

NIH Public Access

Author Manuscript

Biochem Pharmacol. Author manuscript; available in PMC 2010 February 1.

Published in final edited form as:

Biochem Pharmacol. 2009 February 1; 77(3): 285–296. doi:10.1016/j.bcp.2008.09.029.

Catalase and glutathione peroxidase mimics

Brian J. Day*

Department of Medicine, National Jewish Health, Departments of Medicine, Immunology & Pharmaceutical Sciences, University of Colorado Health Sciences Center, Denver, CO, USA

Abstract

Overproduction of the reactive oxygen species (ROS) superoxide (O_2^-) and hydrogen peroxide $(H₂O₂)$ are increasingly implicated in human disease and aging. ROS are also being explored as important modulating agents in a number of cell signaling pathways. Earlier work has focused on development of small catalytic scavengers of O_2^- , commonly referred to as superoxide dismutase (SOD) mimetics. Many of these compounds also have substantial abilities to catalytically scavenge H2O2 and peroxynitrite (ONOO−). Peroxides have been increasingly shown to disrupt cell signaling cascades associated with excessive inflammation associated with a wide variety of human diseases. Early studies with enzymatic scavengers like SOD frequently reported little or no beneficial effect in biologic models unless SOD was combined with catalase or a peroxidase. Increasing attention has been devoted to developing catalase or peroxidase mimetics as a way to treat overt inflammation associated with the pathophysiology of many human disorders. This review will focus on recent development of catalytic scavengers of peroxides and their potential use as therapeutic agents for pulmonary, cardiovascular, neurodegenerative and inflammatory disorders.

Keywords

Catalytic antioxidants; Cell signaling; Drug development; Hydrogen peroxide; Inflammation; Oxidative stress

1. Endogenous catalytic hydrogen peroxide scavengers

Hydrogen peroxide (H₂O₂) is generated directly from superoxide (O₂⁻) through a rapid dismutation reaction that can occur either enzymatically with superoxide dismutases (SOD) or spontaneously. This means that wherever O_2^- is generated there is also formation of H_2O_2 . In addition, H_2O_2 is formed enzymatically as a by-product of lipid metabolism in peroxisomes. H_2O_2 is stable at biological pH and easily crosses lipid membranes. H_2O_2 can participate in hydroxyl radical (HO•) formation in the presence of reduced transition metals. Oxidative stress is traditionally defined as an imbalance between reactive oxygen species (ROS) production and antioxidant defense against these ROS. A consequence of oxidative stress is an increase in the formation of oxidized cellular macromolecules. Critical cysteinethiol groupson proteins are a common site ofoxidation and many of these cysteines are important in maintaining

^{*}Correspondence address: Division of Environmental & Occupational Health Sciences, A439, National Jewish Health, 1400 Jackson Street, Denver, CO 80206, USA. Tel.: +1 303 398 1121; fax: +1 303 270 2263. E-mail address: dayb@njc.org.

Publisher's Disclaimer: This article appeared in a journal published by Elsevier. The attached copy is furnished to the author for internal non-commercial research and education use, including for instruction at the authors institution and sharing with colleagues. Other uses, including reproduction and distribution, or selling or licensing copies, or posting to personal, institutional or third party websites are prohibited.

In most cases authors are permitted to post their version of the article (e.g. in Word or Tex form) to their personal website or institutional repository. Authors requiring further information regarding Elsevier's archiving and manuscript policies are encouraged to visit: <http://www.elsevier.com/copyright>

proteins in a proper conformation for catalytic function. This is a common mechanism proposed for how oxidative stress can disrupt cell signaling pathways leading to unregulated inflammatory responses [1]

SODs and catalase are metalloproteins that catalyze "dismutation" reactions, which detoxify O_2^- and H_2O_2 , respectively. SODs catalyze the formation of oxygen and H_2O_2 from two O_2^- , whereas catalase catalyzes the formation of oxygen and water from two H_2O_2 molecules.

$$
2H_2O_2 \rightarrow O_2 + 2H_2O
$$

Because these efficient reactions do not require additional reducing equivalents, no energy is required from the cell. The overall goal of cellular antioxidant defenses is to reduce ROS to water. Mammalian catalase is a tetramer in which each monomer contains an iron heme (porphyrin) group bound to the catalytic site [2]. The heme groups are protected, being buried in a non-polar pocket with narrow hydrophobic channels to aid in $\rm H_2O_2$ selectivity. It should be noted that catalase also possesses peroxidase activity and is known to oxidize short chain alcohols to their corresponding aldehydes [3]. Overexpression of SOD and catalase in cultured cells and whole animals has provided protection against the deleterious effects of a wide range of oxidative stress paradigms [4].

Another class of endogenous catalytic H_2O_2 scavengers is the selenium-containing peroxidases [5]. This is a broad group of enzymes that utilize H_2O_2 as a substrate along with an endogenous source of reducing equivalence. One of the best studied families of peroxidases are the glutathione peroxidases (GPx). GPxs are tetrameric proteins where each monomer contains one atom of selenium at the catalytic site. The active site of GPx contains a selenocysteine where the sulfur in cysteine has been replaced by selenium (R-SeH). During the catalytic cycle, a selenol (protein-Se⁻) reacts with peroxide (H_2O_2 or lipid peroxide, LOOH) resulting in a selenenic acid (protein-SeOH). The selenenic acid group is reduced back to a selenol by two glutathiones (GSH) which are in turn oxidized to disulfide (GSSG) and LOOH is reduced to its corresponding alcohol (LOH).

 $H_2O_2+2GSH \rightarrow GSSG+2H_2O$

 $LOOH+2GSH \rightarrow GSSG+H_2O+LOH$

The GSSG is converted back to two GSH by glutathione reductase that uses reducing equivalents derived from β-nicotinamide adenine dinucleotide phosphate (NADPH). Overexpression of GPx has been shown to be protective against oxidative stress in cultured cells and whole animals [6,7]. Not all of the peroxidases detoxify H_2O_2 . A number of nonspecific peroxidases, such as myleoperoxidase, eosinophil peroxidase and lactoperoxidase, actually form more reactive produces such as hypochlorite, hypothiocyanite, and hypobromous acid [8].

A new family of proteins have been discovered that are known as the thioredoxin-dependent peroxidases or peroxiredoxins (Prx). These proteins can directly reduce peroxides and the oxidized protein is regenerated indirectly by thioredoxin (Trx) reductase [9,10]. There are at least 13 mammalian Prxs and they are widely abundant in mammalian cells. Prx have been found to be induced in response to oxidative stress [11] and are protective against oxidative

stress when over-expressed in animals [12]. The Prxs and Trxs work in tandom with the GPxs to maintain cellular peroxide steady-state levels and also maintain cellular protein cysteines in a reduced state (Fig. 1). During oxidative stress, where cellular peroxides are elevated, there is an increased level of oxidized protein cysteines that leads to inactivation of phosphatases and transcription factors that are thought to drive dysregulated inflammatory reactions [13]. This is the drug target and rationale for the development of catalase and GPx mimics.

2. Development of catalytic hydrogen peroxide scavengers

There are two main design strategies to detoxify peroxides. One approach models after the catalase dismutation reaction and focuses on compounds with redox-active metal centers that often containing either manganese (Mn) or iron (Fe). Another strategy models after the GPx enzymes using either selenium or tellurium active sites. An ideal mimetic is stable and nontoxic at therapeutically efficacious concentrations. The size and charge of the mimetic is often exploited to target cellular sites of oxidant production, such as the mitochondria, and to improve their pharmacodynamic properties.

Many simple metal chelates readily react with H_2O_2 . However, the rates of reaction with these chelates are generally low and the complexes formed are relatively unstable. Recent developments in the field have yielded more stable and active metal chelates (Fig. 2). These include the salens, metalloporphyrins, and other metal complexes that can dismutate H_2O_2 under highly defined conditions. Likewise, a number of selenium-containing compounds have been developed around the GPx mechanism of H_2O_2 decomposition (Fig. 3).

All of the reported catalytic H_2O_2 scavengers react with a wide range of ROS and are not specific for H_2O_2 . The potencies of these peroxide scavengers are based on rate constants derived under very defined and often non-biologically relevant conditions. For example, most compounds are routinely screened in buffers using very high H_2O_2 (1–10 mM) levels that are two orders of magnitude higher than the 1–100 nM levels that occur physiologically [14]. The findings that many of these diverse compounds are effective in similar oxidative stress models confirms the basic concept that small, efficient, catalytic antioxidants show promise in the treatment of ROS-mediated conditions associated with injury and tissue dysfunction.

