

NIH Public Access

Author Manuscript

Antioxid Redox Signal. Author manuscript; available in PMC 2009 March 25.

Published in final edited form as:

Antioxid Redox Signal. 2008 February ; 10(2): 355–370. doi:10.1089/ars.2007.1916.

Antioxidants as Potential Therapeutics for Lung Fibrosis

BRIAN J. DAY

Division of Environmental and Occupational Health Sciences, Department of Medicine, National Jewish Medical and Research Center, Denver, Colorado

Abstract

Interstitial lung disease encompasses a large group of chronic lung disorders associated with excessive tissue remodeling, scarring, and fibrosis. The evidence of a redox imbalance in lung fibrosis is substantial, and the rationale for testing antioxidants as potential new therapeutics for lung fibrosis is appealing. Current animal models of lung fibrosis have clear involvement of ROS in their pathogenesis. New classes of antioxidant agents divided into catalytic antioxidant mimetics and antioxidant scavengers are being developed. The catalytic antioxidant class is based on endogenous antioxidant enzymes and includes the manganese-containing macrocyclics, porphyrins, salens, and the non–metal-containing nitroxides. The antioxidant scavenging class is based on endogenous antioxidant molecules and includes the vitamin E analogues, thiols, lazaroids, and polyphenolic agents. Numerous studies have shown oxidative stress to be associated with many interstitial lung diseases and that these agents are effective in attenuating fibroproliferative responses in the lung of animals and humans.

LUNG FIBROSIS

Classifications

Human lung fibrosis has been described histopathologically as a group of interstitial pneumonias including usual interstitial pneumonia (UIP), also known as idiopathic lung fibrosis (IPF), desquamative interstitial pneumonia (DIP), respiratory bronchiolitis interstitial lung disease (RB), lymphoid interstitial pneumonia (LIP), cryptogenic organizing pneumonia (OP), diffuse alveolar damage (DAD) or acute interstitial pneumonia (AIP), and nonspecific interstitial pneumonia (NSIP) (108). A large variation is found between the prognosis of the different interstitial pneumonias. NSIP, DIP, RB, and LIP have a 5-year mortality of less than 10% and are usually responsive to corticosteroid treatment. IPF and AIP have poor prognosis with a 5-year mortality $>60\%$ and are generally unresponsive to treatment. Some evidence supports a spectrum of inflammatory and fibrotic mechanisms in interstitial pneumonias (IPs), in which the IP forms that are most responsive to corticosteroids are more inflammatory and the unresponsive IP forms are more fibrotic (199).

IPF is one of the most devastating forms of lung fibrosis and is progressive and fatal (173). IPF occurs most often in people older than 50 years, with a higher occurrence in men than in women and an overall incidence of 13 to 20/100,000 individuals (47). IPF is generally nonresponsive to conventional anti-inflammatory and immunomodulatory therapy and currently is in need of new therapeutic approaches (214). IPF patients are thought to respond poorly to antiinflammatory therapies because little inflammation exists at advanced stages of the disease and the fibrosis is driven by a dysregulated repair process with a loss of epithelial cells and accumulation of mesenchymal cells (199).

Address reprint requests to: Brian J. Day, Ph.D., National Jewish Medical & Research Center, A439, 1400 Jackson St, Denver, CO 80206, E-mail: dayb@njc.org.

Pathogenesis

The pathogenesis of lung fibrosis is complex and is thought to involve a number of processes that lead to an altered alveolar environment and to an abnormal repair process with accumulated fibrosis. Several factors, including age, genetic susceptibility, and environmental agents, are known to contribute to lung fibrosis (13,127,219,222). An increased interest exists in mesenchymal cells and, in particular, in the fibroblastic foci that are associated with disease progression (102). These fibroblastic foci are also associated with increased levels of active transforming growth factor beta (TGF-*β*) in the fibrotic lung. This cytokine is an important mediator of fibroblast differentiation into the myofibroblast phenotype (81). Overex-pression of active TGF-*β* produces lung fibrosis in animals (180). TGF-*β* is a major regulator of wound repair and a stimulant of reactive oxygen species (ROS) production in fibroblasts (212). Oxidative stress is often defined as an imbalance between ROS production and antioxidant defenses. Oxidative stress can dysregulate cell signaling (158) and is a potential target for the development of therapeutics to treat lung fibrosis (110).

OXIDATIVE STRESS AND LUNG FIBROSIS

Reactive oxygen species and antioxidant defenses

Part of the altered alveolar environment in lung fibrosis involves oxidative stress that is driven by an imbalance between oxidant production and antioxidant defenses. Reactive oxygen species (ROS) are normal byproducts of cellular metabolism and are continually produced at low levels under basal conditions. Biologically, the ROS superoxide (O_2^-) is commonly generated from the uncoupling of the cellular electron-transport systems (131). Electrontransport systems that account for a large portion of cellular O_2^- formation are the mitochondrial electron-transport system (21) and various oxidases including NADPH oxidases (35), cytochrome P450 monoxygenases (33), cyclooxygenases (115), lipoxygenases (115), nitric oxide synthases (114), and xanthine oxidoreductase (130). O_2^- can also rapidly react with nitric oxide (NO) to form the strong oxidizing and nitrating agent, peroxynitrite (ONOO[–]). The ROS hydrogen peroxide (H₂O₂) is generated directly from O_2 [–] through a rapid dismutation reaction that can occur either enzymatically with superoxide dismutases (SODs, second-order rate constant of 10^9 M/sec) or spontaneously (second-order rate constant of 10⁵ M/sec). This means that wherever O_2 ⁻ is generated, formation of H_2O_2 also occurs. In addition, H_2O_2 is formed enzymatically as a byproduct of lipid metabolism in peroxisomes (160). H₂O₂ is stable at biologic pH and easily crosses lipid membranes. H₂O₂ can participate in hydroxyl radical (HO[']) formation in the presence of metals (78). H_2O_2 readily reacts with thiol functional groups, and this type of reaction is proposed to be a key mechanism by which ROS modulate cell-signaling events (62).

The impact of ROS may be especially important in the lung because of its large surface area and its exposure to higher oxygen levels than other tissues. The lung counters this with a formable array of antioxidant defense systems, starting with high levels of antioxidants in the epithelial lining fluid (ELF). Glutathione (GSH) is a major water-soluble antioxidant thiol in the lung ELF (27), and its levels are lower in subjects with IPF (26). In addition to GSH, the lung ELF contains other antioxidants such as ascorbate, urate, albumin, mucins, and metalbinding proteins, which are all effective at limiting oxidative damage. The lung also has a number of antioxidant enzyme systems including SODs, catalase, glutathione peroxidases (GPxs), thioredoxin, glutaredoxin, and peroxiredoxins (155). Overexpression of many of these antioxidant enzyme systems is protective against lung fibrosis (74,99,150). Many of these antioxidant systems are upregulated during lung fibrosis *via* the Nrf2 redox-sensitive transcription factor (95), that when deficient, enhances lung fibrotic responses (36). It is likely that inadequate antioxidant adaptive responses play a key role in lung fibrosis.

Oxidative stress and disruption of cell signaling

Several phosphatases contain sensitive thiol residues that are inhibited on oxidation (31). The inactivation of phosphatases is often associated with a perceived activation of their respective kinase(s), many of which play prominent roles in inflammatory responses (154). The phosphorylation state of a protein is a steady-state level set by the relative rates of kinases and phosphatases. As steady-state levels of oxidants increase, an increase occurs in the inactivation of phosphatases and a corresponding increase in the levels and duration of phosphorylated proteins (73,189). An increase in steady-state ROS results in altering the cellular glutathione (GSH)/glutathione disulfide (GSSG) redox couple, which can in turn alter the relative oxidation status of protein thiols. In addition, H_2O_2 can alter cell signaling directly by reacting with specific thiols on transcription factors, such as reducing factor-1, activator protein-1, and inhibiting factor-*κ*B (89,103). Many of these types of mechanisms may contribute to the often observed unregulated repair responses associated with lung fibrosis (Fig. 1).

Evidence of oxidative stress in the pathogenesis of lung fibrosis

When ROS production and antioxidant defenses are mismatched, an increase in ROS steadystate levels leads to an increase in the oxidation of cellular macromolecules. ROS are difficult to measure directly and often are assessed by measuring oxidative footprints in fluids and tissues, such as markers of protein, lipid, and DNA oxidation. Subjects with IPF have increased levels of oxidized proteins in their ELF that correlate with the percentage of neutrophils in the ELF (15,121,166). IPF subjects also have lower antioxidant capacity in their ELF than do healthy subjects (157). IPF subjects have higher levels of exhaled ethane (a marker of lipid peroxidation) than do normal subjects, and these levels are inversely correlated with $PaO₂$ (100). Breath condensate has been investigated as a noninvasive method to assess lung oxidative stress, and IPF subjects have higher levels of H_2O_2 and isoprostanes than do control subjects (151). Evidence also exists of increased nitrosative stress in IPF subjects (196). These data support the concept of increased oxidant formation with decreased antioxidant capacity in IPF, which is the hallmark of oxidative stress and the rationale for antioxidant therapy (Table 1).

ANTIOXIDANTS

Antioxidants have been defined many different ways, and in a very broad sense, they are agents that decrease steady-state ROS levels and protect cellular macromolecules from oxidative modification. The mechanisms by which this is achieved are many, and some are even paradoxic. For instance, some agents can produce a mild oxidative stress that results in a cellular adaptive response that increases endogenous antioxidant defenses. Some agents may inhibit cellular sources of ROS. A classic antioxidant is an agent that can rapidly react with ROS, producing less-reactive species. Catalytic antioxidants are not consumed in the reaction and are regenerated. Regardless of the mechanisms, antioxidants decrease oxidative stress and restore redox balance in biologic systems.

Catalytic antioxidant mimetics

Endogenous antioxidant enzymes are examples of catalytic antioxidants and have been used as models for the development of catalytic antioxidant mimetics. The three most prominent classes are the SOD, catalase, and GPx mimics. Most of these compounds contain a ligated transitional metal or selenium. They are generally broad-spectrum antioxidants that can scavenge O_2^- , H_2O_2 , ONOO⁻, and a variety of lipid peroxides. The SOD and catalase mimic class include macrocyclics, metallo-porphyrins, salens, and nitroxides. The GPx class includes selenium- and tellurium-based compounds.

The potencies of these catalytic antioxidants are often compared by using their rate constants obtained under tightly defined simple chemical systems, which may or may not be relevant in more complex biologic systems. Another important note is that many of these agents can obtain electrons from cellular sources (54). These properties have two important consequences that affect the *in vivo* rate constant for the reaction with ROS and can also result in the inhibition of ROS production. The finding that many of these diverse compounds are effective in similar oxidative stress models confirms the basic concept that small, efficient, catalytic antioxidants show promise in the treatment of ROS-mediated conditions associated with injury and tissue dysfunction.

The pentaazamacrocyclic ligand-based mimetics [M40403 is currently being developed by ActivBiotics [\(http://www.activbiotics.com](http://www.activbiotics.com))] are unique in that they are relatively specific O_2^- scavengers, because the manganese atom (Mn) is held by five coordination points in the macrocyclic structure and is available only for one-electron transfers (Fig. 2). Mn(II) macrocylics function in the dismutation reaction with O_2^- by alternate oxidation and reduction, changing its valence between Mn(II) and Mn(III) (11). This unique aspect of these compounds gives them selectivity toward O_2^- under highly defined conditions. However, in biologic systems, it is unclear whether it is only with O_2 ⁻ that these compounds interact. A number of endogenous compounds also can partake in one-electron reactions beside O_2^- ; these include flavins and ubiquinones. The macrocyclics are effective in many of the same oxidative paradigms in which nonselective catalytic antioxidant mimetics have been used. The different classes of catalytic antioxidant mimetics have not been directly compared in experimental models; therefore, the conditions under which one class holds an advantage over the others are currently not known.

