

Atherosclerosis and Matrix Metalloproteinases: Experimental Molecular MR Imaging in Vivo¹

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Purpose:

To evaluate the capability of P947, a magnetic resonance (MR) imaging contrast agent that molecularly targets matrix metalloproteinases (MMPs), to aid detection and imaging of MMPs in atherosclerotic lesions in vivo; its specificity compared with that of P1135; expression and distribution of MMPs in atherosclerotic vessels; and in vivo distribution and molecular localization of fluorescent europium (Eu) P947.

Materials and Methods:

The Animal Care and Use Committee approved all experiments. P947 was synthesized by attaching a gadolinium chelate (1,4,7,10-tetraazacyclododecane-N,N',N'',N'''-tetraacetic acid) to a peptide that specifically binds MMPs. Scrambled form of P947 (P1135) was synthesized by replacing the targeting moiety of P947 with a scrambled peptide lacking the ability to bind MMPs. P947, P1135, and gadoterate meglumine were injected into atherosclerotic apolipoprotein E-deficient and wild-type mice. The aortic MR imaging enhancement produced by the contrast agents was measured at different times and was compared by using one-way analysis of variance. MMP expression was investigated in the aortas by using MMP immunostaining and in situ MMP zymography. A fluorescent form of P947 (Eu-P947) was synthesized to compare the in vivo distribution of the contrast agent (Eu-P947) with specific MMP immunofluorescent staining.

Results:

MMP-targeted P947 facilitated a 93% increase ($P < .001$) in MR image signal intensity (contrast-to-noise ratio [CNR], 17.7 compared with 7.7; $P < .001$) of atherosclerotic lesions in vivo. Nontargeted P1135 (scrambled P947) provided 33% MR image enhancement (CNR, 10.8), whereas gadoterate meglumine provided 5% (CNR, 6.9). Confocal laser scanning microscopy demonstrated colocalization between fluorescent Eu-P947 and MMPs in atherosclerotic plaques. Eu-P947 was particularly present in the fibrous cap region of plaques.

Conclusion:

P947 improved MR imaging for atherosclerosis through MMP-specific targeting. The results were validated and provide support for further assessment of P947 as a potential tool for the identification of unstable atherosclerosis.

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Supplemental material:

<http://radiology.rsna.org/cgi/content/full/2511080539/DC1>

Despite great advances in cardiovascular medicine, atherosclerosis remains the foremost cause of death in developed industrialized societies (1,2) and has become the leading cause of death globally (3,4). We do not currently possess clinical tools capable of accurately evaluating atherosclerosis directly or at early stages of disease when interventions may have the greatest effect on altering disease progression.

The rapidly evolving field of molecular imaging will ultimately allow us to accurately assess molecular, cellular, and compositional components of atherosclerotic lesions. This information will be clinically vital to identify vulnerable patients (5,6) and facilitate personalized medical management. Molecular magnetic resonance (MR) imaging is especially appealing as a noninvasive modality to evaluate atherosclerosis because it lacks ionizing radiation and possesses the superb spatial resolution necessary to image small-scale pathologic findings such as atherosclerotic lesions (7,8).

Matrix metalloproteinases (MMPs) are implicated in the progression, destabilization, and rupture of atherosclerotic lesions (9–12). Abnormal expression and regulation of MMPs in atherosclerotic

plaques may lead to destabilization, resulting in plaque rupture with subsequent thrombosis, occlusion, and detrimental clinical consequences such as myocardial infarction or stroke (13). MMP overexpression has been demonstrated in the vulnerable shoulder regions of human atheroma (9). Macrophage-derived foam cells that are characteristic of unstable plaques have been shown to be major sources of MMPs in human and experimental atherosclerotic lesions (14). Therefore, MMP expression not only is a part of the pathogenesis of atherosclerosis, but it may be predictive of plaque instability (15). Such a potentially pivotal role in the pathogenesis of atherosclerosis suggests that MMPs are a rewarding target for molecular imaging.

Thus, the molecular MR imaging contrast agent that we investigated in this study was P947, which specifically targets MMPs (16,17). P947 is a newer molecular MR imaging agent made of a gadoterate meglumine (Guerbet, Aulnay-sous-Bois, France) moiety covalently bound, by using a linker, to a peptide that specifically binds MMPs at the enzymatic active site (16,18). For the purpose of demonstrating the specificity of P947 for MMPs in vivo, two other contrast agents were used as controls: P1135 (Guerbet), which is an MR imaging analog of P947 for which the peptide sequence has been scrambled, leading to a loss of affinity for MMPs; and P1191 (Guerbet), which is a europium (Eu) derivative of P947 (Eu-P947) with fluorescence emission properties.

The goals of this investigation were as follows: (a) to study and validate the in vivo capability of P947 (targeted to MMPs) to assess atherosclerosis in apolipoprotein E (*ApoE*) knockout (*ApoE*-deficient) mice by using MR imaging; (b) to evaluate the specificity of P947 for MMPs by comparing its in vivo MR imaging effectiveness to that of P1135 (untargeted, scrambled form of P947 lacking specific binding to MMPs); (c) to demonstrate the expression and examine the distribution of MMPs in the atherosclerotic vessels of the mice we used for the in vivo MR imaging experiments; and (d) to investigate the in vivo distribution of fluorescent Eu-P947 (also known as P1191) in atherosclerosis and to study the in vivo molecular localization of Eu-P947 in relation to specific MMP fluorescent immunostaining in atherosclerotic lesions (examination of

Advances in Knowledge

- P947 facilitated in vivo MR imaging of atherosclerosis in this experimental investigation.
- The in vivo contrast material-enhanced MR imaging effectiveness of P947 was superior to that of P1135 (untargeted, scrambled form of P947 lacking specific binding to matrix metalloproteinases [MMPs]) and to gadoterate meglumine (a standard gadolinium-based contrast agent).
- Data from this study suggest that P947 specifically targets MMPs in atherosclerosis.
- P947 is in its early stages of development; further investigation is needed to explore its potential as a molecular MR imaging contrast agent.

