



Published in final edited form as:

Semin Reprod Med. 2009 May ; 27(3): 229–239. doi:10.1055/s-0029-1216276.

Sex, Sex steroids and Brain injury

Paco S. Herson¹, Ines P. Koerner¹, and Patricia D. Hurn^{1,2,3}

¹ *Departments of Anesthesiology and Perioperative Medicine*

² *Departments of Physiology and Pharmacology*

³ *Department of Neurology*

Abstract

Biologic sex and sex steroids are important factors in clinical and experimental stroke and traumatic brain injury (TBI). Laboratory data strongly show that progesterone treatment after TBI reduces edema, improves outcomes and restores blood brain barrier function. Clinical studies to date agree with these data, and there are ongoing human trials for progesterone treatment after TBI. Estrogen has accumulated an impressive reputation as a neuroprotectant when evaluated at physiologically relevant doses in laboratory studies of stroke, but translation to patients remains to be shown. The role of androgens in male stroke or TBI is understudied and important to pursue given the epidemiology of stroke and trauma in men. To date, male sex steroids remain largely evaluated at the bench rather than the bedside. This review evaluates key evidence and highlights the importance of the platform on which brain injury occurs, i.e. genetic sex and hormonal modulators.

Keywords

stroke; brain ischemia; estradiol; neurosteroids; progesterone; allopreganolone; androgens

1. Sex differences in brain injury

It is now clear that biological sex alters the incidence of, and outcome from, ischemic and traumatic brain injury. For example, male sex is an acknowledged risk factor for stroke, and in most epidemiological series, stroke occurs more frequently in men vs. women. This sexually dimorphic disease pattern remains apparent until ages well beyond the menopausal years^{1, 2}. Nevertheless, stroke risk increases with age in both sexes, and there is broad evidence that outcome from an ischemic event is worse in aged women than in their male counterparts. Knowledge of mechanisms of ischemic cell death and neuroprotection is important for both sexes, but current evidence suggests that these mechanisms are not identical in males and females.

a. Sex differences in ischemic outcomes: animal models

Animal models of brain injury, typically rodents, have been used to evaluate side-by-side outcomes from ischemia or trauma. In most reports, females fare better than do their age-matched male counterparts. Early evidence in female vs. male spontaneously hypertensive, genetically stroke prone rats uncovered the male phenotype of “ischemia-sensitivity”³. This landmark study of 2000 animals showed that life expectancy is longer in the female, and the development of cerebral hemorrhage is delayed until an advanced age³. Subsequently, a

number of studies have shown that outcome from experimental brain injury is clearly sex-linked. Female rats and mice of various inbred and outbred strains experience smaller tissue damage for an equivalent insult from focal or global cerebral ischemia⁴⁻⁷ and improved functional outcome⁸. Similarly, male animals sustain greater injury than do age-matched females after traumatic brain injury⁹. We have explored complicated rodent models with genetic risk factors associated with human stroke, e.g. insulin-dependent genetic diabetes¹⁰, non-insulin dependent diabetes¹¹ and hypertension¹². In each genetic strain and despite deleterious complications from diabetes or hypertension, females are less sensitive to cerebral ischemia than are males.

b. Sex-specific cultures: hormone independent cell death or survival

In vitro data more directly support the concept that cell death mechanisms can be sex-specific. This specificity is typically modeled in primary cell cultures grown without background steroids. Under such conditions, some molecular pathways of cell death or survival diverge based on the genetic sex of the tissue. For example, cultured female dopaminergic neurons tolerate exposure to toxic dopamine concentrations and survive twofold relative to male cells¹³. Similarly, female neurons from the cortical plate or ventricular zone have greater longevity in culture than do male cells, and differentially express higher levels of phosphorylated kinases such as Akt¹⁴. Sensitivity to glutamate, peroxynitrate (ONOO) and staurosporine in neuronal culture is sex specific, with male neurons being more susceptible to glutamate and ONOO than females. In contrast, response to oxidants such as H₂O₂ is gender neutral¹⁵. These observations are not limited to neurons. Cell death resulting from oxygen-glucose deprivation is less in female vs male cultured astrocytes or hippocampal slices^{16, 17}.

In summary, these findings suggest that the response to cerebral injury in vivo and in vitro is partially a function of the sex of the cell. However, this in no way discounts the importance of gonadal steroids or brain-derived neurosteroids as modulators of oxidant, toxic and ischemic challenges to the brain. The role of progesterone, estradiol and testosterone in shaping neuro-injury is reviewed in subsequent sections.

2. Neuroprotective effects of Progesterone

Progesterone is synthesized from cholesterol by the gonads, adrenal gland or placenta. In addition, progesterone can be generated within the brain as a neurosteroid, i.e. steroid hormones that accumulate in the central nervous system (CNS) either by *de novo* synthesis from cholesterol or *in situ* metabolism of precursors from the blood¹⁸. In contrast, the term neuro-active steroid refers to any steroid having a direct effect in the CNS, independent of source of production. Within brain, progesterone is metabolized to the highly neuro-active metabolites 5 α DH-progesterone and 3 α ,5 α TH-progesterone (allopregnanolone; ALLO). Progesterone's neuroprotective actions are mediated, in part, by these neuro-active metabolites.