Metalloporphyrins [AEOL series is currently being developed by Aeolus Pharmaceuticals [\(http://www.aeoluspharma.com\)](http://www.aeoluspharma.com)] are structurally different from endogenous protoporphyrins and are classified as synthetic *meso*-substituted porphyrins (Fig. 3). Metalloporphyrins have been shown to possess at least four distinct antioxidant properties, which include scavenging $O_2^-(149)$, H₂O₂ (53), ONOO⁻ (191), and lipid peroxides (51). Most metalloporphyrins contain either an Fe or Mn that is coordinated by four nitrogen axial ligands. The catalase-like activity of metalloporphyrins is thought to be due to their extensive conjugated ring system that can undergo reversible one-electron transfer in addition to the one-electron transfer on the metal center (70). This mechanism is similar to that proposed for the heme prosthetic groups of endogenous catalase and peroxidases. The two classes of metalloporphyrins include one group in which the SOD activities track with their catalase activities, and another group that has very little SOD activity and high catalase activity. An example of a manganese porphyrin with both high SOD and catalase-like activities is AEOL 10150 (98), whereas an example of a compound with low SOD activity and high catalase activity is AEOL 11207 (122). The compounds with high catalase-like activity still only possess a fraction of the native catalase enzyme activity under chemically defined conditions, yet they can protect cells from H_2O_2 -mediated toxicity (53). This may not be a fair comparison because catalase is hard to saturate with H_2O_2 and has a relative high k_m for H_2O_2 . Under biologically relevant steady-state levels of H_2O_2 , the metalloporphyrins are more comparable to catalase (32). Metalloporphyrins have been shown to be effective in ameliorating oxidative stress, inflammation, and injury in a large number of animal models of human disease (50). Metalloporphyrins have plasma half-lives that range from 4 to 48 hours. Most metalloporphyrins are not extensively metabolized by the body and are largely excreted unchanged in the urine. A previous limitation of the metalloporphyrin class of compounds has been poor oral bioavailability, but several compounds in the AEOL 112 series have good oral bioavailability and longer plasma half-lives that should make them better candidates for treating chronic diseases (122).

The salen class of catalytic antioxidant mimetics (EUK series) is currently being developed by Proteome Systems (<http://www.proteomesystems.com>) (Fig. 4). Generically, salens are aromatic, substituted ethylenediamine metal complexes. The Mn(III)-containing salen complexes have both O_2^- and H_2O_2 dismutation activities (63). However, like the metalloporphyrins, these compounds are not selective and can react with O_2^- and other peroxides, including ONOO⁻ (178). The Mn moiety of the salen is coordinated by four axial ligands. One of the unique features of these compounds is that the metal center is coordinated to oxygen and nitrogen atoms, which is in contrast to the porphyrins, in which the metal is coordinated to nitrogen atoms. The coordination of Mn by four axial ligands results in the formation of several possible valance states that give these compounds their broad ROSscavenging capabilities. The rates at which reported salens scavenge H_2O_2 are similar to those reported for metalloporphyrins, but are many orders less than those documented for catalase under similarly defined conditions (63). Salens have also been shown to protect cells against oxidative stress and are protective in a large number of animal models of human diseases (50). One of the current limitations of the salens is the stability of the parent compounds in biologic matrix, which makes it difficult to determine tissue levels and half-lives.

A number of compounds initially developed as free radical spin traps have been shown to have antioxidant properties in cell and animal systems (136). These compounds react with free radicals and form more-stable free radical products. The most frequently used compounds are the nitroxides and include *α*-phenyl-*tert* -butylnitrone (PBN) and 2,2,6,6-tetramethylpiperidine *N*-oxyl (TEMPO) (Fig. 5). These compounds have also been described as non–metalcontaining SOD mimics (5). The rate of reaction with O_2^- is relatively low and thus requires large amounts (often millimolar levels) of these compounds to be present in the system to be effective (190). Fortunately, these compounds are well tolerated in animals and can achieve high tissue levels (136). These agents have a number of properties other than the reaction with ROS that could also explain some of their protective properties in models of oxidative stress. Many of these compounds can be metabolized to release NO and can inhibit enzymes that are endogenous sources of ROS (34,229).

GPx enzymes are found in every compartment within the cell and tissues and are effective scavengers of cellular peroxides. The GPx mimetic class includes mono- and diselenium– containing compounds (Fig. 6). One of the best-studied GPx-like mimics is 2-phenyl-1,2 benzisoselenazol-3(2H)-one, also known as ebselen or PZ51. Ebselen was one of the first selenium-based GPx mimics developed and catalytically scavenges peroxides in the presence of reducing equivalents such as GSH, *N*-acetylcysteine (NAC), and dihydrolipoate (DHLA) (179). The mechanism by which this occurs is still debated and may differ under different conditions. Ebselen has also been shown to stimulate the decomposition of a number of ROS, including hypochlorous acid (HOCl) (17), singlet oxygen (174), and ONOO⁻ (128). Ebselen can readily bind cellular thiol groups on proteins, which may complicate the interpretation of biologic effects, because many cellular proteins have reactive thiols in their catalytic domains. It has been documented that ebselen can inhibit lipoxygenases (168), NADPH oxidases (46), and nitric oxide synthases (226). All of these enzymes are also potential sources of endogenous ROS. Ebselen has been shown to be protective in a number of cell-culture systems (159,194) and animal models of human disease (179). Ebselen is orally active and appears to be well tolerated in animals and humans. Newer analogues of ebselen have been developed, including BXT-51072, which has increased activity and potency in cell systems. These analogues [BXT series are being developed by Oxis International [\(http://www.oxis.com\)](http://www.oxis.com)] have been shown to be protective in a limited number of cell-culture systems and animal models of human disease (203).

A number of diselenide- and ditelluride-containing compounds have been reported to catalytically scavenge peroxides with higher GPx-like activity than ebselen (75,86,161,162).

Sulfur, selenium, and tellurium belong to group IV of the periodic table and have similar chemical properties. A major difference with these types of compounds is that they usually contain a diselenide bond. Earlier compounds, such as the diphenyl diselenide (DPDS), were electrophilic agents that had cytotoxic, genotoxic, and mutagenic issues (4,163). Many previously reported diselenide compounds release free selenium during the catalytic cycle, and this may be problematic in their development as therapeutic agents. A unique aspect of a newer series of these compounds is the cyclodextrin group, which may help in directing hydrophobic peroxides toward the selenium or tellurium active site. The diselenide, 2,2′-deseleno-bis-*β*cyclodextrin (2-SeCD), can scavenge a variety of peroxides including H_2O_2 , *tert*-butyl hydroperoxide, and cumenyl hydroperoxide by using GSH as a cofactor (124). Only a limited number of cell-culture studies have been reported for these compounds (140,186), and it is still unclear whether these compounds can be successfully used in animal models of lung fibrosis.

Antioxidant scavengers

The largest categories of antioxidants are those that are reactive toward ROS, and the product of the reaction results in a less-toxic species. The naturally occurring vitamins E (*α*-tocopherol) and C (ascorbate) are such examples. Both the ascorbate and *α*-tocopherol radicals are less reactive and can be recycled by cellular reductases. Glutathione is a thiol-containing tripeptide that readily reacts with peroxides and forms a less-toxic disulfide product that is recycled by glutathione reductase. A number of synthetic compounds have been models after these endogenous antioxidants and have been shown to be protective in models of oxidative stress (Fig. 7).

A number of polyphenolic-based antioxidants are known, such as the water-soluble analogue of *α*-tocopherol, known as trolox, hindered phenols that include butylated hydroxytoluene (BHT), and various plant phenolics such as curcumin and flavonoids. These compounds are often chain-breaking antioxidants, and some have been used in the food industry as preservatives (42). In general, they require larger doses or concentrations to produce antioxidant effects in model systems because of their lower rates of reaction with ROS and their limited ability to be recycled endogenously.

Another group of compounds use a steroid nucleus substituted with antioxidant side groups and are known as lazaroids. Lazaroids are very effective at inhibiting iron-dependent lipid peroxidation (39). Lazaroids have been extensively tested as neuroprotective agents, but it is still not clear whether their neuroprotective effects are directly related to their antioxidant properties.

A large class of antioxidants is the thiol-containing compounds. The most extensively studied thiol compound is *N*-acetyl cysteine (NAC). NAC is a direct-acting antioxidant and can scavenge several ROS such as hypochlorous acid, peroxides, `OH, and ONOO[−] (45). NAC can also serve as a cellular source of cysteine for the endogenous synthesis of GSH. NAC can suppress the activation of transcription factors such as NF-*κ*B as a way to modulate cellsignaling pathways. A homocysteine derivative, erdosteine, is a prodrug that, when metabolized, produces an active thiol antioxidant metabolite (56). Erdosteine has beneficial effects in COPD patients (156). Amifostine is another prodrug that, when metabolized, produces an active thiol antioxidant used clinically as a radioprotective agent (37). Thiolcontaining agents can also act as metal chelators and decrease oxidative stress by limiting the ability of transitional metal to participate in ROS formation. Paradoxically, some thiolcontaining agents have the potential to create a more-reactive species when they react with ROS, which is often dependent on availability of oxygen and transitional metals.

ANTIOXIDANTS, OXIDATIVE STRESS AND ANIMAL MODELS OF LUNG FIBROSIS

A large majority of lung fibrosis animal models involve the overproduction of oxidants, and the fibrotic effects are potentiated in antioxidant-deficient animals (Table 2). A number of drugs are known to produce lung fibrosis in humans and animals (105,228). Many of these drugs are chemotherapeutic agents that stimulate oxidative stress. Ionizing radiation is also a well-characterized method of producing lung fibrosis in animals, as well as a known adverse effect of cancer radiation treatment (41). A number of environmental exposures produce lung oxidative stress and fibrosis, including exposure to asbestos and silica (77,220). In addition, known cytokines, such as TGF-*β*, when overproduced, result in lung fibrosis. Most of these models have been shown to stimulate lung-injury responses and oxidative stress. A major limitation with currently available animal models of lung fibrosis is that they do not closely mimic human interstitial pneumonias, and many spontaneously resolve over time (38).

Fibrogenic drugs, antioxidants and oxidative stress

A number of chemotherapeutic agents can elevate intracellular ROS levels (141). The beststudied chemotherapeutic agent associated with oxidative stress and lung fibrosis is bleomycin. Bleomycin is thought to bind transitional metals and, in the presence of oxygen or H_2O_2 , generates a strong oxidant (101,225). The lung is thought to be a target organ because of its low levels of a cysteine protease that degrades bleomycin into an inactive form (117). Bleomycin increases ROS production in lung macrophages and alveolar type II epithelial cells *in vivo* (91). Bleomycin also increases lung epithelial cell apoptosis in a ROS-dependent manner (213). Bleomycin-induced cytotoxicity and lung fibrosis can be modulated by changing the intracellular levels of endogenous antioxidants (61,84,107,112,167).

Both catalytic and scavenger antioxidants have been shown to attenuate bleomycin-induced lung fibrosis in animals (Table 3). Liposomal or lecithinized delivery of SOD with or without catalase decreases bleomycin-induced lung fibrosis in rats and mice (118,119,193,221). The catalytic antioxidant porphyrin MnTBAP attenuates bleomycin-induced lung fibrosis in mice (148). The administration of vitamin E has also been shown to have protective effects, and its deficiency potentiates bleomycin-induced lung fibrosis in animals (57,58,142,188). Thiolcontaining antioxidants have been extensively studied in the bleomycin model of lung fibrosis. The best-studied thiol-containing antioxidant is NAC. Both oral and inhaled NAC decrease lung fibrosis in rats and mice (44,80,129,177). NAC also has been shown to restore lung GSH redox balance and to suppress bleomycin-induced activation of NF-*κ*B (176). Several prodrugs, such as erdosteine and amifostine, that produce active thiol-containing metabolites have also been shown to attenuate bleomycin-induced lung fibrosis in rodents (22,88,144,145,184, 224). In addition, lazaroids are protective against bleomycin-induced lung fibrosis in rats (49,132). A number of natural products that contain polyphenolic compounds, such as *Ginko biloba* extracts and curcumin, have been found to suppress lung oxidative stress and fibrosis in rats treated with bleomycin (48,67,93,152,207).

