Staphylococcus lugdunensis, a Common Cause of Skin and Soft Tissue Infections in the Community $^{\nabla}$

Sidsel Böcher,^{1,2}* Birgitte Tønning,¹ Robert L. Skov,² and Jørgen Prag¹

Department of Clinical Microbiology, Viborg Hospital, Viborg, Denmark,¹ and Staphylococcus Laboratory, Statens Serum Institut, Copenhagen, Denmark²

Received 29 May 2008/Returned for modification 16 August 2008/Accepted 9 February 2009

Staphylococcus lugdunensis, a rare cause of severe infections such as native valve endocarditis, often causes superficial skin infections similar to Staphylococcus aureus infections. We initiated a study to optimize the identification methods in the routine laboratory, followed by a population-based epidemiologic analysis of patients infected with S. lugdunensis in Viborg County, Denmark. Recognition of a characteristic Eikenella corrodens-like odor on Columbia sheep blood agar combined with colony pleomorphism and prominent β-hemolysis after 2 days of incubation, confirmed by API-ID-32 Staph, led to an 11-fold increase in the detection of S. lugdunensis. By these methods we found 491 S. lugdunensis infections in 4 years, corresponding to an incidence of 53 per 100,000 per year, an increase from 5 infections per 100,000 inhabitants in the preceding years. Seventy-five percent of the cases were found in general practice; these were dominated by skin abscesses (36%), wound infections (25%), and paronychias (13%). Fifty-six percent of the infections occurred below the waist, and toes were the most frequently infected site (21%). Only 3% of the patients suffered from severe invasive infections. The median age was 52 years, and the male/female ratio was 0.69. Our study shows that S. lugdunensis is a common cause of skin and soft-tissue infections (SSTI) and is probably underrated by many laboratories. S. lugdunensis should be accepted as a significant pathogen in SSTI and should be looked for in all routine bacteriological examinations, and clinicians should be acquainted with the name and the pathology of the bacterium.

Staphylococcus lugdunensis, one of the coagulase-negative staphylococci (CoNS), was first described by Freney et al. 20 years ago (8). It causes severe infections similar to *Staphylococcus aureus* infections, especially acute endocarditis in prosthetic and native valves (14, 24, 29, 30, 36). Other infections such as osteomyelitis (17, 23), peritonitis (28), intravascular catheter infections (5, 32), prosthetic joint infections (26), and urinary tract infections have also been reported (10). *S. lug-dunensis* has been reported as an important cause also of skin and soft tissue infections (SSTI), primarily in the groin (1, 39) and mammae (13, 22, 25, 37, 41).

S. lugdunensis is part of the normal human skin flora and commonly colonizes the perineal region (1, 37, 39); however, carriage rates of different CoNS are not comprehensively accounted for, and only a few have looked for *S. lugdunensis* carriage (13, 16, 39).

Our study was undertaken after two serious cases of *S. lugdunensis* infection, a fatal case of endocarditis with protracted bacterial identification but correct antibiotic treatment and a case of late-diagnosed osteomyelitis, which prompted an optimization of our routine microbiological methods for identification of *S. lugdunensis*. This subsequently increased the number of *S. lugdunensis* isolates significantly, especially in samples from patients in general practice, and encouraged us to conduct a prospective epidemiologic study.

We here describe the improved method for identification of

* Corresponding author. Mailing address: Staphylococcus Laboratory, National Center for Antimicrobials and Infection Control, Statens Serum Institut, Artillerivej 5, DK-2300 Copenhagen S, Denmark. Phone: 45 32 68 36 84. Fax: 45 32 68 32 31. E-mail: sbc@ssi.dk. *S. lugdunensis*, which led to an 11-fold-increased incidence, and report the epidemiology of the 491 cases identified in Viborg County, Denmark, from July 2002 through June 2006.

MATERIALS AND METHODS

Bacteriology and optimization of microbiological identification. Prior to optimization of laboratory methods to identify *S. lugdunensis*, CoNS were speciated only in cases of invasive infections or if found in pure culture. In order to find a reliable method, we tested different methods from previous studies (3, 8, 11, 19, 20, 28).