3. Antioxidant properties of catalase-like hydrogen peroxide scavengers

3.1. Metalloporphyrins

Metalloporphyrins [AEOL series is currently being developed by Aeolus Pharmaceuticals, Laguna Niguel, CA [\(http://www.aeoluspharma.com\)](http://www.aeoluspharma.com)] are structurally different from endogenous protoporphyrins and are classified as synthetic *meso*-substituted porphyrins. Metalloporphyrins have been shown to possess at least four distinct antioxidant properties, which include scavenging O_2^- [15], H_2O_2 [16], ONOO⁻ [17], and LOOH [18]. Most metalloporphyrins contain either a Fe or Mn moiety that is coordinated by four nitrogen axial ligands. The catalase-like activity of metalloporphyrins is thought to be due to their extensive conjugated ring system that can undergo reversible one-electron transfers in addition to the one-electron transfers on the metal center. This mechanism is similar to that proposed for the heme prosthetic groups of endogenous catalase and peroxidases. There are two classes of metalloporphyrins wherein one group the SOD activities track with their catalase activities and another group that has very little SOD activity with high catalase activity. Examples of a manganese porphyrins with both high SOD and catalase-like activities are the pyridinium and imidazolium-substituted *meso*-porphyrins such, as AEOL 10113 and 10150 [19], whereas examples of compounds with low SOD activity and high catalase activity are MnTBAP [16] and AEOL 11207 [20]. It is still unknown which antioxidant activities are the most important in mediating the protective effects of metalloporphyrins in models of oxidative stress.

Metalloporphyrins have been shown to be effective in ameliorating oxidative stress, inflammation and injury in a large number of *in vitro* [21] and animal models of human disease (Table 1). Metalloporphyrins have plasma half-lives that range from 4 to 48 h. Most metalloporphyrins are not extensively metabolized by the body and are largely excreted unchanged in the urine. A previous limitation of the metalloporphyrin class of compounds has been their poor oral bioavailability, but several compounds in the AEOL 112-series have been shown to have good oral bioavailability and longer plasma half-lives which should make them better candidates for treating chronic diseases [22].

3.2. Salens

The salen class of catalytic antioxidants (EUK series) is currently being developed by Proteome Systems, North Ryde, Australia ([http://www.proteomesystems.com\)](http://www.proteomesystems.com). Generically, salens are aromatic, substituted ethylenediamine metal complexes. The Mn(III)-containing salen complexes have both O_2^- and H_2O_2 dismutation activities [23]. However, like all the small molecular weight scavengers, these compounds are not selective and can react with O_2^- and other peroxides and ONOO−. The Mn moiety of the salen is coordinated by four axial ligands. One of the unique features of these compounds is that the metal center is coordinated to oxygen and nitrogen atoms which is in contrast to the porphyrins where the metal is only coordinated to nitrogen atoms. The coordination of Mn by four axial ligands results in the formation of several possible valence states that give these compounds their broad ROS scavenging capabilities. The rates at which reported salens scavenge H_2O_2 are similar to those reported for metalloporphyrins, but are many orders less than those documented for catalase under similarly defined conditions [23]. Salens have also been shown to protect cells against H_2O_2 mediated injury [24]. Salens have been shown to be efficacious in a large number of animal models of human diseases (Table 1). One of the current limitations of the salens is the stability of the parent compounds in biological matrix which makes it difficult to determine tissue levels and half-lives.

3.3. Other metal complexes

There are a number of other metal containing macrocyclic compounds that have been described as catalase mimics. Iron complexes of 14-membered macrocycles have been shown to catalytically scavenge H_2O_2 to oxygen and water [25]. These compounds have been shown to be effective only in cell culture systems [26]. It is unclear how stable the complexes will be in more complex biological systems. Another group of compounds with H_2O_2 -scavenging activity are the dimanganese complexes. A number of bacteria have catalase enzymes that use a dimanganese center to dismutate H_2O_2 and a number of investigators have tried to emulate this strategy with small molecules and peptides. Some examples of the small molecules are the nitro and chloro-substituted dimanganese complexes of 1,5-bis(5-salicylidenamino) pentan-3 ol (5-NO2-salpent & 5-Cl-sal-pent), 1,5-bis(2-hydroxybenzophylideneamino)pentan-3-ol (2- OH-benzpent), and 1,5-bis(2-hydroxynaphtylideneamino)pentan-3-ol (2-OH-Napthpent) [27]. Very limited data is currently available whether they have protective properties in biological systems. It should be noted that free manganese is an efficient scavenger of H_2O_2 and so stability is an important feature to establish for any claim of catalytic H_2O_2 scavenging by a metal complex.

4. Antioxidant properties of glutathione peroxidase-like hydrogen peroxide scavengers

4.1. Ebselen

One of the best studied GPx-like mimics is 2-phenyl-1,2-benzisoselenazol-3(2H)-one also known as ebselen or PZ51. Ebselen was one of the first selenium-based GPx mimics developed.

It catalytically scavenges peroxides in the presence of reducing equivalents such as GSH, Nacetylcysteine (NAC), and dihydrolipoate (DHLA) [28]. The mechanism by which this occurs is still debated and may differ under different conditions. Ebselen has also been shown to stimulate the decomposition of a number of ROS including hypochlorous acid (HOCl) [29], singlet oxygen [30], and ONOO− [31]. Ebselen can readily bind cellular thiol groups on proteins which may complicate the interpretation of biological effects since many cellular proteins have reactive thiols in their catalytic domains. In fact, it has been documented that ebselen can inhibit lipoxygenases [32], NADPH oxidases [33], and nitric oxide synthases [34]. All of these enzymes are also potential sources of endogenous ROS. Ebselen has been shown to be protective in a number of cell culture systems [28] and animal models of human disease (Table 1). Ebselen is orally active and appears to be well tolerated in animals and humans.

Newer analogs of ebselen have been developed including BXT-51072, which has increased activity and potency in cell systems. These analogs [BXT-series are being developed by Oxis International, Foster City, CA ([http://www.oxis.com\)](http://www.oxis.com)] have been shown to be protective in a limited number of cell culture systems [35] and animal models of human disease (Table 1).

4.2. Diselenide and ditelluride compounds

A number of diselenide and ditelluride containing compounds have been reported to catalytically scavenge peroxides with higher GPx-like activity than ebselen [36]. Sulfur, selenium, and tellurium belong to group VI of the periodic table and have similar chemical properties. Early compounds, such as the diphenyl diselenide (DPDS), were electrophilic agents that have cytotoxic, genotoxic, and mutagenic effects [37,38]. Many previously reported diselenide compounds release free selenium during the catalytic cycle, which may be problematic in their development as therapeutic agents. A unique aspect of a newer series of these compounds is the cyclodextrin (CD) group which may help in directing hydrophobic peroxides towards the selenium or tellurium active site. The diselenide, 2,2′-deseleno-bis-βcyclodextrin (2-SeCD) can scavenge a variety of peroxides including H_2O_2 , *tert*-butyl hydroperoxide, and cumenyl hydroperoxide using GSH as a cofactor [39]. Only a limited number of cell culture studies have been reported for these compounds [40], and it is still unclear whether these compounds can be successfully used in animal models of human disease.

4.3. Peptide compounds

Many proteins and peptides contain cysteine residues that can be readily modified to contain selenium and a number of investigators have examined whether these types of compounds have GPx-like activities. Examples of this include seleno-subtilisin which was produced by chemical modification of the serine protease, substilisin. This compound was found to have GPx-like activity [41]. Another strategy used a phage library of random 15-mer selenopeptides that was screened for GPx activity and generated some active GPx mimics $(15SeP and 15SeP₁)$. The peptides were found to increase the GPx-like activity of treated cultured cells and protected them against H_2O_2 -mediated lipid peroxidation and cytotoxicity [42]. In order to increase the selectivity of the selenoprotein toward GSH, a number of selenium-containing monoclonal antibodies (i.e. Se-4A4 and Se-scFv2F3) were raised against GSH-S-2,4-dinitrophenyl t-butyl ester and found to have GPx-like activity [43]. The selenium-containing antibody Se-4A4 has been shown to protect isolated cardiac mitochondria against xanthine oxidase-induced oxidative modification [44]. Very limited data exist on whether any of these approaches have produced compounds with biological activity in more complex biological systems.