Paraquat is a redox-active herbicide that also is known to produce fatal pulmonary fibrosis in humans (43,202) and animals (181,182). Paraquat is thought to redox cycle with cellular enzymes to produce the paraquat cation radical that rapidly reacts with oxygen to form $\mathrm{O_2}^-$ (3,25,76). Paraquat produces lung oxidative stress in animals (2,23,66,113,218) and humans (94,135). Both catalytic and scavenger antioxidants have been shown to attenuate paraquatinduced lung injury and fibrosis in animals (see Table 3). Administration of SOD has been shown to attenuate paraquat-induced lung injury *in vitro* (90) and *in vivo* (147,215). The manganese-containing porphyrin catalytic antioxidant MnTBAP has been shown to have protective effects against paraquat-induced injury both *in vitro* (55,97) and *in vivo* (52). In

addition, the spin-trap PBN also has protective effects against paraquat-mediated damage (170). The administration of vitamin E has protective effects, and its deficiency potentiates paraquat-induced lung injury and fibrosis in animals (18,187). Thiol-based antioxidants also have protective effects in the paraquat model, including GSH (79,192), NAC (83,216,223), and erdosteine (92). In addition, the polyphenolic compound curcumin has been reported to have protective properties against paraquat-induced lung injury (206).

The antiarrhythmic drug amiodarone produces lung fibrosis in humans (183) and animals (28). Some data suggest a role for oxidative stress in amiodarone-induced lung fibrosis. Amiodarone inhibits mitochondrial complex I and II respiration and produces mitochondrial dysfunction in lung epithelial cells and macrophages (19). In the ventilated perfused rabbit lung system, amiodarone increases the levels of ROS and oxidized glutathione (GSSG) (106). Further studies have revealed that amiodarone is metabolized to an aryl radical that may give rise to other ROS (146,208). Both catalytic and scavenger antioxidants have been shown to attenuate amiodarone-induced lung injury and fibrosis in animals (see Table 3). PBN has been shown to directly scavenge the aryl radical produced by amiodarone (146). Vitamin E supplementation of hamsters attenuated both lung TGF-*β* levels and fibrosis induced by amiodarone (29,30). The thiol-based antioxidant NAC is also effective at attenuating amiodarone-induced lung fibrosis (120). In addition, the phenolic antioxidants BHA and curcumin were effective in limiting amiodarone-induced ROS and lung injury (120,152).

Radiation, antioxidants, and oxidative stress

Ionizing radiation produces fibrotic responses and generates hydrogen atom radical (H), OH, and hydrated electrons (e^-_{aq}) from the ionization of water in tissues. All three of these species are highly reactive and can generate and propagate a cascade of different ROS. Whole-body radiation decreases the levels of endogenous antioxidants and increases markers of lipid oxidation in animals and humans (9,40). Increased oxidative stress has been reported in radiation pneumonitis in humans (96) and in radiation-induced lung injury in rats (72,209). It is interesting to note that oxidative stress is still present even weeks after the radiation exposure (59). Several animal hemithoracic irradiation models of lung fibrosis have been developed and used to screen compounds for antifibrotic effects.

Both catalytic and scavenger antioxidants have been shown to attenuate radiation-induced lung injury and fibrosis in animals (see Table 3). Radiation-induced lung fibrosis is worsened in antioxidant-deficient animals (68,197) and attenuated in SOD-overexpression models (68,99, 126). Manganese-containing porphyrins and salen catalytic antioxidant mimetics have also been shown to have protective effects against radiation-induced lung fibrosis (116,153,210). The results from studies on the administration of vitamin E and its potential to limit radiationinduced lung injury and fibrosis in animals have been controversial, with some negative findings (165,217) and a recent positive result (16). Thiol-based antioxidants also have protective effects in the radiation-induced lung injury and fibrosis, including NAC (143) and amifostine (211). A number of flavonoids have also been used successfully to attenuate radiation-induced lung injury and fibrosis, including curcumin (200) and *Ginko biloba* extracts (175).

Fibrogenic cytokines and oxidative stress

A number of cytokines have been shown to stimulate fibrotic events and include TGF-*β*, tumor necrosis factor (TNF-*α*), platelet-derived growth factor (PDGF), connective tissue growth factor (CTGF), endothelin, granulocyte–macrophage colony-stimulating factor (GM-CSF), interleukin (IL-1*β*), IL-6, IL-10, and IL-13 (10). The best studied of these various cytokines in lung fibrosis is TGF-*β*. TGF-*β* isoforms have a number of effects on cellular responses including modulating cell growth, migration, differentiation, and apoptosis (169). TGF-*β*

induces myofibroblast differentiation, extracellular matrix (ECM) synthesis, and inhibits ECM breakdown (227). TGF-*β*1 is abundant in BALF and present in fibroblastic foci biopsies from IPF subjects (24). Overexpression of TGF-*β*1 in animals induces a progressive lung fibrosis that is largely independent of inflammation (180). TGF-*β*1 produces oxidative stress by the induction of ROS production and a decrease in expression of cellular antioxidants (111). TGF*β*1 induces ROS production by activation of NADPH oxidases (NOXs) and through mitochondrial dysfunction (185,198). TGF-*β*1 has been shown to decrease the expression of both catalase and mitochondrial SOD2 (82). In addition, TGF-*β*1 has been shown to decrease cellular GSH levels through the decreased expression and activity of *γ*-glutamylcysteine ligase (*γ*-GCL), the rate-limiting step in GSH synthesis (8). Interestingly, very few studies have been reported on the effects of antioxidants in this relatively new animal model of lung fibrosis.

Fibrogenic environmental agents and oxidative stress

A number of environmental dust and fiber exposures have been associated with the development of lung fibrosis (137). Both silica and asbestos exposures produce lung fibrosis in animals (64,104) and pneumoconiosis in humans (164). Both silica and asbestos produce injury and oxidative stress in the lungs of animals (1). In humans, a link between oxidative stress and exposure to dusts is supported by the finding of lower levels of glutathione-dependent enzymes in coal workers with pneumoconiosis (69). Numerous reports exist in the literature on the ability of both silica and asbestos exposures to increase ROS production (109,138, 172,204). In addition, beryllium exposure can produce lung granulomatous disease, and beryllium has recently been shown to stimulate increased production of ROS (171). A number of studies have reported an increased risk for developing IPF on various occupational and environmental exposures (13,87,195).

Several asbestos and silica animal models of lung fibrosis have been developed. Silica- and asbestos-induced lung fibrosis are worsened in antioxidant-deficient animals (71,125) and attenuated in catalase-overexpression models (139). Catalytic and scavenger antioxidants have been shown to attenuate asbestos- and silica-induced lung injury and fibrosis in animals (see Table 3). Catalytic antioxidants have been shown to have protective effects against silicainduced injury, including the porphyrin MnTBAP (123) and the nitroxide TEMPO (205). Thiol-based antioxidants also have protective effects in the silica-induced injuries, including NAC and GSH (12), as well as garlic extracts that are rich in thiol-containing compounds (6). The lazaroid compounds U-75412E and U-74389G are also effective against silica-induced injury (7,85).

ANTIOXIDANTS IN HUMAN IPF

Only a few antioxidants have actually been examined in humans with IPF. The thiol class has received the most attention to date. GSH (600 mg, twice daily for 3 days) has been given by inhalation to IPF subjects and found to increase ELF GSH levels and decrease ROS production in airway macrophages (20). NAC is an FDA-approved mucolytic drug that has been extensively used in cystic fibrosis subjects (65). Early studies examined the ability of oral NAC therapy (600 mg, 3 times daily for 5 days) to restore ELF GSH levels in IPF subjects (134). These studies found that this NAC regimen increased ELF GSH levels in IPF subjects 71% and was well tolerated. NAC has also been given to IPF subjects intravenously at 0.6-, 1.6-, and 4.8-g doses and found to elevate ELF GSH levels in IPF but not in normal subjects (133). None of these earlier studies looked at efficacy. NAC has been given to IPF subjects by inhalation (352 mg daily for 12 months), and some improvements were noted in exercise desaturation and high-resolution CT imaging; however, other lung-function tests and qualityof-life scores were not different from those with placebo (201). NAC has been given to a limited number of subjects with fibrosing alveolitis, in addition to immunosuppressive therapy (14). Oral NAC treatment (600 mg, 3 times daily for 12 weeks) increased lung ELF GSH levels 48%

over baseline and was associated with improved pulmonary-function tests. A more recent multicenter randomized trial further investigated the possible benefits of oral NAC therapy in combination with azathioprine and high-dose corticosteroids in a larger cohort of IPF subjects (60). NAC treatment (600 mg, 3 times daily for 12 months) was associated with modestly improved pulmonary-function tests *versus* standard therapy alone. Interestingly, this study found that the NAC-treatment group had a lower rate of myelotoxicity from the immunosuppressive therapy. Although the initial studies with NAC in IPF showed only modest beneficial effects, it sets the stage for testing other antioxidants in IPF.

CONCLUSIONS

The evidence of a redox imbalance in lung fibrosis is substantial, and the rationale for testing antioxidants as potential new therapeutics for lung fibrosis is appealing. All the current animal models of lung fibrosis have clear involvement of ROS in their pathogenesis, and numerous examples of a wide array of different antioxidants attenuating fibroproliferative events are ample in the literature. These factors should continue to drive the investigation and the use of antioxidants to treat the progression of lung fibrosis and other fibroproliferative disorders in humans.

Acknowledgements

This work was supported in part by NIH grants RO1 ES012504, RO1 HL75523, and U54 ES015678, and Research Grants from Aeolus Pharmaceuticals. Dr. Day is a consultant for and holds equity in Aeolus Pharmaceuticals, which is commercially developing catalytic antioxidant mimetics as therapeutic agents.