Exogenous progesterone protects the CNS in a variety of experimental animal models of neurodegeneration, including spinal cord injury¹⁹⁻²⁴, penetrating or diffuse brain injury, traumatic brain injury (TBI)²⁵, transient global and focal cerebral ischemia {Stein, 2008 12702 /id;Schumacher, 2007 12749 /id}. Many of these experimental studies emphasize progesterone's attractive attributes from a clinical perspective, i.e. its relatively long therapeutic window that extends up to 2 hours after middle cerebral artery occlusion²⁷ and to 24 hrs after experimental TBI²⁸. The ability of the active metabolite ALLO to mimic the beneficial effects of progesterone in many models points to metabolism in the CNS as a key factor in the steroid's neuroprotection. In contrast, recent clinical data showed no benefit or even harmful effects on incidence of cerebrovascular disease in postmenopausal women using estrogen/progestin formulations²⁹⁻³¹. One explanation for this apparent discrepancy is that most synthetic progestins cannot be converted to neuro-active metabolites such as ALLO. In fact, the most

commonly used progestin in hormone replacement therapy, medroxyprogesterone acetate (MPA), appears to antagonize the beneficial effects of estrogen against glutamate toxicity in hippocampal neuronal cultures^{32, 33}. In contrast, progesterone protects hippocampal neurons from glutamate toxicity and provides an additive benefit to estrogen-treated neurons^{32, 34}. Similarly, MPA decreases estrogen protection against cerebral ischemia in subcortical brain regions in rat³⁵, while progesterone does not³⁶. Therefore, our emerging knowledge of steroid synthesis and metabolism within the CNS may inform our clinical use of these complex substances.

a. Neurosteroids

Evidence began to emerge in the early 1980's that the brain is a steroidogenic organ, when Baulieu and co-workers found that the steroids dehydroepiandrosterone (DHEA) and pregnenolone, and their sulfated esters, were present in greater concentration in the brain than in circulation^{37, 38}. Importantly, brain concentrations of all these steroids remained very high after adrenalectomy and gonadectomy. This gave rise to the now well accepted concept that steroids are produced in the brain, termed neurosteroids^{18, 22}. The metabolic enzymes required to generate and metabolize steroids are present throughout the rodent and human central nervous system, although not expressed uniformly in all brain regions (For review see 39-41). The non-uniform distribution of the metabolic enzymes points to the possibility of regional differences in generation and metabolism of neurosteroids, although this has not yet been thoroughly investigated.

Synthesis of neurosteroids begins with the conversion of cholesterol to pregnenolone within the mitochondria by the enzyme cytochrome P450scc (cholesterol side-chain cleavage enzyme) (Fig. 1). P450scc is the rate limiting enzyme in neurosteroid biosynthesis. Interestingly, P450scc, and its human counterpart Cyp11a1, is more highly expressed in female brain as compared to male^{42, 43}, indicating the possibility of sexually dimorphic synthesis of neurosteroids. Pregnenolone is the precursor of all neurosteroids, being converted directly into DHEA or progesterone (Fig. 1). Further metabolism of DHEA leads to the production of androgens (testosterone and DHT) as well as 17 β -estradiol and derivatives. On the other hand, progesterone is predominantly converted to its highly active metabolites 5 α -DHP and 3 α ,5 α -THP (ALLO). While the focus of this review is on the beneficial effects of estradiol and progesterone, it is important to note that DHEA and its sulfated ester DHEAS are neuroprotective in a variety of experimental models. DHEA and DHEAS protect rat hippocampal neurons against N-methyl-D-aspartate (NMDA)-induced excitotoxicity^{44, 45}, β -amyloid peptide toxicity⁴⁶ and oxygen-glucose deprivation⁴⁷. Importantly, DHEAS prevents ischemia-induced hippocampal neuron cell loss and impairment of hippocampal long-term potentiation following forebrain ischemia⁴⁸. In addition, the PROG precursor PREG and PREGS also exhibit beneficial effects in experimental models of neurodegeneration^{49, 50}.

b. Progesterone and Stroke

Acute administration of progesterone either immediately before or after transient focal cerebral ischemia decreases infarct volume and improves behavioral outcome in male mice^{51, 52} and rats⁵³⁻⁵⁷ and reduces hippocampal neuron loss following global cerebral ischemia (four vessel occlusion; 4-VO) in male rats⁵⁸. The therapeutic window for acute progesterone in male rats has been shown to be 2 hours after focal cerebral ischemia²⁷ and 24 hr after global ischemia⁵⁸, making it a promising molecule for pharmacological intervention following stroke. Notably, progesterone is an effective neuroprotectant in the more complex cat brain⁵⁹. While most of the studies to date have focused on male animals, progesterone is effective in reproductively senescent aged and young ovariectomized female rats^{53, 54}. However, others have observed either no benefit³⁶ or worse outcome⁶⁰ following chronic progesterone treatment, suggesting that dose and duration of steroid administration is important to outcome.

c. Progesterone and Traumatic Brain Injury

Progesterone has been well-studied in models of TBI, for example controlled contusion of the medial frontal cortex (For review see ²⁵). Combined pre-and post-injury treatment decreases cerebral edema and neuronal loss at 72 hours in both young and aged male and female animals^{61, 62}. Similar results are observed with doses administered up to 48 hours post-injury, although the steroid was most effective when administered within 2 hr of trauma²⁸. Similarly, physiological levels of progesterone provide protection in ovariectomized female rodents⁶³. The main effect observed in these studies is that of restricting post-injury edema. As with ischemic brain injury, progesterone's benefits are dose dependent⁶⁴ and may be lost at very high doses⁶⁵. Importantly, progesterone treatment improves functional recovery following experimental TBI^{62, 66-68}.