5. Catalytic antioxidants are effective in ameliorating oxidative stress *in vitro*

5.1. Cytotoxicity

In vitro models of oxidative stress have proved useful in verifying utility of catalytic antioxidants under more complex biological conditions [21]. Members of all classes of catalytic antioxidants with H_2O_2 -scavenging activities have been shown to be effective in blocking oxidative stress in a variety of *in vitro* cytotoxicity models involving oxidant production [21, 24,28]. The mechanism(s) by which these catalytic antioxidants produce their protective effects are still highly debated and largely unknown. As discussed earlier, many of the compounds are capable of scavenging a number of different ROS and many may also decrease ROS by inhibiting endogenous ROS production [19]. These issues cloud the utility of H_2O_2 -scavenging screens to select potentially biologically potent compounds. Overall, many of the catalytic antioxidant H_2O_2 scavengers at µmolar levels appear non-toxic and show similar efficacy in protecting a wide variety of different types of cultured cells against the toxicity of ROS.

5.2. Apoptosis

Apoptosis is a form of cell death that is biochemically and morphologically distinct from necrosis and has physiological and pathological roles in biological systems. There is an increasing body of literature that supports the involvement of ROS in some apoptotic pathways. The release of pro-apoptotic factors by mitochondria, which are a major source of ROS, lends further credence to this argument. Delivery of catalase and GPx to cells is protective, whereas paucity of either is deleterious, also supporting H_2O_2 involvement in cellular apoptosis [45]. A wide assortment of apoptosis paradigms can be ameliorated by catalytic antioxidants with H2O2-scavenging activity [21,24,28]. Apoptosis can be limited by catalytic antioxidants with catalase or GPx-like activities in a number of different cell types. It is not clear from these studies whether the catalytic antioxidants affect a particular point in the intrinsic and/or extrinsic apoptotic pathways. Some studies suggest that ROS might regulate the expression of pro-apoptotic factors, and antioxidants may directly block apoptosis by increasing the expression of anti-apoptotic factors, such as bcl-2 [46].

5.3. Inflammation

A number of recent studies have suggested that catalytic antioxidants with the ability to scavenge H_2O_2 can attenuate markers of inflammation such as cytokines, chemokines and adhesion molecules. H_2O_2 has been shown to activate a number of transcription factors including NFkB, AP-1 and Nrf2 [47]. In addition, peroxides and other oxidants can inactivate kinase signaling pathways through the inhibition of protein phosphatases [48]. These pathways likely contribute to the protective effects of many catalytic H_2O_2 scavengers in a variety of cell models of inflammation [21,24,28].

6. Catalytic antioxidants are effective in ameliorating oxidative stress *in vivo*

The beneficial effects of catalytic antioxidants with H_2O_2 -scavenging capabilities have been demonstrated in numerous *in vivo* model systems (Table 1). These model systems cover diseases associated with the pulmonary system (such as fibrosis, asthma, acute respiratory distress syndrome (ARDS), chronic obstructive pulmonary disease (COPD), bronchopulmonary dysplasia (BPD), and pleurisy), cardiovascular system (including sepsis, hypertension, and myocardial infarction), neurologic system (including amyotrophic lateral sclerosis (ALS), migraine, spinal cord injury, stroke, Parkinson's disease (PD), and dementia), digestive system (including liver injury, transplantation, hepatitis, and colitis), endocrine system (diabetes), and the renal system (injury, sepsis, and transplantation). The diversity of the various models systems by which these catalytic antioxidants have shown efficacy speaks

to the important role ROS/reactive nitrogen species (RNS) play in animal models of human disease.

6.1. Pulmonary models

The lung functions at higher oxygen tensions than most other organs creating a unique relationship with ROS. There is increasing evidence that ROS have an important role in several lung diseases [49]. A common feature of many lung diseases is an inappropriate or dysfunctional inflammatory response. Catalytic antioxidants decrease airway hyperreactivity and inflammation in antigen-induced mouse models of asthma [50]. This is in contrast to a recent finding which reported high overexpression of catalase (8-fold) in the lung actually increased airway reactivity in a mouse model of asthma [51]. A potential explanation for this paradox is that many catalytic antioxidants have bell-shaped dose-responses and that very high levels can actually give the opposite response. An important goal of antioxidant therapy is to restore redox balance. Given the role of H_2O_2 in cell signaling pathways, it is conceivable that excessive scavenging of H_2O_2 may be detrimental under certain conditions.

COPD, which includes emphysema and bronchitis, is strongly associated with cigarette smoke that is a rich source of ROS [52]. In fact, cigarette smoke has been shown to inhibit catalase activity [53]. The diselenide GPx-like mimic diphenyl diselenide has been shown to protect rat pup lungs from oxidative changes associated with exposure to cigarette smoke [54]. Likewise, the metalloporphyrin AEOL 10150, which has both high SOD and catalase-like activities, attenuated inflammation and protected rat lung epithelium from cigarette smokeinduced precancerous lesions [55]. These data support the development of catalytic antioxidants for the treatment of COPD.

Interstitial lung disease is also associated with oxidative stress and many of the animal models of lung fibrosis use agents that overproduce or stimulate the production of ROS. Bleomycin is a redox-cycling chemotherapeutic agent that produces lung fibrosis in rodents and humans. Bleomycin-induced lung fibrosis can be attenuated by a liposomal mixture of SOD and catalase [56]. The metalloporphyrin AEOL 10201, also known as MnTBAP, has low SOD activity and moderate catalase activities and attenuated bleomycin-induced lung fibrosis in mice [57]. Ionizing radiation is also known to produce lung fibrosis in animals and man. Administration of a mixture of polyethylene glycol-tagged SOD and catalase has been shown to attenuate radiation-induced lung fibrosis in mice without affecting radiation-induced tumor killing [58]. Similarly, a couple of different metalloporphyrins (AEOL 10113 & 10150) have been shown to decrease radiation-induced lung fibrosis in rats [59,60]. The manganese-containing salen EUK-189 also was found to ameliorate early DNA damage in a rat model of lung irradiation [61]. These studies illustrate the potential utility of catalytic antioxidants in the treatment of interstitial lung disease.

ARDS is associated with sepsis and shock and both of these conditions involve the over production of ROS and reactive nitrogen species. A related lung disorder is BPD that occurs in premature infants where the lung is not fully developed and requires supplemental oxygen for adequate gas exchange. Both ARDS and BPD animals models commonly use either hyperoxia or endotoxin exposures, both of which are known to elevate lung ROS production. Both hyperoxia and endotoxin models have been shown to be responsive to modulation of endogenous SOD and catalase lung levels [62,63]. The metalloporphyrin AEOL 10113 was found to be beneficial in a preterm baboon model of BPD [64] and AEOL 10150 was found to attenuate hemorrhage-induced acute lung injury in SOD3 KO mice [65]. The manganese containing salen EUK 8 was found to be protective in an endotoxin-mediated swine model of ARDS [66]. These finding support the further development of catalytic antioxidants for the treatment of ARDS and BPD.

Cardiovascular disease is a major cause of death in humans. A common cause of tissue injury directly related to the cardiovascular system is ischemia-reperfusion (IR). IR is associated with hemorrhage [67], myocardial infarction, arrhythmias, angina, myocardial stunning and transplantation. The role of excessive ROS production during IR and the protective effects of endogenous antioxidants have been well documented. Catalytic antioxidants with H_2O_2 scavenging activities are effective in animal models of heart IR [68,69]. In addition, several other organ systems have been shown to benefit from catalytic antioxidant treatments in IR, including the liver [70,71], lung [65], brain [72–75] and kidney [76].

Hypertension is a well characterized risk factor for cardiovascular disease. A number of systems are involved in the complex regulation of blood pressure and include cardiac output, fluid balance, vasodilatation and renal function. An important system in blood pressure regulation is the renin/angiotensin system which has also been shown to regulate vascular ROS production [77]. A well known anti-hypertensive agent captopril is both an angiotensinconverting enzyme inhibitor and antioxidant [78]. Catalytic antioxidants with H_2O_2 scavenging activities are effective in animal models of hypertension [79,80]. These studies suggest that catalytic antioxidants may be useful antihypertensive agents.

Systemic infection or sepsis frequently results in the overproduction of ROS and RNS that has devastating consequences for the cardiovascular system. A serious consequence of sepsis is the loss of vascular tone and its responsiveness to vasoconstrictive agents in a condition referred to as shock. A number of catalytic antioxidants with H_2O_2 -scavenging activities are effective in animal models of sepsis induced by bacteria or endotoxin [66,81–86]. The well-established role of ROS and RNS in these conditions and the abundance of literature supporting a protective role of the endogenous antioxidant defenses make this a very attractive arena for development of catalytic antioxidants.