ABBREVIATIONS

References

- 1. Abidi P, Afaq F, Arif JM, Lohani M, Rahman Q. Chrysotile-mediated imbalance in the glutathione redox system in the development of pulmonary injury. Toxicol Lett 1999;106:31–39. [PubMed: 10378448]
- 2. Adachi J, Ishii K, Tomita M, Fujita T, Nurhantari Y, Nagasaki Y, Ueno Y. Consecutive administration of paraquat to rats induces enhanced cholesterol peroxidation and lung injury. Arch Toxicol 2003;77:353–357. [PubMed: 12799775]
- 3. Adam A, Smith LL, Cohen GM. An assessment of the role of redox cycling in mediating the toxicity of paraquat and nitro-furantoin. Environ Health Perspect 1990;85:113–117. [PubMed: 2384057]
- 4. Adams WJ Jr, Kocsis JJ, Snyder R. Acute toxicity and urinary excretion of diphenyldiselenide. Toxicol Lett 1989;48:301–310. [PubMed: 2781599]
- 5. Ahmad N, Misra M, Husain MM, Srivastava RC. Metal-independent putative superoxide dismutase mimics in chemistry, biology, and medicine. Ecotoxicol Environ Saf 1996;34:141–144. [PubMed: 8812179]
- 6. Ameen M, Musthapa MS, Abidi P, Ahmad I, Rahman Q. Garlic attenuates chrysotile-mediated pulmonary toxicity in rats by altering the phase I and phase II drug metabolizing enzyme system. J Biochem Mol Toxicol 2003;17:366–371. [PubMed: 14708092]
- 7. Antonini JM, van Dyke K, DiMatteo M, Reasor MJ. Attenuation of acute inflammatory effects of silica in rat lung by 21-aminosteroid, U74389G. Inflammation 1995;19:9–21. [PubMed: 7705890]
- 8. Arsalane K, Dubois CM, Muanza T, Begin R, Boudreau F, As-selin C, Cantin AM. Transforming growth factor-beta1 is a potent inhibitor of glutathione synthesis in the lung epithelial cell line A549: transcriptional effect on the GSH rate-limiting enzyme gamma-glutamylcysteine synthetase. Am J Respir Cell Mol Biol 1997;17:599–607. [PubMed: 9374111]
- 9. Arterbery VE, Pryor WA, Jiang L, Sehnert SS, Foster WM, Abrams RA, Williams JR, Wharam MD Jr, Risby TH. Breath ethane generation during clinical total body irradiation as a marker of oxygenfree-radical-mediated lipid peroxidation: a case study. Free Radic Biol Med 1994;17:569–576. [PubMed: 7867973]
- 10. Ask K, Martin GE, Kolb M, Gauldie J. Targeting genes for treatment in idiopathic pulmonary fibrosis: challenges and opportunities, promises and pitfalls. Proc Am Thorac Soc 2006;3:389–393. [PubMed: 16738206]
- 11. Aston K, Rath N, Naik A, Slomczynska U, Schall OF, Riley DP. Computer-aided design (CAD) of Mn(II) complexes: super-oxide dismutase mimetics with catalytic activity exceeding the native enzyme. Inorg Chem 2001;40:1779–1789. [PubMed: 11312732]

- 12. Barrett EG, Johnston C, Oberdorster G, Finkelstein JN. Anti-oxidant treatment attenuates cytokine and chemokine levels in murine macrophages following silica exposure. Toxicol Appl Pharmacol 1999;158:211–220. [PubMed: 10438654]
- 13. Baumgartner KB, Samet JM, Coultas DB, Stidley CA, Hunt WC, Colby TV, Waldron JA. Occupational and environmental risk factors for idiopathic pulmonary fibrosis: a multicenter casecontrol study collaborating centers. Am J Epidemiol 2000;152:307–315. [PubMed: 10968375]
- 14. Behr J, Maier K, Degenkolb B, Krombach F, Vogelmeier C. Antioxidative and clinical effects of high-dose N-acetylcysteine in fibrosing alveolitis: adjunctive therapy to maintenance immunosuppression. Am J Respir Crit Care Med 1997;156:1897–1901. [PubMed: 9412572]
- 15. Behr J, Maier K, Krombach F, Adelmann-Grill BC. Pathogenetic significance of reactive oxygen species in diffuse fibrosing alveolitis. Am Rev Respir Dis 1991;144:146–150. [PubMed: 2064121]
- 16. Bese NS, Munzuroglu F, Uslu B, Arbak S, Yesiladali G, Sut N, Altug T, Ober A. Vitamin E protects against the development of radiation-induced pulmonary fibrosis in rats. Clin Oncol (R Coll Radiol) 2007;19:260–264. [PubMed: 17433970]
- 17. Biewenga GP, Bast A. Reaction of lipoic acid with ebselen and hypochlorous acid. Methods Enzymol 1995;251:303–314. [PubMed: 7651210]
- 18. Block ER. Potentiation of acute paraquat toxicity by vitamin E deficiency. Lung 1979;156:195–203. [PubMed: 470435]
- 19. Bolt MW, Card JW, Racz WJ, Brien JF, Massey TE. Disruption of mitochondrial function and cellular ATP levels by amiodarone and N-desethylamiodarone in initiation of amiodarone-induced pulmonary cytotoxicity. J Pharmacol Exp Ther 2001;298:1280–1289. [PubMed: 11504831]
- 20. Borok Z, Buhl R, Grimes GJ, Bokser AD, Hubbard RC, Holroyd KJ, Roum JH, Czerski DB, Cantin AM, Crystal RG. Effect of glutathione aerosol on oxidant-antioxidant imbalance in idiopathic pulmonary fibrosis. Lancet 1991;338:215–216. [PubMed: 1676780]
- 21. Boveris A. Mitochondrial production of superoxide radical and hydrogen peroxide. Adv Exp Med Biol 1977;78:67–82. [PubMed: 197811]
- 22. Boyaci H, Maral H, Turan G, Basyigit I, Dillioglugil MO, Yildiz F, Tugay M, Pala A, Ercin C. Effects of erdosteine on bleomycin-induced lung fibrosis in rats. Mol Cell Biochem 2006;281:129–137. [PubMed: 16328965]
- 23. Brigelius R, Dostal LA, Horton JK, Bend JR. Alteration of the redox state of NADPH and glutathione in perfused rabbit lung by paraquat. Toxicol Ind Health 1986;2:417–428. [PubMed: 3590197]
- 24. Broekelmann TJ, Limper AH, Colby TV, McDonald JA. Transforming growth factor beta 1 is present at sites of extracellular matrix gene expression in human pulmonary fibrosis. Proc Natl Acad Sci U S A 1991;88:6642–6646. [PubMed: 1862087]
- 25. Bus JS, Aust SD, Gibson JE. Paraquat toxicity: proposed mechanism of action involving lipid peroxidation. Environ Health Perspect 1976;16:139–146. [PubMed: 1017417]
- 26. Cantin AM, Hubbard RC, Crystal RG. Glutathione deficiency in the epithelial lining fluid of the lower respiratory tract in idiopathic pulmonary fibrosis. Am Rev Respir Dis 1989;139:370–372. [PubMed: 2913886]
- 27. Cantin AM, North SL, Hubbard RC, Crystal RG. Normal alveolar epithelial lining fluid contains high levels of glutathione. J Appl Physiol 1987;63:152–157. [PubMed: 3040659]
- 28. Cantor JO, Osman M, Cerreta JM, Suarez R, Mandl I, Turino GM. Amiodarone-induced pulmonary fibrosis in hamsters. Exp Lung Res 1984;6:1–10. [PubMed: 6734540]
- 29. Card JW, Leeder RG, Racz WJ, Brien JF, Bray TM, Massey TE. Effects of dietary vitamin E supplementation on pulmonary morphology and collagen deposition in amiodarone- and vehicletreated hamsters. Toxicology 1999;133:75–84. [PubMed: 10378474]
- 30. Card JW, Racz WJ, Brien JF, Massey TE. Attenuation of amiodarone-induced pulmonary fibrosis by vitamin E is associated with suppression of transforming growth factor-beta1 gene expression but not prevention of mitochondrial dysfunction. J Pharmacol Exp Ther 2003;304:277–283. [PubMed: 12490602]
- 31. Caselli A, Marzocchini R, Camici G, Manao G, Moneti G, Pieraccini G, Ramponi G. The inactivation mechanism of low molecular weight phosphotyrosine-protein phosphatase by H2O2. J Biol Chem 1998;273:32554–32560. [PubMed: 9829991]

- 32. Castello P, Day BJ, Patel M. Inhibition of mitochondrial hydrogen peroxide production by lipophilic metalloporphyrins. J Pharmacol Exp Ther. submitted
- 33. Cederbaum AI. Microsomal generation of reactive oxygen species and their possible role in alcohol hepatotoxicity. Alcohol Alcohol Suppl 1991;1:291–296. [PubMed: 1669007]
- 34. Chamulitrat W, Mason RP, Riendeau D. Nitroxide metabolites from alkylhydroxylamines and Nhydroxyurea derivatives resulting from reductive inhibition of soybean lipoxygenase. J Biol Chem 1992;267:9574–9579. [PubMed: 1315759]
- 35. Cheson BD, Curnette JT, Babior BM. The oxidative killing mechanisms of the neutrophil. Prog Clin Immunol 1977;3:1–65. [PubMed: 17138]
- 36. Cho HY, Reddy SP, Yamamoto M, Kleeberger SR. The transcription factor NRF2 protects against pulmonary fibrosis. FASEB J 2004;18:1258–1260. [PubMed: 15208274]
- 37. Choi NC. Radioprotective effect of amifostine in radiation pneumonitis. Semin Oncol 2003;30:10– 17. [PubMed: 14727236]
- 38. Chua F, Gauldie J, Laurent GJ. Pulmonary fibrosis: searching for model answers. Am J Respir Cell Mol Biol 2005;33:9–13. [PubMed: 15964990]
- 39. Clark WM, Hazel JS, Coull BM. Lazaroids: CNS pharmacology and current research. Drugs 1995;50:971–983. [PubMed: 8612475]
- 40. Clemens MR, Ladner C, Schmidt H, Ehninger G, Einsele H, Buhler E, Waller HD, Gey KF. Decreased essential antioxidants and increased lipid hydroperoxides following high-dose radiochemotherapy. Free Radic Res Commun 1989;7:227–232. [PubMed: 2684798]
- 41. Coggle JE, Lambert BE, Moores SR. Radiation effects in the lung. Environ Health Perspect 1986;70:261–291. [PubMed: 3549278]
- 42. Conning DM, Phillips JC. Comparative metabolism of BHA, BHT and other phenolic antioxidants and its toxicological relevance. Food Chem Toxicol 1986;24:1145–1148. [PubMed: 3542762]
- 43. Copland GM, Kolin A, Shulman HS. Fatal pulmonary intra-alveolar fibrosis after paraquat ingestion. N Engl J Med 1974;291:290–292. [PubMed: 4407112]
- 44. Cortijo J, Cerda-Nicolas M, Serrano A, Bioque G, Estrela JM, Santangelo F, Esteras A, Llombart-Bosch A, Morcillo EJ. Attenuation by oral N-acetylcysteine of bleomycin-induced lung injury in rats. Eur Respir J 2001;17:1228–1235. [PubMed: 11491169]
- 45. Cotgreave IA. N-acetylcysteine: pharmacological considerations and experimental and clinical applications. Adv Pharmacol 1997;38:205–227. [PubMed: 8895810]
- 46. Cotgreave IA, Duddy SK, Kass GE, Thompson D, Moldeus P. Studies on the anti-inflammatory activity of ebselen: ebselen interferes with granulocyte oxidative burst by dual inhibition of NADPH oxidase and protein kinase C? Biochem Pharmacol 1989;38:649–656. [PubMed: 2537084]
- 47. Coultas DB, Zumwalt RE, Black WC, Sobonya RE. The epidemiology of interstitial lung diseases. Am J Respir Crit Care Med 1994;150:967–972. [PubMed: 7921471]
- 48. Daba MH, Abdel-Aziz AA, Moustafa AM, Al-Majed AA, Al-Shabanah OA, El-Kashef HA. Effects of L-carnitine and *Ginkgo biloba* extract (EG b 761) in experimental bleomycin-induced lung fibrosis. Pharmacol Res 2002;45:461–467. [PubMed: 12162946]
- 49. Dallessio JJ, McLaughlin GE, Frank L. Reduction of bleomycin-induced acute DNA injury in the rat lung by the 21-aminosteroid, U-74389G. Crit Care Med 1997;25:652–656. [PubMed: 9142031]
- 50. Day BJ. Catalytic antioxidants: a radical approach to new therapeutics. Drug Discov Today 2004;9:557–566. [PubMed: 15203091]
- 51. Day BJ, Batinic-Haberle I, Crapo JD. Metalloporphyrins are potent inhibitors of lipid peroxidation. Free Radic Biol Med 1999;26:730–736. [PubMed: 10218663]
- 52. Day BJ, Crapo JD. A metalloporphyrin superoxide dismutase mimetic protects against paraquatinduced lung injury in vivo. Toxicol Appl Pharmacol 1996;140:94–100. [PubMed: 8806874]
- 53. Day BJ, Fridovich I, Crapo JD. Manganic porphyrins possess catalase activity and protect endothelial cells against hydrogen peroxide-mediated injury. Arch Biochem Biophys 1997;347:256–262. [PubMed: 9367533]
- 54. Day BJ, Kariya C. A novel class of cytochrome P450 reductase redox cyclers: cationic manganoporphyrins. Toxicol Sci 2005;85:713–719. [PubMed: 15703263]