In the process we noticed a prominent β -hemolysis on Columbia sheep blood agar (Becton Dickinson [BD] Glostrup, Denmark) after 2 days, whereas no β -hemolysis was seen on 5% horse blood agar (Statens Serum Institute Diagnostica [SSI], Hillerød, Denmark). We further observed that *S. lugdunensis* develops a characteristic odor, similar to that of *Eikenella corrodens*, when cultured on Columbia sheep blood agar. This odor has been described as "hypochlorite bleach-like" (40).

Differences in β -hemolysis and odor were investigated for 35 different isolates from seven staphylococcal species (5 *S. lugdunensis* isolates, 5 *S. warneri* isolates, 5 *S. epidermidis* isolates, 5 *S. haemolyticus* isolates, 5 *S. capitis* isolates, 5 *S. simulans* isolates, and 5 *S. aureus* isolates). Analyses were performed on Columbia agar with 5% sheep blood (BD), Columbia agar with 5% calf blood (SSI), 5% horse blood agar (SSI), 5% calf blood agar (SSI), and 5% human blood agar (SSI). Hemolysis was recorded as clear β -hemolysis, incomplete β -hemolysis, and no β -hemolysis.

To verify the characteristics of the odor, a second study was made, in which 12 blindfolded technologists characterized the odor of 35 staphylococcal isolates (5 *S. lugdunensis* isolates, 4 *S. warneri* isolates, 5 *S. epidermidis* isolates, 5 *S. haemolyticus* isolates, 5 *S. hominis* isolates, 5 *S. capitis* isolates, 1 *S. sinulans* isolate, and 5 *S. aureus* isolates) on Columbia sheep blood agar (BD). Two *E. corrodens* isolates that were positive for the characteristic odor on the basis of 12 independent evaluations of each plate.

New routine identification procedures. Based on the above, new routine procedures were implemented from July 2002. All specimens were inoculated on Columbia agar plates with 5% sheep blood (BD) and read after 24 and 48 h of incubation at 35°C in 5% CO₂. Catalase-positive *Staphylococcus*-like colonies

⁷ Published ahead of print on 25 February 2009.

were examined for colony pleomorphism, β -hemolysis, and the characteristic *Eikenella*-like odor after 48 h of incubation. ID 32 Staph (bioMérieux) was used for confirmatory identification.

Evaluation of PFGE pattern for clonal differences and characteristics. Pulsedfield gel electrophoresis (PFGE) as described by Tenover et al. (33) was performed on 38 *S. lugdunensis* isolates; 19 were from Viborg, Denmark (4 from blood cultures, 13 from SSTI, and 1 each from cases of arthritis and osteitis), and 19 were from Växjö, Sweden (5 from blood cultures, 13 from SSTI, and 1 from a fistula).

Antimicrobial susceptibility testing. Susceptibility testing was performed on Danish blood agar (SSI) using an inoculum giving semiconfluent growth, NEO-Sensitabs (Rosco Diagnostica), and overnight incubation. Antibiotics tested were oxacillin, methicillin (meticillin), penicillin, erythromycin, and fusidic acid. Interpretation of inhibition zones was performed as specified by the manufacturer (25a).

Epidemiologic setting. Viborg County had approximately 235,500 inhabitants, about 5% of the Danish population. One department of clinical microbiology served all five hospitals and all 125 general practitioners.

All patients from whom *S. lugdunensis* was isolated from 1 July 2002 to 30 June 2006 were included in the epidemiological study. The type of infection, localization, and previous *S. lugdunensis* infection were extracted from the laboratory request form.

Additionally, for the period 1 July to 31 December 2002, all patients positive for *S. lugdunensis* and those with SSTI were followed up by telephone interviews by the general practitioners or by reading the hospital records of admitted patients. The following data were extracted: infection site, type of infection, clinical signs and symptoms, treatment, progress of infection, complications, and relapses. For patients with SSTI from which staphylococci other than *S. lugdunensis* were isolated, clinical information was extracted from the laboratory request form only.

Neighboring departments of clinical microbiology were interviewed about their methods and number of *S. lugdunensis* cases identified during 2002.