6.3. Nervous system models

The brain consumes a large amount of oxygen and is particularly sensitive to ROS-mediated damage. Factors that are thought to contribute to this phenomena are the presence of autooxidizable neurotransmitters, the high levels of polyunsaturated fatty acids in neuronal membranes, and modest levels of endogenous antioxidants. Collectively, these factors make catalytic antioxidants with H_2O_2 -scavenging activities good candidates for several acute and chronic neuronal disorders that involve the overproduction of ROS such as PD [87], Huntington's disease (HD) [88], Alzheimer's disease (AD) [89], ALS [90], stroke [91], and trauma [92].

PD is a chronic neurodegenerative disorder where a large body of literature supports a role for ROS and oxidative stress and neuronal loss in the substantia nigra. Much of this data is linked to the mitochondrial dysfunction of complex I associated with this disease [93]. There have been numerous reports of increases in lipid, DNA and protein oxidative changes in PD and animal models of PD [94]. In addition, there are reports of decreases in antioxidants such as GSH, increased iron content and activation of NFkB [95]. Toxins that increase ROS in the substantia nigra such as 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) also produce PD-like symptoms and pathology in man and animals [96]. Catalytic antioxidants with H_2O_2 scavenging activities are effective in animal models of PD [22,97]. Recent studies with the orally active metalloporphyrin AEOL 11207 demonstrated neuroprotection in a mouse MPTP model that was also associated with decreased oxidative stress [22]. Currently available PD therapies are largely limited to palliative treatments with unpleasant side effects and opens PD as an attractive target for the development of catalytic antioxidants.

A number of dementias, such as AD and HD, are memory disorders that have been associated with protein aggregation and increased oxidative stress in the cortex and hippocampus [98]. Some investigators have suggested that amyloid may increase H_2O_2 production through metalcatalyzed reactions [99]. A few studies have shown that catalytic antioxidants can protect neurons from amyloid-induced cytotoxicity [100]. Increased production of ROS and RNS may also be an indirect consequence of amyloid protein deposits stimulating an inflammatory response from microglial cells [101]. In addition, there is some evidence that NO plays a role in memory and that catalytic antioxidants can be protective [102]. Aging is also a factor in the loss of cognitive function and catalytic antioxidants have also shown to be effective at slowing this process [103]. These data suggest that dementias may be a fruitful area for the further development of catalytic anti-oxidants.

ALS is a motor neuron disorder that leads to progressive loss of motor function, muscle atrophy and death within a few years. Most cases of ALS are sporadic, but about 10% are familial and some of these are associated with genetic mutations. Some familial forms of ALS are associated with mutations in SOD1, and transgenic mice overexpressing one of the SOD1 mutations develop a progressive degenerative disease of motor neurons [104]. It is thought that there is a gain of function associated with the mutant SOD1 that leads to the motor neuron loss in these transgenic mice, and the mechanisms by which this occurs are highly debated. The importance of oxidative stress in ALS is also highly debated even though ample evidence exists that it occurs in this disease [90]. A few studies have shown that catalytic antioxidants with H_2O_2 scavenging activities can prolong survival in the SOD1 mutant transgenic mice [105,106]. AEOL 10150 has completed phase I safety testing in ALS patients.

Stroke is a leading cause of death in humans with few treatment options. Stroke is an acute neurodegenerative condition that often involves tissue injury due to IR events. Stroke is associated with increased ROS production, and injury can be enhanced or attenuated by modulation of endogenous antioxidants [107]. A number of catalytic antioxidants with H_2O_2 scavenging activities are effective in animal vessel occlusion models of stroke [72–74]. Catalytic antioxidants are also effective in attenuating cerebral vasoconstriction that is commonly associated with hemorrhagic stroke and migraines [108,109]. Ebselen has had some success in human phase II clinical trials [75] and is currently being evaluated in human phase III clinical trials for stroke in Japan sponsored by Daiichi Sankyo Pharmaceuticals. These studies suggest that stroke may be a potential target for catalytic antioxidants.

7. Implications for the use of catalytic antioxidants as modulators of human disease

The postulated role of ROS as a terminal mediator of tissue injury and dysfunction in diseases of diverse etiologies emphasizes the wide range of therapeutic opportunities for catalytic antioxidant development. Pathologies that are most likely to benefit from catalytic antioxidant therapy include conditions in which a clear role for ROS has been established. Inflammation, which is a pivotal etiological factor in many human pathophysiologic processes involving multiple organ systems, is a major therapeutic opportunity for catalytic antioxidant development given the increasingly important role ROS are being given in modulating cell signaling pathways. The role of ROS has been well documented in host defense mechanisms involving phagocytosis, cytokines, chemokines and immune complex formation, all of which can contribute to auto-immune disorders. Inflammatory lung, intestinal, and cardiovascular diseases all are potentially important targets for catalytic antioxidant therapy.

An important unaddressed issue in the development of catalytic antioxidants is the mechanism by which these agents diminish oxidative stress and injury in animals and humans. To date, catalytic antioxidants are defined by their chemistry under highly defined and largely non-

biological conditions. The relevance of using this classification system is still yet to be verified. In fact, over time the potential number of mechanisms these agents possess in biological systems has increased substantially. A more important fact is that these compounds have potent biological effects that often track with their ability to suppress oxidative stress in a large variety of animal models of human disease. However, definitive proof that these compounds are effective in human disease is still needed.

8. Conclusion

Emerging research is strengthening the role of ROS in redox-mediated cell signaling pathways that have well-established roles in human disease and aging. A novel class of compounds that efficiently scavenge cellular peroxides are being developed to modulate cellular ROS and alter some of the aberrant cell signaling associated with many human disorders. There is promising data in the literature that these compounds may have potential therapeutic use in pulmonary, cardiovascular, neurodegenerative and inflammatory disorders.

Acknowledgements

AP-1

The author wishes to thank Dr. Carl White for his helpful comments and suggestions. This work was supported in part by NIH grants HL075523, HL084469, ES012504, ES015678 and an unrestricted research grant from Aeolus Pharmaceuticals. Dr. Day is a consultant for and holds equity in Aeolus Pharmaceuticals that is commercially developing metalloporphyrins as therapeutic agents.

Abbreviations

References

- 1. Kamata H, Hirata H. Redox regulation of cellular signalling. Cell Signal 1999;11(1):1–14. [PubMed: 10206339]
- 2. Deisseroth A, Dounce AL. Catalase: physical and chemical properties, mechanism of catalysis, and physiological role. Physiol Rev 1970;50(3):319–75. [PubMed: 4912904]
- 3. Oshino N, Oshino R, Chance B. The characteristics of the "peroxidatic" reaction of catalase in ethanol oxidation. Biochem J 1973;131(3):555–63. [PubMed: 4720713]
- 4. Warner HR. Superoxide dismutase, aging, and degenerative disease. Free Radic Biol Med 1994;17(3): 249–58. [PubMed: 7982630]
- 5. Zachara BA. Mammalian selenoproteins. J Trace Elem Electrolytes Health Dis 1992;6(3):137–51. [PubMed: 1483033]
- 6. Mirault ME, Tremblay A, Furling D, Trepanier G, Dugre F, Puymirat J, et al. Transgenic glutathione peroxidase mouse models for neuroprotection studies. Ann N Y Acad Sci 1994;738:104–15. [PubMed: 7832420]
- 7. Comhair SA, Erzurum SC. The regulation and role of extracellular glutathione peroxidase. Antioxid Redox Signal 2005;7(1–2):72–9. [PubMed: 15650397]
- 8. Kettle AJ, van Dalen CJ, Winterbourn CC. Peroxynitrite and myeloperoxidase leave the same footprint in protein nitration. Redox Rep 1997;3(5–6):257–8. [PubMed: 9754322]
- 9. Rhee SG, Chae HZ, Kim K. Peroxiredoxins: a historical overview and speculative preview of novel mechanisms and emerging concepts in cell signalling. Free Radic Biol Med 2005;38(12):1543–52. [PubMed: 15917183]
- 10. Chae HZ, Kang SW, Rhee SG. Isoforms of mammalian peroxiredoxin that reduce peroxides in presence of thioredoxin. Methods Enzymol 1999;300:219–26. [PubMed: 9919524]
- 11. Kim HS, Manevich Y, Feinstein SI, Pak JH, Ho YS, Fisher AB. Induction of 1-cys peroxiredoxin expression by oxidative stress in lung epithelial cells. Am J Physiol Lung Cell Mol Physiol 2003;285 (2):L363–9. [PubMed: 12851211]
- 12. Wang Y, Manevich Y, Feinstein SI, Fisher AB. Adenovirus-mediated transfer of the 1-cys peroxiredoxin gene to mouse lung protects against hyperoxic injury. Am J Physiol Lung Cell Mol Physiol 2004;286(6):L1188–93. [PubMed: 15136296]
- 13. Forman HJ, Torres M. Reactive oxygen species and cell signaling: respiratory burst in macrophage signalling. Am J Respir Crit Care Med 2002;166(12 Part 2):S4–8. [PubMed: 12471082]
- 14. Chance B, Sies H, Boveris A. Hydroperoxide metabolism in mammalian organs. Physiol Rev 1979;59 (3):527–605. [PubMed: 37532]
- 15. Pasternack RF, Skowronek WR. Catalysis of the disproportionation of superoxide by metalloporphyrins. J Inorg Biochem 1979;11:261–7. [PubMed: 229199]
- 16. Day BJ, Fridovich I, Crapo JD. Manganic porphyrins possess catalase activity and protect endothelial cells against hydrogen peroxide-mediated injury. Arch Biochem Biophys 1997;347(2):256–62. [PubMed: 9367533]
- 17. Szabo C, Day BJ, Salzman AL. Evaluation of the relative contribution of nitric oxide and peroxynitrite to the suppression of mitochondrial respiration in immunostimulated macrophages using a