- 55. Day BJ, Shawen S, Liochev SI, Crapo JD. A metalloporphyrin superoxide dismutase mimetic protects against paraquat-induced endothelial cell injury, in vitro. J Pharmacol Exp Ther 1995;275:1227– 1232. [PubMed: 8531085]
- 56. Dechant KL, Noble S. Erdosteine. Drugs 1996;52:875–881. [PubMed: 8957158]discussion 882
- 57. Dede S, Mert H, Mert N, Yur F, Ertekin A, Deger Y. Effects of alpha-tocopherol on serum trace and major elements in rats with bleomycin-induced pulmonary fibrosis. Biol Trace Elem Res 2006;114:175–184. [PubMed: 17206000]
- 58. Deger Y, Yur F, Ertekin A, Mert N, Dede S, Mert H. Protective effect of alpha-tocopherol on oxidative stress in experimental pulmonary fibrosis in rats. Cell Biochem Funct. 2006(Epub ahead of print)
- 59. Delanian S, Lefaix JL. The radiation-induced fibroatrophic process: therapeutic perspective via the antioxidant pathway. Radiother Oncol 2004;73:119–131. [PubMed: 15542158]
- 60. Demedts M, Behr J, Buhl R, Costabel U, Dekhuijzen R, Jansen HM, MacNee W, Thomeer M, Wallaert B, Laurent F, Nicholson AG, Verbeken EK, Verschakelen J, Flower CD, Capron F, Petruzzelli S, De Vuyst P, van den Bosch JM, Rodriguez-Becerra E, Corvasce G, Lankhorst I, Sardina M, Montanari M. High-dose acetylcysteine in idiopathic pulmonary fibrosis. N Engl J Med 2005;353:2229–2242. [PubMed: 16306520]
- 61. Desai VG, Lyn-Cook LE, Aidoo A, Casciano DA, Feuers RJ. Modulation of antioxidant enzymes in bleomycin-treated rats by vitamin C and beta-carotene. Nutr Cancer 1997;29:127–132. [PubMed: 9427975]
- 62. Dickinson DA, Forman HJ. Glutathione in defense and signaling: lessons from a small thiol. Ann N Y Acad Sci 2002;973:488–504. [PubMed: 12485918]
- 63. Doctrow SR, Huffman K, Marcus CB, Musleh W, Bruce A, Baudry M, Malfroy B. Salen-manganese complexes: combined superoxide dismutase/catalase mimics with broad pharmacological efficacy. Adv Pharmacol 1997;38:247–269. [PubMed: 8895812]
- 64. Driscoll KE, Maurer JK, Higgins J, Poynter J. Alveolar macrophage cytokine and growth factor production in a rat model of crocidolite-induced pulmonary inflammation and fibrosis. J Toxicol Environ Health 1995;46:155–169. [PubMed: 7563215]
- 65. Duijvestijn YC, Brand PL. Systematic review of N-acetyl-cysteine in cystic fibrosis. Acta Paediatr 1999;88:38–41. [PubMed: 10090545]
- 66. Dusinska M, Kovacikova Z, Vallova B, Collins A. Responses of alveolar macrophages and epithelial type II cells to oxidative DNA damage caused by paraquat. Carcinogenesis 1998;19:809–812. [PubMed: 9635867]
- 67. El-Khatib AS, Moustafa AM, Abdel-Aziz AA, Al-Shabanah OA, El-Kashef HA. *Ginkgo biloba* extract (EGb 761) modulates bleomycin-induced acute lung injury in rats. Tumori 2001;87:417–422. [PubMed: 11989597]
- 68. Epperly MW, Epstein CJ, Travis EL, Greenberger JS. Decreased pulmonary radiation resistance of manganese superoxide dismutase (MnSOD)-deficient mice is corrected by human manganese superoxide dismutase-plasmid/liposome (SOD2-PL) intratracheal gene therapy. Radiat Res 2000;154:365–374. [PubMed: 11023599]
- 69. Evelo CT, Bos RP, Borm PJ. Decreased glutathione content and glutathione S-transferase activity in red blood cells of coal miners with early stages of pneumoconiosis. Br J Ind Med 1993;50:633–636. [PubMed: 8343424]
- 70. Fajer J, Borg DC, Forman A, Dolphin D, Felton RH. pi-Cation radicals and indications of metalloporphyrins. J Am Chem Soc 1970;92:3451–3459. [PubMed: 5422767]
- 71. Fattman CL, Tan RJ, Tobolewski JM, Oury TD. Increased sensitivity to asbestos-induced lung injury in mice lacking extracellular superoxide dismutase. Free Radic Biol Med 2006;40:601–607. [PubMed: 16458190]
- 72. Fleckenstein K, Zgonjanin L, Chen L, Rabbani Z, Jackson IL, Thrasher B, Kirkpatrick J, Foster WM, Vujaskovic Z. Temporal onset of hypoxia and oxidative stress after pulmonary irradiation. Int J Radiat Oncol Biol Phys 2007;68:196–204. [PubMed: 17448873]
- 73. Forman HJ, Torres M, Fukuto J. Redox signaling. Mol Cell Biochem 2002;234–235:49–62. [PubMed: 12162460]
- 74. Gao F, Kinnula VL, Myllärniemi M, Oury TD. Extracellular superoxide dismutase in pulmonary fibrosis. Antiox Redox Signal 2008;10:000–000.

- 75. Gao SJ, Chen M, Lin W, Luo GM, Shen JC. Kinetic studies of abzyme with glutathione peroxidase activity. Ann N Y Acad Sci 1998;864:280–283. [PubMed: 9928102]
- 76. Gray JP, Heck DE, Mishin V, Smith PJ, Hong JY, Thiruchelvam M, Cory-Slechta DA, Laskin DL, Laskin JD. Paraquat increases cyanide-insensitive respiration in murine lung epithelial cells by activating an NAD(P)H:paraquat oxidoreductase: identification of the enzyme as thioredoxin reductase. J Biol Chem 2007;282:7939–7949. [PubMed: 17229725]
- 77. Gulumian M. The role of oxidative stress in diseases caused by mineral dusts and fibres: current status and future of prophylaxis and treatment. Mol Cell Biochem 1999;196:69–77. [PubMed: 10448904]
- 78. Gutteridge JM. Biological origin of free radicals, and mechanisms of antioxidant protection. Chem Biol Interact 1994;91:133–140. [PubMed: 8194129]
- 79. Hagen TM, Brown LA, Jones DP. Protection against paraquat-induced injury by exogenous GSH in pulmonary alveolar type II cells. Biochem Pharmacol 1986;35:4537–4542. [PubMed: 3790171]
- 80. Hagiwara SI, Ishii Y, Kitamura S. Aerosolized administration of N-acetylcysteine attenuates lung fibrosis induced by bleomycin in mice. Am J Respir Crit Care Med 2000;162:225–231. [PubMed: 10903246]
- 81. Hashimoto S, Gon Y, Takeshita I, Matsumoto K, Maruoka S, Horie T. Transforming growth factorbeta1 induces phenotypic modulation of human lung fibroblasts to myofibroblast through a c-Jun-NH2-terminal kinase-dependent pathway. Am J Respir Crit Care Med 2001;163:152–157. [PubMed: 11208641]
- 82. Herrera B, Murillo MM, Alvarez-Barrientos A, Beltran J, Fernandez M, Fabregat I. Source of early reactive oxygen species in the apoptosis induced by transforming growth factor-beta in fetal rat hepatocytes. Free Radic Biol Med 2004;36:16–26. [PubMed: 14732287]
- 83. Hoffer E, Avidor I, Benjaminov O, Shenker L, Tabak A, Tamir A, Merzbach D, Taitelman U. Nacetylcysteine delays the infiltration of inflammatory cells into the lungs of paraquat-intoxicated rats. Toxicol Appl Pharmacol 1993;120:8–12. [PubMed: 8390113]
- 84. Hoshino T, Nakamura H, Okamoto M, Kato S, Araya S, Nomiyama K, Oizumi K, Young HA, Aizawa H, Yodoi J. Redox-active protein thioredoxin prevents proinflammatory cytokine- or bleomycininduced lung injury. Am J Respir Crit Care Med 2003;168:1075–1083. [PubMed: 12816738]
- 85. Huang SH, Leonard S, Shi X, Goins MR, Vallyathan V. Anti-oxidant activity of lazaroid (U-75412E) and its protective effects against crystalline silica-induced cytotoxicity. Free Radic Biol Med 1998;24:529–536. [PubMed: 9559864]
- 86. Huang X, Dong Z, Liu J, Mao S, Xu J, Luo G, Shen J. Selenium-mediated micellar catalyst: an efficient enzyme model for glutathione peroxidase-like catalysis. Langmuir 2007;23:1518–1522. [PubMed: 17241082]
- 87. Hubbard R, Lewis S, Richards K, Johnston I, Britton J. Occupational exposure to metal or wood dust and aetiology of cryptogenic fibrosing alveolitis. Lancet 1996;347:284–289. [PubMed: 8569361]
- 88. Hyde DM, Giri SN, Schiedt MJ, Krishna GA. Effects of three cysteine pro-drugs on bleomycininduced lung fibrosis in hamsters. Pathology 1990;22:93–101. [PubMed: 1700359]
- 89. Iles KE, Dickinson DA, Watanabe N, Iwamoto T, Forman HJ. AP-1 activation through endogenous H(2)O(2) generation by alveolar macrophages. Free Radic Biol Med 2002;32:1304–1313. [PubMed: 12057768]
- 90. Ilizarov AM, Koo HC, Kazzaz JA, Mantell LL, Li Y, Bhapat R, Pollack S, Horowitz S, Davis JM. Overexpression of manganese superoxide dismutase protects lung epithelial cells against oxidant injury. Am J Respir Cell Mol Biol 2001;24:436–441. [PubMed: 11306437]
- 91. Inghilleri S, Morbini P, Oggionni T, Barni S, Fenoglio C. In situ assessment of oxidant and nitrogenic stress in bleomycin pulmonary fibrosis. Histochem Cell Biol 2006;125:661–669. [PubMed: 16307278]
- 92. Inglesi M, Nicola M, Fregnan GB, Bradamante S, Pagani G. Synthesis and free radical scavenging properties of the enantiomers of erdosteine. Farmaco 1994;40:703–708. [PubMed: 7832972]
- 93. Iraz M, Erdogan H, Kotuk M, Yagmurca M, Kilic T, Ermis H, Fadillioglu E, Yildirim Z. *Ginkgo biloba* inhibits bleomycin-induced lung fibrosis in rats. Pharmacol Res 2006;53:310–316. [PubMed: 16459098]
- 94. Ishii K, Adachi J, Tomita M, Kurosaka M, Ueno Y. Oxysterols as indices of oxidative stress in man after paraquat ingestion. Free Radic Res 2002;36:163–168. [PubMed: 11999384]