RESULTS

Optimization of identification methods. Of the examined staphylococcal isolates only *S. lugdunensis* and *S. aureus* isolates showed prominent β -hemolysis around the colonies after 48 h incubation on Columbia agar with 5% sheep blood. The other staphylococcal species showed weak or no hemolysis. *S. simulans* and *S. haemolyticus* produced hemolysis on Columbia agar with 5% calf blood. *S. lugdunensis* did not produce β -hemolysis on human, horse, or calf blood agar without Columbia agar base. The characteristic *Eikenella*-like odor was present when *S. lugdunensis* isolates grew on sheep, calf, and human blood agar but not when they were cultured on horse blood agar.

The characteristic odor was present in 93% of the blinded odor tests of *S. lugdunensis* cultured on 5% Columbia sheep blood agar (Table 1). Other CoNS species were suspected to be *S. lugdunensis* in 3% to 17% of test situations.

PFGE of the 19 isolates from Viborg County, Denmark, and 19 isolates from Växjö County, Sweden, showed that the same two clones dominated in both countries, and there was no difference between invasive and noninvasive strains regarding PFGE pattern (Fig. 1).

Epidemiology. By using the improved methods from the pilot studies, it was determined that the incidence of *S. lug-dunensis* was 53 infections per 100,000 inhabitants per year, in contrast to the value of 5 infections per 100,000 inhabitants per year previously determined (representing an increased rate of detection). Seventy-five percent of the patients were from general practice (Fig. 2). The departments of clinical microbiology in three neighboring counties, all using plating on 5% horse blood plates (SSI) for swabs from SSTI, found from zero to four *S. lugdunensis* infections per 100,000 inhabitants.

 TABLE 1. Odor evaluation by 12 blindfolded laboratory technologists of 35 staphylococcus and 2

 Eikenella corrodens isolates

Species tested	No. of tests resulting in detection of characteristic odor ⁴ / total no. of tests		
Staphylococcus lugdunensis			
Staphylococcus capitis			
Staphylococcus warneri			
Staphylococcus haemolyticus			
Staphylococcus aureus			
Staphylococcus hominis			
Staphylococcus epidermidis			
Staphylococcus simulans			
Eikenella corrodens			

^a The characteristic odor is a hypochlorite bleach-like odor, similar to that characteristic of *Eikenella corrodens*.

During a 6-month period, *S. lugdunensis* was found in 13% of 159 abscesses from general practice. In these specimens, *S. lugdunensis* was found as the only pathogen, i.e., "monoculture" in 70% (Table 2). This was similar to results for abscess specimens containing *S. aureus* isolates, where 86% were found in monoculture, and was significantly more frequent than the frequency for abscess specimens containing monoculture of other CoNS (20%). In swabs from infections other than abscesses, *S. lugdunensis* strains were found from 0.09% of 1,087 patients with impetigo, 0.16% of 616 with ear infections, 0.6% of 1,614 with unspecified wound infections, 0.7% of 587 with unspecified skin infections, 1.4% of 204 with incisional surgical wound infections, 4.5% of 22 with pustules, and 15% of 13 with infected cutaneous atheromas, respectively.

SSTI caused by *S. lugdunensis* were clinically indistinguishable from *S. aureus* infections. Ninety-six percent of *S. lugdunensis* patients were treated with surgical incision and/or antibiotics. Cases not needing antibiotic or surgical treatment were all superficial wound infections.

During 2002 to 2006 a total of 491 cases were found. The distribution of infectious sites is shown in Table 3. In 45% of the 491 samples *S. lugdunensis* was found in mixed cultures; however, other significant pathogens (e.g., *S. aureus* and hemolytic streptococci) were found in 4% of these cultures only.

S. lugdunensis infections were primarily seen in middle-aged and elderly people (median age, 52 years) except for otitis media, where a median age of 1 year was found. The overall male/female ratio was 0.69.

Recurrent infections were seen in seven patients: three mamma infections, two genital abscesses, one wound infection, and one dialysis catheter infection.

Invasive *S. lugdunensis* infections were seen in two patients from general practice and 12 hospitalized patients. Six of these had necrotic toe infections, and among these three had diabetes. One of the patients with toe infection developed gangrene and died following a crus amputation. The other deep infections were two cases of knee arthritis, two deep-seated groin infections, one perineal fistula, one case of bursitis in the elbow, and two lung infections, one a case of empyema and one a case of aspergillosis with *S. lugdunensis* coinfection.