manganese mesoporphyrin superoxide dismutase mimetic and peroxynitrite scavenger. Fed Eur Biochem Soc Lett 1996;381(1–2):82–6.

- 18. Day BJ, Batinic-Haberle I, Crapo JD. Metalloporphyrins are potent inhibitors of lipid peroxidation. Free Radic Biol Med 1999;26(5–6):730–6. [PubMed: 10218663]
- 19. Kachadourian R, Johnson CA, Min E, Spasojevic I, Day BJ. Flavin-dependent antioxidant properties of a new series of meso-N,N ′-dialkyl-imidazolium substituted manganese(III) porphyrins. Biochem Pharmacol 2004;67(1):77–85. [PubMed: 14667930]
- 20. Castello PR, Drechsel DA, Day BJ, Patel M. Inhibition of mitochondrial hydrogen peroxide production by lipophilic metalloporphyrins. J Pharmacol Exp Ther 2008;324(3):970–6. [PubMed: 18063723]
- 21. Day BJ. Catalytic antioxidants: a radical approach to new therapeutics. Drug Discov Today 2004;9 (13):557–66. [PubMed: 15203091]
- 22. Liang LP, Huang J, Fulton R, Day BJ, Patel M. An orally active catalytic metalloporphyrin protects against 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine neurotoxicity in vivo. J Neurosci 2007;27(16): 4326–33. [PubMed: 17442816]
- 23. Doctrow SR, Huffman K, Marcus CB, Musleh W, Bruce A, Baudry M, et al. Salen-manganese complexes: combined superoxide dismutase/catalase mimics with broad pharmacological efficacy. Adv Pharmacol 1997;38:247–69. [PubMed: 8895812]
- 24. Doctrow SR, Huffman K, Marcus CB, Tocco G, Malfroy E, Adinolfi CA, et al. Salen-manganese complexes as catalytic scavengers of hydrogen peroxide and cytoprotective agents: structure-activity relationship studies. J Med Chem 2002;45(20):4549–58. [PubMed: 12238934]
- 25. Sicking W, Korth HG, Jansen G, de Groot H, Sustmann R. Hydrogen peroxide decomposition by a non-heme iron(III) catalase mimic: a DFT study. Chemistry 2007;13(15):4230–45. [PubMed: 17323385]
- 26. Rauen U, Li T, Sustmann R, De Groot H. Protection against iron- and hydrogen peroxide-dependent cell injuries by a novel synthetic iron catalase mimic and its precursor, the iron-free ligand. Free Radic Biol Med 2004;37(9):1369–83. [PubMed: 15454276]
- 27. Moreno D, Palopoli C, Daier V, Shova S, Vendier L, Sierra MG, et al. Synthesis, structure and catalase-like activity of dimanganese(III) complexes of 1,5-bis(X-salicylidenamino)pentan-3-ol (X = 3- and 5-methyl). Influence of phenyl-ring substituents on catalytic activity. Dalton Trans 2006;43:5156–66. [PubMed: 17077889]
- 28. Sies H. Ebselen, a selenoorganic compound as glutathione peroxidase mimic. Free Radic Biol Med 1993;14(3):313–23. [PubMed: 8458589]
- 29. Biewenga GP, Bast A. Reaction of lipoic acid with ebselen and hypochlorous acid. Methods Enzymol 1995;251:303–14. [PubMed: 7651210]
- 30. Scurlock R, Rougee M, Bensasson RV, Evers M, Dereu N. Deactivation of singlet molecular oxygen by organo-selenium compounds exhibiting glutathione peroxidase activity and by sulfur-containing homologs. Photochem Photobiol 1991;54(5):733–6. [PubMed: 1798750]
- 31. Masumoto H, Sies H. The reaction of ebselen with peroxynitrite. Chem Res Toxicol 1996;9(1):262– 7. [PubMed: 8924601]
- 32. Safayhi H, Tiegs G, Wendel A. A novel biologically active seleno-organic compound–V. Inhibition by ebselen (PZ 51) of rat peritoneal neutrophil lipoxygenase. Biochem Pharmacol 1985;34(15):2691– 4. [PubMed: 2990494]
- 33. Cotgreave IA, Duddy SK, Kass GE, Thompson D, Moldeus P. Studies on the anti-inflammatory activity of ebselen. Ebselen interferes with granulocyte oxidative burst by dual inhibition of NADPH oxidase and protein kinase C? Biochem Pharmacol 1989;38(4):649–56. [PubMed: 2537084]
- 34. Zembowicz A, Hatchett RJ, Radziszewski W, Gryglewski RJ. Inhibition of endothelial nitric oxide synthase by ebselen, Prevention by thiols suggests the inactivation by ebselen of a critical thiol essential for the catalytic activity of nitric oxide synthase. J Pharmacol Exp Ther 1993;267(3):1112– 8. [PubMed: 7505326]
- 35. Moutet M, d'Alessio P, Malette P, Devaux V, Chaudiere J. Glutathione peroxidase mimics prevent TNFalpha- and neutrophil-induced endothelial alterations. Free Radic Biol Med 1998;25(3):270–81. [PubMed: 9680172]

