

Repeated independent evolution of obligate pollination mutualism in the Phyllanthae–*Epicephala* association

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The well-known fig–fig wasp and yucca–yucca moth mutualisms are classic examples of obligate mutualisms that have been shaped by millions of years of coevolution. Pollination systems involving obligate seed parasites are only expected to evolve under rare circumstances where their positive effects are not swamped by abundant co-pollinators and heavy costs resulting from seed destruction. Here, we show that, in Phyllanthae, specialization to pollination by *Epicephala* moths evolved at least five times, involving more than 500 Phyllanthae species in this obligate association. Active pollination behaviour evolved once in *Epicephala*, 10–20 Myr after the initial divergence of their host plants. The pollinating *Epicephala* moths thus radiated on an already-diverged host lineage and successively colonized new Phyllanthae hosts, thereby giving rise to repeated independent evolution of the specialized pollination system in Phyllanthae. The present evolutionary success of this association rests entirely upon active pollination by *Epicephala*, making this a distinct example of an evolutionary key innovation. Overall, our findings provide a clear empirical demonstration of how a combination of evolutionary innovation and partner shifts facilitates the spread of mutualism in a coevolving species interaction.

Keywords: active pollination behaviour; *Epicephala*; key innovation; obligate pollination mutualism; Phyllanthae

1. INTRODUCTION

The well-known obligate fig–fig wasp and yucca–yucca moth mutualisms have become principal model systems for the study of coevolution (Weiblen 2002; Cook & Rasplus 2003; Pellmyr 2003). Female fig wasps and yucca moths pollinate the flowers in which they lay eggs, and the plants sacrifice a fraction of their developing seeds to nourish pollinator larvae. Despite a wealth of documented examples of specialized pollination systems in angiosperms, however, pollination by obligate seed parasites is rare. This is because seed parasitism inflicts a heavy cost on plants, while abundant co-pollinators swamp the mutualistic effect of pollination by seed parasites (Thompson & Pellmyr 1992; Thompson & Cunningham 2002). Consequently, there are only a handful of mutualisms wherein obligate seed parasites act as effective pollinators of their host plants (Pellmyr 1989; Pellmyr *et al.* 1996a; Fleming & Holland 1998).

A novel example of such a coevolved obligate pollination mutualism has recently been reported between Phyllanthae plants (Phyllanthaceae) and *Epicephala* moths (Gracillariidae) (Kato *et al.* 2003; Kawakita & Kato 2004a,b). Phyllanthae is a pantropical tribe of more than 1200 species having a remarkable diversity of growth forms, ranging from annual and perennial herbs, shrubs and trees to climbers, succulents, rheophytes and aquatics

(Hoffmann *et al.* 2006; Kathriarachchi *et al.* 2006). The plants have minute, unisexual flowers that are borne as clusters on leaf axils (figure 1). Production of nectar is variable across taxa, which probably reflects differences in pollination systems. Species belonging to *Glochidion* (more than 300 spp.), *Breynia* (35 spp.) and *Phyllanthus* subgenus *Gomphidium* (more than 150 spp.) are each pollinated nocturnally by a species-specific *Epicephala* moth whose larvae feed on the seeds (figure 1). The female moths have evolved to actively collect and transport pollen between flowers using specialized proboscides equipped with numerous sensilla (Kawakita & Kato 2006). They lay eggs in the flowers they pollinate, and the larvae consume a fraction of the developing seeds, leaving others viable for plant reproduction.

Although the phylogenetic classification of Phyllanthae has been the subject of previous investigations (Kathriarachchi *et al.* 2005, 2006), virtually nothing is known about pollination biology in the remaining groups of Phyllanthae. We therefore studied pollination systems and associations with *Epicephala* in 26 species of Phyllanthae during 2002–2007 in Southeast Asia, New Caledonia, Australia, Madagascar, Guinea and North America (table 1). Based on the results of the pollination study, we explored the origin of the Phyllanthae–*Epicephala* mutualism by reconstructing robust phylogenies for 46 species of Phyllanthae and associated *Epicephala* moths. While the present study focuses on only a small proportion of the global diversity of Phyllanthae, the sampled species cover the entire range of taxonomic diversity within the tribe (Hoffmann *et al.* 2006; Kathriarachchi *et al.* 2006),

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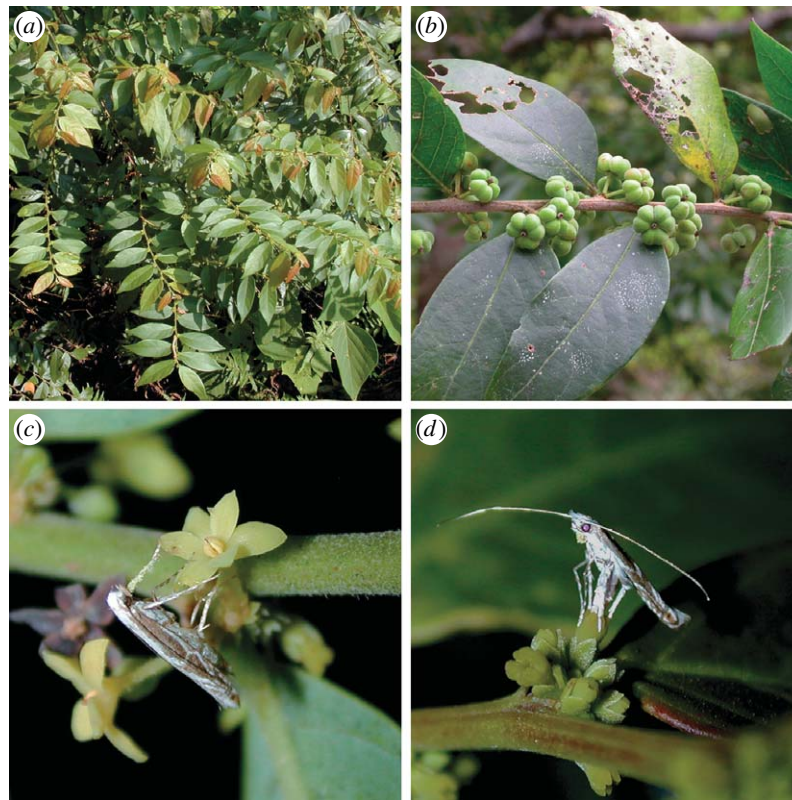