These experimental observations have been extended to clinical trials involving patients with TBI⁶⁹. Patients of both sexes (n=100) with moderate to severe brain damage from blunt head trauma were enrolled in a prospective, randomized, placebo controlled study. Progesterone was continuously administered by intravenous infusion for 3 days post-injury, and mortality was reduced in patients receiving progesterone as compared to placebo-treatment. Functional recovery was also significantly better in moderately injured patients who received progesterone. Similar positive results have been recently reported with severe traumatic brain injury⁷⁰. While these results must be confirmed in a larger, fully powered trial, they are the first evidence that progesterone may be effective for the treatment of clinical TBI.

3. Mechanisms of Progesterone Neuroprotection

Studies of experimental TBI again provide the best clues as to progesterone's mechanisms of protection. For example, PROG decreases vasogenic and cytotoxic cerebral edema following TBI²⁸ and focal stroke^{51, 52, 71}. Recent evidence suggests that the steroid regulates expression of aquaporin-4, a water permeable channel that likely plays a significant role in edema formation following brain injury⁷². Additional evidence implicates progesterone's transcriptional actions, by increasing expression of anti-apoptotic genes such as Bcl-2, and decreasing pro-apoptotic genes such as bax, bad and caspase-3 activity^{73, 74}. Inhibition of sigma1 receptors has also been implicated⁷⁵, as has anti-oxidant actions as revealed by a reduction in lipid peroxidation after TBI⁷⁶. Progesterone is thought to have anti-inflammatory actions by reducing microglia activation, toll-like receptor expression⁶⁸ and pro-inflammatory cytokines^{77, 78, 79, 80}.

In addition to neuroprotection, progesterone may play a role in post-injury remyelination and repair²⁶. The steroid increases the rate of myelin formation in cultured Schwann cells⁸¹ and improves regeneration of lesioned mouse sciatic nerve⁸². Within the central nervous system, progesterone stimulated oligodendrocyte proliferation and subsequent myelination^{20, 83-85}, possibly by progesterone receptor dependent mechanisms⁸³. Interestingly, the latter is the only effect to date directly attributed to classical nuclear progesterone receptor signaling.

4. Allopreganolone

The ability of the nervous system to metabolize progesterone complicates our understanding of its mechanism of protection. To date few studies have demonstrated that progesterone, rather than a metabolite, is the active molecule responsible for protection in brain ischemia and injury. In fact, Sayeed et al. recently demonstrated that ALLO is more effective than progesterone in transient ischemic injury, suggesting that it is the metabolite that is of greatest potency⁵⁵. Further, we have observed that progesterone protection of cerebellar Purkinje cells treated with oxygen and glucose deprivation requires metabolism to ALLO⁸⁶.

ALLO is synthesized in the brain by sequential reductions by 5 α -R and 3 α -HSD. 5 α -R reduces progesterone to 5 α DH-PROG in a uni-directional reaction that is rate-limiting in the production of ALLO. 3 α -HSD converts 5 α DH-PROG into 3 α ,5 α TH-PROG (ALLO), and is also capable of oxidizing ALLO into 5 α DH-PROG. ALLO is neuroprotective in several animal models of neurodegeneration and injury, including Alzheimers⁸⁷, Niemann-Pick C disease⁸⁸, kainate excitotoxicity⁸⁹, focal cerebral ischemia⁵⁵, and TBI⁹⁰. ALLO improves post-injury neurogenesis⁹¹, decrease apoptosis via activation of PKB/Akt kinase⁹², and like progesterone, increases anti-apoptotic gene transcription^{93, 94} and decreases inflammation⁷⁹. Because ALLO is among the most abundant and potent endogenous positive modulators of GABA_A receptors, most physiological effects of ALLO are attributed to this mechanism (For review see⁹⁵). Interaction with GABA_A receptors may also hold the key to ALLO's neuroprotection, as we have recently demonstrated in cultured neurons treated with *in vitro* "ischemic" insults⁸⁶.

5. Summary

The brain is a steroidogenic organ, possessing the metabolic enzymes necessary to produce most steroid hormones. Progesterone, among the most abundant neurosteroids in the brain, is neuroprotective in a variety of experimental animal settings. This protection may be mediated by altering specific gene expression via binding to the classical progesterone receptor (PR). Alternatively, neuro-active progesterone metabolites interact directly with neuronal membrane receptors and intracellular signaling cascades. In fact, the active metabolite ALLO mimics PROG neuroprotection in most experimental settings. Few studies have attempted to determine the relation between the parent hormone progesterone and its conversion to metabolites in animal studies of brain injury, but our recent data suggests that progesterone neuroprotection is completely mediated by metabolism to active metabolites, likely ALLO.