The antibiotic susceptibilities of the 491 isolates (% resistant) were as follows: penicillin, 20%; fusidic acid, 5%; eryth-



FIG. 1. PFGE on 38 S. lugdunensis isolates from blood cultures, SSTI, and two deep infections seen in Viborg County, Denmark, and in Växjö County, Sweden, 1991 to 2006.



FIG. 2. Numbers of *Staphylococcus lugdunensis* infections per quarter, Viborg County, Denmark, January 1998 through June 2006.

romycin, 2%; methicillin-oxacillin, 0%. There was no difference in antimicrobial susceptibility between isolates from hospitalized patients and isolates from general practice.

DISCUSSION

CoNS from SSTI are seldom routinely diagnosed to species level. Although definitive identification of S. lugdunensis is well described and reliable (3, 8, 19), this study indicates that one problem is failure to suspect S. lugdunensis. Phenotypic characteristics such as colony pleomorphism and β -hemolysis are useful when detecting S. lugdunensis, in contrast to clumping and synergistic hemolysis, which characterize several CoNS species (7). In this study we found that S. lugdunensis gives a prominent β -hemolysis and a characteristic *Eikenella*-like odor after 2 days of incubation on Columbia agar with 5% sheep blood, which, combined with colony pleomorphism, helps in the initial recognition of S. lugdunensis. This is not described in textbooks such as the Manual of Clinical Microbiology and Topley and Wilson's Microbiology and Microbial Infections (35, 40). The hemolytic peptides with delta-toxin-like activity (SLUSH-A, SLUSH-B, and SLUSH-C) lyse certain mammalian blood cells (38), and this might be an explanation for the differences found in the evaluation of blood agars.

The characteristic odor when isolates were cultured on Columbia sheep blood agar was also present when they were cultured on calf and human blood agar, but not on horse blood

TABLE 2. Swab sample results for *S. lugdunensis*, other CoNS, and *S. aureus*^a

Organism(s)	Total no. of patients (incidence ^b)	No. of abscesses	% of abscesses where bacteria were found in pure culture (CI ^c)	No. of other SSTI
Staphylococcus lugdunensis Other CoNS Staphylococcus aureus	43 (18) 932 (398) 2,760 (1,178)	20 69 56	70 (46–88) 20 (12–32) 86 (74–94)	23 863 2,704

^{*a*} Swabs were from soft-tissue infections and were taken in general practice in Viborg County, Denmark, during a 6-month sampling period in 2002.

^b Number of patients/100,000 persons/6 months

^c CI, confidence interval.

TABLE 3. Infection type and demographic data on 491Staphylococcus lugdunensis infections, Viborg County,July 2002 through June 2006

Infection type	No. of patients	Median age (yr) (range)	Male/female ratio	
Total	491	52 (0-96)	0.69	
Abscesses	179	44 (3–90)	0.50	
Wound infections	122	64 (0–93)	0.61	
Paronychia	63	44 (7–90)	1.42	
Incisional surgical wound	49	54 (6-84)	0.81	
Otitis media	17	1 (0-8)	2.4	
Otitis externa	8	54 (22-83)	0.15	
Urinary tract infections	15	71 (9–89)	0.50	
Deep infections	14	77 (31–96)	0.75	
Other	24	49 (0–93)	0.5	

agar. The decomposition of certain fatty acids in some mammal erythrocytes might be the cause of this odor (31).

When using the improved methods, we discovered 11 times more *S. lugdunensis*, infections than before, primarily SSTI. This in combination with the much lower rate of identification of *S. lugdunensis* found in three neighboring counties indicates that *S. lugdunensis* is vastly underrated as a cause of infection.

Clinical information supported the findings that *S. lugdunen*sis predominantly causes abscesses and that *S. lugdunensis* infections are clinically indistinguishable from infections caused by *S. aureus*. The ability of *S. lugdunensis* to cause abscesses has also been described in mouse model studies (6, 18).