Implications for Patient Care

- Specific molecular imaging of atherosclerosis will give clinicians the unprecedented ability to detect pathologic findings at early stages before irreversible damage has taken place.
- P947 may facilitate specific noninvasive assessment of atherosclerosis.
- P947 could potentially also be used to evaluate atherosclerotic plaque instability.

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Abbreviations:

CNR = contrast-to-noise ratio
 MMP = matrix metalloproteinase
 NER = normalized enhancement ratio
 SI = signal intensity
 WT = wild type

Author contributions:

Guarantors of integrity of entire study, J.G.S.A., E.L., Z.A.F.; study concepts/study design or data acquisition or data analysis/interpretation, all authors; manuscript drafting or manuscript revision for important intellectual content, all authors; manuscript final version approval, all authors; literature research, V.A., J.G.S.A., S.A., F.H., E.V., M.S., D.B.W., E.L., C.C., Z.S.G., Z.A.F.; clinical studies, V.A., J.G.S.A., M.S., E.L., Z.A.F.; experimental studies, V.A., J.G.S.A., S.A., F.H., E.V., M.S., D.B.W., S.L.G., E.L., C.C., Z.S.G., Z.A.F.; statistical analysis, J.G.S.A., E.V., M.S., Z.A.F.; and manuscript editing, V.A., J.G.S.A., S.A., F.H., E.V., M.S., D.B.W., C.C., Z.S.G., Z.A.F.

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See Materials and Methods for pertinent disclosures.

See also Science to Practice in this issue.

possible colocalization between fluorescent Eu-P947 and specific MMPs). A limited part of this work was presented at the American Heart Association scientific meeting (17), but the images in this article are not the same as those published previously (17).

Materials and Methods

This study was partially funded by Guerbet. All contrast agents were provided by Guerbet. The authors who were not employees of Guerbet (V.A., J.G.S.A., S.A., F.H., E.V., M.S., D.B.W., E.A.F., Z.S.G., Z.A.F.) had full control of the data; those who were employees (S.L.G., E.L., C.C.) did not have control of the data.

Synthesis and Characterization of Contrast Agents

P947 was synthesized by Guerbet through the coupling of an MMP-binding peptide to the gadolinium chelate 1,4,7,10-tetraazacyclododecane-N,N',N'',N'''-tetraacetic acid, as described in Appendix E1 (<http://radiology.rsnajnl.org/cgi/content/full/2511080539/DC1>). P1135 (Guerbet) was synthesized by using the same steps as were used for P947 except that the peptide that was attached was scrambled in sequence so that the ability to bind MMPs was lost (the molecular weight and size were identical to those of P947). Eu-P947 (fluorescent P947) was synthesized by using the same steps as those for P947 except that europium was used in place of gadolinium in the 1,4,7,10-tetraazacyclododecane-N,N',N'',N'''-tetraacetic acid moiety. Gadoterate meglumine (Dotarem; Guerbet) is a standard nonspecific gadolinium-based contrast agent that was used as one of the reference compounds.

Animal Protocol

Atherosclerotic *ApoE*-deficient mice (C57BL/6) (Jackson Laboratory, Bar Harbor, Me) and matched normal (non-atherosclerotic) wild-type (WT) mice (Jackson Laboratory) were used in the experiments, as described in Appendix E1 (<http://radiology.rsnajnl.org/cgi/content/full/2511080539/DC1>). The Animal Care and Use Committee for

Mount Sinai School of Medicine, New York, NY, approved all experiments.

In Vivo MR Imaging by Using P947 and Various Controls

In vivo MR imaging was performed (19) with a 9.4-T, 89-mm bore MR imaging system operating at a proton frequency of 400 MHz (Bruker Instruments, Billerica, Mass), as described in Appendix E1 (<http://radiology.rsnajnl.org/cgi/content/full/2511080539/DC1>). After preinjection baseline MR imaging, *ApoE*-deficient mice were injected, via a tail-vein injection catheter (MTV no. 06; Strategic Applications, Libertyville, Ill), with either MMP-targeted P947 ($n = 16$) or the untargeted scrambled control P1135 ($n = 5$) at 100 μ mol of gadolinium per kilogram. For the control group, *ApoE*-deficient mice ($n = 6$) were injected (at 100 μ mol of gadolinium per kilogram) with the standard nonspecific gadolinium-based MR contrast agent gadoterate meglumine. In addition, as another control group, WT mice were injected with equivalent doses of P947 ($n = 4$). MR imaging was performed at 1, 2, 3, and 22 hours after injection. Slices were precisely anatomically matched (V.A., J.G.S.A., Z.A.F., M.S.) to the sections obtained on the preinjection baseline image (7).

Analysis of MR Imaging Results and Pathologic Correlation

After the 22-hour postinjection image, the animals were sacrificed. The aortas were carefully isolated by using microscopic dissection and were fixed for standard pathologic analysis. The histologic slides from the atherosclerotic aortas were then matched to the MR image sections for comparison and correlation. To quantitatively analyze the MR imaging results, signal intensity (SI) measurements were obtained by using regions of interest on the aortic wall, with four points in four Cartesian quadrants of the aorta on each section, as described in Appendix E1 (<http://radiology.rsnajnl.org/cgi/content/full/2511080539/DC1>). The contrast-to-noise (CNR) ratio of the aortic wall (W) to the lumen (L)

($CNR_{W/L}$) was calculated for each section by using Equation (1):

$$CNR_{W/L} = \frac{[(W_1 + W_2 + W_3 + W_4)/4] - L_{SI}}{SD \text{ of Noise}}, \quad (1)$$

where W_n is the SI value of the aortic wall in quadrant n , L_{SI} is the SI of the lumen, and SD is the standard deviation.

