data from the 20 samples were grouped as follows to perform statistical analyses: uninfected controls (three samples), samples infected with the virulent strain LCMV-WE taken during the pre-viremic stage (four samples), samples infected with LCMV-WE during the viremic stage (five samples), samples infected with LCMV-WE during the post-viremic

stage (two samples), and six samples that were infected but not diseased (two monkeys infected with LCMV-ARM and four infected with both LCMV-ARM and LCMV-WE) (Table 1). Cluster analysis justified grouping LCMV-ARM samples with LCMV-ARM+WE samples since heat maps of gene expression from LCMV-ARM-infected animals were most similar to those from animals doubly-infected with LCMV-ARM and LCMV-WE (see Additional File 1). Mean *n*-fold changes were calculated using a division of normalized expression values between experimental sample and uninfected control. False discovery rates [20] of the pairwise comparisons were calculated using *p*-values from the Linear Models for Microarray Data (LIMMA) package [designed for analysis of Affymetrix array data; <http://bioinf.wehi.edu.au/affylmGUI/>]. Differentially-regulated genes were selected using a 2-fold cut-off and a false discovery rate (FDR) of < 0.05.

Table 1: Macaque liver tissues used for transcriptome analyses

Liver samples	AST/ALT ^d	Glucose ^e	Triglycerides ^f	Virus titers ^g
Uninfected controls				
Rh Ctrl7	38/65	74	65	<10 ²
Rh Ctrl8	24/53	79	61	<10 ²
Rh Ctrl9	47/71	87	77	<10 ²
LCMV-WE1-Pre-viremic				
Rh 1WE (day 1) ^a	44/61	53	27	<10 ²
Rh 2WE (day 1)	63/54	58	30	<10 ²
Rh 3WE (day 2)	59/37	60	18	<10 ²
Rh 5WE (day 3)	27/71	62	37	<10 ²
LCMV-WE2-Viremic^b				
Rh 6WE (day 4)	96/137	71	33	<10 ²
Rh 7WE (day 4)	24/38	67	18	<10 ²
Rh 8WE (day 6)	26/56	54	32	1.4 × 10⁴
Rh 9WE (day 6)	125/82	70	31	5.0 × 10³
Rh 10WE (day 7)	190/325	66	30	1.5 × 10⁶
LCMV-WE3 Terminal				
Rh 11WE (day 11)	642/587	30	235	5.5 × 10⁵
Rh 12WE (day 12)	1504/681	40	1402	1.0 × 10⁶
LCMV-not diseased^c				
Rh 3ARM (day 3)	45/74	70	75	<10 ²
Rh 5ARM (day 5)	18/40	78	62	<10 ²
Rh 1ARM/WE-1	52/24	65	79	<10 ²
Rh 1ARM/WE-5	Not done	Not done	ND	<10 ²
Rh 2ARM/WE-6	56/34	63	80	<10 ²
Rh 2ARM/WE-2	Not done	Not done	ND	<10 ²

^a (day 1) is the day of necropsy after infection. Each sample is from the liver of a different rhesus macaque.

^b Pre-viremic means liver was harvested day 1, day 2 or day 3 after infection; Viremic means liver was harvested day 4 to day 7, and Terminal means liver was harvested day 11 or 12 during the terminal stage of disease. ND = Not done.

^c Animals that were LCMV-infected but not diseased were either infected with LCMV-ARM and sacrificed day 3 or day 5 after infection, or they were infected with LCMV-ARM and LCMV-WE (10³ pfu each virus) and sacrificed on day 30 after infection.

^d AST/ALT means the ratio of plasma levels of aspartate aminotransferase to alanine aminotransferase in international units per liter (IU/L). Normal ranges are AST/ALT = 18–82/22–87 IU/L. **Values outside the reference range are bolded.**

^e Serum glucose levels are in mg/dL with a reference range of 55–80 mg/dL (Animal Diagnostics Laboratory, Baltimore, MD.)

^f Serum triglyceride levels have a reference range of 50–82 mg/dL. Other parameters derived from blood analysis include red blood cells, hemoglobin levels and cholesterol levels which were all within normal ranges for all samples.

^g Virus was titered on Vero cell monolayers as described [2], and given as plaque forming units per gram of liver tissue. Samples in the viremic group that had undetectable titers (<10² pfu/gm) were positive for virus nucleic acid by RT-PCR and virus co-cultivation as described previously [3].

Pairwise comparisons were performed to identify 3,125 differentially-regulated genes (n -fold of at least 2 and FDR $P < 0.05$) between infected samples ($n = 9$) and uninfected control samples ($n = 3$) or between infected samples ($n = 9$) and samples from animals that were infected but not diseased ($n = 6$). Six separate pair-wise comparisons were also performed to compare the three groups of infected samples: pre-viremic ($n = 4$), viremic ($n = 5$) and post-viremic ($n = 2$) to the two groups; controls ($n = 3$) or non-diseased samples ($n = 6$). These six pairwise comparisons resulted in identification of 4,482 differentially-regulated genes (n -fold of at least 2 and FDR $P < 0.05$) amongst any one pairwise comparison. KEGG software was used to identify key functions and metabolic pathways differentially-regulated between uninfected and LCMV-infected liver samples. The raw gene expression data and the related experimental information from this study can be found at <http://www.ncbi.nlm.nih.gov/geo>, platform number GSE12254.

Quantitative real-time reverse-transcriptase PCR

Genes for microarray analysis were validated by quantitative real-time PCR as we described [3] to determine the extent to which gene expression was up- or down-regulated as a result of infection. Monkey and human-specific primers were used to validate expression levels for selected host genes. Although we ordinarily used primers derived from GAPDH, Actin, or 18S RNA for baseline expression, we found that the expression of these three genes varied in infected versus uninfected liver, so we searched for genes that were expressed in liver but at a steady level for all infection time-points. We found that expression of the protein kinase C substrate 80 K-H gene (PRKCSH, or hepatocystin) had a less than 0.04-fold standard deviation over all the samples used for this liver transcriptome analysis. Baseline expression of PRKCSH could be determined with primers for the *Macaca mulatta* sequence (first, CGT-TAGGCAGCCGTGC and second, GGCCGTGGAGGT-CAAGAGGC).

Results

Identification of differentially-regulated genes in LCMV-infected rhesus macaque liver

Liver RNA from LCMV-infected macaques was used for transcriptome profiling with the goal of identifying gene expression that differentiated infected from uninfected liver and that differentiated virulently-infected (diseased) from mildly-infected (non-diseased) liver. As described previously [3], we characterized two major stages of infection, pre-viremic (day 1 to day 3) and viremic (day 4 to day 7) and compared them to uninfected samples or samples from monkeys infected with LCMV that did not have disease. Three types of results are presented here: 1) genes with the greatest differential expression after pair-wise comparisons of infected and uninfected liver samples, 2)

gene expression associated with major pathways in the liver such as gluconeogenic, glycolytic, lipogenic and coagulation pathways, and 3) genes with the greatest differential expression after pair-wise comparison of virulent and mildly-infected samples. In the latter category we especially noted gene expression changes that occurred before the viremic stage of disease, and could potentially serve as biomarkers that discriminate between virulent and benign infections.

The liver is highly vascularized and sensitive to changes in blood composition. Since liver tissue contains a mixture of cells including hematopoietic cells found in the circulation, we examined the overlap of gene expression in liver and in blood for the virulently-infected monkeys (Figure 1; Additional Files 2, 3, 4, 5, 6, 7). Among the subset of genes displaying comparably elevated expression in blood and liver were interferon-induced transmembrane protein 1 (IFITM1), tumor necrosis factor (ligand) superfamily, member 10 (TNFSF10, or TRAIL), ubiquitin-conjugating enzyme E2L6 (UBE2L6), BCL2-related protein A1 (BCL2A1), profilin 1 (PFN1), some coagulation pathway genes, and some JAK-STAT/toll-like receptor signaling pathway genes.

Additional File 8 lists differentially-regulated genes in macaque liver ($n = 4482$), which include 1234 significantly ($p < 0.05$) up-regulated genes and 123 down-regulated genes (the remainder were up-regulated at some time points and down-regulated at others). Differential gene expression is presented in two ways: 1) LCMV-WE-infected (diseased) liver compared to uninfected liver, and 2) LCMV-WE-infected (diseased) liver compared to LCMV-infected non-diseased liver. Differentially-expressed genes were defined as those that had at least a 2-fold alteration in LCMV-infected livers compared to the uninfected controls.

Genes with the highest differential expression between LCMV-infected and uninfected livers (53 are listed in Figure 2) encode several heat shock proteins, ribosomal proteins, energy-generating enzymes, and proteins known to be involved in anti-microbial defense [e. g. Shwachman-Bodian-Diamond syndrome (SBDS), complement (C1S), and interferon-inducible genes (IFITM1, IFITM2)]. Prominent genes associated with energy-generation include HPD and GSL2 encoding enzymes that break down amino acids. Also notable are two gene products with anti-inflammatory functions: 1) HSP60 (encoded by HSPD1) that down-regulates T-bet, NF-kappaB, and NFAT and up-regulates GATA-3, leading to decreased secretion of TNF-alpha [21], and 2) LTB4DH that encodes a dehydrogenase which inactivates several eicosanoids such as leukotriene B4 [22].