6. Estradiol: The best studied sex steroid in neuroprotection

The estrogens, particularly E2, have been widely studied in experimental ischemic and hemorrhagic brain injury. Almost without exception, these studies report that E2 reduces tissue damage, improves functional recovery and may stimulate repair processes. Interest in the estrogen steroid family arose, in part, from clinical use of contraceptives and of perimenopausal hormone replacement therapy in women with potential cerebrovascular disease. In addition, two key observations fueled ongoing interest in estrogens as neuroprotectants. First, outcome from experimental stroke in female animals is influenced by the stage of the estrous cycle. Proestrus is associated with smaller tissue damage (quantified as infarct size) after focal cerebral ischemia in rat, and this association is generally attributed to the high levels of E2 produced during this stage. In contrast, metestrus (low E2 production) is associated with greater brain damage⁹⁶. Second, female animals sustain less tissue damage than do males after similar ischemic or traumatic insults^{5, 6, 53, 97}. The benefit of female gender is lost in reproductively senescent animals or after ovariectomy but can be restored by estrogen supplementation^{5, 53}. These data emphasize that estrogens are highly important to outcome from ischemic brain injury in the female.

a. Basis for estrogen as a neuroprotective therapy

Subsequently, a large number of laboratories focused on the estrogens as a form of endogenous neuroprotection or as a potential therapy for stroke⁹⁸. Estrogen therapy of a variety of types and formulations has been shown to be beneficial in the male ischemic brain⁹⁹⁻¹⁰¹. While most of these animal studies emphasized tissue damage quantified as infarct size or cell loss early after the insult, chronic E2 supplementation also improves functional outcome^{8, 102}.

Despite the mass of evidence that E2 is neuroprotective, it should be noted that some investigators have found no effect or even detrimental effects of E2 in experimental stroke. Such findings may be related to the dose of E2 used since neuroprotection may be lost or detrimental effects may occur at higher doses^{12,103}. E2 may also be less beneficial with increased severity of injury, e.g., prolonged or permanent vessel occlusion as opposed to transient occlusion^{104, 105}. Age may also be a confounding factor, as steroid receptor expression is altered with reproductive senescence¹⁰⁶. A recent study showed that E2 replacement reduced infarct size, systemic and brain inflammation only if it was initiated immediately after ovariectomy, but not after a prolonged period of hypoestrogenicity¹⁰⁷. This observation, if translated to the human, may have clinical implications for postmenopausal women. Overall, experimental data supports a beneficial effect of E2, however dissenting findings suggest that E2 mediated neuroprotection may depend on the specifics of the experimental and clinical situation.

One final consideration is that acute steroid injection has been protective in some studies, while in others only chronic therapy produces beneficial results^{108, 109}. The latter issue influenced subsequent investigations into E2's protective mechanisms, i.e. if such mechanisms are mediated by the steroid's cognate estrogen receptors (ER) of either subtype and if transcriptional processes are required for benefit. A profound understanding of these mechanisms is required if we are to develop drugs that mirror estrogen's neuroprotection without the undesirable hormonal effects.

7. Mechanisms of Estrogenic Neuroprotection

It is now well-recognized that E2 elicits both genomic and non-genomic protective actions following an ischemic insult (**Fig.2**). These actions include stabilizing the blood-brain barrier and subsequently reducing brain edema^{110,111}, increasing blood flow during and after the ischemic insult^{112, 113}, engaging anti-inflammatory molecules^{114, 115}, and increasing expression of cell-survival mediators such as bcl2 and the novel cocaine and amphetamine regulated transcript (CART)^{116, 117}. In addition, E2 can act as a concentration-dependent antioxidant and anti-lipid peroxidation agent¹¹⁸⁻¹²⁰. The steroid also interacts with N-methyl-D-aspartate (NMDA) receptors, producing receptor activation at low doses (Connell BJ et al., 2007), but inhibiting receptors and subsequently blunting excitotoxicity at higher E2 doses¹²¹. Lastly, E2 not only acts as an acute protectant after ischemia but can also improve regeneration and plasticity of new neurons¹⁰⁷. This may contribute to improved post-ischemic memory after ischemia that is seen in estrogen-supplemented animals¹⁰².

A plethora of literature documents E2 mechanisms that are transcriptional, involving genes that do or do not carry estrogen response elements (EREs), as well as non-transcriptional rapid signaling action through interaction with membrane-bound G-proteins, modification of protein phosphorylation and of intracellular second messenger levels such as cAMP or calcium. The relative importance of these mechanisms in explaining E2's ability to improve ischemic and traumatic neuronal injury is unclear. For example, whether classical nuclear ERs are essential for neuroprotection has remained controversial for some time, and there is disagreement amongst studies that use pharmacological ER agonists vs. genetically ER deficient mice. It is known that both ER- α and ER- β are widely expressed in all cell types throughout the brain, i.e., neurons, glia, and endothelial cells, and are present in ischemia-sensitive areas such as neocortex and hippocampus¹²²⁻¹²⁴. Not surprisingly, concentrations of ER- α and ER- β are higher in adult females as compared to males¹²². We showed in a very early study that generalized pharmacological blockade of ERs with ICI182,780 exacerbated ischemic brain injury in female mice, suggesting that nuclear receptors play a role¹²⁵. However, ER α gene deletion in female mice was paradoxically associated with decreased infarct size, which may be related to the enhanced brain tissue perfusion observed in ER α knockout mice¹²⁶. In a

model of permanent cerebral vessel occlusion, E2 failed to protect cortical tissue in ER α , but not ER β , knockout mice, suggesting that E2 may signal through ER α to reduce stroke damage¹²⁷. However, a selective ER β , but not ER α , agonist reduced hippocampal ischemic damage in ovariectomized mice¹², suggesting that different receptor subtypes may mediate E2 signaling in a model-specific and possible brain-region specific manner. Clearly, further work is required to translate our extensive understanding of E2 signaling under physiological conditions to those of brain injury.