For each matched section, the normalized enhancement ratio (NER) and percentage NER were used to measure the normalized SI enhancement of the aortic wall, relative to muscle, after contrast agent injection by using Equations (2) and (3), as follows:

$$NER = \frac{\left[\frac{(W_{1POST} + W_{2POST} + W_{3POST} + W_{4POST})/4}{SI_{MPost}} \right]}{\left[\frac{(W_{1PRE} + W_{2PRE} + W_{3PRE} + W_{4PRE})/4}{SI_{MPre}} \right]}, \quad (2)$$

$$\text{Percentage NER} = (NER - 1) \cdot 100, \quad (3)$$

where W_{nPOST} is the SI of the aortic wall in quadrant n after injection of contrast agent, W_{nPRE} is the SI of the aortic wall in quadrant n before injection of contrast agent, SI_{MPost} is the SI of muscle after contrast agent injection, and SI_{MPre} is the SI of muscle before contrast agent injection.

MMP Immunohistochemical and Pathologic Analysis

Abdominal aortas matching the MR imaging area were removed, sectioned, and stained by using Masson trichrome staining, as described in Appendix E1 (<http://radiology.rsnajnl.org/cgi/content/full/2511080539/DC1>). Immunostaining intended for light microscopy was performed, targeting MMP-2, MMP-3, and MMP-9 by using standard established techniques, as described in Appendix E1 (<http://radiology.rsnajnl.org/cgi/content/full/2511080539/DC1>). In situ gelatin zymography was performed by using standard established techniques, as described in Appendix E1

(<http://radiology.rsnajnl.org/cgi/content/full/2511080539/DC1>), and imaging was performed by using a laser scanning confocal microscope.

Confocal Laser Scanning Fluorescence Microscopy

Abdominal aortas matching the MR imaging area from *ApoE*-deficient mice were fluorescently immunostained for MMP-2, MMP-3, or MMP-9 by using standard established techniques, as described in Appendix E1 (<http://radiology.rsnajnl.org/cgi/content/full/2511080539/DC1>). Confocal laser scanning fluorescence microscopy was performed.

Data and Statistical Analysis

To determine the significance of the SI changes at different times, a paired *t* test analysis was performed by using the NER or percentage NER from matched

sections in the same animals at different times. A one-way analysis of variance, with Bonferroni post hoc multiple comparison tests, was used to compare the NER or percentage NER values among P947-injected, P1135-injected, and gadoterate meglumine-injected *ApoE*-deficient mouse groups, as well as the P947-injected WT mouse group. For all statistical analyses, *P* values less than .05 were considered significant. The analysis was performed by using software (Number Crunching Statistical System, 2001; NCS, Kaysville, Utah).

Results

P947 MR Imaging for in Vivo Atherosclerotic Lesions

Prior to injection of the contrast agent, MR imaging was performed in all mice

at baseline. Before injection of the contrast agent, MR imaging of *ApoE*-deficient mice showed a heterogeneous distribution of aortic wall thickening. Aortic wall thickening correlates strongly with the presence of atherosclerotic lesions, a finding that is consistent with results in our previously published studies (7). As anticipated, baseline MR imaging of WT mice showed no regions of aortic wall thickening.

After injection of P947, designed to specifically target MMPs, we (V.A., J.G.S.A., Z.A.F., M.S., S.A., F.H.) observed very strong and significant heterogeneous enhancement of the aortic wall in *ApoE*-deficient mice (Fig 1a). The change in the MR image SI of the aorta was measured by using the NER and percentage NER. In particular, by using *ApoE*-deficient mice, at 1 hour after injection, P947 provided an average