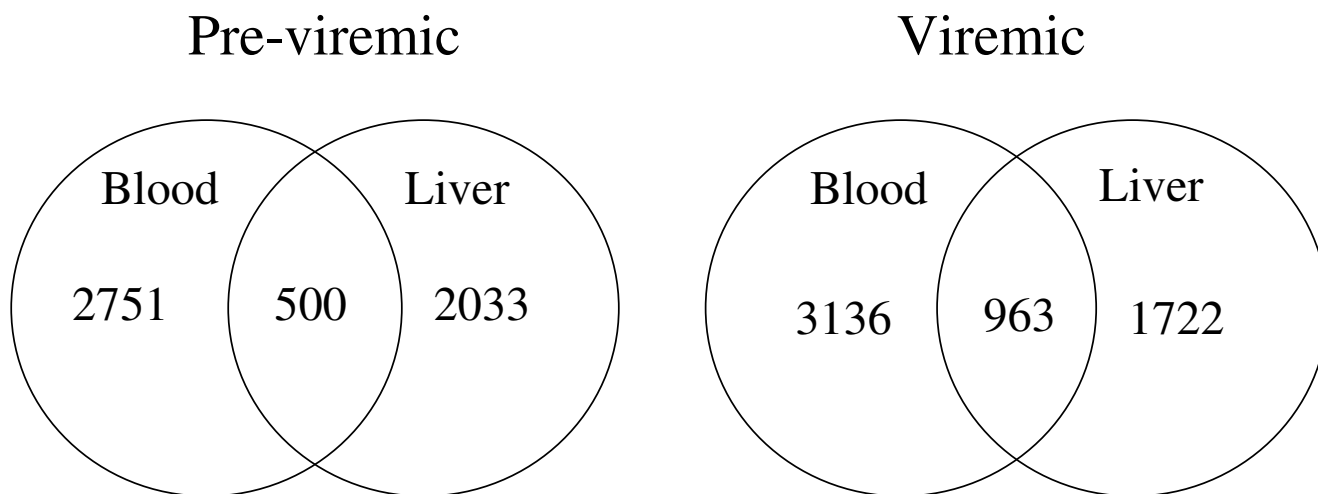


Figure 1

Venn diagram of gene expression in virulently-infected liver compared to virulently-infected blood. Genes identified as being differentially-regulated in both blood and liver can be found in Additional Files 2, 3, 4, 5, 6, 7.

Metabolic pathway gene expression in LCMV-infected rhesus macaques

To understand the metabolic responses to LCMV infection, changes in gene expression in macaque livers were examined in relation to their associated metabolic pathways using the KEGG pathways analysis. Additional File 9 shows 32 of the most significantly affected pathways in LCMV-infected macaque liver with respect to uninfected liver for both pre-viremic and viremic stages of infection. Four of these pathways are described here and shown in (Tables 3,4,5 and 6).

The largest number of genes with altered expression contribute to lipid and glucose metabolism, and hence to the generation of energy for the organism. Key regulators of cholesterol synthesis, the mRNA for 3-hydroxy-3-methylglutaryl-CoA synthase 1 and 2 (HMGCS1, HMGCS2), were up-regulated during LCMV infection. Nevertheless, the liver-specific transporter gene APOAV was down-regulated (Additional File 8) suggesting that transport of triglycerides and cholesterol (via HDL/LDL) would be decreased in LCMV-infected liver. This is corroborated by lowered serum triglycerides during the pre-viremic and viremic stages (averaging 28 mg/dL when the norm is 50–82 mg/dL; Table 1). However blood chemistry showed that cholesterol levels were not affected by LCMV infection, so several transcriptional changes were not obviously related to effects on downstream metabolic products. A possibility is that increased energy-generating activity was defeated by poor transport, or that post-transcriptional regulation over-shadowed the rise in transcript levels.

Effect of LCMV infection on glucose metabolism

Monkeys undergoing virulent infection had normal blood glucose levels (except at the terminal stage; Table 1) up-regulated numerous genes involved in carbohydrate metabolism (Additional File 9, tables 3, 4, 5 and 6). During the pre-viremic stage of infection several genes were up-regulated for enzymes involved in gluconeogenesis, i.e. the breakdown of fatty acids, amino acids and pyruvate to make glucose (Figure 2, Tables 3, 4, 5, and 6). For example, glucose-6-phosphatase (G6Pase) was up-regulated in both pre-viremic and viremic stages, and fructose 1,6-bis phosphatase (FBP1) was four-fold up-regulated only at the pre-viremic stage (Table 3). The increased gluconeogenesis during LCMV infection resembled that seen during starvation [23] or cachexia [24], however the infection also up-regulated mRNA encoding glycolytic enzymes (PGK1, GCK, GAPDH and ALDOB; Table 3) that are usually down-regulated during a fast [25]. Simultaneous up-regulation of glycolytic and gluconeogenic enzymes in infected animals could mean that sugar metabolism is in a futile cycle and that the majority of energy production during LCMV infection comes from beta-oxidation of fatty acids (Tables 3, 4 and 5, Figure 3).

LCMV-infected liver up-regulated expression of glycogen synthase 2 (GYS2) in both pre-viremic and viremic stages of infection (Table 3) suggesting the possibility that glucose storage was enhanced. Glycogen in the LCMV-infected liver could also be derived from hydrolysis of triglycerides in fat cells since several lipases and beta-oxidation enzymes were up-regulated (Table 7) and triglycerides in blood were unusually low (Table 1).

WE1-uninf	WE2-uninf	GenBank Acc. #	Gene symbol	Description
15.56	23.10	NM_001134	AFP	Alpha-fetoprotein
39.95	21.71	D16931	ALB	Albumin
37.01	22.78	NM_005589	ALDH6A1	Aldehyde dehydrogenase 6 family, member A1
-4.79	-5.46	AF202889	APOA5	Apolipoprotein A-V
85.04	224.41	AW188940	B2M	Beta-2-microglobulin
53.45	50.21	BC007010	C1S	Complement component 1, s subcomponent
17.27	18.64	AF367476	CCNL1	Cyclin L1
41.93	31.56	W80357	CPS1	Carbamoyl-phosphate synthetase 1, mitochondrial
16.11	10.93	NM_000099	CST3	Cystatin C (amyloid angiopathy and cerebral hemorrhage)
-4.53	-5.13	AA889653	DES	Desmin
15.35	12.13	AW083133	EIF3S12	Eukaryotic translation initiation factor 3, subunit 12
25.63	20.82	NM_000120	EPHX1	Epoxide hydrolase 1, microsomal (xenobiotic)
49.87	45.57	BG545288	FGB	Fibrinogen beta chain
17.88	13.09	AK022172	FMO5	Flavin containing monooxygenase 5
22.47	20.68	AI743037	FMR1	Fragile X mental retardation 1
30.06	11.31	AF110329	GLS2	Glutaminase 2 (liver, mitochondrial)
17.39	5.82	AB046780	GPAM	Glycerol-3-phosphate acyltransferase, mitochondrial
15.67	14.22	U56250	GSTO1	Glutathione S-transferase omega 1
15.56	19.97	NM_015987	HEBP1	Heme binding protein 1
20.11	17.88	N32864	HINT1	Histidine triad nucleotide binding protein 1
110.66	74.54	NM_002150	HPD	4-hydroxyphenylpyruvate dioxygenase
15.24	28.05	BG612458	HSP90AB1	Heat shock protein 90kDa alpha class B member 1
82.14	114.56	AB034951	HSPA8	Heat shock 70kDa protein 8
106.89	106.89	BE256479	HSPD1	Heat shock 60kDa protein 1 (chaperonin)
19.43	198.09	NM_006417	IFI44	Interferon-induced protein 44
11.88	144.01	NM_001548	IFIT1	Interferon-induced protein with tetratricopeptide repeats 1
100.43	68.59	NM_000597	IGFBP2	Insulin-like growth factor binding protein 2, 36kDa
23.92	8.46	BE566894	LTB4DH	Leukotriene B4 12-hydroxydehydrogenase
24.76	68.12	NM_002356	MARCKS	Myristoylated alanine-rich protein kinase C substrate
11.71	14.72	BE880828	MCFD2	Multiple coagulation factor deficiency 2
12.21	7.78	NM_002415	MIF	Macrophage migration inhibitory factor
14.32	15.45	NM_014078	MRPL13	Mitochondrial ribosomal protein L13
18.00	13.27	BF215996	MYO1B	Myosin IB
-4.89	-5.03	AW172570	NMNAT3	Nicotinamide nucleotide adenyltransferase 3
14.42	8.06	NM_018441	PECR	Peroxisomal trans-2-enoyl-CoA reductase
25.28	9.99	N26005	PPP1R3C	Protein phosphatase 1, regulatory (inhibitor) subunit 3C
18.13	22.16	NM_002787	PSMA2	Proteasome (prosome, macropain) subunit, alpha type, 2
10.63	10.93	BC003005	PTGES3	prostaglandin E synthase 3 (cytosolic)
26.91	20.97	NM_007104	RPL10A	Ribosomal protein L10a
39.95	35.26	NM_000991	RPL28	Ribosomal protein L28
49.87	39.67	NM_001008	RPS4Y1	Ribosomal protein S4, Y-linked 1
22.63	13.93	NM_001012	RPS8	Ribosomal protein S8
20.39	18.00	BC010183	SBDS	Shwachman-Bodian-Diamond syndrome
20.11	24.59	AF328864	SELS	Selenoprotein S
18.51	17.39	NM_016275	SELT	Selenoprotein T
30.70	19.29	BC022309	SERPINC1	Serpin peptidase inhibitor, clade C (antithrombin), member 1
-3.76	-3.63	AI734001	SEZ6L2	Seizure related 6 homolog (mouse)-like 2
23.92	20.25	NM_007108	TCEB2	Transcription elongation factor B (SIII), polypeptide 2 (elongin B)
32.90	33.82	BG104571	TM9SF3	Transmembrane 9 superfamily member 3
-4.79	-4.47	BF508371	TNFRSF1A	Tumor necrosis factor receptor superfamily, member 1A
-3.25	-3.46	AI885873	TNPO2	Transportin 2 (importin 3, karyopherin beta 2b)
12.30	15.56	NM_016146	TRAPPC4	Trafficking protein particle complex 4
23.26	15.67	BF591611	ZADH2	Zinc binding alcohol dehydrogenase, domain containing 2