It has been suggested that S. lugdunensis primarily causes infections below the waist (12, 37), in particular the inguinal area (1, 39). Our results indicate a broader distribution of infected sites, which also was reported in a study by Herchline and Ayers in 1991 (13). The localization of infection seems to be age related. In small children, S. lugdunensis was predominantly found in otitis media, whereas in middle-aged patients infections were most common in armpits, groin, buttocks, mamma, and incisional sites. Among elderly people most infections were in fingers, toes, and leg ulcers. Women were more frequently infected than men, which is in contrast to S. aureus infections but in accordance with a higher carriage rate of S. lugdunensis among women (12, 39). Areas that possess apocrine sweat glands appear to the sites most frequently infected. As has been suggested for S. aureus and Staphylococcus sciuri (4), S. lugdunensis might be a commensal in areas that possesses apocrine sweat glands, i.e., axilla, anogenital region, auditory canals, eyelids, and mammary areolae.

Most of our cases with toe infections were elderly patients (median age, 60 years), and some had diabetes and fungal toe infections. Both elderly people and diabetics already have a higher prevalence of toe infections (2, 9, 27). Previous studies have described the dynamics of toe web infections, where fungal infections induce damage to the stratum corneum, which is followed by a bacterial superinfection. This might also be the case for *S. lugdunensis* toe infections (15, 21).

To our knowledge only one report has previously described a case of *S. lugdunensis* mastoiditis in relation to otitis media in a child (34). This contrasts with the 37 cases of ear infections found in the present series.

It is important that microbiologists make the general practitioners and hospital physicians acquainted with *S. lugdunen*- *sis*, its pathology, and risk of serious infections. In Viborg County, findings of *S. lugdunensis* are now followed by the comment that *S. lugdunensis* infections should be considered as serious as *S. aureus* infections.

ACKNOWLEDGMENTS

We thank Maiken Jensen and Karen Marie Søby at the Department of Clinical Microbiology in Viborg for laboratory assistance during the study. We thank Gunnar Kahlmeter at the Department of Clinical Microbiology, Växjö, Sweden, for *S. lugdunensis* isolates and associated clinical information. We thank Dominique L. Monnet and Niels Frimodt-Møller for their helpful comments regarding the manuscript.

REFERENCES

- Bellamy, R., and T. Barkham. 2002. *Staphylococcus lugdunensis* infection sites: predominance of abscesses in the pelvic girdle region. Clin. Infect. Dis. 35:E32–E34.
- Crawford, F., and J. Ferrari. 2006. Fungal toenail infections. Clin. Evid. 2212–2220.
- De Paulis, A. N., S. C. Predari, C. D. Chazarreta, and J. E. Santoianni. 2003. Five-test simple scheme for species-level identification of clinically significant coagulase-negative staphylococci. J. Clin. Microbiol. 41:1219–1224.
- Eady, E. A., and J. H. Cove. 2003. Staphylococcal resistance revisited: community-acquired methicillin resistant *Staphylococcus aureus*—an emerging problem for the management of skin and soft tissue infections. Curr. Opin. Infect. Dis. 16:103–124.