Figure 1

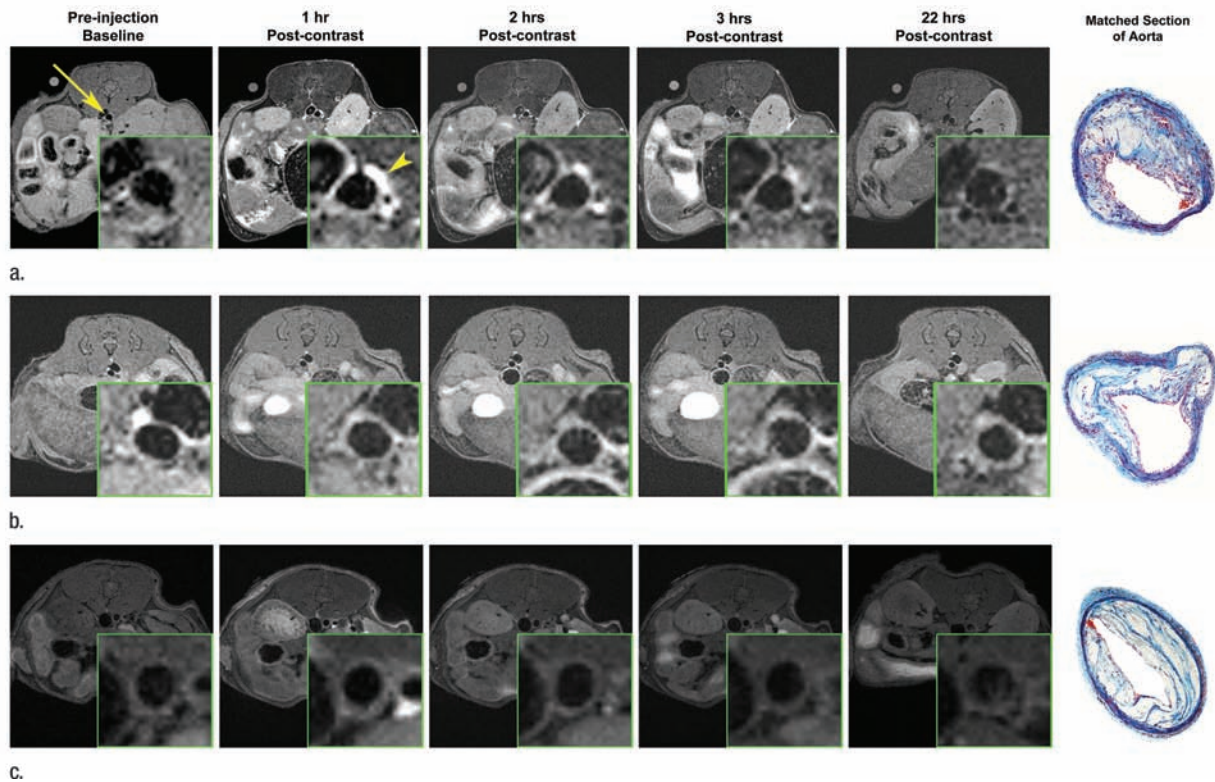
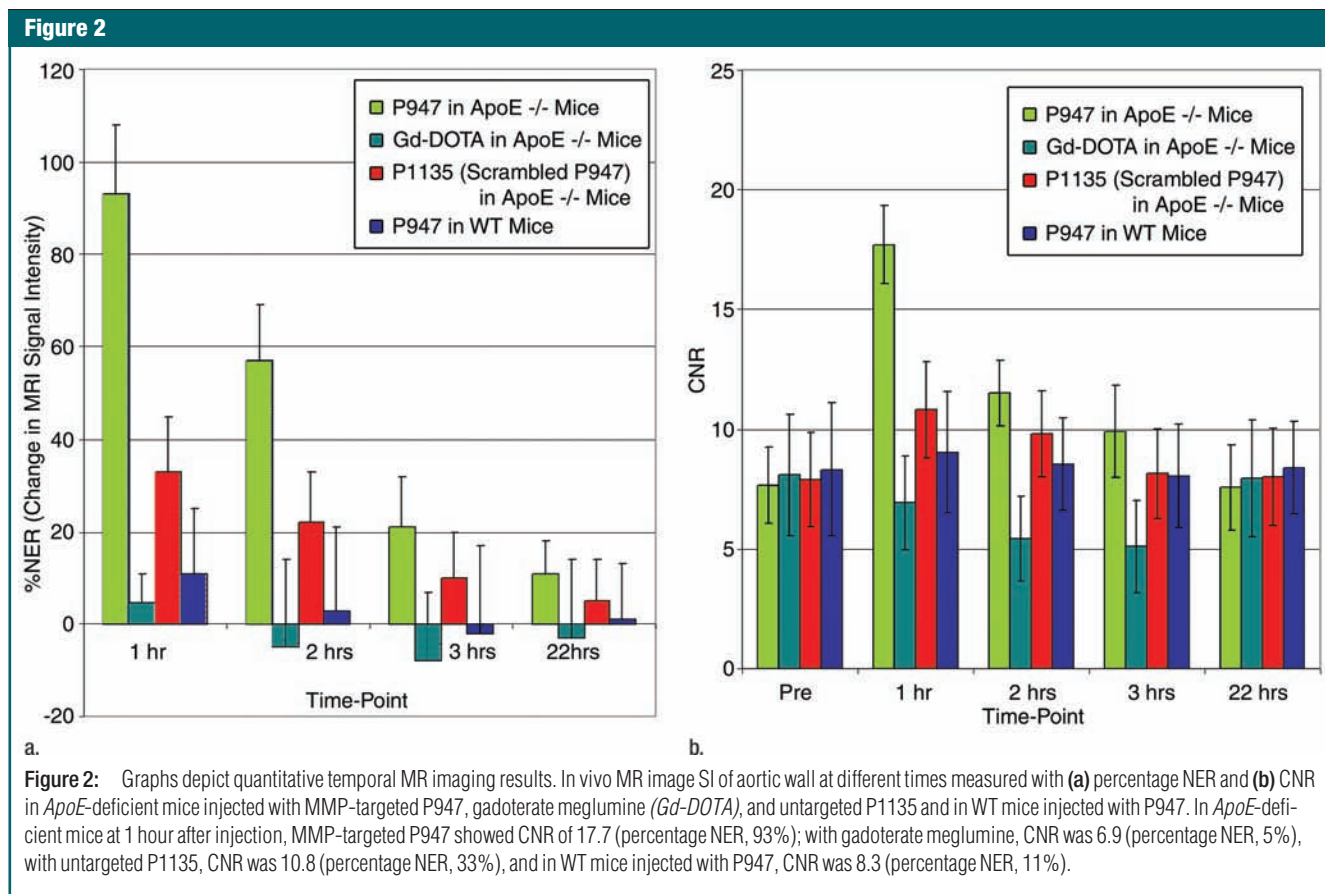


Figure 1: In vivo MR images acquired in atherosclerotic *ApoE*-deficient mice. (a) At baseline (before injection) and up to 22 hours after injection of MMP-targeted P947. (b) With untargeted P1135 (scrambled form of P947). (c) With standard nontargeted gadoterate meglumine. Insets are enlargements of aorta (arrow). Arrowhead points to MR inset of atherosclerotic aorta after injection with P947. (a–c) Farthest right images show hematoxylin-eosin–stained sections of aorta at identical anatomic level as MR images from same animal (17).



aortic MR image signal enhancement of 127% (percentage NER), corresponding to a mean NER of 2.27 ± 0.15 (standard deviation) when standardized to an externally placed standard solution of gadolinium and percentage NER of 93% (mean NER, 1.93 ± 0.15), when standardized to adjacent muscle tissue, as shown in Figures 1 and 2. In comparison, the percentage NER was 33% (mean NER, 1.33 ± 0.12) when we used P1135 and 4.6% (mean NER, 1.05 ± 0.06) with gadoterate meglumine (both also standardized to muscle). As demonstrated in Figures 1 and 2, these results translate into a three-fold increase in the MR image SI in the atherosclerotic aortic wall when we used P947 compared with P1135, a permuted derivative of P947 with the amino acid sequence scrambled such that specific binding to MMPs is lost, supporting an MMP-specific image enhancement mechanism. In addition,

the SI increase with P947 was 15-fold higher when compared with the standard nonspecific gadolinium-based MR imaging contrast agent gadoterate meglumine ($P < .001$) (Fig 2).