- 95. Itoh K, Ishii T, Wakabayashi N, Yamamoto M. Regulatory mechanisms of cellular response to oxidative stress. Free Radic Res 1999;31:319–324. [PubMed: 10517536]
- 96. Jack CI, Cottier B, Jackson MJ, Cassapi L, Fraser WD, Hind CR. Indicators of free radical activity in patients developing radiation pneumonitis. Int J Radiat Oncol Biol Phys 1996;34:149–154. [PubMed: 12118544]
- 97. Kachadourian R, Flaherty MM, Crumbliss AL, Patel M, Day BJ. Synthesis and in vitro antioxidant properties of manganese(III) beta-octabromo-meso-tetrakis(4-carboxyphenyl)porphyrin. J Inorg Biochem 2003;95:240–248. [PubMed: 12818794]
- 98. Kachadourian R, Johnson CA, Min E, Spasojevic I, Day BJ. Flavin-dependent antioxidant properties of a new series of meso-N,N ′-dialkyl-imidazolium substituted manganese(III) porphyrins. Biochem Pharmacol 2004;67:77–85. [PubMed: 14667930]
- 99. Kang SK, Rabbani ZN, Folz RJ, Golson ML, Huang H, Yu D, Samulski TS, Dewhirst MW, Anscher MS, Vujaskovic Z. Overexpression of extracellular superoxide dismutase protects mice from radiation-induced lung injury. Int J Radiat Oncol Biol Phys 2003;57:1056–1066. [PubMed: 14575837]
- 100. Kanoh S, Kobayashi H, Motoyoshi K. Exhaled ethane: an in vivo biomarker of lipid peroxidation in interstitial lung diseases. Chest 2005;128:2387–2392. [PubMed: 16236899]
- 101. Kappus H, Mahmutoglu I, Kostrucha J, Scheulen ME. Liver nuclear NADPH-cytochrome P-450 reductase may be involved in redox cycling of bleomycin-Fe(III), oxy radical formation and DNA damage. Free Radic Res Commun 1987;2:271–277. [PubMed: 2462531]
- 102. Kasai H, Allen JT, Mason RM, Kamimura T, Zhang Z. TGF-beta1 induces human alveolar epithelial to mesenchymal cell transition (EMT). Respir Res 2005;6:56. [PubMed: 15946381]
- 103. Kaul N, Forman HJ. Activation of NF kappa B by the respiratory burst of macrophages. Free Radic Biol Med 1996;21:401–405. [PubMed: 8855453]
- 104. Kawanami O, Jiang HX, Mochimaru H, Yoneyama H, Kudoh S, Ohkuni H, Ooami H, Ferrans VJ. Alveolar fibrosis and capillary alteration in experimental pulmonary silicosis in rats. Am J Respir Crit Care Med 1995;151:1946–1955. [PubMed: 7767544]
- 105. Kehrer JP, Lee YC, Smith RD. Effect of intratracheally administered anticancer drugs on lung hydroxyproline content. Toxicol Lett 1986;30:63–70. [PubMed: 2420036]
- 106. Kennedy TP, Gordon GB, Paky A, McShane A, Adkinson NF Jr, Peters SP, Friday K, Jackman W, Sciuto AM, Gurtner GH. Amiodarone causes acute oxidant lung injury in ventilated and perfused rabbit lungs. J Cardiovasc Pharmacol 1988;12:23–36. [PubMed: 2459531]
- 107. Kilinc C, Ozcan O, Karaoz E, Sunguroglu K, Kutluay T, Karaca L. Vitamin E reduces bleomycininduced lung fibrosis in mice: biochemical and morphological studies. J Basic Clin Physiol Pharmacol 1993;4:249–269. [PubMed: 8679519]
- 108. Kim DS, Collard HR, King TE Jr. Classification and natural history of the idiopathic interstitial pneumonias. Proc Am Thorac Soc 2006;3:285–292. [PubMed: 16738191]
- 109. Kinnula VL. Oxidant and antioxidant mechanisms of lung disease caused by asbestos fibres. Eur Respir J 1999;14:706–716. [PubMed: 10543297]
- 110. Kinnula VL, Fattman CL, Tan RJ, Oury TD. Oxidative stress in pulmonary fibrosis: a possible role for redox modulatory therapy. Am J Respir Crit Care Med 2005;172:417–422. [PubMed: 15894605]
- 111. Koli K, Myllärniemi M, Keski-Oja J, Kinnula VL. Transforming growth factor-*β* activation in the lung: focus on fibrosis and reactive oxygen species. Antioxid Redos Signal 2008;10:000–000.
- 112. Kondo Y, Woo ES, Michalska AE, Choo KH, Lazo JS. Metallothionein null cells have increased sensitivity to anticancer drugs. Cancer Res 1995;55:2021–2023. [PubMed: 7743495]
- 113. Kornbrust DJ, Mavis RD. The effect of paraquat on microsomal lipid peroxidation in vitro and in vivo. Toxicol Appl Pharmacol 1980;53:323–332. [PubMed: 7394773]
- 114. Kotsonis P, Frohlich LG, Shutenko ZV, Horejsi R, Pfleiderer W, Schmidt HH. Allosteric regulation of neuronal nitric oxide synthase by tetrahydrobiopterin and suppression of auto-damaging superoxide. Biochem J 2000;346:767–776. [PubMed: 10698705]
- 115. Kukreja RC, Kontos HA, Hess ML, Ellis EF. PGH synthase and lipoxygenase generate superoxide in the presence of NADH or NADPH. Circ Res 1986;59:612–619. [PubMed: 3028671]

NIH-PA Author Manuscript

NIH-PA Author Manuscript

- 116. Langan AR, Khan MA, Yeung IW, Van Dyk J, Hill RP. Partial volume rat lung irradiation: the protective/mitigating effects of Eukarion-189, a superoxide dismutase-catalase mimetic. Radiother Oncol 2006;79:231–238. [PubMed: 16675053]
- 117. Lazo JS, Humphreys CJ. Lack of metabolism as the biochemical basis of bleomycin-induced pulmonary toxicity. Proc Natl Acad Sci U S A 1983;80:3064–3068. [PubMed: 6190169]
- 118. Ledwozyw A. Protective effect of liposome-entrapped superoxide dismutase and catalase on bleomycin-induced lung injury in rats, I: antioxidant enzyme activities and lipid peroxidation. Acta Vet Hung 1991;39:215–224. [PubMed: 1723845]
- 119. Ledwozyw A. Protective effect of liposome-entrapped superoxide dismutase and catalase on bleomycin-induced lung injury in rats, II: phospholipids of the lung surfactant. Acta Physiol Hung 1991;78:157–162. [PubMed: 1725569]
- 120. Leeder RG, Brien JF, Massey TE. Investigation of the role of oxidative stress in amiodarone-induced pulmonary toxicity in the hamster. Can J Physiol Pharmacol 1994;72:613–621. [PubMed: 7954092]
- 121. Lenz AG, Costabel U, Maier KL. Oxidized BAL fluid proteins in patients with interstitial lung diseases. Eur Respir J 1996;9:307–312. [PubMed: 8777969]
- 122. Liang LP, Huang J, Fulton R, Day BJ, Patel M. An orally active catalytic metalloporphyrin protects against 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine neurotoxicity in vivo. J Neurosci 2007;27:4326–4333. [PubMed: 17442816]
- 123. Liu H, Zhang H, Forman HJ. Silica induces macrophage cytokines through phosphatidylcholinespecific phospholipase C with hydrogen peroxide. Am J Respir Cell Mol Biol 2007;36:594–599. [PubMed: 17158358]
- 124. Liu J, Luo G, Ren X, Mu Y, Bai Y, Shen J. A bis-cyclodextrin diselenide with glutathione peroxidaselike activity. Biochim Biophys Acta 2000;1481:222–228. [PubMed: 11018712]
- 125. Lombard-Gillooly K, Hubbard AK. Modulation of silica-induced lung injury by reducing lung nonprotein sulfhydryls with buthionine sulfoximine. Toxicol Lett 1993;66:305–315. [PubMed: 8386403]
- 126. Malaker K, Das RM. Effect of superoxide dismutase on early radiation injury of lungs in the rat. Mol Cell Biochem 1988;84:141–145. [PubMed: 3231220]
- 127. Mannino DM, Etzel RA, Parrish RG. Pulmonary fibrosis deaths in the United States, 1979–1991: an analysis of multiple-cause mortality data. Am J Respir Crit Care Med 1996;153:1548–1552. [PubMed: 8630600]
- 128. Masumoto H, Sies H. The reaction of ebselen with peroxynitrite. Chem Res Toxicol 1996;9:262– 267. [PubMed: 8924601]
- 129. Mata M, Ruiz A, Cerda M, Martinez-Losa M, Cortijo J, Santangelo F, Serrano-Mollar A, Llombart-Bosch A, Morcillo EJ. Oral N-acetylcysteine reduces bleomycin-induced lung damage and mucin Muc5ac expression in rats. Eur Respir J 2003;22:900–905. [PubMed: 14680076]
- 130. McCord JM. Oxygen-derived free radicals in postischemic tissue injury. N Engl J Med 1985;312:159–163. [PubMed: 2981404]
- 131. McCord JM, Fridovich I. The biology and pathology of oxygen radicals. Ann Intern Med 1978;89:122–127. [PubMed: 208444]
- 132. McLaughlin GE, Frank L. Effects of the 21-aminosteroid, U74389F, on bleomycin-induced pulmonary fibrosis in rats. Crit Care Med 1994;22:313–319. [PubMed: 7508358]
- 133. Meyer A, Buhl R, Kampf S, Magnussen H. Intravenous N-acetylcysteine and lung glutathione of patients with pulmonary fibrosis and normals. Am J Respir Crit Care Med 1995;152:1055–1060. [PubMed: 7663783]
- 134. Meyer A, Buhl R, Magnussen H. The effect of oral N-acetyl-cysteine on lung glutathione levels in idiopathic pulmonary fibrosis. Eur Respir J 1994;7:431–436. [PubMed: 8013597]
- 135. Minakata K, Suzuki O, Saito S, Harada N. Ascorbate radical levels in human sera and rat plasma intoxicated with paraquat and diquat. Arch Toxicol 1993;67:126–130. [PubMed: 8386923]
- 136. Mitchell JB, Krishna MC, Kuppusamy P, Cook JA, Russo A. Protection against oxidative stress by nitroxides. Exp Biol Med (Maywood) 2001;226:620–621. [PubMed: 11444094]
- 137. Mossman BT, Janssen YM, Marsh JP, Sesko A, Shatos MA, Doherty J, Adler KB, Hemenway D, Mickey R, Vacek, et al. Development and characterization of a rapid-onset rodent inhalation model of asbestosis for disease prevention. Toxicol Pathol 1991;19:412–418. [PubMed: 1667555]