- 36. Huang X, Dong Z, Liu J, Mao S, Xu J, Luo G, et al. Selenium-mediated micellar catalyst: an efficient enzyme model for glutathione peroxidase-like catalysis. Langmuir 2007;23(3):1518–22. [PubMed: 17241082]
- 37. Adams WJ Jr, Kocsis JJ, Snyder R. Acute toxicity and urinary excretion of diphenyldiselenide. Toxicol Lett 1989;48(3):301–10. [PubMed: 2781599]
- 38. Rosa RM, Sulzbacher K, Picada JN, Roesler R, Saffi J, Brendel M, et al. Genotoxicity of diphenyl diselenide in bacteria and yeast. Mutat Res 2004;563(2):107–15. [PubMed: 15364277]
- 39. Liu J, Luo G, Ren X, Mu Y, Bai Y, Shen J. A bis-cyclodextrin diselenide with glutathione peroxidaselike activity. Biochim Biophys Acta 2000;1481(2):222–8. [PubMed: 11018712]
- 40. Sun Y, Mu Y, Ma S, Gong P, Yan G, Liu J, et al. The molecular mechanism of protecting cells against oxidative stress by 2-selenium-bridged beta-cyclodextrin with glutathione peroxidase activity. Biochim Biophys Acta 2005;1743(3):199–204. [PubMed: 15843033]
- 41. Bell IM, Fisher ML, Wu ZP, Hilvert D. Kinetic studies on the peroxidase activity of selenosubtilisin. Biochemistry 1993;32(14):3754–62. [PubMed: 8385489]
- 42. Sun Y, Li T, Chen H, Zhang K, Zheng K, Mu Y, et al. Selenium-containing 15-mer peptides with high glutathione peroxidase-like activity. J Biol Chem 2004;279(36):37235–40. [PubMed: 15148324]
- 43. Ren X, Gao S, You D, Huang H, Liu Z, Mu Y, et al. Cloning and expression of a single-chain catalytic antibody that acts as a glutathione peroxidase mimic with high catalytic efficiency. Biochem J 2001;359(Part 2):369–74. [PubMed: 11583583]
- 44. Ding L, Liu Z, Zhu Z, Luo G, Zhao D, Ni J. Biochemical characterization of selenium-containing catalytic antibody as a cytosolic glutathione peroxidase mimic. Biochem J 1998;332(Part 1):251–5. [PubMed: 9576875]
- 45. Kahl R, Kampkotter A, Watjen W, Chovolou Y. Antioxidant enzymes and apoptosis. Drug Metab Rev 2004;36(3–4):747–62. [PubMed: 15554245]
- 46. Hildeman DA, Mitchell T, Aronow B, Wojciechowski S, Kappler J, Marrack P. Control of Bcl-2 expression by reactive oxygen species. Proc Natl Acad Sci U S A 2003;100(25):15035–40. [PubMed: 14657380]
- 47. Muller JM, Rupec RA, Baeuerle PA. Study of gene regulation by NF-kappa B and AP-1 in response to reactive oxygen intermediates. Methods 1997;11(3):301–12. [PubMed: 9073573]
- 48. Caselli A, Marzocchini R, Camici G, Manao G, Moneti G, Pieraccini G, et al. The inactivation mechanism of low molecular weight phosphotyrosine-protein phosphatase by H2O2. J Biol Chem 1998;273(49):32554–60. [PubMed: 9829991]
- 49. Rahman I, Biswas SK, Jimenez LA, Torres M, Forman HJ. Glutathione, stress responses, and redox signaling in lung inflammation. Antioxid Redox Signal 2005;7(1–2):42–59. [PubMed: 15650395]
- 50. Chang LY, Crapo JD. Inhibition of airway inflammation and hyperreactivity by an antioxidant mimetic. Free Radic Biol Med 2002;33(3):379–86. [PubMed: 12126760]
- 51. Reynaert NL, Aesif SW, McGovern T, Brown A, Wouters EF, Irvin CG, et al. Catalase overexpression fails to attenuate allergic airways disease in the mouse. J Immunol 2007;178(6):3814–21. [PubMed: 17339480]
- 52. Pryor WA, Dooley MM, Church DF. The inactivation of alpha-1-proteinase inhibitor by gas-phase cigarette smoke: protection by antioxidants and reducing species. Chem Biol Interact 1986;57(3): 271–83. [PubMed: 3486047]
- 53. Mendez-Alvarez E, Soto-Otero R, Sanchez-Sellero I, Lopez-Rivadulla Lamas M. In vitro inhibition of catalase activity by cigarette smoke: relevance for oxidative stress. J Appl Toxicol 1998;18(6): 443–8. [PubMed: 9840751]
- 54. Luchese C, Stangherlin EC, Ardais AP, Nogueira CW, Santos FW. Diphenyl diselenide prevents oxidative damage induced by cigarette smoke exposure in lung of rat pups. Toxicology 2007;230(2– 3):189–96. [PubMed: 17178183]
- 55. Smith KR, Uyeminami DL, Kodavanti UP, Crapo JD, Chang LY, Pinkerton KE. Inhibition of tobacco smoke-induced lung inflammation by a catalytic antioxidant. Free Radic Biol Med 2002;33(8):1106– 14. [PubMed: 12374622]

- 56. Ledwozyw A. Protective effect of liposome-entrapped superoxide dismutase and catalase on bleomycin-induced lung injury in rats. I Antioxidant enzyme activities and lipid peroxidation. Acta Vet Hung 1991;39(3–4):215–24. [PubMed: 1723845]
- 57. Oury TD, Thakker K, Menache M, Chang LY, Crapo JD, Day BJ. Attenuation of bleomycin-induced pulmonary fibrosis by a catalytic antioxidant metalloporphyrin. Am J Respir Cell Mol Biol 2001;25 (2):164–9. [PubMed: 11509325]
- 58. Machtay M, Scherpereel A, Santiago J, Lee J, McDonough J, Kinniry P, et al. Systemic polyethylene glycol-modified (PEGylated) superoxide dismutase and catalase mixture attenuates radiation pulmonary fibrosis in the C57/bl6 mouse. Radiother Oncol 2006;81(2):196–205. [PubMed: 17069914]
- 59. Vujaskovic Z, Batinic-Haberle I, Rabbani ZN, Feng QF, Kang SK, Spasojevic I, et al. A small molecular weight catalytic metalloporphyrin antioxidant with superoxide dismutase (SOD) mimetic properties protects lungs from radiation-induced injury. Free Radic Biol Med 2002;33(6):857–63. [PubMed: 12208373]
- 60. Rabbani ZN, Batinic-Haberle I, Anscher MS, Huang J, Day BJ, Alexander E, et al. Long-term administration of a small molecular weight catalytic metalloporphyrin antioxidant, AEOL 10150, protects lungs from radiation-induced injury. Int J Radiat Oncol Biol Phys 2007;67(2):573–80. [PubMed: 17236973]
- 61. Langan AR, Khan MA, Yeung IW, Van Dyk J, Hill RP. Partial volume rat lung irradiation: the protective/mitigating effects of Eukarion-189, a superoxide dismutase-catalase mimetic. Radiother Oncol 2006;79(2):231–8. [PubMed: 16675053]
- 62. White CW, Jackson JH, Abuchowski A, Kazo GM, Mimmack RF, Berger EM, et al. Polyethylene glycol-attached antioxidant enzymes decrease pulmonary oxygen toxicity in rats. J Appl Physiol 1989;66(2):584–90. [PubMed: 2540139]
- 63. Olson NC, Grizzle MK, Anderson DL. Effect of polyethylene glycol-superoxide dismutase and catalase on endotoxemia in pigs. J Appl Physiol 1987;63(4):1526–32. [PubMed: 2826378]
- 64. Chang LY, Subramaniam M, Yoder BA, Day BJ, Ellison MC, Sunday ME, et al. A catalytic antioxidant attenuates alveolar structural remodeling in bronchopulmonary dysplasia. Am J Respir Crit Care Med 2003;167(1):57–64. [PubMed: 12502477]
- 65. Bowler RP, Arcaroli J, Abraham E, Patel M, Chang LY, Crapo JD. Evidence for extracellular superoxide dismutase as a mediator of hemorrhage-induced lung injury. Am J Physiol Lung Cell Mol Physiol 2003;284(4):L680–7. [PubMed: 12618426]
- 66. Gonzalez PK, Zhuang J, Doctrow SR, Malfroy B, Benson PF, Menconi MJ, et al. Role of oxidant stress in the adult respiratory distress syndrome: evaluation of a novel antioxidant strategy in a porcine model of endotoxin-induced acute lung injury. Shock 1996;6(Suppl 1):S23–6. [PubMed: 8828094]
- 67. Izumi M, McDonald MC, Sharpe MA, Chatterjee PK, Thiemermann C. Superoxide dismutase mimetics with catalase activity reduce the organ injury in hemorrhagic shock. Shock 2002;18(3): 230–5. [PubMed: 12353923]
- 68. Blum S, Asaf R, Guetta J, Miller-Lotan R, Asleh R, Kremer R, et al. Haptoglobin genotype determines myocardial infarct size in diabetic mice. J Am Coll Cardiol 2007;49(1):82–7. [PubMed: 17207726]
- 69. Lubbers NL, Polakowski JS, Crapo JD, Wegner CD, Cox BF. Preischemic and postischemic administration of AEOL10113 reduces infarct size in a rat model of myocardial ischemia and reperfusion. J Cardiovasc Pharmacol 2003;41(5):714–9. [PubMed: 12717101]
- 70. Hines IN, Hoffman JM, Scheerens H, Day BJ, Harada H, Pavlick KP, et al. Regulation of postischemic liver injury following different durations of ischemia. Am J Physiol Gastrointest Liver Physiol 2003;284(3):G536–45. [PubMed: 12444015]
- 71. Ozaki M, Nakamura M, Teraoka S, Ota K. Ebselen, a novel anti-oxidant compound, protects the rat liver from ischemia-reperfusion injury. Transpl Int 1997;10(2):96–102. [PubMed: 9089992]
- 72. Sheng H, Enghild JJ, Bowler R, Patel M, Batinic-Haberle I, Calvi CL, et al. Effects of metalloporphyrin catalytic antioxidants in experimental brain ischemia. Free Radic Biol Med 2002;33(7):947–61. [PubMed: 12361805]
- 73. Baker K, Marcus CB, Huffman K, Kruk H, Malfroy B, Doctrow SR. Synthetic combined superoxide dismutase/catalase mimetics are protective as a delayed treatment in a rat stroke model: a key role

for reactive oxygen species in ischemic brain injury. J Pharmacol Exp Ther 1998;284(1):215–21. [PubMed: 9435181]