In nonatherosclerotic WT mice, at 1 hour after injection, P947 provided minimal change of the average aortic MR imaging SI that was homogeneous in nature and was not significant from baseline (Fig 2). At 2 and 3 hours after injection, no significant changes were seen ($P > .05$) of the aortic MR image SI values in WT mice (Fig 2).

Expression and Activity of MMPs in Mice at MR Imaging

After the MR imaging experiments were completed, the mice were sacrificed for various types of pathologic analyses of the atherosclerotic aortas (all animals). Pathologic analysis of aortic tissue sections was performed to validate expression of MMPs in atherosclerotic plaques

in *ApoE*-deficient mice used for the MR imaging experiments in this study. Tissue sections prepared from *ApoE*-deficient mouse aortas were immunostained for MMP-2, MMP-3, and MMP-9 for immunohistopathologic analysis. Atherosclerotic sections of *ApoE*-deficient mouse aortas revealed intense MMP-3 staining, as shown in Figure 3, consistent with substantial expression of MMP-3. Aortic sections immunostained for MMP-9 also showed high-level staining (Fig 3) that indicates a substantial level of MMP-9 in atherosclerotic lesions of *ApoE*-deficient mice. Finally, MMP-2-immunostained sections were also obtained (Fig 3) but showed less robust staining as compared with that for sections immunostained with MMP-3 and MMP-9.

To further assess MMPs in atherosclerosis, we performed (F.H., E.V., J.G.S.A., V.A., D.B.W.) in situ MMP zymography on atherosclerotic aortic

Figure 3

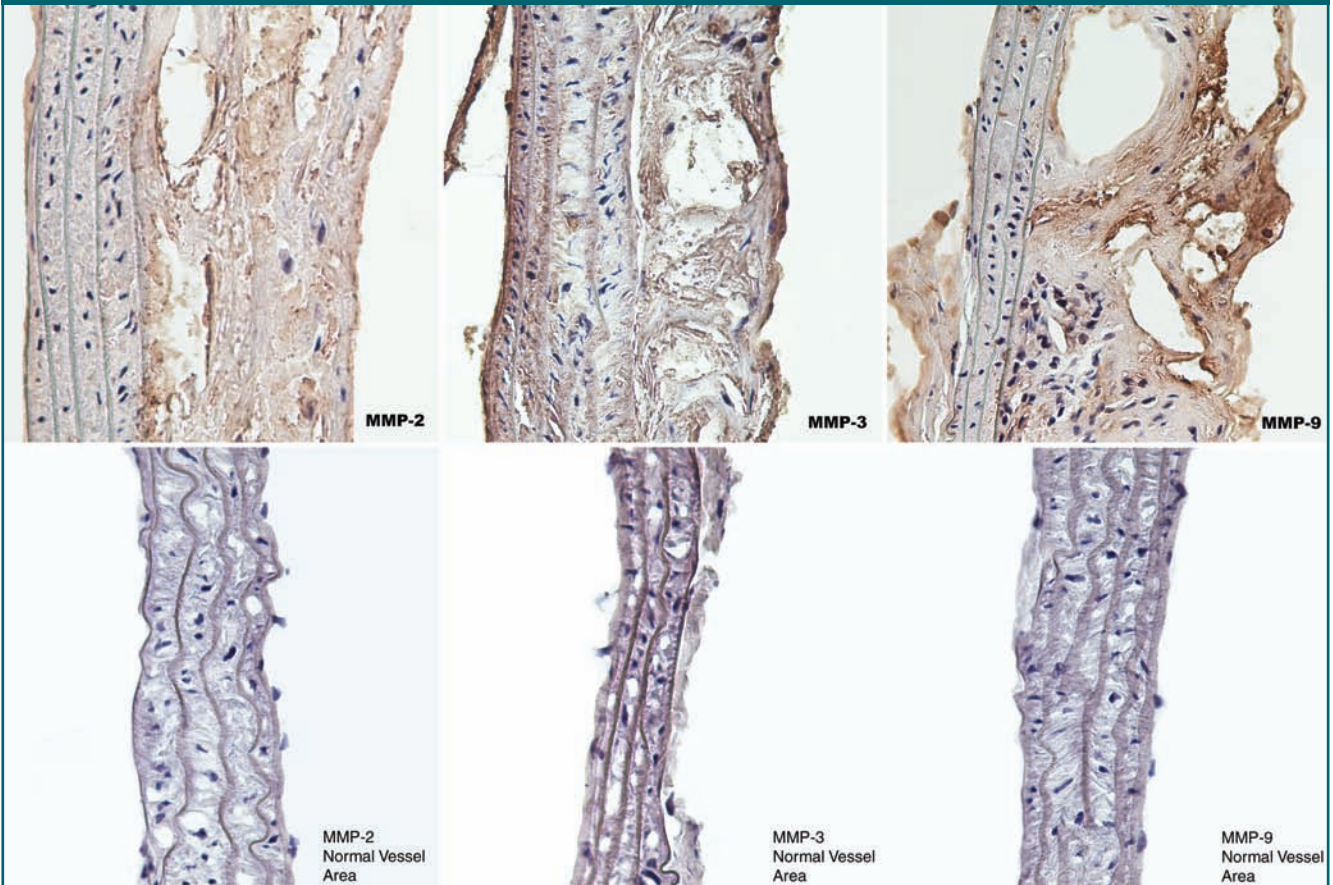


Figure 3: Light microscopic images show specific MMP immunostaining of atherosclerotic aortic lesions from *ApoE*-deficient mice. Top: Increased expression of MMP-2, MMP-3, and MMP-9. Bottom: Normal vessel areas. Sections were taken from aortas of mice used in in vivo MR imaging experiments.