Figure 1. Phyllanthaceae–*Epicephala* obligate pollination mutualism. (a) *Glochidion acuminatum* is pollinated by a species-specific *Epicephala* moth that infests (b) the fruit as larvae. The adult female moth (approx. 5 mm in body length) (c) actively collects pollen on male flowers and (d) pollinates the female flower in which she lays an egg.

allowing a sound investigation into broad coevolutionary history of the Phyllanthaceae–*Epicephala* association. Overall, our results reveal an unexpectedly complex origin of the Phyllanthaceae–*Epicephala* pollination mutualism and provide important general insights into how a combination of evolutionary innovation and partner shifts shapes the evolutionary dynamics of mutualism in coevolving species interactions.

2. MATERIAL AND METHODS

(a) *Determination of pollination systems*

We determined whether a species is *Epicephala*-pollinated or not by examining the presence/absence of *Epicephala* eggs and larvae in female flowers and fruits, respectively. We judged that a species has a non-*Epicephala* pollination system if no larvae were found in the sampled fruits (table S1 in the electronic supplementary material). Whenever possible, this was further confirmed by the absence of eggs in pollinated female flowers under a light microscope. Species that had *Epicephala* larvae in fruits were further examined for eggs in pollinated female flowers (tables S1 and S2 in the electronic supplementary material). If the pollinated status of a female flower and the presence of *Epicephala* eggs are coupled, it is likely that the species is *Epicephala*-pollinated because the female moth invariably lays an egg in the flower that she pollinated (Kato *et al.* 2003; Kawakita & Kato 2004a,b). In turn, a lack of association or absence of eggs in female flowers indicates that *Epicephala* moths are not involved in pollination. Pollination status was determined by checking for the presence of pollen grains on the stigma under a light microscope. Overall, we dissected 19–461 fruits representing 1–31 plants for each of 19 species to examine the presence of seed feeding by *Epicephala* larvae. Also, we looked for

Epicephala eggs in 25–393 female flowers representing 2–13 plants in 24 species.

Inference of pollination systems based on the above methods was further substantiated by field observations of diurnal and nocturnal flower visitors (table S3 in the electronic supplementary material). We paid special attention to the behaviour of *Epicephala* to determine whether the moths exhibited pollinating behaviour. In herbaceous species of *Phyllanthus* that belong to subgenera *Isocladius* and *Phyllanthus*, ants were the predominant flower visitors (table S3 in the electronic supplementary material). Because antibiotic substances secreted from ant metapleural glands can inhibit pollen germination (Beattie *et al.* 1984), we experimentally tested the effectiveness of ant pollination in *Phyllanthus lepidocarpus* (see the electronic supplementary material).

Although our inference of pollination system does not rely on pollinator exclusion or pollen deposition experiments, which provide a more direct evidence of pollinator contributions, the above approach has proven to be reliable in previous studies (Kato *et al.* 2003; Kawakita & Kato 2004a,b) and consistently leads to unambiguous inference of whether or not a species is *Epicephala*-pollinated (table 1; tables S1–S3 in the electronic supplementary material); thus, pollination systems inferred as such can be reliably combined with phylogenetic results to explore an overall evolutionary pattern of *Epicephala* pollination in Phyllanthaceae.

(b) *Quantification of style spreading*

Of the 46 Phyllanthaceae species sampled in this study, information on the pollination systems for 40 species was available from previous literature and field data collected as described above. To infer pollination systems for the other six species, we quantified the degree of style spreading, which

Table 1. List of species studied.