Figure 2

The most differentially regulated genes in liver after infection of macaques with LCMV-WE. WE1 refers to pre-viremic samples (day 1–3) and WE2 refers to viremic samples (day 4–7). Both WE1 and WE2 are compared with uninfected samples (WE1 vs uninf) using the sample numbers shown in Table 1. Nfold refers to fold up- or down- regulation of each gene expressed in a virulently-infected liver in relation to expression in an uninfected liver. The *p* values for these genes are all less than 0.05. This is a subset of Additional File 8.

Table 2: Validation of microarray results by real-time PCR^a

Gene Symbol (GenBank ID)	WE vs uninfected		WE vs no disease	
	Pre-viremic	Viremic Stage	Pre-viremic	Viremic stage
ACACB (AI057637)	-3.58/-5	-4.38/-5	-2.16/-2	-2.64/-2
ACVRI (NM_001105)	10.85/15	7.62/12	7.46/11	5.24/10
ARG2 (U75667)	1.02/2	1.04/1	-12.13/-9	-11.88/-5
CIS (BC007010)	53.45/25	50.21/22	6.73/4.2	6.32/3.7
CYP26A1 (NM_000783)	9.85/13	2.11/1.2	10.56/14	2.27/2.9
FGB (BG545288)	49.87/32	45.57/29	3.36/2.4	3.07/2.0
FST (NM_013409)	-1.67/1.8	1.09/1.3	-16.8/-19	-9.25/-16
GOT1 (BC000498)	3.92/3.0	2.0/1.6	-6.54/-13	-12.82/-18
HBA1/HBA2 (V00489)	2.79/2	1.2/3.1	-12.91/-11	-30.06/-25
HSPB1 (NM_001540)	28.64/22	36.76/43	9.38/7.2	12.04/9.7
IFI44 (NM_006417)	19.43/14	198.1/166	3.71/4.0	37.79/21
INHBA (M13436)	1.71/1.9	-1.13/1.3	11.16/15	5.78/8.7
PPARGCIA (NM_013261)	-1.22/1.0	3.01/1.9	-21.41/-9	-5.82/-3
PPPIR3C (N26005)	25.28/32	9.99/12	156.5/87	61.82/45
SDS (NM_006843)	-1.97/1.7	-2.07/1.2	-32.22/-28	-33.82/-29
TGFBRI (AA604375)	4.29/4.8	3.1/2.5	1.33/1.6	-1.04/1.5
PRKCSH (AI815793)	<2/- ^b	<2/-	<2/-	<2/-

^a The numbers represent N-folds from Affychip data (most in Figure 4, all in Additional file 8) and N-fold by real-time PCR ($N_{\text{aff}}/N_{\text{PCR}}$). N-folds by real-time PCR make use of liver RNA from uninfected, pre-viremic LCMV-WE-infected, viremic LCMV-WE-infected, and infected-but-not-diseased monkeys.

^b PRKCSH is the gene symbol for protein kinase C substrate 80 K-H to which the PCR data are normalized. It is expressed in liver and has a less than 0.04 standard deviation over all samples used for this liver transcriptome analysis. (-) means it is the reference value.

Effect of virulent infection on lipogenic pathway (fatty acid metabolism)

Mobilization of fat (lipolysis) is favored under conditions of increased energy need, such as exercise, fasting, hypothermia [23], or cachexia associated with cancer and AIDS [24]. When triglycerides in adipose tissue are broken down by lipase, then the fatty acids and glycerol are released to the bloodstream. Genes involved in fatty acid transport, albumin (ALB), myosin (MYO1B), and the thrombospondin receptor (CD36), were up-regulated during infection (Figure 2). In the mitochondria, fatty acids undergo β -oxidation into two-carbon acetyl groups attached to CoA [23]. During LCMV infection, genes for fatty acid metabolic enzymes were strongly up-regulated: thiolase (ACAT2), hydroxyacyl-CoA dehydrogenase (HADHA), enoyl-CoA hydratase (ECHS1), and stearoyl-CoA desaturase (SCD) (Tables 4 and 5). The only down-regulated gene in the fatty acid metabolic group was acetyl-CoA carboxylase (ACACB), a major regulator of fatty acid synthesis [23]. This would predict that free fatty acids arose more frequently by β -oxidation than by fatty acid synthesis via ACACB (Figure 3; Table 5). In support of that prediction, several cytochrome P450 isoforms (CYP2A, -3A, -2B) were up-regulated that are known to break down steroids and fats during the course of LCMV-infection (Table 4) [26,27]. The down-regulation of ACACB and the up-regulation of CYP26A1 have been validated by quantitative PCR (Table 2).

Free fatty acids (FFA) stimulate key gluconeogenic enzymes [28,29]. For example the enzyme encoded by PCK is up-regulated by FFA and peroxisome proliferator-activated receptor (encoded by PPAR) in hepatoma cells [30]. However both PPAR and PCK are more up-regulated in the mild (non-diseased) infections than in the virulent infections (Additional File 8), contributing to the picture that these changes in intermediary metabolism are a protective host response that is stronger in the non-diseased cases.

The aldehyde dehydrogenases clear the blood of toxic aldehydes by generating acetate, which can be converted to acetyl CoA. Several aldehyde dehydrogenases are up-regulated during infection, especially ALDH6A1, a liver mitochondrial enzyme that catalyzes the breakdown of malonate to propionyl – and acetyl-CoA, (up-regulated 37- and 22-fold in pre-viremic and viremic stages, Additional File 8). Levels of acetyl-CoA are central to the balance between carbohydrate and fat metabolism, so in the LCMV-infected liver, it appears that the product of ALDH6A1 could drive acetyl CoA into the cholesterol or citric acid cycles to generate energy but not into fatty acid synthesis due to down-regulated ACACB acting as a bottleneck (Figure 3).

Ketone-body formation (ketogenesis) occurs during high rates of fatty acid oxidation through generation of large

Table 3: Expression of genes involved with TCA cycle and glucose metabolism in LCMV-WE infected macaque liver