Figure 4

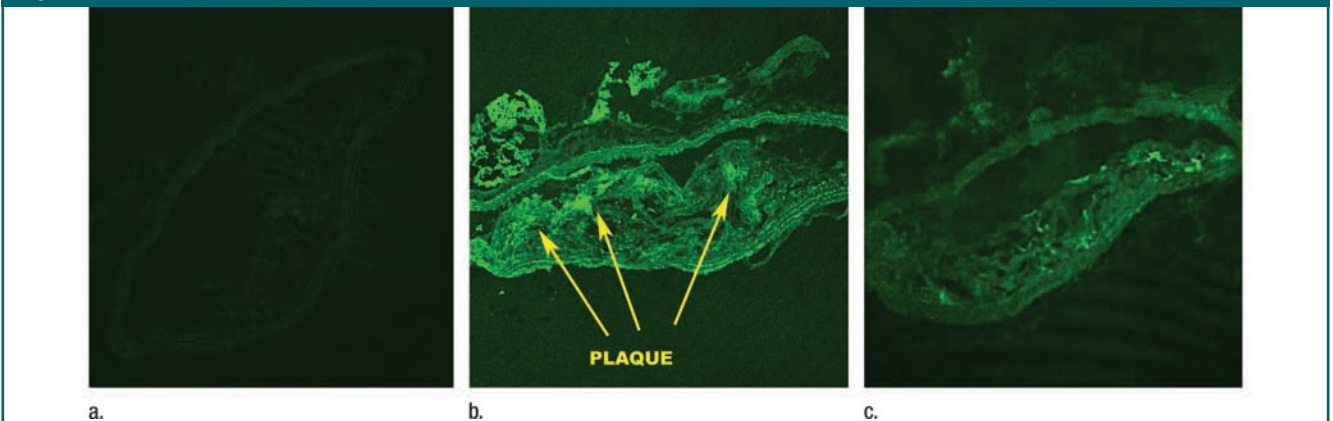


Figure 4: Fluorescence microscopic images from MMP in situ zymography of mouse aortas demonstrate increased MMP gelatinolytic activity in atherosclerotic sections. **(a)** Control section that was incubated with gel alone shows that there is negligible intrinsic fluorescence. **(b)** Atherosclerotic aortic section incubated with gel and dye that fluoresced only after cleavage by MMPs (arrows point to atherosclerotic plaque). **(c)** Atherosclerotic section that was incubated with gel, dye, and ethylenediaminetetraacetic acid, which blocks MMP activity, demonstrates that most of fluorescence seen on **(b)** was caused by active MMP cleavage of dye. All sections taken from mice used in in vivo MR imaging experiments.

sections from *ApoE*-deficient mice that were used in the imaging experiments. In contrast to MMP immunohistochemical staining, which is used to assess expression of both active and inactive MMPs, the information gathered by using MMP zymography represents MMP enzymatic activity. To perform the in situ zymography, we used an MMP substrate that becomes fluorescent only after cleavage by active MMPs. The in situ MMP zymography revealed substantial MMP activity in the aortic atherosclerotic lesions, as shown in Figure 4b. After broad-spectrum blockage of MMP activity by using ethylenediaminetetraacetic acid, there was a large decrease in the fluorescent activity seen in the atherosclerotic sections (Fig 4c). This behavior indicates that most of the fluorescence seen during MMP zymography is caused by MMP enzymatic activity. Sections incubated as controls did not show fluorescence (Fig 4a).

Distribution of Fluorescent P947 in Atherosclerotic Sections and Colocalization with MMPs

To confirm the in vivo distribution and localization of P947, we (E.L., C.C., S.L.G.) synthesized Eu-P947, replacing gadolinium on a one-to-one basis with europium (a lanthanide that is chemically similar to gadolinium), which fluoresces when excited at the appropriate wavelength, allowing detection by using confocal laser scanning microscopy. After intravenous injection of Eu-P947 (fluorescent P947), *ApoE*-deficient mice were sacrificed, and the aortas were dissected out to prepare tissue sections. The atherosclerotic aortic sections were then imaged by using confocal laser scanning fluorescence microscopy. The images obtained showed that Eu-P947 localized within areas of aortic plaque, as shown in Figure 5a. Most notably, in sections where a fibrous cap was apparent, there was significant Eu-P947 fluorescence signal emanating from the fibrous cap (far left and left images of Fig 5a).

To investigate the distribution of Eu-P947 in relation to MMPs, we (D.B.W., J.G.S.A., S.A., V.A.) performed specific fluorescent MMP immunostaining of aortic sections from *ApoE*-deficient

mice previously injected with Eu-P947. Specifically, we performed fluorescent immunostaining for MMP-2, MMP-3, and MMP-9 at 1 hour after injection (when the MR image signal was maximal with P947). Confocal laser scanning microscopy revealed substantial overlap in the distribution of MMP-3 fluorescence and Eu-P947 fluorescence, as shown in Figure 5c (combined overlaid image, right). Similarly, coregistration was present between the fluorescent signal emitted by Eu-P947 and MMP-9, as well as by MMP-2 (Fig 5d and 5b, respectively). Overall, the results of the confocal microscopy indicate that there is colocalization between fluorescent P947 (Eu-P947) and MMPs.