- Elliott, S. P., R. Yogev, and S. T. Shulman. 2001. *Staphylococcus lugdunensis*: an emerging cause of ventriculoperitoneal shunt infections. Pediatr. Neurosurg. 35:128–130.
- Ferguson, K. P., D. W. Lambe, Jr., J. L. Keplinger, and J. H. Kalbfleisch. 1991. Comparison of the pathogenicity of three species of coagulase-negative *Staphylococcus* in a mouse model with and without a foreign body. Can. J. Microbiol. 37:722–724.
- Fleurette, J., M. Bes, Y. Brun, J. Freney, F. Forey, M. Coulet, M. E. Reverdy, and J. Etienne. 1989. Clinical isolates of *Staphylococcus lugdunensis* and *S. schleiferi*: bacteriological characteristics and susceptibility to antimicrobial agents. Res. Microbiol. 140:107–118.
- Freney, J., Y. Brun, M. Bes, M. Meugnier, et al. 1988. Staphylococcus lugdunensis sp. nov. and Staphylococcus schleiferi sp. nov., two species from human clinical specimens. Int. J. Syst. Bacteriol. 38:168–172.
- Gupta, A. K., N. Konnikov, P. MacDonald, P. Rich, N. W. Rodger, M. W. Edmonds, R. McManus, and R. C. Summerbell. 1998. Prevalence and epidemiology of toenail onychomycosis in diabetic subjects: a multicentre survey. Br. J. Dermatol. 139:665–671.
- Haile, D. T., J. Hughes, E. Vetter, P. Kohner, R. Snyder, R. Patel, and F. R. Cockerill III. 2002. Frequency of isolation of *Staphylococcus lugdunensis* in consecutive urine cultures and relationship to urinary tract infection. J. Clin. Microbiol. 40:654–656.
- Hebert, G. A. 1990. Hemolysins and other characteristics that help differentiate and biotype *Staphylococcus lugdunensis* and *Staphylococcus schleiferi*. J. Clin. Microbiol. 28:2425–2431.
- Hellbacher, C., E. Tornqvist, and B. Soderquist. 2006. Staphylococcus lugdunensis: clinical spectrum, antibiotic susceptibility, and phenotypic and genotypic patterns of 39 isolates. Clin. Microbiol. Infect. 12:43–49.
- Herchline, T. E., and L. W. Ayers. 1991. Occurrence of *Staphylococcus lugdunensis* in consecutive clinical cultures and relationship of isolation to infection. J. Clin. Microbiol. 29:419–421.
- Jones, R. M., M. A. Jackson, C. Ong, and G. K. Lofland. 2002. Endocarditis caused by *Staphylococcus lugdunensis*. Pediatr. Infect. Dis. J. 21:265–268.
- Kates, S. G., K. M. Nordstrom, K. J. McGinley, and J. J. Leyden. 1990. Microbial ecology of interdigital infections of toe web spaces. J. Am. Acad. Dermatol. 22:578–582.
- Koziol-Montewka, M., A. Szczepanik, I. Baranowicz, L. Jozwiak, A. Ksiazek, and D. Kaczor. 2006. The investigation of *Staphylococcus aureus* and coagulase-negative staphylococci nasal carriage among patients undergoing haemodialysis. Microbiol. Res. 161:281–287.
- Kragsbjerg, P., J. Bomfim-Loogna, E. Tornqvist, and B. Soderquist. 2000. Development of antimicrobial resistance in *Staphylococcus lugdunensis* during treatment-report of a case of bacterial arthritis, vertebral osteomyelitis and infective endocarditis. Clin. Microbiol. Infect. 6:496–499.
- Lambe, D. W., Jr., K. P. Ferguson, J. L. Keplinger, C. G. Gemmell, and J. H. Kalbfleisch. 1990. Pathogenicity of *Staphylococcus lugdunensis*, *Staphylococ-*

