Letter to the Editor

Routine Culturing of Stool Specimens for Yersinia enterocolitica

The sweeping conclusion of Kachoris et al. (3) that routine culturing of stool specimens for *Yersinia enterocolitica* is not a cost-effective procedure concerns us. We wish to recommend that the word procedure be followed by "from our patient population in Boston, Mass." The authors mentioned the variability of isolation rates reported by three groups of investigators and referred to Weissfeld's comments that geographical variations of *Y. enterocolitica* isolation rates do occur but failed to elaborate upon this phenomenon in their population.

Since June 1982, our laboratory has used cefsulodin-Irgasan-novobiocin (CIN) agar (Oxoid Ltd., Basingstoke, England; or Prepared Media Laboratories, Richmond, British Columbia, Canada) and cold enrichment to isolate Yersinia spp. After examining 6,209 fecal specimens, 415 Yersinia spp. have been recovered. More than 7% of the samples and 10.8% of symptomatic patients have been positive. On a yearly basis, our isolation rate has been consistent since we began culturing. Although the majority of our isolates have come from cold enrichment, many of them appear to be clinically relevant (2, 3a–5). In comparison, 4% of all specimens and 5% of all patients have been culture positive with Campylobacter spp.

Our methodology differs from that of Kachoris et al. (3) in that CIN plates are incubated at 35°C, not at 28°C; and for only 18 to 24 h, not 48 h or longer. After 18 to 24 h of incubation our technologists have no difficulty in picking small (1- to 1.5-mm) typical red bull's-eye colonies for identification. Other enteric organisms are larger at this temperature and generally would not be picked. All of our biochemical identification tests are incubated at 28°C. Yersinia spp. are more active metabolically between 22 and 28°C; thus, biochemical interpretations are most reliable when incubation is at 28°C rather than at 37°C (1). Although we have no firsthand experience with the Vitek system, as the authors do, we suspect that any system which requires a 37°C incubation for identification may possibly yield unreliable results for Yersinia spp.

Prior to the development of CIN medium, only 48 isolates of *Yersinia* spp. were reported from British Columbia, Canada, from 1966 to 1978 (6). In fact, regional differences in the isolation rate of *Yersinia* spp. still exist, both within the province of British Columbia and even the city of Vancouver, despite a uniformity of technique. For our primarily adult patient population, we can only conclude that the routine culturing of feces specimens for *Y. enterocolitica* is both clinically relevant and cost-effective. Other laboratories should be encouraged, not discouraged as Kachoris et al. (3) suggest, to search for this organism in their own populations and draw their own conclusions as to the cost-effectiveness of a CIN plate in their routine stool-screening procedure.

LITERATURE CITED

- 1. Bottone, E. J. 1977. Yersinia enterocolitica: a panoramic view of a charismatic microorganism. Crit. Rev. Microbiol. 5:211-241.
- Fletcher, K. M., C. M. Morris, and M. A. Noble. 1988. Human coproantibody secretory immunoglobulin A response to *Yersinia* species. J. Clin. Microbiol. 26:287–292.
- Kachoris, M., K. L. Ruoff, K. Welch, W. Kallas, and M. J. Ferraro. 1988. Routine culture of stool specimens for Yersinia

- enterocolitica is not a cost-effective procedure. J. Clin. Microbiol. 26:582-583.
- 3a. Noble, M. A., and R. L. Barteluk. 1988. Species and serotypes of yersiniae recovered in British Columbia. Can. J. Public Health 79:130-133.
- 4. Noble, M. A., R. L. Barteluk, H. J. Freeman, R. Subramaniam, and J. B. Hudson. 1987. Clinical significance of virulence-related assay of *Yersinia* species. J. Clin. Microbiol. 25:802–807.
- Simmonds, S. D., M. A. Noble, and H. J. Freeman. 1987. Gastrointestinal features of culture positive *Yersinia enterocolitica* infection. Gastroenterology 92:112-117.
- 6. Toma, S., and L. Lafleur. 1981. Yersinia enterocolitica infections in Canada 1966 to August 1978, p. 183–191. In E. J. Bottone (ed.), Yersinia enterocolitica. CRC Press, Boca Raton, Fla.

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Author's Reply

We agree that our findings on isolation of Yersinia entero-colitica from stool specimens may not be applicable to other geographical areas. While variation in reported isolation rates of Y. enterocolitica is a well-documented phenomenon, a second major cost-related point made in our article (appearing in the abstract and text) is that when the expected prevalence of Yersinia species is low additional planting of all stools on a medium selective for Yersinia species is not warranted due to the small increase in yield with cefsulodin-Irgasan-novobiocin (CIN) agar versus a MacConkey plate incubated first at 35°C and then at room temperature. We continue to screen all stool specimens for Yersinia species by this more cost-effective method and use CIN agar only when Yersinia cultures are specifically requested.

The Vitek system has been reported to be an accurate method for Y. enterocolitica identification (3). In our study, all colonies from 3,122 of the 3,622 cultures examined were screened on nutrient agar at 35°C, and all of the small colony-forming strains of the family Enterobacteriaceae examined by this method were identified as Y. enterocolitica by the Vitek system. Thus, we feel confident that no Yersinia isolates were missed due to misidentification.

While the authors of the letter note that Yersinia spp. are metabolically more active at lower temperatures, they fail to note that the optimum growth temperature for these organisms is also less than 35°C (1). This explains the recommendations of the manufacturer of the CIN agar used in our study and the observations of Head et al. (2) and Schiemann (4) concerning lowered incubation temperatures for CIN agar plates used as a primary isolation medium for Yersinia spp.

Decisions on whether to implement or discontinue any laboratory procedure should be based on the diagnostic value of the test. Each laboratory should consider parameters such as sensitivity, specificity, prevalence, speed, and the costs associated with false-negative or false-positive results in making its decision.

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LITERATURE CITED

- Bercovier, H., and H. H. Mollaret. 1984. Genus XIV. Yersinia Van Loghem 1944, 15^{AL}, p. 498-506. In N. R. Krieg and J. G. Holt (ed.), Bergey's manual of systematic bacteriology, vol. 1. The Williams & Wilkins Co., Baltimore.
- Head, C. B., D. A. Whitty, and S. Ratnam. 1982. Comparative study of selective media for recovery of *Yersinia enterocolitica*. J. Clin. Microbiol. 16:615-621.
- 3. Isenberg, H. D., T. L. Gavan, P. B. Smith, A. Sonnenwirth, W. Taylor, W. J. Martin, D. Rhoden, and A. Balows. 1980.
- Collaborative investigation of the AutoMicrobic System *Enterobacteriaceae* biochemical card. J. Clin. Microbiol. 11:694–702.
- Schiemann, D. A. 1979. Synthesis of a selective agar medium for Yersinia enterocolitica. Can. J. Microbiol. 25:1298-1304.

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