Discussion

A major result of this investigation was that MMP-targeted P947 facilitated significant MR image enhancement (increase of 93%) of the atherosclerotic vessel wall of *ApoE*-deficient mice in vivo. Our findings were specific because nontargeted P1135 (the scrambled form of P947 that has no specific binding to MMPs) delivered only one-third of the enhancement (33%) of P947, whereas gadoterate meglumine (a clinically available nonspecific MR imaging contrast agent) alone enhanced atherosclerotic lesions by only 4.6%, about 1/15 that of P947. These results suggest that the bulk of atherosclerosis enhancement seen with P947 is caused by specific binding, accumulation, or retention of P947, making P947 substantially better than any of the MR imaging contrast agents that we tested in this study.

The MR imaging results of this study correlated well with the standard pathologic findings in the regions of vessels that we imaged. As compared with normal vessels and controls, the atherosclerotic vessels from *ApoE*-deficient mice used in the MR imaging experiments showed significant overexpression of MMP-2, MMP-3, and MMP-9. This finding is consistent with findings in previously published data (9,20–23) and supports our hypothesis that MMP expression is largely responsible for P947-

mediated MR imaging enhancement of atherosclerosis in *ApoE*-deficient mice. As an additional mode of validation, in situ MMP zymography was performed on sections of atherosclerotic aortas from the mice used in the experiments. As compared with MMP immunostaining, zymography is a qualitative approach to examining functional MMP activity. The atherosclerotic vessel lesions that we examined showed substantial functional MMP activity. Control sections did not show significant fluorescence. Moreover, sections that were first incubated with a broad-spectrum MMP inhibitor showed dramatically less fluorescence. This indicates that the bulk of fluorescence seen at in situ zymography was caused by functional MMP activity.

The results of confocal laser scanning microscopy strongly suggest colocalization between specific MMPs and fluorescently tagged P947 (Eu-P947). There appeared to be large regions of strong MMP-3 staining that significantly overlapped with Eu-P947 distribution. Similarly, there was good overlap between the distributions of Eu-P947 and MMP-2, as well as MMP-9, which is thought to play a highly detrimental pathogenic role in atherosclerosis of *ApoE*-deficient mice (23). Overall, there were large areas of coregistration between the distribution of specific MMPs and fluorescent Eu-P947. This colocalization further supports the hypothesis that P947 accumulates and gets retained in atherosclerotic plaques largely because of binding by MMPs. We also examined the distribution of fluorescent Eu-P947 alone in atherosclerotic plaques. Notably, in atherosclerotic plaques where a fibrous cap was apparent, we noticed substantial Eu-P947 fluorescent signal by using confocal laser scanning microscopy. MMPs have been previously implicated in atherosclerotic fibrous cap thinning and degradation (9,20–22,24), a process thought to lead to plaque rupture and thrombosis.

The work presented in this study represents an innovative approach to imaging MMPs by using positive (increasing SI)-contrast agent molecular