species sampled ^a	abbreviation	study site/sampling locality	<i>Epicephala</i> as the pollinator	criteria for pollinator determination ^b	style spreading
<i>Margaritaria</i>					
<i>M. discoidea</i> (Baill.) G. L. Webster	Mdis	Guinea: Bossou	no	E (fr), M	7.07
<i>M. indica</i> (Dalziel) Airy Shaw	Mind	Japan: Okinawa Island	no	M	5.46
<i>Flueggea</i>					
<i>F. jullienii</i> (Beille) G. L. Webster	Fjul	Laos: Mahaxai	no	M	4.99
<i>F. suffruticosa</i> Baill.	Fsuf	Japan: Hyogo/Hiroshima/ Amami Islands	no	E (fv, fr, fl), M	4.82
<i>F. virosa</i> (Willd.) Voigt	Fvir	Laos: Vieng Xai/Taiwan: Fangliao	no	E (fv, fr), M	4.47
<i>Phyllanthus</i>					
<i>P. (Is.) ussuriensis</i> Rupr. & Maxim.	Puss	Japan: Tokyo/Kyoto	no	E (fv, fr, fl), M	7.01
<i>P. (Is.) virgatus</i> G. Forst	Pvir	Laos: Vientiane	no	E (fv, fr), M	5.97
<i>P. (Er.) liukiensis</i> Hayata	Pliu	Japan: Okinawa Island	no	E (fr, fl), M	7.87
<i>P. (Er.) pulcheroides</i> Beille	Ppul	Laos: Mahaxai	no	E (fr, fl), M	8.69
<i>P. (Ki.) reticulatus</i> Poir.	Pret	Taiwan: HENCHUN	yes	E (fv, fr, fl), M	0.45
<i>P. (Ki.)</i> sp.	Psp	Laos: Laksao	yes	E (fl), M	0.50
<i>P. (Ki.) flexuosus</i> Müll. Arg.	Pfle	Japan: Kyoto/Hyogo/Miyazaki	no	E (fv, fr, fl), M	4.87
<i>P. (Ki.) oligospermus</i> Hayata	Poli	Japan: Yonaguni Island	no	E (fv, fr, fl), M	4.96
<i>P. (Ki.) tenellus</i> Roxb.	Pten	Japan: Okinawa Island	no	E (fr, fl), M	6.41
<i>P. (Ph.) amarus</i> Schumach. & Thonn.	Pama	Japan: Ishigaki Island/Laos: Thakhaek	no	E (fv, fr, fl), M	4.09
<i>P. (Ph.) debilis</i> Willd.	Pdeb	Japan: Ishigaki Island	no	E (fv, fr, fl), M	4.32
<i>P. (Ph.) lepidocarpus</i> Siebold & Zucc.	Plep	Japan: Kyoto/Miyako Island/ Ishigaki Island	no	E (fv, fr, fl), M	3.12
<i>P. (Go.) aeneus</i> Baill.	Paen	New Caledonia: Cap Bocage	yes	L, M	1.08
<i>P. (Go.) gneissicus</i> S. Moore	Pgne	New Caledonia: Mt Panié	yes	L	n.a.
<i>P. (Go.) guillauminii</i> Däniker	Pgui	New Caledonia: Tiébaghi	yes	L	n.a.
<i>P. (Go.) vulcani</i> Guillaumin	Pvul	New Caledonia: Riviere Bleue	yes	L, M	0.62
<i>P. (Go.) bourgeoisii</i> Baill.	Pbou	New Caledonia: Cap Bocage	yes	L, M	0.38
<i>P. (Go.) chamaecerasus</i> Baill.	Pcha	New Caledonia: Chutes de Ba	yes	L	n.a.
<i>P. (Go.) caudatus</i> Müll. Arg.	Pcau	New Caledonia: Riviere Bleue	yes	L	n.a.
<i>P. (Go.) koniamboensis</i> M. Schmid	Pkon	New Caledonia: Tinip	yes	L	n.a.
<i>P. (Go.) mangenotii</i> M. Schmid	Pman	New Caledonia: Cap Bocage	yes	L, M	0.49
<i>P. (Ci.) acidus</i> (L.) Skeels	Paci	Laos: Vientiane (cultivated)	no	L, E (fr), M	2.50
<i>P. (Em.) emblica</i> L.	Pemb	Laos: Ban Chomesy	no	L, E (fr)	n.a.
<i>P. (Pd.) roseus</i> Beille	Pros	Laos: Phialat	no	E (fv, fr, fl), M	1.99
<i>P. marojejiensis</i> (Leandri) Petra Hoffm. & McPherson	Pmar	Madagascar: Mt Marojeji	yes	E (fv, fr, fl), M	0.18
<i>P. humbertii</i> (Leandri) Petra Hoffm. & McPherson	Phum	Madagascar: Mt Marojeji	yes	E (fr), M	0.39
<i>Reverchonia</i>					
<i>R. arenaria</i> A. Gray	Rare	USA: New Mexico	no	E (fv, fr), M	1.87
<i>Sauropus</i>					
<i>S. androgynus</i> Merr.	Sand	Laos: Thakhaek	no	E (fr, fl), M	2.03
<i>S. brevipes</i> Müll. Arg.	Sbre	Laos: Vientiane	no	E (fr, fl), M	2.14
<i>S. granulosus</i> Airy Shaw	Sgra	Laos: Vientiane	no	E (fv, fl), M	2.04
<i>S. quadrangularis</i> Müll. Arg.	Squa	Laos: Vientiane	no	E (fv, fr, fl), M	2.53
<i>Breynia</i>					
<i>B. disticha</i> Forst.	Bdis	New Caledonia: Koumac	yes	M	0.25
<i>B. fruticosa</i> (Müll. Arg.) Hook.f.	Bfru	Laos: Vientiane	yes	L, M	1.45
<i>B. oblongifolia</i> Müll. Arg.	Bobl	Australia: Windsor Tableland	yes	M	0.20
<i>B. retusa</i> Alston	Bret	Laos: Vientiane	no	E (fv, fr, fl), M	3.02
<i>B. vitis-idaea</i> (Burm.f.) C. E. C. Fisch.	Bvit	Japan: Amami Islands	yes	L, M	0.43
<i>Glochidion</i>					
<i>G. acuminatum</i> Müll. Arg.	Gacu	Japan: Amami Islands	yes	L, M	0.86
<i>G. lanceolatum</i> Hayata	Glan	Japan: Ishigaki Island	yes	L, M	0.31
<i>G. obovatum</i> Siebold & Zucc.	Gobo	Japan: Wakayama	yes	L, M	0.93

(Continued.)

Table 1. (Continued.)

species sampled ^a	abbreviation	study site/sampling locality	<i>Epicephala</i> as the pollinator	criteria for pollinator determination ^b	style spreading
<i>G. rubrum</i> Blume	Grub	Japan: Ishigaki Island	yes	L, M	0.87
<i>G. zeylanicum</i> (Gaertn.) A. Juss.	Gzey	Japan: Okinawa Island	yes	L, M	0.24

^a *Phyllanthus* subgenera are abbreviated as follows: *Is.*, *Isocladus*; *Er.*, *Eriococcus*; *Ki.*, *Kirganelia*; *Ph.*, *Phyllanthus*; *Ci.*, *Cicca*; *Em.*, *Embllica*; *Pd.*, *Phyllanthodendron*. *Phyllanthus marojejiensis* and *Phyllanthus humbertii* have not been assigned to any subgenus.

^b Each species was judged as either *Epicephala* or non-*Epicephala* pollinated based on literature information (L), ecological data (E) and/or style morphology of the female flower (M). Ecological data consisted of direct observation of flower visitors (fv) and/or examination of *Epicephala* larvae/eggs in fruits (fr)/flowers (fl), respectively.

reflects a syndrome associated with *Epicephala* pollination. In species pollinated by the moths, styles are reduced and fused to form a narrow apical cavity into which moths actively deposit pollen (figure 2; Kato *et al.* 2003; Kawakita & Kato 2004a,b). By contrast, species diurnally pollinated by various nectar-seeking insects usually have bifid styles that are spread horizontally, which facilitates passive pollen receipt from insect bodies (figure 2). Style spreading is a continuous, quantitative measure that does not by itself provide information about pollination system. However, species with different pollination syndromes had non-overlapping distributions of style spreading, and the six species with unknown pollination systems were nested well within either of the cohorts (see §3); thus, we were able to reliably assign their pollination modes despite lack of ecological data.