GenBank Accession no.	Symbol	Gene description	Pre-viremic		Viremic	
			Fold change ^a	p ^b	Fold change ^a	p ^b
Citric acid cycle						
NM_001096	ACLY	ATP citrate lyase	2.14	0.0809	3.58	0.0040
AI363836	FH	Fumarate hydratase	5.54	0.0045	3.05	0.0356
AI826060	IDH3A	Isocitrate dehydrogenase 3 (NAD+) alpha	1.51	0.4150	2.29	0.0440
AF023266	IDH3B	Isocitrate dehydrogenase 3 (NAD+) beta	2.41	0.0219	2.19	0.0290
NM_004135	IDH3G	Isocitrate dehydrogenase 3 (NAD+) gamma	2.14	0.0262	2.29	0.0122
NM_005917	MDH1	Malate dehydrogenase 1, NAD (soluble)	9.51	0.0056	9.25	0.0039
BC001917	MDH2	Malate dehydrogenase 2, NAD (mitochondrial)	2.77	0.0739	3.48	0.0221
NM_004168	SDHA	Succinate dehydrogenase complex, A, flavoprotein	4.17	0.0108	4.03	0.0087
NM_003000	SDHB	Succinate dehydrogenase complex, B, iron sulfur	5.98	0.0120	6.68	0.0056
AF080579	SDHC	Succinate dehydrogenase complex, C, membrane protein	14.12	0.0003	11.63	0.0003
AL050226	SUCLG2	Succinate-CoA ligase, GDP-forming, beta	4.23	0.0108	2.83	0.0442
Glycolysis						
NM_000034	ALDOA	Aldolase A, fructose-bisphosphate	2.03	0.0136	3.05	0.0004
AK026411	ALDOB	Aldolase B, fructose-bisphosphate	14.83	0.0002	6.02	0.0027
AK026525	GAPDH	Glyceraldehyde-3-phosphate dehydrogenase	6.59	0.0034	4.89	0.0066
M69051	GCK	Glucokinase (hexokinase 4)	3.92	0.0010	1.01	0.9980
S81916	PGKI	Phosphoglycerate kinase I	23.59	0.0009	23.43	0.0005
NM_002633	PGMI	Phosphoglucosmutase I	6.28	0.0038	5.03	0.0058
Glycogen metabolism						
S70004	GYS2	Glycogen synthase 2, liver	9.65	0.0004	5.54	0.0020
NM_002863	PYGL	Phosphorylase glycogen, liver	4.96	0.0120	3.14	0.0485
NM_002633	PGMI	Phosphoglucosmutase I	6.28	0.0038	5.03	0.0058
Gluconeogenesis						
D26054	FBPI	Fructose-1,6-bisphosphatase I	4.08	0.0061	1.59	0.1559
BC020700	G6PC	Glucose-6-phosphatase, catalytic	12.13	0.0034	5.17	0.0265
Others						
NM_000284	PDHAI	Pyruvate dehydrogenase (lipoamide) alpha I	3.48	0.0769	4.53	0.0249

^a Mean fold changes were calculated using a division of raw expression values between experimental sample and uninfected control.

^b All samples were analyzed separately. Changes in gene expression with a cutoff of 2.0-fold increased or decreased expression was used and, the *p*-value was calculated by Student's *t*-test. Data are displayed only where the, $p \leq 0.05$. This *p*-value was used as a measure of the magnitude of the difference between groups and to determine significance of the modulation. The modulated genes in the pathway are listed in alphabetic order.

amounts of acetyl-CoA in liver [23]. Ketone bodies, consisting of acetoacetate, acetone and β -hydroxybutyrate, serve as an alternative source of energy at low glucose levels. Ketogenesis was likely higher in LCMV-WE infected rhesus macaques than in the mildly-infected macaques since fatty acid synthesis via acetyl-coenzyme A carboxylase beta (ACACB) was more down-regulated in the virulent infection (Figure 2; Table 5). Genes involved with mitochondrial fatty acid oxidation, cholesterol synthesis and ketogenesis (Table 3; Additional File 8), for example, the hydroxy-3-methylglutaryl-CoA synthases (HMGCS1 and 2) were induced by LCMV infection. They condense acetyl-CoA and acetoacetyl-CoA to form HMG-CoA, an intermediate in the pathway for synthesis of ketone bodies, a likely source of energy during LCMV-WE infection (Table 5).

The expression of several lipase genes is strongly up-regulated during LCMV-WE infection, suggesting a relation-

ship between up-regulation of beta-oxidation enzymes and lipolysis in the infected livers (Table 6). In general, we found that genes involved in degradation of lipids and fatty acids were up-regulated while a key gene involved in fatty acid synthesis was down-regulated.

Effect of LCMV infection on gene expression related to coagulation and complement cascade pathways

The complement and coagulation cascades interact and modulate each other; both are affected during LCMV infection. Although we do not present coagulation data here, platelet defects and a mild thrombocytopenia were documented during severe Lassa fever [11,12], and macaques with virulent LCMV disease showed a marked thrombocytopenia [4]. The transcriptome data (Table 7) show over-expression of complement component 1 (C1S), an anti-thrombin peptide (SERPINC1), and fibrinogen beta chain (FGB) during virulent infection. Over-expression of C1S and FGB have been validated by quan-

Table 4: Expression of genes involved with fatty acid oxidation in LCMV-WE infected macaque liver

GenBank Accession no.	Symbol	Gene description	Pre-viremic		Viremic	
			Fold change ^a	p ^b	Fold change ^a	p ^b
Mitochondrial {beta}-oxidation						
<u>BE897866</u>	ACADSB	Acyl-CoA dehydrogenase, short/branched chain	1.77	0.6240	-2.93	0.0190
<u>NM_004453</u>	ETFDH	Electron-transferring flavoprotein dehydrogenase	3.10	0.0019	1.49	0.1412
<u>NM_004092</u>	ECHS1	Enoyl CoA hydratase, short chain, I, mitochondrial	7.06	0.0110	4.79	0.0260
<u>U04627</u>	HADHA	Hydroxyacyl-CoA dehydrogenase/3-ketoacyl-CoA thiolase/enoyl-CoA hydratase (trifunctional protein), alpha	3.43	0.0180	3.75	0.0086
Microsomal oxidation						
<u>AF182275</u>	CYP2A6	Cytochrome P450, family 2, subfamily A, polypeptide 6	17.88	0.0008	7.21	0.0009
<u>X06399</u>	CYP2B6	Cytochrome P450, family 2, subfamily B, polypeptide 6	3.16	0.0079	-1.02	0.9980
<u>NM_030878</u>	CYP2C8	Cytochrome P450, family 2, subfamily C, polypeptide 8	3.51	0.0020	2.35	0.0147
<u>NM_000106</u>	CYP2D6	Cytochrome P450, family 2, subfamily D, polypeptide 6	7.78	0.0065	2.66	0.1530
<u>AF182276</u>	CYP2E1	Cytochrome P450, family 2, subfamily E, polypeptide 1	4.06	0.0150	2.87	0.0459
<u>NM_000777</u>	CYP3A5	Cytochrome P450, family 3, subfamily A, polypeptide 5	7.57	0.0005	6.36	0.0005
<u>AF315325</u>	CYP3A7	Cytochrome P450, family 3, subfamily A, polypeptide 7	11.39	0.0006	5.28	0.0050
Ketogenesis						
<u>BC000408</u>	ACAT2	Acetyl-CoA acetyltransferase 2 (acetoacetyl CoA thiolase)	6.10	0.0065	4.78	0.0107
<u>BG035985</u>	HMGCS1	3-hydroxy-3-methylglutaryl-CoA synthase 1 (soluble)	3.10	0.0198	4.69	0.0467
<u>NM_005518</u>	HMGCS2	3-hydroxy-3-methylglutaryl-CoA synthase 2 (mitochondrial)	5.16	0.0135	2.91	0.0748
<u>BF224073</u>	TCPI	T-complex I	3.43	0.0332	2.50	0.0977
Peroxisomal oxidation						
<u>NM_003500</u>	ACOX2	Acyl-CoA oxidase 2, branched chain	6.63	0.0019	3.12	0.0267
<u>NM_004092</u>	ECHS1	Enoyl CoA hydratase, short chain, I, mitochondrial	7.06	0.0110	4.79	0.0260
<u>U04627</u>	HADHA	Hydroxyacyl-CoA dehydrogenase/3-ketoacyl-CoA thiolase/enoyl-CoA hydratase (trifunctional protein), alpha	3.43	0.0180	3.75	0.0086
<u>NM_005525</u>	HSD11B1	Hydroxysteroid (11-beta) dehydrogenase 1	2.80	0.0325	2.20	0.0874
<u>AL031228</u>	HSD17B8	Hydroxysteroid (17-beta) dehydrogenase 8	2.80	0.0258	2.20	0.0620
Other peroxisomal proteins						
<u>AU147084</u>	CAT	Catalase	8.63	0.0028	5.78	0.0069
Others						
<u>NM_000072</u>	CD36	CD36 molecule (thrombospondin receptor)	48.18	0.0001	16.11	0.0008

^a Mean fold changes were calculated using a division of raw expression values between experimental sample and uninfected control.

^b All samples were analyzed separately. Changes in gene expression with a cutoff of 2.0-fold increased or decreased expression was used and, the p-value was calculated by Student's t-test. Data are displayed only where the, $p \leq 0.05$. This p-value was used as a measure of the magnitude of the difference between groups and to determine significance of the modulation. The modulated genes in the pathway are listed in alphabetic order.

titative PCR (Table 2). These 3 genes alone would predict problems with coagulation: high C1S should drive the classical pathway of the complement cascade towards more phagocyte recruitment and cell lysis, high serpin should inhibit the intrinsic pathway of coagulation, and high fibrinogen should overwhelm the fibrinolysis that occurs during clot formation [31]. Additionally, the multiple coagulation factor deficiency 2 (MCFD2) gene is up-regulated 11-fold in pre-viremic and 15-fold during the

viremic phase and is important in the secretion of coagulation factors V and VIII [32]. In addition to these highly over-expressed genes, there is modest up-regulation of coagulation factors II, V, and X which should promote coagulation and has been associated with consumptive coagulopathy and disseminated intravascular coagulation (DIC); although DIC has not been associated with Lassa [13] as it has with other hemorrhagic virus infections like Ebola fever [33]. We also observed down-modulation of a

Table 5: Expression of genes involved with fatty acid, cholesterol and amino acid metabolism in LCMV-WE infected macaque liver.