Figure 5

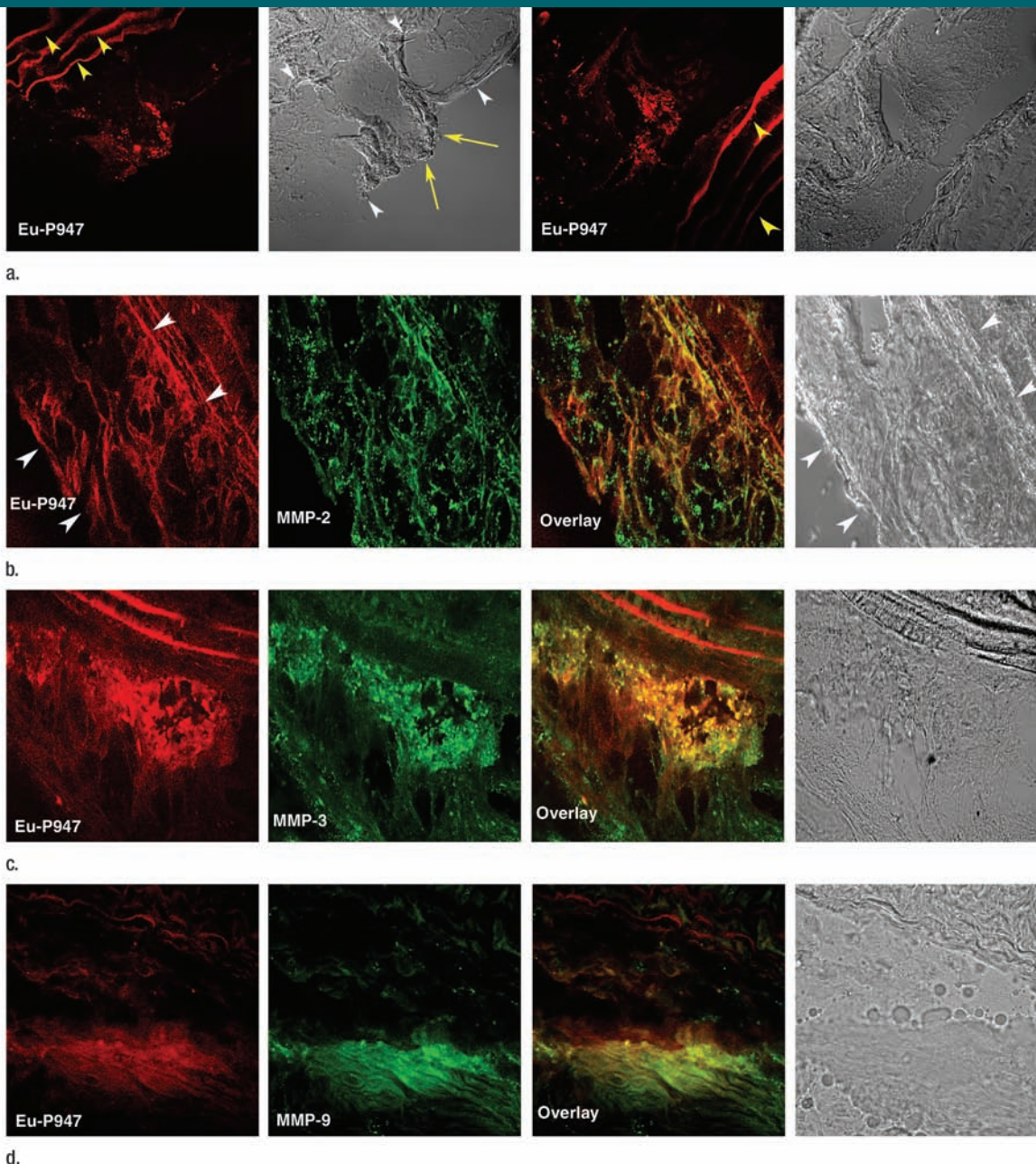


Figure 5: Confocal laser scanning fluorescence microscopic images show Eu-P947 localization alone and colocalization with MMPs. Aortic sections from *ApoE*-deficient mice injected in vivo with Eu-P947 show localization of fluorescent Eu-P947 (red) in atherosclerotic plaques (far left and right images on **a**), alternating with their corresponding differential interference contrast light microscopic images (left and far right images on **a**). Yellow arrowheads indicate elastic lamina. Arrows point to fibrous cap in light microscopic image and show possible accumulation of P947 in fibrous cap (far left image on **a**). Fluorescent immunostaining for MMP-2, MMP-3, and MMP-9 (green on **b–d**, respectively) in aortic sections obtained from *ApoE*-deficient mice injected with fluorescent Eu-P947 (red on **b–d**). Overlaid images at right on **b**, **c**, and **d** show substantial overlap (yellow areas) between Eu-P947 and MMP-2, MMP-3, and MMP-9, respectively. On **b** and **c**, differential interference contrast light microscopic images at right are of respective sections at left. White arrowheads indicate general borders of atherosclerotic plaques on **a** and **b**.

MR imaging. In previous work, researchers attempted to image MMPs with nuclear imaging, including positron emission tomography (PET) (25,26). Researchers used scintigraphy with a radiolabeled broad-spectrum MMP inhibitor to assess MMPs (26). In that study, the issue of the inherent low resolution of scintigraphy was circumvented by inducing a highly localized MMP and foam cell-rich lesion by using carotid artery ligation in *ApoE*-deficient mice fed a high-cholesterol diet (27); however, native atherosclerosis was not imaged in vivo. In vivo imaging of native atherosclerosis by using scintigraphy in mice or in human coronary arteries has yet to be demonstrated. However, nuclear imaging (including PET and single photon emission computed tomography [SPECT]) by using targeted radiotracers will likely offer superb sensitivity because of the very high signal-to-noise ratios possible with radiotracers.

In comparison, MR imaging offers superb anatomic resolution that is orders of magnitude higher than that of scintigraphy, including PET or SPECT. Molecular MR imaging, such as P947-enhanced MR imaging, will offer molecular information combined with superior anatomic resolution, which is vital for imaging small-scale pathologic findings such as atherosclerosis. Optical techniques have also been used to image MMPs. One group imaged gelatinases and MMPs by using a near-infrared fluorescence probe (28), whereas another group used optical techniques to image cathepsin K (a protease with some functions similar to those of MMPs) activity in atherosclerosis (29). Optical techniques offer good sensitivity and, with the use of near-infrared fluorescence probes, the specificity is also quite high, but direct translation of optical techniques into applications in humans remains technically unrealized for now.

At this time, it is not possible to use optical techniques to image structures that are deeper than a couple of centimeters beyond the surface. As such, application for noninvasive imaging in patients is currently not possible. Molecular MR imaging of atherosclerosis has

been investigated by using targeted imaging of neovascularization through the molecular target $\alpha_v\beta_3$ integrin (30), whereas others have targeted adhesion molecules, including vascular adhesion molecule 1 (31). To evaluate atherosclerosis with MR imaging, we used both gadolinium-carrying micelles (32,33) and molecularly targeted gadolinium immunomicelles to image macrophages through the macrophage scavenger receptor (19). An interesting observation made by Deguchi et al (28) was that there was an absence of MMP activity in atherosclerotic areas without a high content of macrophages (the presence of MMPs seemed to be a surrogate marker for the presence of macrophages). As such, molecular imaging of MMPs may offer a way to indirectly assess plaque macrophage content and macrophage activity. Although in our investigation we did not find gadoterate meglumine to be very helpful for imaging atherosclerosis, researchers have used gadopentetate dimeglumine to image fibrous components of atherosclerotic plaques with intravascular MR imaging, a relatively invasive imaging approach (34).

There were limitations to this study. P947 is a newer contrast agent, and we currently do not have substantial data regarding potential toxicity of this agent, including data in regard to tissue retention of gadolinium and nephrogenic systemic fibrosis. The goals of this study did not and could not realistically include full characterization of toxic effects, pharmacokinetics, and pharmacodynamics of this agent. These topics will require careful future investigation in multiple staged studies. Another potential limitation is translation to MR imaging for atherosclerosis in humans. Although it is entirely possible to perform excellent MR imaging of carotid artery and aortic atherosclerosis in patients, technical barriers still remain that make adequate MR imaging of the coronary arteries difficult.

In summary, we showed that P947 targeted to MMPs facilitated specific in vivo assessment of atherosclerosis at a molecular pathologic level. Knowledge about MMP content and activity may aid

in the prediction of the vulnerability of plaque to rupture and thrombosis. Therefore, with proper development and investigation, imaging with P947 (or similar MMP imaging approaches) may contribute substantially to preclinical and clinical evaluation of atherosclerosis.

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