We quantified style spreading as the ratio of apical-to-basal style width using 3–45 flowers representing 1–10 plants for each species in 40 *Phyllanthaceae* taxa. Measurements of style width were done under a light microscope equipped with an eyepiece micrometer using either fresh plant materials or specimens preserved in 70 per cent ethanol. Because flowers of *Phyllanthaceae* are trimerous, the apical styler width was represented by the diameter of the minimum circle circumscribing the triangle formed by the three styles. The average lengths of the three sides were used to calculate the diameter by approximating the triangle to be equilateral. The left arms of the styles were used for measurements in species with bifid styles.

(c) Molecular phylogenetic analysis

DNA extraction and sequencing followed previously described methods (Kawakita *et al.* 2004; Kawakita & Kato 2006). The phylogenetic analysis of the 46 species of *Phyllanthaceae* was based on sequences of the chloroplast *matK*, *ndhF*, *atpB* and nuclear *PHYC* genes. In addition to the species sampled here, we included 46 sequences of other members of *Phyllanthaceae* and four outgroup taxa representing *Picrodendraceae* and *Putranjivaceae* that were available from a previous study (Kathriarachchi *et al.* 2005) to allow fossil calibrations in the following divergence time estimation. Species with more than one missing gene in Kathriarachchi *et al.* (2005) were not included, to avoid potential confounding effects of large amounts of missing data on phylogenetic reconstruction and divergence time estimation (Douzery *et al.* 2004; Wiens 2006). We also reconstructed the phylogeny of the associated 26 *Epicephala* species, which were each specific to one, or rarely two, *Phyllanthaceae* species. These moths are currently all undescribed but can be clearly distinguished based on male genital morphology. The phylogenetic analysis of *Epicephala* was

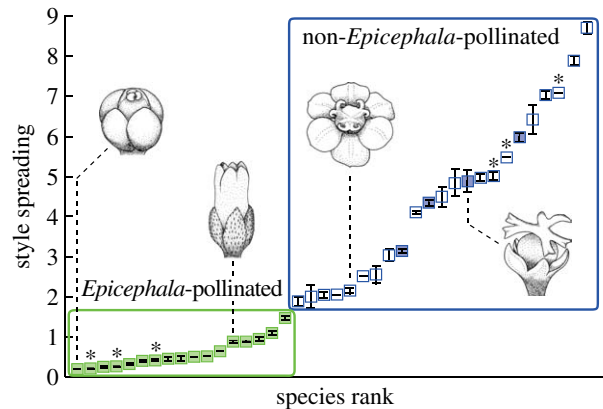


Figure 2. Distribution of style spreading in *Phyllanthaceae*. Species pollinated by *Epicephala* have reduced styles that are medially fused, whereas non-*Epicephala*-pollinated species have horizontally spread, bifid styles. Filled and open boxes indicate species with and without associations with *Epicephala*, respectively. Ecological data were not available for species with asterisks. Female flowers are drawn for *Phyllanthus marojejiensis*, *G. acuminatum*, *Sauropus brevipes* and *Flueggea suffruticosa* (from left to right). Error bars, ± 1 s.e.

based on sequences of the mitochondrial *COI*, nuclear arginine kinase (*ArgK*), elongation factor-1 alpha (*EF-1 α*), wingless (*Wg*) and 18S *rDNA* genes. For the moth outgroups, we sampled three gracillariid moths: *Cuphodes diospyrosella*, *Stomphastis labyrinthica* and *Melanocercops ficuvorella*. The last species was included as the farthest outgroup based on a larger phylogenetic analysis of the subfamily based on *EF-1 α* (A. Kawakita & M. Kato 2007, unpublished data; also see the electronic supplementary material for details of sampling). Polymerase chain reactions were done using primers and protocols provided previously (Brower & DeSalle 1998; Heraty *et al.* 2004; Kathriarachchi *et al.* 2005; Kawakita & Kato 2006) and additional *matK* primers (listed in the electronic supplementary material, table S4). Available plant and moth voucher specimens are preserved in the Kyoto University Museum, and all newly obtained sequences have been deposited in the GenBank database under accession numbers FJ235206–FJ235514 (see tables S5 and S6 in the electronic supplementary material for the full list of accession numbers).

Phylogenetic analysis was conducted using maximum-parsimony, maximum-likelihood (ML) and Bayesian methods. Maximum-parsimony heuristic searches were done in PAUP* v. 4.0.b10 (Swofford 2002) with 100 random addition analyses and tree bisection–reconnection branch swapping, and the robustness of trees was assessed by non-parametric bootstrapping with 1000 replicates. We also performed ML heuristic searches using PAUP* with 10 random addition analyses and subtree pruning–regrafting

branch swapping. The ApproxLim parameter was adjusted to 2 to accelerate the tree search (Morrison 2007). Appropriate models of base substitution were selected using MODELTEST v. 3.06 (Posada & Crandall 1998). The robustness of ML trees was validated with non-parametric bootstrapping using PHYML (Guindon & Gascuel 2003) with 500 replicates. We performed a Bayesian analysis using MRBAYES v. 3.1.2 (Ronquist & Huelsenbeck 2003), with unlinked models for each partition. The analysis consisted of two million generations, sampling trees every 1000 generations for a total of 2001 trees. We plotted ln-likelihood of the sampled trees against generation time to identify the region of the analysis in which the parameter estimates were stable, and accordingly discarded the initial 1001 trees as burn-in.

Because *Epicephala*-pollinated Phyllanthae species were not recovered as monophyletic (see §3), we tested the robustness of this reconstruction using an approximately unbiased test (Shimodaira 2002). Similarly, we tested the monophyly of the actively pollinating *Epicephala* species, which were also recovered as non-monophyletic (see §3). The tests were conducted using CONSEL (Shimodaira & Hasegawa 2001), with alternative hypotheses obtained by constraining the monophyly of *Epicephala*-pollinated Phyllanthae species or that of actively pollinating *Epicephala*, and performing ML searches as described above.