GenBank Accession no.	Symbol	Gene description	Pre-viremic		Viremic	
			Fold change ^a	p ^b	Fold change ^a	p ^b
Fatty acid metabolism						
AI057637	ACACB	Acetyl-Coenzyme A carboxylase beta	-3.58	0.0103	-4.38	0.0027
NM_001096	ACLY	ATP citrate lyase	2.14	0.0809	3.58	0.0040
AB032261	SCD	Stearoyl-CoA desaturase (delta-9-desaturase)	2.06	0.0448	4.69	0.0420
Cholesterol metabolism						
BC000408	ACAT2	Acetyl-CoA acetyltransferase 2 (acetoacetyl CoA thiolase)	6.10	0.0065	4.78	0.0107
U40053	CYP51A1	Cytochrome P450, family 51, subfamily A, polypeptide 1	3.68	0.0622	12.04	0.0010
NM_014762	DHCR24	24-dehydrocholesterol reductase	5.28	0.0006	3.66	0.0021
BC003573	FDFT1	Farnesyl-diphosphate farnesyltransferase 1	5.70	0.0195	6.02	0.0011
NM_002004	FDPS	Farnesyl diphosphate synthase	3.63	0.0218	5.03	0.0040
BG035985	HMGCS1	3-hydroxy-3-methylglutaryl-CoA synthase 1 (soluble)	3.01	0.0198	4.69	0.0467
NM_005336	HDLBP	High density lipoprotein binding protein (vigilin)	4.96	0.0007	3.14	0.0042
AF478446	NR1H4	Nuclear receptor subfamily 1, group H, member 4	11.31	0.0003	10.48	0.0002
BF530535	SPCS2	Signal peptidase complex subunit 2	10.34	0.0009	8.28	0.0011
Amino acid metabolism						
NM_000687	AHCY	S-adenosylhomocysteine hydrolase	7.46	0.0005	4.03	0.0048
AF110329	GLS2	Glutaminase 2	30.06	0.0039	11.31	0.0201
NM_002080	GOT2	Glutamic-oxaloacetic transaminase 2, mitochondrial (aspartate aminotransferase 2)	4.53	0.0036	3.12	0.0140
NM_000531	OTC	Ornithine carbamoyltransferase	4.47	0.0182	2.58	0.1090

^a Mean fold changes were calculated using a division of raw expression values between experimental sample and uninfected control.

^b All samples were analyzed separately. Changes in gene expression with a cutoff of 2.0-fold increased or decreased expression was used and, the *p*-value was calculated by Student's *t*-test. Data are displayed only where the, $p \leq 0.05$. This *p*-value was used as a measure of the magnitude of the difference between groups and to determine significance of the modulation. The modulated genes in the pathway are listed in alphabetic order.

Table 6: Expression of various lipase genes and genes involved with lipase activity in LCMV-WE infected macaque liver.

GenBank Accession no.	Symbol	Gene description	Pre-viremic		Viremic	
			Fold change ^a	p ^b	Fold change ^a	p ^b
NM_000483	APOC2	Apolipoprotein C-II	5.50	0.0003	3.53	0.0019
AK023348	GRN	Granulin	1.84	0.0448	3.20	0.0007
NM_006854	KDEL2	KDEL (Lys-Asp-Glu-Leu) endoplasmic reticulum protein retention receptor 2	7.46	0.0056	7.36	0.0038
NM_000235	LIPA	Lipase A, lysosomal acid, cholesterol esterase	2.17	0.1350	4.08	0.0059
NM_006033	LIPG	Lipase, endothelial	2.20	0.4480	2.55	0.2950
AF077198	LYPLA1	Lysophospholipase I	7.16	0.0006	3.81	0.0008
BC006230	MGLL	Monoglyceride lipase	3.27	0.0090	2.73	0.0162
D83485	PDIA3	Protein disulfide isomerase family A, member 3	6.36	0.0028	7.46	0.0010
AL542253	PLA1A	Phospholipase A1 member A	1.37	0.0035	3.01	0.0124
NM_000300	PLA2G2A	Phospholipase A2, group IIA (platelets)	2.19	0.7700	26.17	0.0218
AF145020	PLAA	Phospholipase A2-activating protein	3.51	0.0026	2.69	0.0077
NM_002937	RNASE4	Ribonuclease, RNase A family, 4	45.25	0.00008	25.81	0.0001

^a Mean fold changes were calculated using a division of raw expression values between experimental sample and uninfected control.

^b All samples were analyzed separately. Changes in gene expression with a cutoff of 2.0-fold increased or decreased expression was used and, the *p*-value was calculated by Student's *t*-test. Data are displayed only where the, $p \leq 0.05$. This *p*-value was used as a measure of the magnitude of the difference between groups and to determine significance of the modulation. The modulated genes in the pathway are listed in alphabetic order.

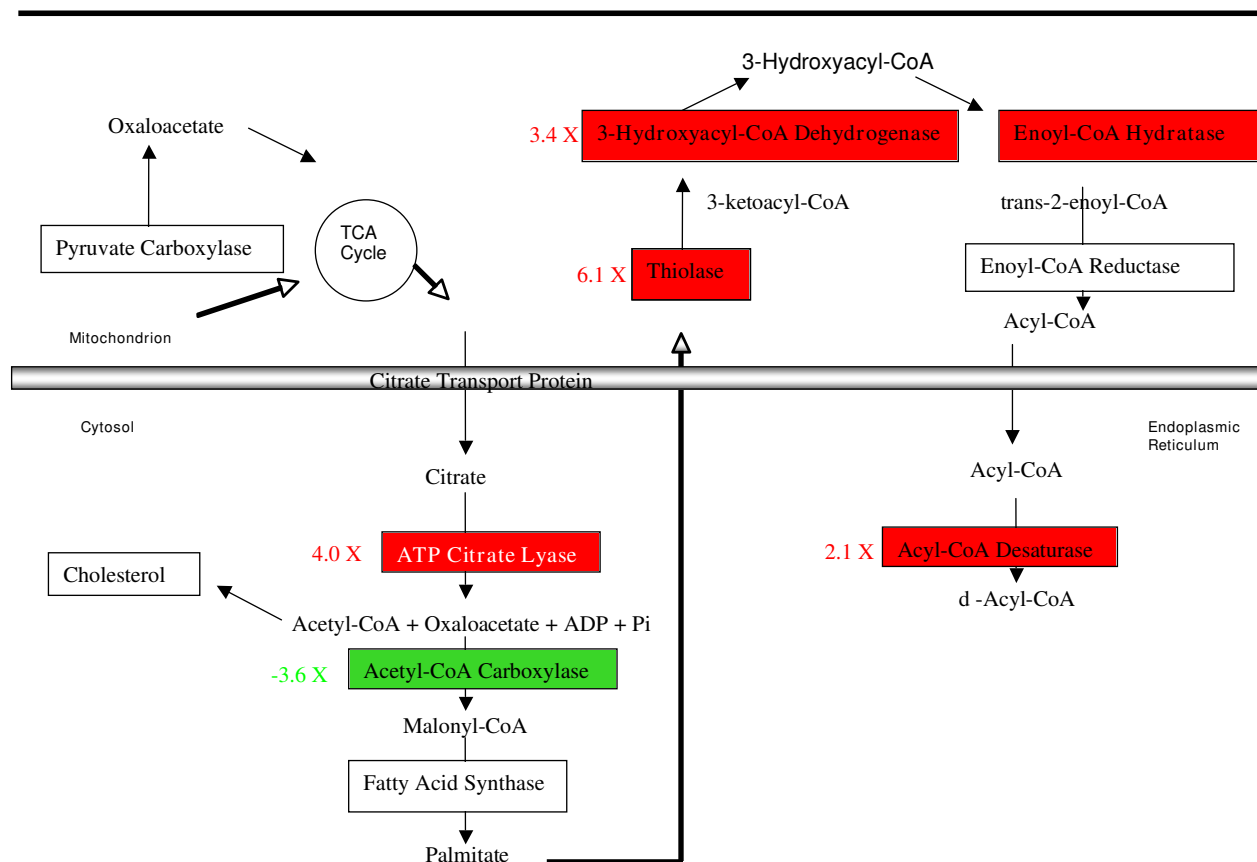


Figure 3
Subcellular localization and metabolic pathways of significantly-modulated genes in the fatty acid synthesis pathway during the pre-viremic stage of LCMV-WE infection in macaque liver tissues. Red or green color corresponds to up- or down-regulation of gene expression, respectively. ATP citrate lyase is encoded by ACLY (Tables 3 and 5), Acetyl-CoA carboxylase is encoded by ACACB (Table 4), thiolase is encoded by ACAT2 (Tables 3 and 4), 3-hydroxyacyl-CoA dehydrogenase is encoded by HADHA (Table 4), enoyl-CoA hydratase is encoded by ECHS1 (Table 4) and acyl-CoA desaturase is encoded by SCD (Table 5).

gene encoding kallikrein (KLKB1) that dilates blood vessels and should prevent vessel leakage.