8. Androgens and Brain Injury in the Male

Despite the relative paucity of data that address “male sensitivity” to brain injury, emerging evidence suggests that androgens strongly impact outcome and mechanisms of cell death. In men, low testosterone levels have been associated with poor outcome after acute ischemic events¹²⁸. Androgen levels are inversely associated with stroke severity, infarct size, and 6-month mortality; and total and free testosterone levels tend to normalize within 6 months following stroke. These data do not necessarily suggest a direct causal relationship because brain injury provokes an acute stress reaction that causes a reduction in plasma testosterone. Animal studies are starting to address this issue. In male rats, androgen replacement in castrates increases histological damage from stroke^{100, 129, 130}, while stressors that reduce pre-ischemic testosterone level also reduce stroke damage¹³¹. In contrast, testosterone replacement in male castrates *after* stroke accelerates functional recovery⁶⁷. Beneficial effects of androgens following peripheral nerve damage or brain trauma have also been reported in animals¹³²⁻¹³⁴. These apparently conflicting results may be reconciled by the hypothesis that androgens are deleterious during acute injury but beneficial during the recovery phase. Potential mechanisms by which androgens could enhance post-stroke recovery include normalization of reperfusion, promotion of axonal regeneration, synaptogenesis, and neurogenesis¹³⁵.

9. Conclusion

Biologic sex and sex steroids are important factors in clinical and experimental brain injury. Our understanding of these factors is relatively recent, in part, because many pre-clinical and mechanistic studies have been carried out only in male animals and mixed sex cell cultures. Estrogen has accumulated an impressive reputation as a neuroprotectant in physiologically relevant doses in laboratory studies, but translation to patients remains to be shown. Laboratory data strongly show that progesterone treatment after TBI reduces edema, improves outcomes and restores blood brain barrier function. Clinical studies to date agree with these data, and there are ongoing human trials for progesterone treatment after TBI. The role of androgens in male stroke or TBI is understudied and important to pursue given the epidemiology of stroke and trauma in men. To date, male sex steroids remain largely evaluated at the bench rather than the bedside. Further studies of genetic sex and of the hormonal platform on which brain injury occurs are needed and should provide new insights for science and health care.

Acknowledgements

Support for this research was provided by NIH grants NS049210, NS 058792, and the Medical Research Foundation of Oregon. The authors gratefully acknowledge Executive Specialist Ashley Branch for expert manuscript preparation.

References

1. Giroud M, Milan C, Beuriat P, et al. Incidence and survival rates during a two-year period of intracerebral and subarachnoid haemorrhages, cortical infarcts, lacunes and transient ischaemic attacks. the stroke registry of dijon: 1985–1989. *Int J Epidemiol* 1991;20(4):892–899. [PubMed: 1800427]