Major clades of *Epicephala* were generally associated with well-defined taxonomic groups of Phyllanthae (see §3), but relationships at higher levels were not congruent between the two phylogenies. We therefore tested the cophylogenetic association between plant and moth phylogenies at higher taxonomic levels using PARAFIT (Legendre *et al.* 2002). To focus on phylogenetic relationships at higher levels, terminal taxa in the plant phylogeny were pruned to major plant taxonomic groups, and those in the moth phylogeny to well-defined *Epicephala* clades having exclusive associations with major host groups (see the electronic supplementary material).

(d) Local ML ancestral state reconstruction

To investigate the evolutionary history of the Phyllanthae–*Epicephala* mutualism, we conducted local ML ancestral character state reconstructions using BAYESMULTISTATE (Pagel *et al.* 2004) and ML phylogenies of Phyllanthae and *Epicephala*. We first tested for a difference in transition rates between the two states (presence/absence of *Epicephala* pollination in Phyllanthae and pollination behaviour in *Epicephala*) by comparing likelihoods between the one-rate and the two-rate model. We used a likelihood ratio test to determine statistical significance following a χ^2 -distribution with one degree of freedom. Because there was no significant difference between the two rates in either the plant or moth phylogeny, the one-rate model was used in ancestral state reconstructions (Pagel 1999). A likelihood ratio of greater than 2 was considered significant evidence for the occurrence of either state at ancestral nodes (Pagel 1999). The common ancestor of Phyllanthae and its sister outgroup clade was assumed to have a non-*Epicephala* pollination system because none of the plants outside Phyllanthae are known to have associations with *Epicephala*. Similarly, the common ancestor of *Epicephala* and its sister clade was assumed to be a non-pollinator, because the rest of the gracillariid moths are generally leaf miners and do not visit flowers. The robustness of the results to phylogenetic uncertainty was validated using 100 subset trees from the Bayesian analysis and performing the reconstructions as described above, which indicated that

all significant reconstructions were stable in at least 70 per cent of the trees tested (results not shown).

(e) Divergence time estimation

The above analysis of ancestral character state reconstruction indicated that *Epicephala*-pollinated Phyllanthae plants evolved multiple times independently (see §3). However, because our taxon sampling was limited to 46 species amid the global diversity of Phyllanthae, results of ancestral state reconstruction might change with the addition of more taxa. We therefore estimated divergence times for the Phyllanthae and *Epicephala* phylogenies to test whether the multiple-origins hypothesis is in fact the preferred scenario. If the age of the most recent common ancestor (MRCA) of moth-pollinated plants is contemporary with that of *Epicephala*, a single origin of the mutualism followed by multiple losses would still be a viable hypothesis. Alternatively, evolution of pollinating behaviour post-dating initial host divergence would provide strong support for the multiple-origins hypothesis.

A likelihood ratio test rejected the assumption of a strict molecular clock in both the plant and moth phylogenies ($p < 0.0001$); thus, we used the penalized-likelihood (Sanderson 2002) and Bayesian methods (Thorne & Kishino 2002) to estimate divergence times using the ML phylogenies for each group. For the penalized-likelihood approach, we used the program r8s (Sanderson 2002) with smoothing parameters of 10 and 5 obtained by cross-validation for the plant and moth phylogenies, respectively. Confidence intervals were obtained by means of non-parametric bootstrapping, which consisted of generating 100 bootstrap datasets with SEQBOOT (Felsenstein 1993), estimating branch lengths of the ML topology with each of these datasets under the original substitution model and estimating divergence times as described. The Bayesian relaxed-clock approach used was that implemented in MULTIDIVTIME (Thorne & Kishino 2002), with model parameters obtained using PAML (Yang 1997). The analysis consisted of 100 000 burn-in cycles and 1 million post-burn-in cycles, with sampling at every 100th tree. Prior distributions on parameters for the Bayesian analysis are provided in the electronic supplementary material.

We used the following four non-redundant fossils as minimum age constraints for the Phyllanthaceae phylogeny (table S7 in the electronic supplementary material): *Bischofia*-type pollen from Bartonian, Middle Eocene (37.2 Myr ago); *Actephila*-type pollen from Late Eocene (33.9 Myr ago); *Phyllanthus*-type pollen from Early Eocene (48.6 Myr ago; Gruas-Cavagnetto & Köhler 1992); and *Glochidion* leaf impressions from Middle Miocene (11.6 Myr ago; Prasad 1994; Antal & Prasad 1996). We initially constrained the root node (i.e. the node splitting Phyllanthaceae and Picrodendraceae) to be no older than 125 Myr ago, a well-established maximum age of the eudicot clade based on the earliest known occurrence of tricolpate pollen (Magallón *et al.* 1999). However, because Phyllanthaceae is nested well within the phylogeny of eudicots, we also used a more conservative maximum age of 108 Myr ago for our basal node, which is the oldest estimate of the corresponding node in a previous study of Malpighiales radiation (Davis *et al.* 2005). Because attribution of some of the Phyllanthaceae fossils may still need refinements (Gruas-Cavagnetto & Köhler 1992), caution may be necessary when taking the precise dates resulting from our analysis. For the details of Phyllanthaceae fossils, see the electronic supplementary material.