Gene expression profiling that discriminates between virulent and non-virulent virus infection in LCMV-infected liver

Genes that discriminate between virulent and non-virulent infection, especially if they do so during the pre-viremic stage, have the greatest potential as prognostic biomarkers for severe arenaviral disease. Virulent and non-virulent liver transcriptomes were compared by pooling results from nine animals (LCMV-WE-infected) to compare with six samples from animals that were LCMV-infected but not diseased (two animals infected with

LCMV-ARM and four infected with both LCMV-WE and LCMV-ARM that did not develop disease). In our study, the transcriptome of non-diseased animals closely resembled the transcriptome of animals infected with non-virulent LCMV-ARM (see clustering in Additional File 1). Liver gene expression profiles showed significant differences between virulent and non-virulent infections (Figure 4). We determined that approximately 244 genes significantly discriminated non-virulent and virulent LCMV infections, and of these, approximately 30 did so during the pre-viremic stage of infection (Figure 4 and Additional File 8). Notable are large drops in expression of hemoglobin alpha and beta genes, despite up-regulation of transferrin (TF) the product of which promotes iron

Table 7: Expression of genes involved with complement and coagulation cascades in LCMV-WE-infected macaque liver

GenBank Accession no.	Symbol	Gene description	Pre-viremic		Viremic	
			Fold change ^a	p ^b	Fold change ^a	p ^b
NM_000014	A2M	Alpha-2-macroglobulin	3.31	0.0219	2.71	0.0417
NM_000491	CIQB	Complement component I, q subcomponent, B chain	3.65	0.1140	12.2	0.0029
AI184968	CIQC	Complement component I, q subcomponent, C chain	1.47	0.5402	3.27	0.0140
AL573058	CIR	Complement component I, r subcomponent	2.20	0.0287	2.73	0.0054
BC007010	CIS	Complement component I, s subcomponent	53.44	0.0006	50.21	0.0004
NM_000592	C4A/C4B	Complement component 4A (Rodgers blood group)//complement component 4B (Childo blood group)	5.20	0.0002	6.32	0.0001
NM_000716	C4BPB	Complement component 4 binding protein, beta	3.22	0.0401	5.65	0.0032
NM_001735	C5	Complement component 5	8.75	0.0015	7.83	0.0013
J05064	C6	Complement component 6	3.24	0.0332	2.63	0.0624
NM_000587	C7	Complement component 7	2.34	0.1850	4.28	0.0152
M17263	C8G	Complement component 8, gamma polypeptide	4.72	0.0051	4.56	0.0038
AL570661	CD46	CD46 molecule, complement regulatory protein	4.72	0.0015	3.24	0.0060
X04697	CFH	Complement factor H	2.90	0.0124	2.86	0.0091
NM_000506	F2	Coagulation factor II (thrombin)	2.11	0.0329	1.65	0.1360
AA910306	F5	Coagulation factor V (proaccelerin, labile factor)	3.16	0.0218	2.47	0.0071
NM_000504	F10	Coagulation factor X	2.42	0.0070	1.85	0.0513
NM_000128	F11	Coagulation factor XI (plasma thromboplastin antecedent)	-2.28	0.0150	-2.75	0.0029
NM_021871	FGA	Fibrinogen alpha chain	3.86	0.0035	3.48	0.0039
BG545288	FGB	Fibrinogen beta chain	49.86	0.0001	45.56	0.0001
NM_000509	FGG	Fibrinogen gamma chain	2.67	0.0355	2.56	0.0325
M74220	LPA/PLG	Lipoprotein, Lp(a)//plasminogen	4.69	0.0256	3.81	0.0396
NM_000892	KLKB1	Kallikrein B, plasma (Fletcher factor) I	-1.70	0.1730	-2.32	0.0207
AI274095	MASPI	Mannan-binding lectin serine peptidase 1 (C4/C2 activating component of Ra-reactive factor)	3.07	0.0004	2.78	0.0005
AB008047	MASP2	Mannan-binding lectin serine peptidase 2	4.75	0.0144	3.16	0.0489
NM_000242	MBL2	Mannose-binding lectin (protein C) 2, soluble (opsonic defect)	3.18	0.0162	1.37	0.5910
BE880828	MCFD2	Multiple coagulation factor deficiency 2	11.71	0.0004	14.72	0.0001
M74220	PLG	Plasminogen	3.86	0.0059	3.60	0.0054
NM_000295	SERPINA1	Serpin peptidase inhibitor, clade A (alpha-1 antitrypsin, antitrypsin), I	8.20	0.0003	7.21	0.0004
BC022309	SERPINC1	Serpin peptidase inhibitor, clade C (antithrombin), member 1	30.65	0.0005	19.29	0.0009
NM_000062	SERPING1	Serpin peptidase inhibitor, clade G (C1 inhibitor), member 1	2.77	0.0336	3.07	0.0296
BF511231	TFPI	Tissue factor pathway inhibitor (lipoprotein-associated coagulation inhibitor)	2.05	0.0312	1.45	0.2590

^a Mean fold changes were calculated using a division of raw expression values between experimental sample and uninfected control.

^b All samples were analyzed separately. Changes in gene expression with a cutoff of 2.0-fold increased or decreased expression was used and, the p-value was calculated by Student's t-test. Data are displayed only where the, $p \leq 0.05$. This p-value was used as a measure of the magnitude of the difference between groups and to determine significance of the modulation. The modulated genes in the pathway are listed in alphabetic order.

absorption and hemoglobin production [34]. Down-regulation of hemoglobin is concomitant with up-regulation of IL-6-inducible hepcidin (HAMP), an iron regulatory hormone known to block iron export and uptake [35]. Down-regulation of hemoglobin mRNA may also be related to a block in erythropoiesis caused by interferon [36]; since interferon levels are higher in virulent than in benign infection of primates [3]. The down-regulation of

hemoglobin genes has been validated by quantitative PCR (Table 2).

As mentioned earlier, LCMV infection is predicted to promote significant amino acid breakdown by gluconeogenesis. Three amino acid catabolic enzymes, arginase (ARG2), serine dehydratase (SDS), and glutamic-oxaloacetic transaminase (GOT1), were not altered in

WE1-uninf	WE2-uninf	WE3-uninf	WE1-nodis	WE2-nodis	WE3-nodis	GenBank Acc. #	Gene symbol	Description
10.85	7.62	18.51	7.46	5.24	12.73	NM_001105	ACVR1	Activin A receptor, type I
-4.79	-5.46	-12.82	-5.24	-5.98	-14.03	AF202889	APOA5	Apolipoprotein A-V
1.02	1.04	1.03	-12.13	-11.88	-12.04	U75667	ARG2	Arginase, type II
1.08	1.93	2.46	-18.64	-10.41	-8.17	BG435404	ARL4	CADP-ribosylation factor-like 4C
1.12	2.66	8.69	-23.92	-10.06	-3.07	NM_012118	CCRN4L	CCR4 carbon catabolite repression 4-like
1.82	13.64	88.65	-5.28	1.42	9.25	BE221212	COL1A1	Collagen, type I, alpha 1
4.47	5.98	20.68	3.84	5.13	17.75	A1246687	CTSC	Cathepsin C
9.85	2.11	24.76	10.56	2.27	26.54	NM_000783	CYP26A1	Cytochrome P450, family 26, subfamily A, polypeptide 1
12.13	69.07	120.26	2.91	16.56	28.84	NM_001443	FABP1	Fatty acid binding protein 1, liver
2.68	2.68	1.28	-2.77	-2.77	-5.78	AK001699	FBXO21	F-box protein 21
-1.34	1.04	-1.57	-4.69	-3.39	-5.50	AW080845	FOXP1	Forkhead box P1
-1.67	1.09	-1.40	-16.80	-9.25	-14.12	NM_013409	FST	Follistatin
12.13	5.17	2.07	-2.46	-5.78	-14.42	BC020700	G6PC	Glucose-6-phosphatase, catalytic subunit
3.92	2.00	1.55	-6.54	-12.82	-16.56	BC000498	GOT1	Glutamic-oxaloacetic transaminase 1, soluble
7.84	11.24	7.06	15.35	22.01	13.83	NM_021175	HAMP	Hepcidin antimicrobial peptide
3.73	1.71	1.62	-15.78	-34.54	-36.25	NM_000558	HBA1	Hemoglobin, alpha 1
2.79	1.20	1.22	-12.91	-30.06	-29.45	V00489	HBA1-2	Hemoglobin, alpha 1 /// hemoglobin, alpha 2
2.27	1.06	-1.20	-13.64	-29.04	-37.01	BC005931	HBA1-2	Hemoglobin, alpha 1 /// hemoglobin, alpha 2
-1.10	-4.29	-8.94	-5.46	-21.26	-44.32	M25079	HBB	Hemoglobin, beta
-1.21	-4.17	-9.85	-4.03	-13.83	-32.67	AF059180	HBB	Hemoglobin, beta
6.11	40.50	40.50	6.77	44.94	44.94	NM_017912	HERC6	Hect domain and RLD 6
28.64	36.76	18.38	9.38	12.04	6.02	NM_001540	HSPB1	Heat shock 27kDa protein 1
19.43	198.09	95.67	3.71	37.79	18.25	NM_006417	IFI44	Interferon-induced protein 44
1.71	-1.13	1.97	11.16	5.78	12.91	M13436	INHBA	Inhibin, beta A (activin A)
6.59	5.39	7.89	2.01	1.65	2.41	NM_005538	INHBC	Inhibin, beta C
3.58	5.86	1.53	-5.35	-3.27	-12.55	AF073310	IRS2	Insulin receptor substrate 2
2.71	25.46	8.11	2.53	23.75	7.57	NM_005101	ISG15	ISG15 ubiquitin-like modifier
2.20	2.55	1.80	-6.87	-5.94	-8.40	NM_006033	LIPG	Lipase, endothelial
11.71	14.72	6.11	2.27	2.85	1.18	BE880828	MCFD2	Multiple coagulation factor deficiency 2
2.69	2.71	1.85	-6.63	-6.59	-9.65	NM_005863	NET1	Neuroepithelial cell transforming gene 1
-1.37	1.09	-7.89	-4.96	-3.34	-28.64	NM_002591	PCK1	Phosphoenolpyruvate carboxykinase 1
-1.22	3.01	-1.35	-21.41	-5.82	-23.59	NM_013261	PPARGC1A	Peroxisome proliferator-activated receptor gamma, coactivator 1 a
25.28	9.99	6.73	156.50	61.82	41.64	N26005	PPP1R3C	Protein phosphatase 1, regulatory subunit 3C
-1.97	-2.07	-2.48	-32.22	-33.82	-40.50	NM_006843	SDS	Serine dehydratase
6.82	49.52	49.87	5.03	36.50	36.76	BC002704	STAT1	Signal transducer and activator of transcription 1, 91kDa
7.46	6.77	11.79	2.62	2.38	4.14	BC004188	TUBB2C	Tubulin, beta 2C
3.32	28.25	26.17	2.08	17.75	16.45	NM_004223	UBE2L6	Ubiquitin-conjugating enzyme E2L 6