- 74. Imai H, Masayasu H, Dewar D, Graham DI, Macrae IM. Ebselen protects both gray and white matter in a rodent model of focal cerebral ischemia. Stroke 2001;32(9):2149–54. [PubMed: 11546910]
- 75. Yamaguchi T, Sano K, Takakura K, Saito I, Shinohara Y, Asano T, et al. Ebselen in acute ischemic stroke: a placebo-controlled, double-blind clinical trial. Ebselen Study Group Stroke 1998;29(1):12– 7.
- 76. Chatterjee PK, Patel NS, Kvale EO, Brown PA, Stewart KN, Mota-Filipe H, et al. EUK-134 reduces renal dysfunction and injury caused by oxidative and nitrosative stress of the kidney. Am J Nephrol 2004;24(2):165–77. [PubMed: 14752229]
- 77. Rajagopalan S, Kurz S, Munzel T, Tarpey M, Freeman BA, Griendling KK, et al. Angiotensin IImediated hypertension in the rat increases vascular superoxide production via membrane NADH/ NADPH oxidase activation. Contribution to alterations of vasomotor tone. J Clin Invest 1996;97(8): 1916–23. [PubMed: 8621776]
- 78. Bagchi D, Prasad R, Das DK. Direct scavenging of free radicals by captopril, an angiotensin converting enzyme inhibitor. Biochem Biophys Res Commun 1989;158(1):52–7. [PubMed: 2536279]
- 79. Kastenbauer S, Koedel U, Becker BF, Pfister HW. Pneumococcal meningitis in the rat: evaluation of peroxynitrite scavengers for adjunctive therapy. Eur J Pharmacol 2002;449(1–2):177–81. [PubMed: 12163122]
- 80. Sui H, Wang W, Wang PH, Liu LS. Effect of glutathione peroxidase mimic ebselen (PZ51) on endothelium and vascular structure of stroke-prone spontaneously hypertensive rats. Blood Press 2005;14(6):366–72. [PubMed: 16403691]
- 81. McDonald MC, d'Emmanuele di Villa Bianca R, Wayman NS, Pinto A, Sharpe MA, Cuzzocrea S, et al. A superoxide dismutase mimetic with catalase activity (EUK-8) reduces the organ injury in endotoxic shock. Eur J Pharmacol 2003;466(1–2):181–9. [PubMed: 12679155]
- 82. Wang W, Jittikanont S, Falk SA, Li P, Feng L, Gengaro PE, et al. Interaction among nitric oxide, reactive oxygen species, and antioxidants during endotoxemia-related acute renal failure. Am J Physiol Renal Physiol 2003;284(3):F532–7. [PubMed: 12556364]
- 83. van Empel VP, Bertrand AT, van Oort RJ, van der Nagel R, Engelen M, van Rijen HV, et al. EUK-8, a superoxide dismutase and catalase mimetic, reduces cardiac oxidative stress and ameliorates pressure overload-induced heart failure in the harlequin mouse mutant. J Am Coll Cardiol 2006;48 (4):824–32. [PubMed: 16904556]
- 84. Nin N, Cassina A, Boggia J, Alfonso E, Botti H, Peluffo G, et al. Septic diaphragmatic dysfunction is prevented by Mn(III)porphyrin therapy and inducible nitric oxide synthase inhibition. Intensive Care Med 2004;30(12):2271–8. [PubMed: 15349724]
- 85. Wendel A, Tiegs G. A novel biologically active seleno-organic compound—VI. Protection by ebselen (PZ 51) against galactosamine/endotoxin-induced hepatitis in mice. Biochem Pharmacol 1986;35 (13):2115–8. [PubMed: 3729968]
- 86. Bianca RV, Wayman NS, McDonald MC, Pinto A, Shape MA, Chatterjee PK, et al. Superoxide dismutase mimetic with catalase activity, EUK-134, attenuates the multiple organ injury and dysfunction caused by endotoxin in the rat. Med Sci Monit 2002;8(1):BR1–7. [PubMed: 11796957]
- 87. Przedborski S, Ischiropoulos H. Reactive oxygen and nitrogen species: weapons of neuronal destruction in models of Parkinson's disease. Antioxid Redox Signal 2005;7(5–6):685–93. [PubMed: 15890013]
- 88. Browne SE, Beal MF. Oxidative damage in Huntington's disease pathogenesis. Antioxid Redox Signal 2006;8(11–12):2061–73. [PubMed: 17034350]
- 89. Forero DA, Casadesus G, Perry G, Arboleda H. Synaptic dysfunction and oxidative stress in Alzheimer's disease: emerging mechanisms. J Cell Mol Med 2006;10(3):796–805. [PubMed: 16989739]
- 90. Barber SC, Mead RJ, Shaw PJ. Oxidative stress in ALS: a mechanism of neurodegeneration and a therapeutic target. Biochim Biophys Acta 2006;1762(11–12):1051–67. [PubMed: 16713195]
- 91. Crack PJ, Taylor JM. Reactive oxygen species and the modulation of stroke. Free Radic Biol Med 2005;38(11):1433–44. [PubMed: 15890617]

- 92. Shohami E, Beit-Yannai E, Horowitz M, Kohen R. Oxidative stress in closed-head injury: brain antioxidant capacity as an indicator of functional outcome. J Cereb Blood Flow Metab 1997;17(10): 1007–19. [PubMed: 9346425]
- 93. Mortiboys HJ, Schaefer J, Reichmann H, Jackson S. Mitochondrial dysfunction in Parkinson's disease —revisited. Neurol Neurochir Pol 2007;41(2):150–9. [PubMed: 17530578]
- 94. Przedborski S, Vila M. The 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model: a tool to explore the pathogenesis of Parkinson's disease. Ann N Y Acad Sci 2003;991:189–98. [PubMed: 12846987]
- 95. Bharath S, Hsu M, Kaur D, Rajagopalan S, Andersen JK. Glutathione, iron and Parkinson's disease. Biochem Pharmacol 2002;64(5–6):1037–48. [PubMed: 12213603]
- 96. Sun F, Kanthasamy A, Anantharam V, Kanthasamy AG. Environmental neurotoxic chemicalsinduced ubiquitin proteasome system dysfunction in the pathogenesis and progression of Parkinson's disease. Pharmacol Ther 2007;114(3):327–44. [PubMed: 17521740]
- 97. Moussaoui S, Obinu MC, Daniel N, Reibaud M, Blanchard V, Imperato A. The antioxidant ebselen prevents neurotoxicity and clinical symptoms in a primate model of Parkinson's disease. Exp Neurol 2000;166(2):235–45. [PubMed: 11085889]
- 98. Moreira PI, Honda K, Liu Q, Santos MS, Oliveira CR, Aliev G, et al. Oxidative stress: the old enemy in Alzheimer's disease pathophysiology. Curr Alzheimer Res 2005;2(4):403–8. [PubMed: 16248845]
- 99. Huang X, Atwood CS, Hartshorn MA, Multhaup G, Goldstein LE, Scarpa RC, et al. The A beta peptide of Alzheimer's disease directly produces hydrogen peroxide through metal ion reduction. Biochemistry 1999;38(24):7609–16. [PubMed: 10386999]
- 100. Bruce AJ, Malfroy B, Baudry M. beta-Amyloid toxicity in organotypic hippocampal cultures: protection by EUK-8, a synthetic catalytic free radical scavenger. Proc Natl Acad Sci U S A 1996;93 (6):2312–6. [PubMed: 8637869]
- 101. Yan SD, Chen X, Fu J, Chen M, Zhu H, Roher A, et al. RAGE and amyloid-beta peptide neurotoxicity in Alzheimer's disease. Nature 1996;382(6593):685–91. [PubMed: 8751438]
- 102. Klann E. Cell-permeable scavengers of superoxide prevent long-term potentiation in hippocampal area CA1. J Neurophysiol 1998;80(1):452–7. [PubMed: 9658063]
- 103. Liu R, Liu IY, Bi X, Thompson RF, Doctrow SR, Malfroy B, et al. Reversal of age-related learning deficits and brain oxidative stress in mice with superoxide dismutase/catalase mimetics. Proc Natl Acad Sci U S A 2003;100(14):8526–31. [PubMed: 12815103]
- 104. Tu PH, Raju P, Robinson KA, Gurney ME, Trojanowski JQ, Lee VM. Transgenic mice carrying a human mutant superoxide dismutase transgene develop neuronal cytoskeletal pathology resembling human amyotrophic lateral sclerosis lesions. Proc Natl Acad Sci U S A 1996;93(7):3155–60. [PubMed: 8610185]
- 105. Crow JP, Calingasan NY, Chen J, Hill JL, Beal MF. Manganese porphyrin given at symptom onset markedly extends survival of ALS mice. Ann Neurol 2005;58(2):258–65. [PubMed: 16049935]
- 106. Jung C, Rong Y, Doctrow S, Baudry M, Malfroy B, Xu Z. Synthetic superoxide dismutase/catalase mimetics reduce oxidative stress and prolong survival in a mouse amyotrophic lateral sclerosis model. Neurosci Lett 2001;304(3):157–60. [PubMed: 11343826]
- 107. Chan PH, Epstein CJ, Li Y, Huang TT, Carlson E, Kinouchi H, et al. Transgenic mice and knockout mutants in the study of oxidative stress in brain injury. J Neurotrauma 1995;12(5):815–24. [PubMed: 8594209]
- 108. Niwa K, Carlson GA, Iadecola C. Exogenous A beta1-40 reproduces cerebrovascular alterations resulting from amyloid precursor protein overexpression in mice. J Cereb Blood Flow Metab 2000;20(12):1659–68. [PubMed: 11129782]
- 109. Aladag MA, Turkoz Y, Sahna E, Parlakpinar H, Gul M. The attenuation of vasospasm by using a sod mimetic after experimental subarachnoidal haemorrhage in rats. Acta Neurochir (Wien) 2003;145(8):673–7. [PubMed: 14520547]
- 110. Xu Y, Armstrong SJ, Arenas IA, Pehowich DJ, Davidge ST. Cardioprotection by chronic estrogen or superoxide dismutase mimetic treatment in the aged female rat. Am J Physiol Heart Circ Physiol 2004;287(1):H165–71. [PubMed: 14988070]