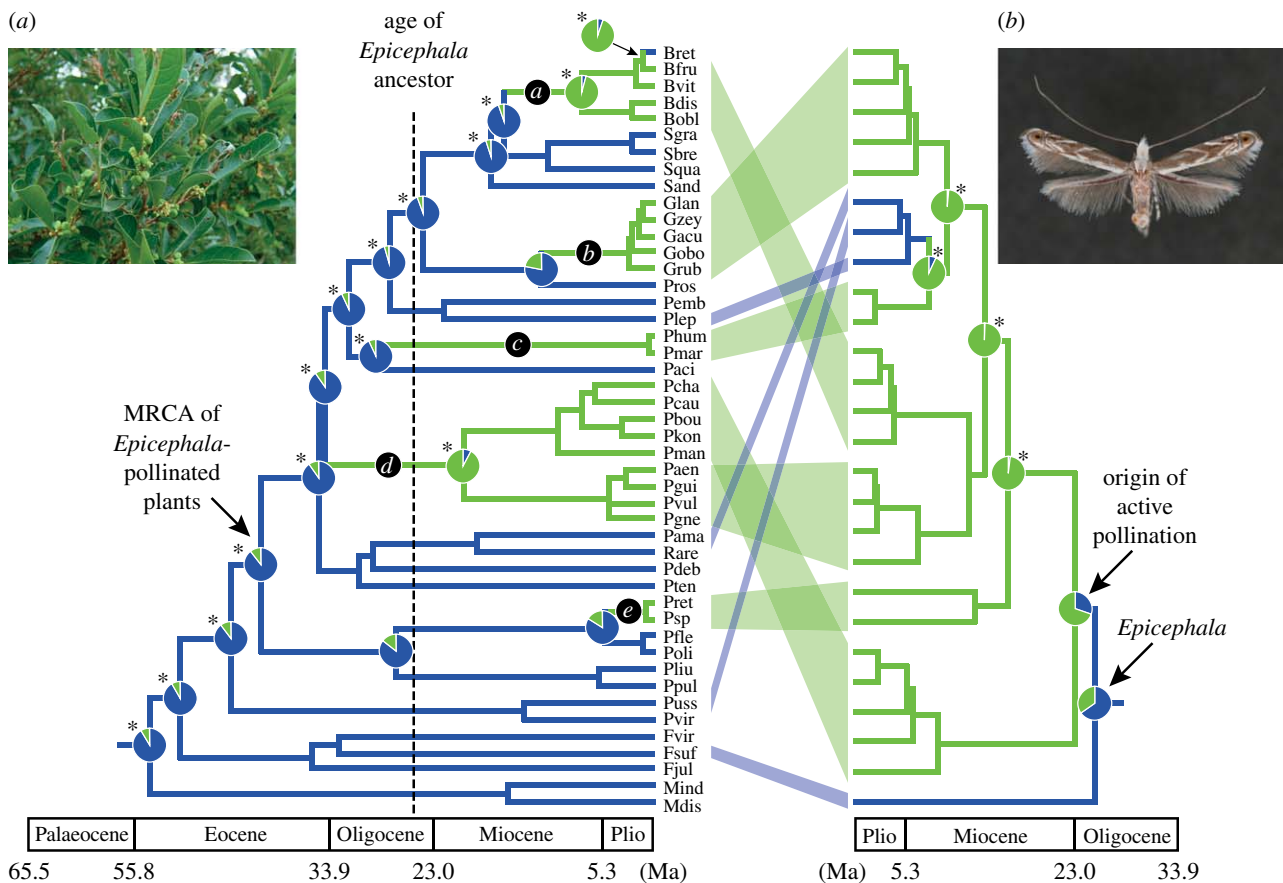


Figure 3. Phylogenetic analysis and the origin of the Phyllanthae–*Epicephala* mutualism. ML chronograms for (a) Phyllanthae plants (Phyllanthaceae root node constrained to be no older than 108 Myr ago) and (b) associated *Epicephala* moths. Pie charts indicate the probability of *Epicephala* and non-*Epicephala* pollination systems (Phyllanthae) or the presence/absence of active pollination behaviour (*Epicephala*) occurring at ancestral nodes. Asterisks indicate significant differences in likelihoods. The *Epicephala* tree is calibrated with 25 Myr ago at the root node. Mutualism is represented in green, and associations between major plant and moth clades are indicated. See table 1 for terminal taxon names.

Because gracillariid moths are extremely scarce in the fossil record (Lopez-Vaamonde *et al.* 2006), *Epicephala* divergence times were obtained by estimating relative divergence times for the *Epicephala* phylogeny and calibrating node ages based on the substitution rate of the *COI* gene. We used only the *COI* substitution rate because it is generally conserved across arthropod taxa (Gaunt & Miles 2002), has been widely used for dating in insects (Kandul *et al.* 2004; Quek *et al.* 2007; Ueda *et al.* 2008) and clusters at approximately $1.5\% \text{ Myr}^{-1}$ in several arthropod groups (Farrell 2001; Quek *et al.* 2004; Sota & Hayashi 2007). To obtain relative divergence times for the *Epicephala* phylogeny, we forced the root node (i.e. the split between *Epicephala* and *C. diospyrosella*) to have one arbitrary time unit in both penalized-likelihood and Bayesian analyses. Every node on the obtained ultrametric tree was then individually used as a calibration point to obtain the range of age estimates for the MRCA of *Epicephala*. *COI* distances were corrected using the appropriate substitution model (GTR+ Γ +I) and model parameters inferred by MODELTEST, and the average pairwise *COI* distance across each node was transformed to absolute age using the $1.5\% \text{ Myr}^{-1}$ substitution rate.

In addition to using relative node ages inferred from relaxed-clock methods, we also estimated the age of the *Epicephala* root node by calculating branch lengths of the ML tree with the *COI* data under the assumption of a strict clock. Branch lengths were calculated in PAUP* with the above GTR+ Γ +I substitution model, and node height of the

Epicephala root node on the resulting ultrametric tree was transformed to absolute age using the $1.5\% \text{ Myr}^{-1}$ substitution rate. We obtained 95% credibility interval of the estimated age by generating 100 bootstrap datasets with SEQBOOT and calculating the root age as described. Although a likelihood ratio test rejected a strict molecular clock in the *Epicephala* phylogeny (see above), we conducted this analysis as complementary to the relaxed-clock methods because the use of constant *COI* substitution rate inherently assumes an underlying molecular clock.