2. Sacco RL, Boden-Albala B, Gan R, et al. Stroke incidence among white, black, and hispanic residents of an urban community: The northern manhattan stroke study. *Am J Epidemiol* 1998;147(3):259–268. [PubMed: 9482500]
3. Yamagata K, Tagami M, Ikeda K, Yamori Y, Nara Y. Altered gene expressions during hypoxia and reoxygenation in cortical neurons isolated from stroke-prone spontaneously hypertensive rats. *Neurosci Lett* 2000;284(3):131–134. [PubMed: 10773416]
4. Hall ED, Pazara KE, Braughler JM. Effects of tirilazad mesylate on postischemic brain lipid peroxidation and recovery of extracellular calcium in gerbils. *Stroke* 1991;22:361–366. [PubMed: 2003306]
5. Alkayed NJ, Harukuni I, Kimes AS, London ED, Traystman RJ, Hurn PD. Gender-linked brain injury in experimental stroke. *Stroke* 1998;29:159–166. [PubMed: 9445346]
6. Carswell HV, Anderson NH, Clark JS, et al. Genetic and gender influences on sensitivity to focal cerebral ischemia in the stroke-prone spontaneously hypertensive rat. *Hypertension* 1999;33(0194–911 2):681–685. [PubMed: 10024327]
7. Alkayed NJ, Murphy SJ, Traystman RJ, Hurn PD, Miller VM. Neuroprotective effects of female gonadal steroids in reproductively senescent female rats. *Stroke* 2000;31(0039–2499 1):161–168. [PubMed: 10625733]
8. Li X, Blizzard KK, Zeng Z, DeVries AC, Hurn PD, McCullough LD. Chronic behavioral testing after focal ischemia in the mouse: Functional recovery and the effects of gender. *Exp Neurol* 2004;187(1):94–104. [PubMed: 15081592]
9. Bramlett HM, Dietrich WD. Neuropathological protection after traumatic brain injury in intact female rats versus males or ovariectomized females. *J Neurotrauma* 2001;18(0897–7151 9):891–900. [PubMed: 11565601]
10. Toung TK, Hurn PD, Traystman RJ, Sieber FE. Estrogen decreases infarct size after temporary focal ischemia in a genetic model of type 1 diabetes mellitus. *Stroke* 2000;31(1524–4628 11):2701–2706. [PubMed: 11062297]
11. Vannucci SJ, Willing LB, Goto S, et al. Experimental stroke in the female diabetic, db/db, mouse. *J Cereb Blood Flow Metab* 2001;21(0271–678 1):52–60. [PubMed: 11149668]
12. Carswell HV, Macrae IM, Gallagher L, Harrop E, Horsburgh KJ. Neuroprotection by a selective oestrogen receptor {beta} agonist in a mouse model of global ischaemia. *Am J Physiol Heart Circ Physiol*. 2004;(0363–6135)
13. Lieb K, Andrae J, Reisert I, Pilgrim C. Neurotoxicity of dopamine and protective effects of the NMDA receptor antagonist AP-5 differ between male and female dopaminergic neurons. *Exp Neurol* 1995;134(0014–4886 2):222–229. [PubMed: 7556542]
14. Zhang L, Li PP, Feng X, Barker JL, Smith SV, Rubinow DR. Sex-related differences in neuronal cell survival and signaling in rats. *Neurosci Lett* 2003;337(0304–3940 2):65–68. [PubMed: 12527389]
15. Du L, Bayir H, Lai Y, et al. Innate gender-based proclivity in response to cytotoxicity and programmed cell death pathway. *J Biol Chem*. 2004;(1083–351)
16. Liu M, Hurn PD, Roselli CE, Alkayed NJ. Role of P450 aromatase in sex-specific astrocytic cell death. *J Cereb Blood Flow Metab* 2007;27(1):135–141. [PubMed: 16736049]10.1038/sj.jcbfm.9600331
17. Li H, Pin S, Zeng Z, Wang MM, Andreasson KA, McCullough LD. Sex differences in cell death. *Ann Neurol* 2005;58(0364–5134 2):317–321. [PubMed: 15988750]
18. Baulieu EE, Robel P, Schumacher M. Neurosteroids: Beginning of the story. *Int Rev Neurobiol* 2001;46(0074–7742):1–32. [PubMed: 11599297]
19. Schumacher M, Baulieu EE. Neurosteroids: Synthesis and functions in the central and peripheral nervous systems. *Ciba Found Symp* 1995;191(0300–5208):90–106. [PubMed: 8582208]
20. Jung-Testas I, Schumacher M, Robel P, Baulieu EE. The neurosteroid progesterone increases the expression of myelin proteins (MBP and CNPase) in rat oligodendrocytes in primary culture. *Cell Mol Neurobiol* 1996;16(0272–4340 3):439–443. [PubMed: 8818411]
21. Schumacher M, Robel P, Baulieu EE. Development and regeneration of the nervous system: A role for neurosteroids. *Dev Neurosci* 1996;18(1–2):6–21. [PubMed: 8840083]
22. Baulieu EE. Neurosteroids: Of the nervous system, by the nervous system, for the nervous system. *Recent Prog Horm Res* 1997;52(0079–9963):1–32. [PubMed: 9238846]