- 111. Melov S, Schneider JA, Day BJ, Hinerfeld D, Coskun P, Mirra SS, et al. A novel neurological phenotype in mice lacking mitochondrial manganese superoxide dismutase. Nat Genet 1998;18(2): 159–63. [PubMed: 9462746][see comments]
- 112. Tong JJ, Schriner SE, McCleary D, Day BJ, Wallace DC. Life extension through neurofibromin mitochondrial regulation and antioxidant therapy for neurofibromatosis-1 in *Drosophila melanogaster*. Nat Genet 2007;39(4):476–85. [PubMed: 17369827]
- 113. Leski ML, Bao F, Wu L, Qian H, Sun D, Liu D. Protein and DNA oxidation in spinal injury: neurofilaments—an oxidation target. Free Radic Biol Med 2001;30(6):613–24. [PubMed: 11295359]
- 114. Kalayci M, Coskun O, Cagavi F, Kanter M, Armutcu F, Gul S, et al. Neuroprotective effects of ebselen on experimental spinal cord injury in rats. Neurochem Res 2005;30(3):403–10. [PubMed: 16018585]
- 115. Sheng H, Spasojevic I, Warner DS, Batinic-Haberle I. Mouse spinal cord compression injury is ameliorated by intrathecal cationic manganese(III) porphyrin catalytic antioxidant therapy. Neurosci Lett 2004;366(2):220–5. [PubMed: 15276251]
- 116. Malassagne B, Ferret PJ, Hammoud R, Tulliez M, Bedda S, Trebeden H, et al. The superoxide dismutase mimetic MnTBAP prevents Fas-induced acute liver failure in the mouse. Gastroenterology 2001;121(6):1451–9. [PubMed: 11729124]
- 117. Kono H, Arteel GE, Rusyn I, Sies H, Thurman RG. Ebselen prevents early alcohol-induced liver injury in rats. Free Radic Biol Med 2001;30(4):403–11. [PubMed: 11182296]
- 118. Wasser S, Lim GY, Ong CN, Tan CE. Anti-oxidant ebselen causes the resolution of experimentally induced hepatic fibrosis in rats. J Gastroenterol Hepatol 2001;16(11):1244–53. [PubMed: 11903743]
- 119. Zhang HJ, Doctrow SR, Oberley LW, Kregel KC. Chronic antioxidant enzyme mimetic treatment differentially modulates hyperthermia-induced liver HSP70 expression with aging. J Appl Physiol 2006;100(4):1385–91. [PubMed: 16254069]
- 120. Choudhary S, Keshavarzian A, Yong S, Wade M, Bocckino S, Day BJ, et al. Novel antioxidants zolimid and AEOL11201 ameliorate colitis in rats. Dig Dis Sci 2001;46(10):2222–30. [PubMed: 11680601]

NIH-PA Author Manuscript

Fig. 1.

Endogenous scavenging of cellular peroxide. The cell's steady-state peroxide (ROOH) levels are largely maintained by the activities of catalase, glutathione peroxidases (GPx) and the thioredoxin-assisted peroxidases (peroxiredoxins, Prx). This system also maintains cellular protein cysteines in a reduced (red) state. During oxidative stress, where cellular peroxides are elevated, there is an increased level of oxidized (ox) protein cysteines that leads to inactivation of phosphatases and transcription factors and dysregulated inflammatory reactions. This is the drug target and rationale for the development of catalase and GPx mimics.

Fig. 2.

Examples of catalase-like mimics chemical structures: (A) metalloporphyrins; (B) salens; and (C) other metal complexes.

Examples of glutathione peroxidase-like mimics chemical structures: (A) mono-selenium mimics; and (B) di-selenium mimics.

Table 1

Examples of catalytic antioxidants with H_2O_2 -scavenging activity that are effective in attenuating oxidative stress in *in vivo* models..

Model system	Species	Active site	Compound(s)	Reference
Pulmonary				
Bleomycin fibrosis	Mice	Mn	AEOL 10201	$[57]$
Radiation fibrosis	Rats	Mn	AEOL 10113	$[59]$
		Mn	AEOL 10150	$[60]$
		Mn	EUK 189	$[61]$
Cigarette smoke	Rats	Se	DPDS	$[54]$
		Mn	AEOL 10150	$[55]$
Endotoxin	Pigs	Mn	EUK 8	[66]
Bronchopulmonary dysplasia	Preterm baboons	Mn	AEOL 10113	$[64]$
Antigen-induced asthma	Mice	Mn	AEOL 10113	$[50]$
Hemorrhage	SOD3 KO mice	Mn	AEOL 10150	$[65]$
Cardiovascular				
Heart ischemia/reperfusion	Mice	Se	BXT 51072	[68]
	Aged rats	Mn	EUK 8	$[110]$
	Rats	Mn	AEOL 10113	$[69]$
Dilated cardiomyopathy	SOD2 KO mice	Mn	AEOL 10201	$[111]$
	Harlequin mutant mice	Mn	EUK 8	$[83]$
Sepsis	Rats	Mn	AEOL 10113	$[84]$
		Mn	EUK 8	[81]
Hemorrhage	Rats	Mn	EUK 8	$[67]$
		Mn	EUK 134	
Nervous system				
Neurofibromatosis	Flies	Mn	AEOL 10201	$[112]$
		Mn	AEOL 10150	
MPTP	Monkeys	Se	Ebselen	$[97]$
	Mice	Mn	AEOL 11207	$[22]$
ALS	Mutant SOD1	Mn	AEOL 10150	[105]
	Tg mice	Mn	EUK 8	$[106]$
		Mn	EUK 134	
Stroke	Rats	Se	Ebselen	$[74]$
		Mn	EUK 134	$[73]$
	Mice	Mn	AEOL 10113	$[72]$
		Mn	AEOL 10150	
	Humans	Se	Ebselen	$[75]$
Spinal cord trauma	Rats	Mn	AEOL 10201	[113]
		Se	Ebselen	[114]
	Mice	Mn	AEOL 10150	$[115]$
Cerebral vasoconstriction	Amyloid Tg mice	Mn	AEOL 10201	[108]
	Rats	Mn	AEOL 10201	[109]
Cognitive function	Aged mice	Mn	EUK 189	[103]