Figure 4
Listing of the most differentially-modulated genes in liver with respect to infected-but not-diseased samples.
 There are clear differences between infected and uninfected samples, but less frequently, there are also differences between virulently-infected and mildly-infected samples and the most prominent genes in this latter category are highlighted in this figure. WE1 vs uninf (or vs nodisease) refers to pre-viremic samples (day 1–3) and WE2 vs uninf (or vs nodisease) refers to viremic samples (day 4–7). N-fold refers to fold up- or down-regulation of each gene expressed in a virulently-infected liver in relation to expression in an uninfected or non-diseased liver. Gene accession number, gene symbol and description of each gene are shown. Red or green color corresponds to up- or down-regulation of gene expression, respectively. This figure is a subset of Additional File 8.

their expression when comparing infected versus uninfected samples, yet there was a significant down-regulation of these genes when comparing LCMV-WE-infected to LCMV-non-diseased samples (Figure 4). This means that transcripts from mildly-infected samples are up-regulated with respect to transcripts from uninfected liver. The up-regulation of these samples in mild infections has been validated by quantitative PCR (Table 2). A similar pattern has been observed for peroxisome proliferator-activated receptor gamma (PPARGC1A), hemoglobin α 1/ α 2 (HBA 1–2) and follistatin (FST) (Figure 4), and has been validated by quantitative PCR (Table 2). It is likely

that increased expression of these genes protects animals with mild infections from a more severe disease.

PPP1R3C is the most dramatically up-regulated transcript in virulently-infected livers with respect to mildly-infected liver samples (156-fold in the pre-viremic stage), and this up-regulation has been validated by quantitative PCR (Table 2). PPP1R3C encodes an inhibitory subunit of protein phosphatase and is known to bind liver glycogen in response to high levels of insulin. It belonging to a family of pro-inflammatory genes with short-lived-mRNA controlled by AU-rich elements in their 3'UTR [37]. Express-

sion of PPP1R3C has been associated with encephalopathy [38].

Additional notable differences between virulent and mild infections are large increases in expression of genes for activin receptor (ACVR1) and inhibin (INHBA, INHBC), along with a decrease in expression of follistatin (FST); all of which belong to the transforming growth factor beta (TGF- β) gene family. Expression of these TGF- β family genes was validated by real-time quantitative PCR (Table 2). Though the gene for TGF- β was not differentially expressed during LCMV infection, the protein was detectable in plasma during the viremic stage [3]. Follistatin is known to bind and inactivate inhibin, and the down-regulation of FST is related to the rise in INHB and ACVR1 expression [39]. The inhibins act as negative regulators of B cell development [40], which may contribute to LCMV-mediated suppression of anti-viral immune responses.

Discussion

The LCMV-infected monkey model for Lassa fever was investigated with the goal of finding transcriptome changes that could eventually be used as prognostic biomarkers to discriminate virulent and mild infections. Transcriptome analysis of rhesus macaque liver showed that both mild and virulent LCMV infections had tremendous effects on glucose, amino acid and fatty acid metabolism, and on the complement and coagulation cascades. As with our previous transcriptome analysis using blood [3], prominent expression of inflammatory-response genes was also seen in liver.

A transcriptome study using the HuH-7 liver cell line [41] showed that Lassa infection up-regulated only laminin-a and ribosomal protein L28 with respect to gene expression in uninfected cells. It is frequently observed that studies of single cell types will show few differences in comparison to studies of complex tissues. Although our study did not see changes in expression of the laminin-a gene, several ribosomal protein genes were up-regulated in LCMV-infected liver: approximately 30 were moderately up-regulated, and 20 were highly up-regulated, with L28 being the most up-regulated of all (Additional File 8). None of the ribosomal gene expression changes observed in LCMV-infected liver differed significantly when comparing the virulent and mild infections. It is possible that L28 is a co-factor of arenavirus replication, since replicating LCMV is associated with ribosomal protein complexes [42-44].

Other liver transcriptome studies have identified major molecular events associated with viral infection, with higher levels of interferon-responsive genes being a common theme. We observed significant up-regulation of ISG15, IFIT1, and STAT1 in LCMV-WE-infected (diseased)

monkey liver with respect to non-diseased liver, and similar increased expression was seen in human liver infected with hepatitis C virus [45]. As noted in a recent paper on the Ebola virus transcriptome [46], the majority of cells profiled are uninfected bystanders and are responding to plasma interferon despite the interferon-antagonistic activity of viral genes within infected cells [47]. In two liver transcriptome studies of human beings infected with hepatitis C virus, MHC class I and II genes were up-regulated in infected versus uninfected liver [45,48]; in our studies, several MHC genes are up-regulated in liver, with concomitant down-regulation in PBMC [3], possibly due to migration of activated monocytes from the circulation to infected sites. A notable difference between the hepatitis C liver transcriptome and the LCMV-liver transcriptome is that Serpin D1 and C genes are down-regulated by hepatitis C, whereas Serpin C is 30-fold up-regulated in the LCMV liver (Figure 2) and would be expected to inhibit the intrinsic coagulation pathway.

Two crucial liver functions are glycogen breakdown and gluconeogenesis from non-sugars like fatty acids, pyruvate, lactate, and amino acids, with both functions beginning a few hours after the onset of fasting. Elevated rates of fat breakdown (lipolysis) increase the release of free fatty acids that simultaneously stimulate gluconeogenesis and inhibit glycogenolysis [49,50]. The oxidation of fatty acids by liver mitochondria leads to the generation of ketone bodies, which appear in the blood in inverse proportion to glucose and provide an important alternative fuel source [51,52]. In LCMV-infected monkey liver there was a significant increase in transcripts promoting gluconeogenesis in both stages of infection and a decrease in transcripts promoting glycogenolysis (e. g. G6PC and FBP1) in the early stage of infection. It is likely that gluconeogenesis was driven by hydrolysis of triglycerides since transcripts for β -oxidation enzymes and lipases were up-regulated in LCMV-infected livers (Table 6). Glycolysis and gluconeogenesis constituted a "futile cycle" that would be predicted to waste energy during LCMV infection (Table 2). During the early stage of infection the macaque liver increased the usage of fatty acids and ketone bodies, and later in the infection, the liver functioned in a glucose-producing role, using amino acids and fatty acids as an energy source.