- 138. Mossman BT, Marsh JP. Evidence supporting a role for active oxygen species in asbestos-induced toxicity and lung disease. Environ Health Perspect 1989;81:91–94. [PubMed: 2667992]
- 139. Mossman BT, Marsh JP, Sesko A, Hill S, Shatos MA, Doherty J, Petruska J, Adler KB, Hemenway D, Mickey R, Vacek P, Kagan E. Inhibition of lung injury, inflammation, and interstitial pulmonary fibrosis by polyethylene glycol-conjugated catalase in a rapid inhalation model of asbestosis. Am Rev Respir Dis 1990;141:1266–1271. [PubMed: 2160214]
- 140. Mu Y, Lv S, Ren X, Jin G, Liu J, Yan G, Li W, Shen J, Luo G. UV-B induced keratinocyte apoptosis is blocked by 2-selenium-bridged beta-cyclodextrin, a GPX mimic. J Photochem Photobiol B 2003;69:7–12. [PubMed: 12547491]
- 141. Muller I, Niethammer D, Bruchelt G. Anthracycline-derived chemotherapeutics in apoptosis and free radical cytotoxicity [Review]. Int J Mol Med 1998;1:491–494. [PubMed: 9852255]
- 142. Nakamura H, Sato S, Takahashi K. Effects of vitamin E deficiency on bleomycin-induced pulmonary fibrosis in the hamster. Lung 1988;166:161–176. [PubMed: 2454380]
- 143. Neal R, Matthews RH, Lutz P, Ercal N. Antioxidant role of N-acetyl cysteine isomers following high dose irradiation. Free Radic Biol Med 2003;34:689-695. [PubMed: 12633746]
- 144. Nici L, Calabresi P. Amifostine modulation of bleomycin-induced lung injury in rodents. Semin Oncol 1999;26:28–33. [PubMed: 10348257]
- 145. Nici L, Santos-Moore A, Kuhn C, Calabresi P. Modulation of bleomycin-induced pulmonary toxicity in the hamster by the antioxidant amifostine. Cancer 1998;83:2008–2014. [PubMed: 9806661]
- 146. Nicolescu AC, Comeau JL, Hill BC, Bedard LL, Takahashi T, Brien JF, Racz WJ, Massey TE. Aryl radical involvement in amiodarone-induced pulmonary toxicity: investigation of protection by spintrapping nitrones. Toxicol Appl Pharmacol 2007;220:60–71. [PubMed: 17316728]
- 147. Ogata H, Luo XX, Xu X. Comparison of effects of margarite extract and recombinant human superoxide dismutase on paraquat-induced superoxide anion radicals in rat lung. Circ Shock 1994;43:161–165. [PubMed: 7895320]
- 148. Oury TD, Thakker K, Menache M, Chang LY, Crapo JD, Day BJ. Attenuation of bleomycin-induced pulmonary fibrosis by a catalytic antioxidant metalloporphyrin. Am J Respir Cell Mol Biol 2001;25:164–169. [PubMed: 11509325]
- 149. Pasternack RFSW. Catalysis of the disproportionation of super-oxide by metalloporphyrins. J Inorgan Biochem 1979;11:261–267.
- 150. Phan SH, Fantone JC. Inhibition of bleomycin-induced pulmonary fibrosis by lipopolysaccharide. Lab Invest 1984;50:587–591. [PubMed: 6201676]
- 151. Psathakis K, Mermigkis D, Papatheodorou G, Loukides S, Panagou P, Polychronopoulos V, Siafakas NM, Bouros D. Exhaled markers of oxidative stress in idiopathic pulmonary fibrosis. Eur J Clin Invest 2006;36:362–367. [PubMed: 16634841]
- 152. Punithavathi D, Venkatesan N, Babu M. Curcumin inhibition of bleomycin-induced pulmonary fibrosis in rats. Br J Pharmacol 2000;131:169–172. [PubMed: 10991907]
- 153. Rabbani ZN, Batinic-Haberle I, Anscher MS, Huang J, Day BJ, Alexander E, Dewhirst MW, Vujaskovic Z. Long-term administration of a small molecular weight catalytic metalloporphyrin antioxidant, AEOL 10150, protects lungs from radiation-induced injury. Int J Radiat Oncol Biol Phys 2007;67:573–580. [PubMed: 17236973]
- 154. Rahman I, Adcock IM. Oxidative stress and redox regulation of lung inflammation in COPD. Eur Respir J 2006;28:219–242. [PubMed: 16816350]
- 155. Rahman I, Biswas SK, Kode A. Oxidant and antioxidant balance in the airways and airway diseases. Eur J Pharmacol 2006;533:222–239. [PubMed: 16500642]
- 156. Rahman I, Kilty I. Antioxidant therapeutic targets in COPD. Curr Drug Targets 2006;7:707–720. [PubMed: 16787173]
- 157. Rahman I, Skwarska E, Henry M, Davis M, O'Connor CM, FitzGerald MX, Greening A, MacNee W. Systemic and pulmonary oxidative stress in idiopathic pulmonary fibrosis. Free Radic Biol Med 1999;27:60–68. [PubMed: 10443920]
- 158. Rahman I, Yang SR, Biswas SK. Current concepts of redox signaling in the lungs. Antioxid Redox Signal 2006;8:681–689. [PubMed: 16677111]
- 159. Ramakrishnan N, Kalinich JF, McClain DE. Ebselen inhibition of apoptosis by reduction of peroxides. Biochem Pharmacol 1996;51:1443–1451. [PubMed: 8630085]

- 160. Reddy JK, Rao MS. Oxidative DNA damage caused by persistent peroxisome proliferation: its role in hepatocarcinogenesis. Mutat Res 1989;214:63–68. [PubMed: 2671702]
- 161. Ren X, Liu J, Luo G, Zhang Y, Luo Y, Yan G, Shen J. A novel selenocystine-beta-cyclodextrin conjugate that acts as a glutathione peroxidase mimic. Bioconjug Chem 2000;11:682–687. [PubMed: 10995212]
- 162. Ren X, Xue Y, Zhang K, Liu J, Luo G, Zheng J, Mu Y, Shen J. A novel dicyclodextrinyl ditelluride compound with antioxidant activity. FEBS Lett 2001;507:377–380. [PubMed: 11696375]
- 163. Rosa RM, Sulzbacher K, Picada JN, Roesler R, Saffi J, Brendel M, Henriques JA. Genotoxicity of diphenyl diselenide in bacteria and yeast. Mutat Res 2004;563:107–115. [PubMed: 15364277]
- 164. Ross MH, Murray J. Occupational respiratory disease in mining. Occup Med (Lond) 2004;54:304– 310. [PubMed: 15289586]
- 165. Rostock RA, Stryker JA, Abt AB. Evaluation of high-dose vitamin E as a radioprotective agent. Radiology 1980;136:763–766. [PubMed: 7403557]
- 166. Rottoli P, Magi B, Cianti R, Bargagli E, Vagaggini C, Nikiforakis N, Pallini V, Bini L. Carbonylated proteins in bronchoalveolar lavage of patients with sarcoidosis, pulmonary fibrosis associated with systemic sclerosis and idiopathic pulmonary fibrosis. Proteomics 2005;5:2612–2618. [PubMed: 15924291]
- 167. Russo A, Mitchell JB, McPherson S, Friedman N. Alteration of bleomycin cytotoxicity by glutathione depletion or elevation. Int J Radiat Oncol Biol Phys 1984;10:1675–1678. [PubMed: 6207156]
- 168. Safayhi H, Tiegs G, Wendel A. A novel biologically active seleno-organic compound V: inhibition by ebselen (PZ 51) of rat peritoneal neutrophil lipoxygenase. Biochem Pharmacol 1985;34:2691– 2694. [PubMed: 2990494]
- 169. Sanchez-Capelo A. Dual role for TGF-beta1 in apoptosis. Cytokine Growth Factor Rev 2005;16:15– 34. [PubMed: 15733830]
- 170. Sata T, Kubota E, Misra HP, Mojarad M, Pakbaz H, Said SI. Paraquat-induced lung injury: prevention by N-tert-butyl-alpha-phenylnitrone, a free-radical spin-trapping agent. Am J Physiol 1992;262:L147–L152. [PubMed: 1311515]
- 171. Sawyer RT, Dobis DR, Goldstein M, Velsor L, Maier LA, Fontenot AP, Silveira L, Newman LS, Day BJ. Beryllium-stimulated reactive oxygen species and macrophage apoptosis. Free Radic Biol Med 2005;38:928–937. [PubMed: 15749389]
- 172. Schapira RM, Ghio AJ, Effros RM, Morrisey J, Almagro UA, Dawson CA, Hacker AD. Hydroxyl radical production and lung injury in the rat following silica or titanium dioxide instillation in vivo. Am J Respir Cell Mol Biol 1995;12:220–226. [PubMed: 7865220]
- 173. Schwartz DA, Van Fossen DS, Davis CS, Helmers RA, Dayton CS, Burmeister LF, Hunninghake GW. Determinants of progression in idiopathic pulmonary fibrosis. Am J Respir Crit Care Med 1994;149:444–449. [PubMed: 8306043]
- 174. Scurlock R, Rougee M, Bensasson RV, Evers M, Dereu N. Deactivation of singlet molecular oxygen by organo-selenium compounds exhibiting glutathione peroxidase activity and by sulfur-containing homologs. Photochem Photobiol 1991;54:733–736. [PubMed: 1798750]
- 175. Sener G, Kabasakal L, Atasoy BM, Erzik C, Velioglu-Ogunc A, Cetinel S, Gedik N, Yegen BC. *Ginkgo biloba* extract protects against ionizing radiation-induced oxidative organ damage in rats. Pharmacol Res 2006;53:241–252. [PubMed: 16412663]
- 176. Serrano-Mollar A, Closa D, Prats N, Blesa S, Martinez-Losa M, Cortijo J, Estrela JM, Morcillo EJ, Bulbena O. In vivo anti-oxidant treatment protects against bleomycin-induced lung damage in rats. Br J Pharmacol 2003;138:1037–1048. [PubMed: 12684259]
- 177. Shahzeidi S, Sarnstrand B, Jeffery PK, McAnulty RJ, Laurent GJ. Oral N-acetylcysteine reduces bleomycin-induced collagen deposition in the lungs of mice. Eur Respir J 1991;4:845–852. [PubMed: 1720100]
- 178. Sharpe MA, Ollosson R, Stewart VC, Clark JB. Oxidation of nitric oxide by oxomanganese-salen complexes: a new mechanism for cellular protection by superoxide dismutase/catalase mimetics. Biochem J 2002;366:97–107. [PubMed: 11994046]
- 179. Sies H. Ebselen, a selenoorganic compound as glutathione peroxidase mimic. Free Radic Biol Med 1993;14:313–323. [PubMed: 8458589]

- 180. Sime PJ, Xing Z, Graham FL, Csaky KG, Gauldie J. Aden-ovector-mediated gene transfer of active transforming growth factor-beta1 induces prolonged severe fibrosis in rat lung. J Clin Invest 1997;100:768–776. [PubMed: 9259574]
- 181. Smith P. A light- and electron-microscope study of the pulmonary lesions induced in rats by paraquat. J Pathol 1971;104:vii. [PubMed: 5124998]
- 182. Smith P, Heath D, Kay JM. The pathogenesis and structure of paraquat-induced pulmonary fibrosis in rats. J Pathol 1974;114:57–67. [PubMed: 4443850]
- 183. Sobol SM, Rakita L. Pneumonitis and pulmonary fibrosis associated with amiodarone treatment: a possible complication of a new antiarrhythmic drug. Circulation 1982;65:819–824. [PubMed: 7060263]
- 184. Sogut S, Ozyurt H, Armutcu F, Kart L, Iraz M, Akyol O, Ozen S, Kaplan S, Temel I, Yildirim Z. Erdosteine prevents bleomycin-induced pulmonary fibrosis in rats. Eur J Pharmacol 2004;494:213– 220. [PubMed: 15212977]
- 185. Sturrock A, Cahill B, Norman K, Huecksteadt TP, Hill K, Sanders K, Karwande SV, Stringham JC, Bull DA, Gleich M, Kennedy TP, Hoidal JR. Transforming growth factor-beta1 induces Nox4 NAD (P)H oxidase and reactive oxygen species-dependent proliferation in human pulmonary artery smooth muscle cells. Am J Physiol Lung Cell Mol Physiol 2006;290:L661–L673. [PubMed: 16227320]
- 186. Sun Y, Mu Y, Ma S, Gong P, Yan G, Liu J, Shen J, Luo G. The molecular mechanism of protecting cells against oxidative stress by 2-selenium-bridged beta-cyclodextrin with glutathione peroxidase activity. Biochim Biophys Acta 2005;1743:199–204. [PubMed: 15843033]
- 187. Suntres ZE, Shek PN. Intratracheally administered liposomal alpha-tocopherol protects the lung against long-term toxic effects of paraquat. Biomed Environ Sci 1995;8:289–300. [PubMed: 8719170]
- 188. Suntres ZE, Shek PN. Protective effect of liposomal alpha-tocopherol against bleomycin-induced lung injury. Biomed Environ Sci 1997;10:47–59. [PubMed: 9099426]
- 189. Suzuki YJ, Forman HJ, Sevanian A. Oxidants as stimulators of signal transduction. Free Radic Biol Med 1997;22:269–285. [PubMed: 8958153]
- 190. Swartz HM. Principles of the metabolism of nitroxides and their implications for spin trapping. Free Radic Res Commun 1990;9:399–405. [PubMed: 2167277]
- 191. Szabo C, Day BJ, Salzman AL. Evaluation of the relative contribution of nitric oxide and peroxynitrite to the suppression of mitochondrial respiration in immunostimulated macrophages using a manganese mesoporphyrin superoxide dismutase mimetic and peroxynitrite scavenger. FEBS Lett 1996;381:82–86. [PubMed: 8641445]
- 192. Szabo L, Matkovics B, Barabas K, Oroszlan G. Effects of various thiols on paraquat toxicity. Comp Biochem Physiol C 1986;83:149–153. [PubMed: 2869890]
- 193. Tamagawa K, Taooka Y, Maeda A, Hiyama K, Ishioka S, Yamakido M. Inhibitory effects of a lecithinized superoxide dismutase on bleomycin-induced pulmonary fibrosis in mice. Am J Respir Crit Care Med 2000;161:1279–1284. [PubMed: 10764324]
- 194. Tamasi V, Jeffries JM, Arteel GE, Falkner KC. Ebselen augments its peroxidase activity by inducing nrf-2-dependent transcription. Arch Biochem Biophys 2004;431:161–168. [PubMed: 15488464]
- 195. Taskar VS, Coultas DB. Is idiopathic pulmonary fibrosis an environmental disease? Proc Am Thorac Soc 2006;3:293–298. [PubMed: 16738192]
- 196. Terasaki Y, Akuta T, Terasaki M, Sawa T, Mori T, Okamoto T, Ozaki M, Takeya M, Akaike T. Guanine nitration in idiopathic pulmonary fibrosis and its implication for carcinogenesis. Am J Respir Crit Care Med 2006;174:665–673. [PubMed: 16741153]
- 197. Thanislass J, Raveendran M, Devaraj H. Buthionine sulfoximine-induced glutathione depletion: its effect on antioxidants, lipid peroxidation and calcium homeostasis in the lung. Biochem Pharmacol 1995;50:229–234. [PubMed: 7632167]
- 198. Thannickal VJ, Fanburg BL. Activation of an H2O2-generating NADH oxidase in human lung fibroblasts by transforming growth factor beta 1. J Biol Chem 1995;270:30334–30338. [PubMed: 8530457]
- 199. Thannickal VJ, Toews GB, White ES, Lynch JP 3rd, Martinez FJ. Mechanisms of pulmonary fibrosis. Annu Rev Med 2004;55:395–417. [PubMed: 14746528]