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cus schleiferi, and three other coagulase-negative staphylococci in a mouse model and possible virulence factors. Can. J. Microbiol. **36**:455–463.

- Leung, M. J., N. Nuttall, M. Mazur, T. L. Taddei, M. McComish, and J. W. Pearman. 1999. Case of *Staphylococcus schleiferi* endocarditis and a simple scheme to identify clumping factor-positive staphylococci. J. Clin. Microbiol. 37:3353–3356.
- Leung, M. J., N. Nuttall, T. M. Pryce, G. W. Coombs, and J. W. Pearman. 1998. Colony variation in *Staphylococcus lugdunensis*. J. Clin. Microbiol. 36:3096–3098.
- Leyden, J. J. 1993. Progression of interdigital infections from simplex to complex. J. Am. Acad. Dermatol. 28:S7–S11.
- Lina, B., F. Vandenesch, M. E. Reverdy, T. Greenland, J. Fleurette, and J. Etienne. 1994. Non-puerperal breast infections due to *Staphylococcus lugdunensis*. Eur. J. Clin. Microbiol. Infect. Dis. 13:686–687.
- Murdoch, D. R., R. J. Everts, S. T. Chambers, and I. A. Cowan. 1996. Vertebral osteomyelitis due to *Staphylococcus lugdunensis*. J. Clin. Microbiol. 34:993–994.
- Patel, R., K. E. Piper, M. S. Rouse, J. R. Uhl, F. R. Cockerill III, and J. M. Steckelberg. 2000. Frequency of isolation of *Staphylococcus lugdunensis* among staphylococcal isolates causing endocarditis: a 20-year experience. J. Clin. Microbiol. 38:4262–4263.
- Ros, M. J., A. Ramirez, E. Arteaga, C. Alberto, J. Gil, and J. Reina. 1999. Infection by *Staphylococcus lugdunensis*: clinico-microbiologic characterization of 25 cases. Enferm. Infecc. Microbiol. Clin. 17:223–226.
- 25a.Rosco Diagnostica. 2002. Rosco's user's guide 15th ed. Rosco Diagnostica, Taastrup, Denmark.
- Sampathkumar, P., D. R. Osmon, and F. R. Cockerill III. 2000. Prosthetic joint infection due to *Staphylococcus lugdunensis*. Mayo Clin. Proc. 75:511– 512.
- Saunte, D. M., J. B. Holgersen, M. Haedersdal, G. Strauss, M. Bitsch, O. L. Svendsen, M. C. Arendrup, and E. L. Svejgaard. 2006. Prevalence of toe nail onychomycosis in diabetic patients. Acta Derm. Venereol. 86:425–428.
- Schnitzler, N., R. Meilicke, G. Conrads, D. Frank, and G. Haase. 1998. Staphylococcus lugdunensis: report of a case of peritonitis and an easy-toperform screening strategy. J. Clin. Microbiol. 36:812–813.
- Schonheyder, H. C., V. K. Hansen, P. Asschenfeldt, and V. T. Rosdahl. 1993. Staphylococcus lugdunensis: an important cause of endocarditis. A case report. APMIS 101:802–804.
- Sotutu, V., J. Carapetis, J. Wilkinson, A. Davis, and N. Curtis. 2002. The "surreptitious staphylococcus": *Staphylococcus lugdunensis* endocarditis in a child. Pediatr. Infect. Dis. J. 21:984–986.
- Stoakes, L., M. A. John, R. Lannigan, B. C. Schieven, M. Ramos, D. Harley, and Z. Hussain. 1994. Gas-liquid chromatography of cellular fatty acids for identification of staphylococci. J. Clin. Microbiol. 32:1908–1910.
- Tee, W. S., S. Y. Soh, R. Lin, and L. H. Loo. 2003. *Staphylococcus lugdunensis* carrying the *mecA* gene causes catheter-associated bloodstream infection in premature neonate. J. Clin. Microbiol. 41:519–520.
- 33. Tenover, F. C., R. D. Arbeit, R. V. Goering, P. A. Mickelsen, B. E. Murray, D. H. Persing, and B. Swaminathan. 1995. Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing. J. Clin. Microbiol. 33:2233–2239.
- Thomas, S., C. Hoy, and R. Capper. 2006. Osteomyelitis of the ear canal caused by *Staphylococcus lugdunensis*. J. Infect. 53:e227–e229.
- Topley, W. W. C., G. S. Wilson, and L. Collier. 1998. Topley and Wilson's microbiology and microbial infections, 9th ed., vol. 2. Hodder Arnold, London, United Kingdom.
- Vandenesch, F., J. Etienne, M. E. Reverdy, and S. J. Eykyn. 1993. Endocarditis due to *Staphylococcus lugdunensis*: report of 11 cases and review. Clin. Infect. Dis. 17:871–876.
- Vandenesch, F., S. J. Eykyn, J. Etienne, and J. Lemozy. 1995. Skin and post-surgical wound infections due to *Staphylococcus lugdunensis*. Clin. Microbiol. Infect. 1:73–74.
- Vandenesch, F., M. J. Storrs, F. Poitevin-Later, J. Etienne, P. Courvalin, and J. Fleurette. 1991. Delta-like haemolysin produced by *Staphylococcus lugdunensis*. FEMS Microbiol. Lett. 62:65–68.
- van der Mee-Marquet, N., A. Achard, L. Mereghetti, A. Danton, M. Minier, and R. Quentin. 2003. *Staphylococcus lugdunensis* infections: high frequency of inguinal area carriage. J. Clin. Microbiol. 41:1404–1409.
- 40. von Gravenitz, A., R. Zbinden, and R. Mutters. 2007. Actinobacillus, Capnocytophaga, Eikenella, Kingella, Pasteurella, and other fastidious or rarely encountered gram-negative rods, p. 621–635. In P. R. Murray, E. J. Baron, J. H. Jorgensen, M. L. Landry, and M. A. Pfaller (ed.), Manual of clinical microbiology, 9th ed. ASM Press, Washington, DC.
- Waghorn, D. J. 1994. Staphylococcus lugdunensis as a cause of breast abscess. Clin. Infect. Dis. 19:814–815.