3. RESULTS AND DISCUSSION

We found two additional Phyllanthae lineages that are obligately pollinated by host-specific *Epicephala* moths (see the electronic supplementary material), namely *Phyllanthus* section *Anisonema* with approximately 20 species throughout the Palaetropics and an unclassified group of *Phyllanthus* endemic to Madagascar (approx. 10 species). The remaining species were pollinated by diurnal insects that visited flowers for nectar and pollen, and did not have associations with pollinating *Epicephala* (tables S1 and S3 in the electronic supplementary material). However, *Flueggea suffruticosa* and three herbaceous *Phyllanthus* species were parasitized by seed-parasitic *Epicephala* species that did not pollinate the flowers. These plants were effectively pollinated by bees, flies, beetles or ants (table S8 in the electronic supplementary material); thus, the associated *Epicephala* are truly

parasitic on their hosts. Overall, species with different pollination syndromes had non-overlapping degrees of style spreading (figure 2); thus, we were able to reliably assign pollination systems to plant species for which sufficient ecological data were not available.

To investigate the origin of the Phyllanthae–*Epicephala* mutualism, we reconstructed the phylogeny of 46 Phyllanthae species and mapped whether each species is pollinated by *Epicephala*. Data matrix of the combined chloroplast *matK*, *ndhF*, *atpB* and nuclear *PHYC* genes consisted of 4059 bp aligned sequences. Maximum-parsimony, likelihood and Bayesian analyses all produced a highly resolved and well-supported phylogeny for Phyllanthae (fig. S1 in the electronic supplementary material). Species pollinated by *Epicephala* were not monophyletic (approximately unbiased test; $p < 0.0001$), indicating that there have been multiple shifts in pollination systems. To determine the direction of pollinator shifts, we reconstructed ancestral character states for pollination systems using the local maximum likelihood. The results clearly indicated five independent origins of the obligate pollination mutualism in Phyllanthae, with a single reversal to non-*Epicephala* pollination in *Breynia retusa* (figure 3).

We also reconstructed the phylogeny of 26 *Epicephala* species associated with the above Phyllanthae species. Analysis of the combined 2933 bp aligned sequences of the mitochondrial *COI*, nuclear *ArgK*, *EF-1 α* , *Wg* and *18S rDNA* genes produced a well-resolved phylogeny, although the earliest branching lineage within *Epicephala* remained ambiguous (fig. S2 in the electronic supplementary material). The pollinator species were non-monophyletic (approximately unbiased test; $p < 0.0001$), and ancestral character state reconstruction indicated a likely single origin of pollination behaviour with a single event of secondary loss (figure 3). Major clades of *Epicephala*, generally, had specific associations with well-defined taxonomic groups of Phyllanthae (figure 3; fig. S2 in the electronic supplementary material), but relationships at higher levels were not different from random (PARAFIT; $p = 0.39$), indicating that host shifts have occurred repeatedly.

Divergence time estimates using relaxed-clock methods and fossil calibrations indicated that the MRCA of *Epicephala*–pollinated plants occurred 41.0 Myr ago (penalized likelihood; 95% credibility interval, 39.3–48.3 Myr ago) or 40.4 Myr ago (Bayesian method; 33.4–48.2 Myr ago), when constraining the root node at 108 Myr ago (fig. S3 in the electronic supplementary material). When using the older maximum age for the root node (125 Myr ago), the corresponding estimates were 47.4 Myr ago (penalized likelihood; 45.0–55.2 Myr ago) and 42.2 Myr ago (Bayesian method; 34.2–51.8 Myr ago). By contrast, estimated ages of the *Epicephala* root node clustered within a time frame between 20 and 35 Myr ago (figure 4; also see the electronic supplementary material and fig. S4 therein) and converged at 20–30 Myr ago as deeper nodes were used to calibrate the root age. Because fixing ages at shallowly placed nodes for deep extrapolations can be prone to large systematic errors (Ho *et al.* 2005; Sota & Hayashi 2007), the range of 20–30 Myr ago probably provides a robust time frame for the *Epicephala* root node, which is consistent with the estimates obtained from the *COI* molecular clock phylogeny (optimal, 22.76 Myr ago; 95% confidence interval, 18.6–29.1 Myr ago). These

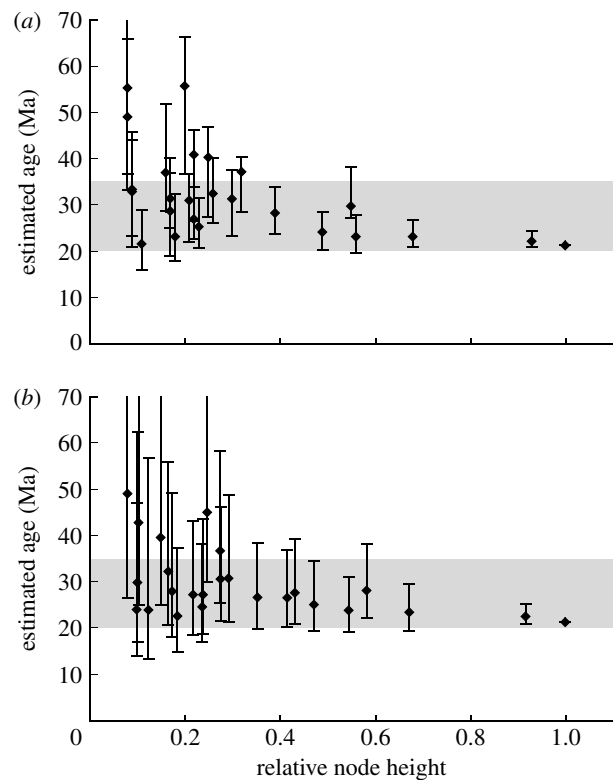


Figure 4. Estimated ages for the most recent common ancestor of *Epicephala*. Optimal ages and 95% credibility intervals are plotted against relative node heights of those used to calibrate the *Epicephala* chronogram. The majority of estimates fall within a time window of 20–35 Myr ago (shaded). (a) Penalized-likelihood and (b) Bayesian estimation.

estimates for the age of *Epicephala* post-date initial host divergence by roughly 10–20 Myr, which is consistent with delayed radiation of *Epicephala* and hence multiple origins of the obligate pollination mutualism in Phyllanthae. Although our estimates of the timing of *Epicephala* divergence depend largely on the accuracy of the *COI* substitution rate, the assumed 1.5% Myr⁻¹ is among the slowest of known substitution rates for the arthropod *COI* gene (1.3–2.3% Myr⁻¹; Brower 1994; Quek *et al.* 2004), and using higher rates would only give younger estimates for the age of the *Epicephala* root node; thus, our method is conservative with respect to providing young ages.