23. De Nicola AF, Gonzalez SL, Labombarda F, et al. Progesterone treatment of spinal cord injury: Effects on receptors, neurotrophins, and myelination. *J Mol Neurosci* 2006;28(0895–8696 1):3–15. [PubMed: 16632872]
24. Labombarda F, Gonzalez S, Gonzalez Deniselle MC, et al. Progesterone increases the expression of myelin basic protein and the number of cells showing NG2 immunostaining in the lesioned spinal cord. *J Neurotrauma* 2006;23(0897–7151 2):181–192. [PubMed: 16503802]
25. Stein DG. Progesterone exerts neuroprotective effects after brain injury. *Brain Res Rev* 2008;57(0165–0173 2):386–397. [PubMed: 17826842]
26. Schumacher M, Guennoun R, Stein DG, De Nicola AF. Progesterone: Therapeutic opportunities for neuroprotection and myelin repair. *Pharmacol Ther* 2007;116(0163–7258 1):77–106. [PubMed: 17659348]
27. Jiang N, Chopp M, Stein D, Feit H. Progesterone is neuroprotective after transient middle cerebral artery occlusion in male rats. *Brain Res* 1996;735(0006–8993 1):101–107. [PubMed: 8905174]
28. Roof RL, Duvdevani R, Heyburn JW, Stein DG. Progesterone rapidly decreases brain edema: Treatment delayed up to 24 hours is still effective. *Exp Neurol* 1996;138(0014–4886 2):246–251. [PubMed: 8620923]
29. Rossouw JE, Anderson GL, Prentice RL, et al. Risks and benefits of estrogen plus progestin in healthy postmenopausal women: Principal results from the women's health initiative randomized controlled trial. *JAMA* 2002;288(0098–7484 3):321–333. [PubMed: 12117397]
30. Simon JA, Hsia J, Cauley JA, et al. Postmenopausal hormone therapy and risk of stroke: The heart and estrogen-progestin replacement study (HERS). *Circulation* 2001;103(1524–4539 5):638–642. [PubMed: 11156873]
31. Hulley S, Grady D, Bush T, et al. Randomized trial of estrogen plus progestin for secondary prevention of coronary heart disease in postmenopausal women. heart and Estrogen/progestin replacement study (HERS) research group. *JAMA* 1998;280(0098–7484 7):605–613. [PubMed: 9718051]
32. Nilsen J, Brinton RD. Divergent impact of progesterone and medroxyprogesterone acetate (provera) on nuclear mitogen-activated protein kinase signaling. *Proc Natl Acad Sci U S A* 2003;100(0027–8424 18):10506–10511. [PubMed: 12925744]
33. Nilsen J, Morales A, Brinton RD. Medroxyprogesterone acetate exacerbates glutamate excitotoxicity. *Gynecol Endocrinol* 2006;22(0951–3590 7):355–361. [PubMed: 16864144]
34. Nilsen J, Brinton RD. Impact of progestins on estradiol potentiation of the glutamate calcium response. *Neuroreport* 2002;13(0959–4965 6):825–830. [PubMed: 11997695]
35. Littleton-Kearney MT, Klaus JA, Hurn PD. Effects of combined oral conjugated estrogens and medroxyprogesterone acetate on brain infarction size after experimental stroke in rat. *J Cereb Blood Flow Metab* 2005;25(0271–678 4):421–426. [PubMed: 15689957]
36. Toung TJ, Chen TY, Littleton-Kearney MT, Hurn PD, Murphy SJ. Effects of combined estrogen and progesterone on brain infarction in reproductively senescent female rats. *J Cereb Blood Flow Metab* 2004;24(0271–678 10):1160–1166. [PubMed: 15529016]
37. Corpechot C, Robel P, Axelson M, Sjoval J, Baulieu EE. Characterization and measurement of dehydroepiandrosterone sulfate in rat brain. *Proc Natl Acad Sci U S A* 1981;78(0027–8424 8):4704–4707. [PubMed: 6458035]
38. Corpechot C, Synguelakis M, Talha S, et al. Pregnenolone and its sulfate ester in the rat brain. *Brain Res* 1983;270(0006–8993 1):119–125. [PubMed: 6223687]
39. Mellon SH, Vaudry H. Biosynthesis of neurosteroids and regulation of their synthesis. *Int Rev Neurobiol* 2001;46(0074–7742):33–78. [PubMed: 11599305]
40. Baulieu EE, Robel P. Dehydroepiandrosterone (DHEA) and dehydroepiandrosterone sulfate (DHEAS) as neuroactive neurosteroids. *Proc Natl Acad Sci U S A* 1998;95(0027–8424 8):4089–4091. [PubMed: 9539693]
41. Stoffel-Wagner B. Neurosteroid biosynthesis in the human brain and its clinical implications. *Ann N Y Acad Sci* 2003;1007(0077–8923):64–78. [PubMed: 14993041]
42. Watzka M, Bidlingmaier F, Schramm J, Klingmuller D, Stoffel-Wagner B. Sex- and age-specific differences in human brain CYP11A1 mRNA expression. *J Neuroendocrinol* 1999;11(12):901–905. [PubMed: 10583724]

43. Lavaque E, Mayen A, Azcoitia I, Tena-Sempere M, Garcia-Segura LM. Sex differences, developmental changes, response to injury and cAMP regulation of the mRNA levels of steroidogenic acute regulatory protein, cytochrome p450scc, and aromatase in the olivocerebellar system. *J Neurobiol* 2006;66(0022–3034 3):308–318. [PubMed: 16329132]
44. Kimonides VG, Khatibi NH, Svendsen CN, Sofroniew MV, Herbert J. Dehydroepiandrosterone (DHEA) and DHEA-sulfate (DHEAS) protect hippocampal neurons against excitatory amino acid-induced neurotoxicity. *Proc Natl Acad Sci U S A* 1998;95(0027–8424 4):1852–1857. [PubMed: 9465106]
45. Mao X, Barger SW. Neuroprotection by dehydroepiandrosterone-sulfate: Role of an NFkappaB-like factor. *Neuroreport* 1998;9(0959–4965 4):759–763. [PubMed: 9559952]
46. Cardounel A, Regelson W, Kalimi M. Dehydroepiandrosterone protects hippocampal neurons against neurotoxin-induced cell death: Mechanism of action. *Proc Soc Exp Biol Med* 1999;222(0037–9727 2):145–149. [PubMed: 10564538]
47. Kaasik A, Kalda A, Jaako K, Zharkovsky A. Dehydroepiandrosterone sulphate prevents oxygen-glucose deprivation-induced injury in cerebellar granule cell culture. *Neuroscience* 2001;102(2):427–432. [PubMed: 11166128]
48. Li Z, Zhou R, Cui S, et al. Dehydroepiandrosterone sulfate prevents ischemia-induced impairment of long-term potentiation in rat hippocampal CA1 by up-regulating tyrosine phosphorylation of NMDA receptor. *Neuropharmacology* 2006;51(0028–3908 5):958–966. [PubMed: 16895729]
49. Weaver CE Jr, Marek P, Park-Chung M, Tam SW, Farb DH. Neuroprotective activity of a new class of steroidal inhibitors of the N-methyl-D-aspartate receptor. *Proc Natl Acad Sci U S A* 1997;94(0027–8424 19):10450–10454. [PubMed: 9294231]
50. Gursoy E, Cardounel A, Kalimi M. Pregnenolone protects mouse hippocampal (HT-22) cells against glutamate and amyloid beta protein toxicity. *Neurochem Res* 2001;26(0364–3190 1):15–21. [PubMed: 11358277]
51. Gibson CL, Murphy SP. Progesterone enhances functional recovery after middle cerebral artery occlusion in male mice. *J Cereb Blood Flow Metab* 2004;24(0271–678 7):805–813. [PubMed: 15241189]
52. Gibson CL, Constantin D, Prior MJ, Bath PM, Murphy SP. Progesterone suppresses the inflammatory response and nitric oxide synthase-2 expression following cerebral ischemia. *Exp Neurol* 2005;193(0014–4886 2):522–530. [PubMed: 15869954]
53. Alkayed NJ, Murphy SJ, Traystman RJ, Hurn PD, Miller VM. Neuroprotective effects of female gonadal steroids in reproductively senescent female rats. *Stroke* 2000;31(0039–2499 1):161–168. [PubMed: 10625733]
54. Murphy SJ, Littleton-Kearney MT, Hurn PD. Progesterone administration during reperfusion, but not preischemia alone, reduces injury in ovariectomized rats. *J Cereb Blood Flow Metab* 2002;22(10):1181–1188. [PubMed: 12368656]
55. Sayeed I, Guo Q, Hoffman SW, Stein DG. Allopregnanolone, a progesterone metabolite, is more effective than progesterone in reducing cortical infarct volume after transient middle cerebral artery occlusion. *Ann Emerg Med* 2006;47(1097–6760 4):381–389. [PubMed: 16546625]
56. Chen Z, Yuhanna IS, Galcheva-Gargova Z, Karas RH, Mendelsohn ME, Shaul PW. Estrogen receptor alpha mediates the nongenomic activation of endothelial nitric oxide synthase by estrogen. *J Clin Invest* 1999;103(0021–9738 3):401–406. [PubMed: 9927501]
57. Kumon Y, Kim SC, Tompkins P, Stevens A, Sakaki S, Loftus CM. Neuroprotective effect of postischemic administration of progesterone in spontaneously hypertensive rats with focal cerebral ischemia. *J Neurosurg* 2000;92(0022–3085 5):848–852. [PubMed: 10794300]
58. Morali G, Letechipia-Vallejo G, Lopez-Loeza E, Montes P, Hernandez-Morales L, Cervantes M. Post-ischemic administration of progesterone in rats exerts neuroprotective effects on the hippocampus. *Neurosci Lett* 2005;382(0304–3940 3):286–290. [PubMed: 15885907]
59. Cervantes M, Gonzalez-Vidal MD, Ruelas R, Escobar A, Morali G. Neuroprotective effects of progesterone on damage elicited by acute global cerebral ischemia in neurons of the caudate nucleus. *Arch Med Res* 2002;33(0188–4409 1):6–14. [PubMed: 11825624]