- 200. Thresiamma KC, George J, Kuttan R. Protective effect of curcumin, ellagic acid and bixin on radiation induced toxicity. Indian J Exp Biol 1996;34:845–847. [PubMed: 9014516]
- 201. Tomioka H, Kuwata Y, Imanaka K, Hashimoto K, Ohnishi H, Tada K, Sakamoto H, Iwasaki H. A pilot study of aerosolized N-acetylcysteine for idiopathic pulmonary fibrosis. Respirology 2005;10:449–455. [PubMed: 16135167]
- 202. Toner PG, Vetters JM, Spilg WG, Harland WA. Fine structure of the lung lesion in a case of paraquat poisoning. J Pathol 1970;102:182–185. [PubMed: 5508915]
- 203. Tuvlin JA, Kane SV. Novel therapies in the treatment of ulcerative colitis. Expert Opin Investig Drugs 2003;12:483–490.
- 204. Vallyathan V, Castranova V, Pack D, Leonard S, Shumaker J, Hubbs AF, Shoemaker DA, Ramsey DM, Pretty JR, McLaurin JL, et al. Freshly fractured quartz inhalation leads to enhanced lung injury and inflammation: potential role of free radicals. Am J Respir Crit Care Med 1995;152:1003–1009. [PubMed: 7663775]
- 205. Vallyathan V, Leonard S, Kuppusamy P, Pack D, Chzhan M, Sanders SP, Zweir JL. Oxidative stress in silicosis: evidence for the enhanced clearance of free radicals from whole lungs. Mol Cell Biochem 1997;168:125–132. [PubMed: 9062901]
- 206. Venkatesan N. Pulmonary protective effects of curcumin against paraquat toxicity. Life Sci 2000;66:PL21–L28. [PubMed: 10666014]
- 207. Venkatesan N, Punithavathi V, Chandrakasan G. Curcumin protects bleomycin-induced lung injury in rats. Life Sci 1997;61:PL51–PL58. [PubMed: 9250724]
- 208. Vereckei A, Blazovics A, Gyorgy I, Feher E, Toth M, Szenasi G, Zsinka A, Foldiak G, Feher J. The role of free radicals in the pathogenesis of amiodarone toxicity. J Cardiovasc Electrophysiol 1993;4:161–177. [PubMed: 8269288]
- 209. Vujaskovic Z, Anscher MS, Feng QF, Rabbani ZN, Amin K, Samulski TS, Dewhirst MW, Haroon ZA. Radiation-induced hypoxia may perpetuate late normal tissue injury. Int J Radiat Oncol Biol Phys 2001;50:851–855. [PubMed: 11429211]
- 210. Vujaskovic Z, Batinic-Haberle I, Rabbani ZN, Feng QF, Kang SK, Spasojevic I, Samulski TV, Fridovich I, Dewhirst MW, Anscher MS. A small molecular weight catalytic metalloporphyrin antioxidant with superoxide dismutase (SOD) mimetic properties protects lungs from radiationinduced injury. Free Radic Biol Med 2002;33:857–863. [PubMed: 12208373]
- 211. Vujaskovic Z, Feng QF, Rabbani ZN, Samulski TV, Anscher MS, Brizel DM. Assessment of the protective effect of amifostine on radiation-induced pulmonary toxicity. Exp Lung Res 2002;28:577–590. [PubMed: 12396250]
- 212. Waghray M, Cui Z, Horowitz JC, Subramanian IM, Martinez FJ, Toews GB, Thannickal VJ. Hydrogen peroxide is a diffusible paracrine signal for the induction of epithelial cell death by activated myofibroblasts. FASEB J 2005;19:854–856. [PubMed: 15857893]
- 213. Wallach-Dayan SB, Izbicki G, Cohen PY, Gerstl-Golan R, Fine A, Breuer R. Bleomycin initiates apoptosis of lung epithelial cells by ROS but not by Fas/FasL pathway. Am J Physiol Lung Cell Mol Physiol 2006;290:L790–L796. [PubMed: 16306138]
- 214. Walter N, Collard HR, King TE Jr. Current perspectives on the treatment of idiopathic pulmonary fibrosis. Proc Am Thorac Soc 2006;3:330–338. [PubMed: 16738197]
- 215. Wasserman B, Block ER. Prevention of acute paraquat toxicity in rats by superoxide dismutase. Aviat Space Environ Med 1978;49:805–809. [PubMed: 656008]
- 216. Wegener T, Sandhagen B, Chan KW, Saldeen T. N-acetyl-cysteine in paraquat toxicity: toxicological and histological evaluation in rats. Ups J Med Sci 1988;93:81–89. [PubMed: 3376355]
- 217. Wiegman EM, van Gameren MM, Kampinga HH, Szabo BG, Coppes RP. Post-irradiation dietary vitamin E does not affect the development of radiation-induced lung damage in rats. Radiother Oncol 2004;72:67–70. [PubMed: 15236876]
- 218. Wilhelm J, Frydrychova M, Vizek M. Hydrogen peroxide in the breath of rats: the effects of hypoxia and paraquat. Physiol Res 1999;48:445–449. [PubMed: 10783909]
- 219. Xaubet A, Marin-Arguedas A, Lario S, Ancochea J, Morell F, Ruiz-Manzano J, Rodriguez-Becerra E, Rodriguez-Arias JM, Inigo P, Sanz S, Campistol JM, Mullol J, Picado C. Transforming growth factor-beta1 gene polymorphisms are associated with disease progression in idiopathic pulmonary fibrosis. Am J Respir Crit Care Med 2003;168:431–435. [PubMed: 12746254]

- 220. Yamano Y, Kagawa J, Hanaoka T, Takahashi T, Kasai H, Tsugane S, Watanabe S. Oxidative DNA damage induced by silica in vivo. Environ Res 1995;69:102–107. [PubMed: 8608769]
- 221. Yamazaki C, Hoshino J, Hori Y, Sekiguchi T, Miyauchi S, Mizuno S, Horie K. Effect of lecithinizedsuperoxide dismutase on the interstitial pneumonia model induced by bleomycin in mice. Jpn J Pharmacol 1997;75:97–100. [PubMed: 9334891]
- 222. Yang IV, Burch LH, Steele MP, Savov JD, Hollingsworth JW, McElvania-Tekippe E, Berman KG, Speer MC, Sporn TA, Brown KK, Schwarz MI, Schwartz DA. Gene expression profiling of familial and sporadic interstitial pneumonia. Am J Respir Crit Care Med 2007;175:45–54. [PubMed: 16998095]
- 223. Yeh ST, Guo HR, Su YS, Lin HJ, Hou CC, Chen HM, Chang MC, Wang YJ. Protective effects of N-acetylcysteine treatment post acute paraquat intoxication in rats and in human lung epithelial cells. Toxicology 2006;223:181–190. [PubMed: 16713667]
- 224. Yildirim Z, Kotuk M, Iraz M, Kuku I, Ulu R, Armutcu F, Ozen S. Attenuation of bleomycin-induced lung fibrosis by oral sulfhydryl containing antioxidants in rats: erdosteine and N-acetylcysteine. Pulmon Pharmacol Ther 2005;18:367–373.
- 225. Yu TW, Anderson D. Reactive oxygen species-induced DNA damage and its modification: a chemical investigation. Mutat Res 1997;379:201–210. [PubMed: 9357549]
- 226. Zembowicz A, Hatchett RJ, Radziszewski W, Gryglewski RJ. Inhibition of endothelial nitric oxide synthase by ebselen: prevention by thiols suggests the inactivation by ebselen of a critical thiol essential for the catalytic activity of nitric oxide synthase. J Pharmacol Exp Ther 1993;267:1112– 1118. [PubMed: 7505326]
- 227. Zhang K, Phan SH. Cytokines and pulmonary fibrosis. Biol Signals 1996;5:232–239. [PubMed: 8891199]
- 228. Zimmerman MS, Ruckdeschel JC, Hussain M. Chemotherapy-induced interstitial pneumonitis during treatment of small cell anaplastic lung cancer. J Clin Oncol 1984;2:396–405. [PubMed: 6726294]
- 229. Zollner S, Haseloff RF, Kirilyuk IA, Blasig IE, Rubanyi GM. Nitroxides increase the detectable amount of nitric oxide released from endothelial cells. J Biol Chem 1997;272:23076–23080. [PubMed: 9287307]

FIG. 1. Role of ROS in the dysregulation of tissue repair

Several phosphatases contain sensitive thiol residues that are inhibited on oxidation. As steadystate levels of oxidants increase, an increase in the inactivation of phosphatases and a corresponding increase in the levels and duration of phosphorylated proteins occur. An increase in steady-state ROS results in altering the cellular glutathione (GSH)/glutathione disulfide (GSSG) redox couple, which can in turn alter the relative oxidation status of protein thiols. In addition, ROS can alter cell signaling directly by reacting with specific thiols on transcription factors. Many of these types of mechanisms are thought to contribute to the often observed unregulated tissue-repair responses associated lung fibrosis.

 NIH-PA Author ManuscriptNIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH-PA Author Manuscript

Structures of Catalytic Antioxidants with SOD and Catalase Activity

FIG. 3.

Chemical structures of metalloporphyrin catalytic antioxidant mimetics with SOD and catalase activities.

Structures of Catalytic Antioxidants with SOD and Catalase Activity

Structures of Catalytic Antioxidants with SOD Activity

Nitroxides

Chemical structures of nitroxide catalytic antioxidant mimetics with SOD activity.

Structures of Catalytic Antioxidants with Glutathione Peroxidase Activity

Chemical structures of selenium-containing catalytic antioxidant mimetics with glutathione peroxidase activity.

Structures of Antioxidant Scavengers

Table 1

Evidence of Oxidative Stress in IPF

 \overline{a}

Table 2

Models of Lung Fibrosis