Because our taxon sampling was limited to 20 per cent of the global diversity of Phyllanthae at the section level (Kathriarachchi *et al.* 2006) and less than 5 per cent at the species level, the entire picture of the evolutionary history of *Epicephala* pollination in Phyllanthae is probably much more complex than as presented here. However, inclusion of other lineages would probably only strengthen our conclusion of repeated independent evolution because these plants have bifid, horizontally spread styles that are characteristic of non-*Epicephala*–pollinated plants (figure 2). An exception is the New World *Phyllanthus* subgenus *Xylophylla*, which consists of approximately 60 species having reduced, columnar styles (Webster 1958) and thus may represent an additional origin of *Epicephala* moth pollination in Phyllanthae (figure 5).

Our finding that the obligate pollination mutualism arose repeatedly in Phyllanthae is in stark contrast with the situations in the fig–fig wasp and yucca–yucca moth mutualisms. Recent coevolutionary analyses in the fig and

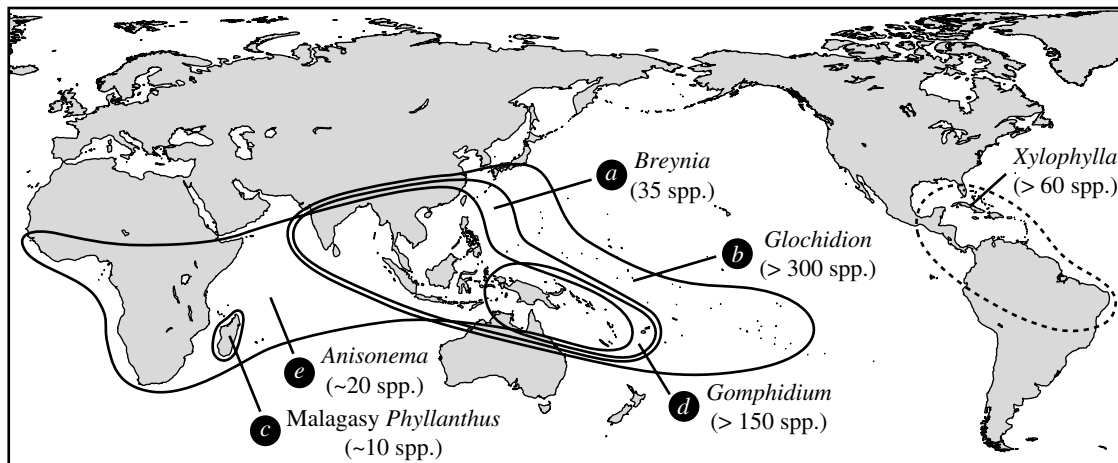


Figure 5. Five independent origins of the Phyllanthae–*Epicephala* obligate pollination mutualism. The New World subgenus *Xylophylla* shares the reduced, columnar style morphology with other *Epicephala*-pollinated plants and may represent an additional origin of the mutualism. Letters in circles correspond to clade names in figure 3.

yucca systems have indicated that these associations arose only once in each partner lineage 40–60 Myr ago (Pellmyr & Leebens-Mack 1999; Rønsted *et al.* 2005). An exception is *Hesperoyucca whipplei*, which is phylogenetically distant from the rest of the yuccas and independently established the mutualism with a yucca moth (Bogler *et al.* 1995; Pellmyr *et al.* 2007; Smith *et al.* 2008). In the Phyllanthae–*Epicephala* system, major lineages of Phyllanthae had already emerged when *Epicephala* colonized these plants *ca* 30 Myr ago. Sequential radiation of *Epicephala* on an already-diverged host lineage has probably provided opportunities for the moth pollinators to establish new mutualistic associations in distant host lineages. Thus, specialization to moth pollination occurred multiple times independently in Phyllanthae as *Epicephala* evolved to use a broad range of the Phyllanthae lineage.

Our results also indicate that colonization of new host lineages by the pollinators sometimes results in a loss of mutualistic traits. A derived clade of *Epicephala* has completely lost the pollinating behaviour after colonizing herbaceous species of *Phyllanthus*. These plants regularly attain full seed set through ant pollination (table S8 in the electronic supplementary material); thus, time and energetic costs required during pollination probably outweighed the benefit of assuring seed set in these moth lineages. At the same time, effective pollination by ants probably swamped the mutualistic effect of pollination by moths; thus, selection did not favour these *Phyllanthus* to specialize to moth pollination.

Taken together, this study provides two general implications for the coevolutionary dynamics of mutualisms. First, although species associations are phylogenetically conserved in most coevolving interactions (Thompson 2005), rare shifts by a partner possessing the mutualistic trait can give rise to new mutualisms in phylogenetically distant partner lineages. In this sense, the active pollination behaviour in *Epicephala* has been of critical importance for the establishment and maintenance of the Phyllanthae–*Epicephala* mutualism and thus represents a key innovation in this association. Second, the outcome of a species interaction can vary greatly depending on the community context in which it occurs (Thompson & Pellmyr 1992; Thompson & Cunningham 2002; Westerbergh 2004); thus, transitions between

mutualism and antagonism can occur repeatedly within a single phylogenetic lineage. This parallels recent findings in other mutualisms where derived parasitic taxa are nested within ancestrally mutualistic clades (Pellmyr *et al.* 1996b; Machado *et al.* 2001; Als *et al.* 2004). Of particular relevance to future studies is our finding that the mutualism arose independently in several Phyllanthae lineages, which provides outstanding opportunities for comparative analyses of character evolution, diversification rates and factors affecting mutualism stability. Thus, the Phyllanthae–*Epicephala* association is a promising new model system for studies of mutualism and the coevolutionary process.

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