Other viral infections are also inter-twined with glucose, amino acid and fatty-acid metabolism. The transcription of hepatitis B virus, a DNA virus, requires a key regulatory enzyme of gluconeogenesis, peroxisome proliferator-activated receptor γ C1 (PPARGC1[53]. This enzyme acts through nuclear receptors (the glucocorticoid receptor, the forkhead transcription factor, and the nuclear receptor 4a) to stimulate transcription of hepatitis B. PPARGC1 is known to be up-regulated by fasting, cold temperature or

stress [53], and, remarkably, it is significantly up-regulated in the mild LCMV infection but not in the virulent infection; this has been validated by quantitative PCR (Table 2). Another glycolytic enzyme, phosphoglycerate kinase (PGK), stimulates Sendai virus transcription through its interaction with tubulin in the initiation complex [54]. Vero cells infected with the alphavirus, Mayaro, have altered glucose metabolism and increased glucose consumption [55], but it is unclear whether this is directly related to virus replication. The key glycolytic enzyme, glyceraldehyde 3-phosphate dehydrogenase (GAPDH) is involved in the life cycle of parainfluenza virus type 3 (HPIV3) [56]. Interestingly, the genes for both PGK and GAPDH were strongly up-regulated in both mild and virulent LCMV infections. Hepatitis C virus (HCV) infection also affects hepatic glucose metabolism. HCV down-regulates insulin-receptor substrate genes (IRS1 and IRS2) through up-regulation of suppressor of cytokine signaling (SOCS) [57]. Similarly in the LCMV-infected primate, IRS1 expression is decreased and expression of IRS2 is moderate, but SOCS2 is up-regulated (7.2-fold) at the pre-viremic stage of infection (Additional File 8).

The acute LCMV disease has characteristics of starvation that resemble both cachexia (TNF- α or IL-6-mediated wasting) and anorexia (appetite loss). High levels of IL-6 were detected in LCMV-WE-infected liver [6]. IL-6 is known to stimulate protein catabolism in order to maintain glucose levels via gluconeogenesis [24], whereas anorexia, is largely driven by IL-1 β [58] and is known to mobilize fat stores in lieu of protein stores [24]. Here, though we failed to detect increases in IL-1 β , we observed increased expression of both lipid-oxidation and protein catabolism genes. We also observed an early decrease in blood triglycerides (Table 1) that corroborates our transcriptome result of suppressed ACACB. The low triglycerides we observe also contrast with the profile of bacterial sepsis in which high liver IL-6 coincides with high triglycerides [59]. Although high IL-6 is detected by day 7 [6] high circulating triglycerides are not seen until day 11 or 12 in the LCMV-WE-infected macaques (Table 1).

It is difficult to relate changes in intermediary metabolism of LCMV-infected liver to a lethal disease that resembles Lassa fever. Although the liver transcriptome may resemble that of a starving organism, and such a profile is supported by the dehydration and appetite-loss observed in diseased monkeys, it is known that primates can survive weeks of starvation [51]; so it is unlikely that starvation alone explains such a rapid disease. Since most of the transcriptome changes that affected intermediary metabolism were seen in both the diseased and non-diseased animals it is unlikely that those changes were the primary cause of disease. Importantly, several genes involved in intermediary metabolism (ARG2, GOT1, G6PC, IRS2, LIPG, PCK1,

SDS, PPARGC1) were considerably more up-regulated in the mild than in the fatal infections (Figure 4) leading to the possibility that the altered energy metabolism is primarily a self-preserving rather than a pathogenic activity.

Some of the differences between virulently – and mildly-infected liver could be ascribed to the over-expression of interferon-response genes (e.g. IFI44, Figure 4). A large drop in hemoglobin gene expression in virulent infections was likely due to interferon inhibition of erythropoiesis [36]. Paradoxically, circulating levels of red blood cells and hemoglobin were unaffected by LCMV-WE infection (Table 1), but events in the liver may precede what is detectable in the circulation. A recent paper describes lethal hemorrhagic anemia in LCMV-infected mice that did not occur in IFN- α/β knockout mice [60]. In the murine model, both LCMV-WE and LCMV-ARM caused platelet dysfunction and life-threatening anemia. The murine studies used much higher doses of virus than we used for our primate pathogenesis studies, i. e. mice were given 10^6 pfu virus, whereas the monkeys were given 10^3 pfu. Also, the disease in mice is generally considered immunopathological, i.e. alleviated by immunosuppression, unlike the disease in guinea pigs and primates [61]. In the monkey model, only LCMV-WE replicates well in monkey liver whereas LCMV-ARM replicates poorly [4], and only LCMV-WE elicits detectable levels of interferon in plasma [8]. Nevertheless, it is quite reasonable that in the primate, IFN- α/β initiates platelet dysfunction, and it might be reasonable to treat the primate disease with platelet transfusions, as was done in the murine model.

Conclusion

Microarray analysis identified several potential gene markers of LCMV-WE-associated liver disease. By investigating changes in gene expression during the early stages of disease we identified pathways most likely to be instigating the disease signs observed during viral hemorrhagic fever. Alterations in intermediary metabolism are more likely a sign of active resistance than of pathogenesis. Virus-mediated cytokine production could be responsible for curtailed erythropoiesis and platelet dysfunction. Changes in expression of genes in the coagulation cascade could be directly responsible for capillary leakage and thrombocytopenia in virulent LCMV infection.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

MD participated in study design, processed the liver samples, oversaw the workflow, participated in bioinformatics analyses and wrote the first drafts of the manuscript. ORC contributed to study design, and oversaw the bioinformatic analyses of the chip hybridizations. YZ per-

formed much of the bioinformatics analyses and uploaded the data to the GEO database, JCZ contributed to study design and was responsible for animal sampling and sample organization. BS oversaw the workflow at the Virginia Bioinformatics Institute where ORC and YZ conducted analyses. MGL validated liver gene expression using real-time RT-PCR and contributed edits to the final manuscript. JB oversaw the animal model and contributed to study design. HD cared for the animals and oversaw the sampling. MS conceived of the study, participated in design and edited the final drafts of the manuscript. All authors read and approved the final manuscript.

Additional material

Additional File 1

Cluster analysis of differentially-expressed genes. Cluster analysis of differentially expressed genes representing 4482 probe sets from uninfected or LCMV-infected liver samples filtered by having $p < 0.05$ and at least 20 percent of present call. Each row represents the indicated gene. Each column corresponds to the experimental liver sample as listed at the top. Red indicates an up-regulation of gene expression and green indicates a down-regulation of expression in rhesus macaque liver. The origin of each sample is described in Table 1.

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Additional File 2

Lists genes from the Venn diagram of Figure 1 in the form of 2751 probe sets. "Pre" means they were expressed in the pre-viremic stage (day 1–3) in blood, but not in liver.

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[<http://www.biomedcentral.com/content/supplementary/1743-422X-6-20-S2.xls>]

Additional File 3

Lists genes from the Venn diagram of Figure 1 in the form of 500 probe sets. "Pre" means they were expressed in the pre-viremic stage (day 1–3) in both blood and liver.

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[<http://www.biomedcentral.com/content/supplementary/1743-422X-6-20-S3.xls>]

Additional File 4

Lists genes from the Venn diagram of Figure 1 in the form of 2033 probe sets. "Pre" means they were expressed in the pre-viremic stage (day 1–3) in liver, but not in blood.

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[<http://www.biomedcentral.com/content/supplementary/1743-422X-6-20-S4.xls>]

Additional File 5

Lists genes from the Venn diagram of Figure 1 in the form of 3136 probe sets. "Vir" means they were expressed in the viremic stage (day 4–7) in blood, but not in liver.

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[<http://www.biomedcentral.com/content/supplementary/1743-422X-6-20-S5.xls>]

Additional File 6

Lists genes from the Venn diagram of Figure 1 in the form of 963 probe sets. "Vir" means they were expressed in the viremic stage (day 4–7) in blood and liver.

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[<http://www.biomedcentral.com/content/supplementary/1743-422X-6-20-S6.xls>]

Additional File 7

Lists genes from the Venn diagram of Figure 1 in the form of 1722 probe sets. "Vir" means they were expressed in the viremic stage (day 4–7) in liver, but not in blood.

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Additional File 8

List of differentially-expressed genes in LCMV-infected liver. This list of human probe sets identified in hybridization of rhesus macaque liver cRNA includes those transcripts differentially expressed with respect to uninfected (uninf) control samples ($n = 4484$), or those transcripts differentially expressed with respect to samples from monkeys that were LCMV-infected but not diseased (nodis). These were identified as differentially expressed using LIMMA analysis. WE1 refers to 4 liver samples taken day 1 to day 3 after infection (pre-viremic stage). WE2 refers to 5 liver samples taken day 4 to day 7 after infection (viremic stage), and WE3 refers to 2 liver samples taken day 11 and day 12 after infection (terminal stage). WEx vs uninf refers to virulent samples being compared to data from 3 uninfected samples. WEx vs nodis refers to virulent samples being compared to data from 6 infected but not diseased samples. Samples are more fully described in Table 1 of the manuscript. Mean fold changes were calculated using a division of $\log(2)$ -normalized expression values between experimental sample and uninfected control as described in Materials & Methods. False discovery rate of the pair-wise comparisons were calculated using p-values from LIMMA. Significantly regulated genes were selected using a 2-fold cut-off and a false discovery rate of < 0.05 .

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Additional File 9

Pathway gene expression in LCMV-WE-infected macaque liver tissues. Significantly affected pathway gene expression in LCMV-WE-infected macaque liver tissues. KEGG software was used to identify groupings of genes with pathways based on published data. False discovery rate of the pair-wise comparisons were calculated using p-values from LIMMA. Significantly regulated genes were selected using a 2-fold cut-off and a false discovery rate of < 0.05 .

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