60. Murphy SJ, Traystman RJ, Hurn PD, Duckles SP. Progesterone exacerbates striatal stroke injury in progesterone-deficient female animals. *Stroke* 2000;31(0039–2499 5):1173–1178. [PubMed: 10797182]
61. Roof RL, Duvdevani R, Stein DG. Gender influences outcome of brain injury: Progesterone plays a protective role. *Brain Res* 1993;607(0006–8993 1–2):333–336. [PubMed: 8481809]
62. Roof RL, Duvdevani R, Braswell L, Stein DG. Progesterone facilitates cognitive recovery and reduces secondary neuronal loss caused by cortical contusion injury in male rats. *Exp Neurol* 1994;129(0014–4886 1):64–69. [PubMed: 7925843]
63. Robertson CL, Puskar A, Hoffman GE, Murphy AZ, Saraswati M, Fiskum G. Physiologic progesterone reduces mitochondrial dysfunction and hippocampal cell loss after traumatic brain injury in female rats. *Exp Neurol* 2006;197(0014–4886 1):235–243. [PubMed: 16259981]
64. Wright DW, Bauer ME, Hoffman SW, Stein DG. Serum progesterone levels correlate with decreased cerebral edema after traumatic brain injury in male rats. *J Neurotrauma* 2001;18(0897–7151 9):901–909. [PubMed: 11565602]
65. Goss CW, Hoffman SW, Stein DG. Behavioral effects and anatomic correlates after brain injury: A progesterone dose-response study. *Pharmacol Biochem Behav* 2003;76(0091–3057 2):231–242. [PubMed: 14592674]
66. Shear DA, Galani R, Hoffman SW, Stein DG. Progesterone protects against necrotic damage and behavioral abnormalities caused by traumatic brain injury. *Exp Neurol* 2002;178(0014–4886 1):59–67. [PubMed: 12460608]
67. Pan DS, Liu WG, Yang XF, Cao F. Inhibitory effect of progesterone on inflammatory factors after experimental traumatic brain injury. *Biomed Environ Sci* 2007;20(0895–3988 5):432–438. [PubMed: 18188998]
68. Chen G, Shi J, Jin W, et al. Progesterone administration modulates TLRs/NF-kappaB signaling pathway in rat brain after cortical contusion. *Ann Clin Lab Sci* 2008;38(0091–7370 1):65–74. [PubMed: 18316784]
69. Wright DW, Kellermann AL, Hertzberg VS, et al. ProTECT: A randomized clinical trial of progesterone for acute traumatic brain injury. *Ann Emerg Med*. 2006;(1097–6760)
70. Xiao G, Wei J, Yan W, Wang W, Lu Z. Improved outcomes from the administration of progesterone for patients with acute severe traumatic brain injury: A randomized controlled trial. *Crit Care* 2008;12(1466–609 2):R61. [PubMed: 18447940]
71. Betz AL, Coester HC. Effect of steroids on edema and sodium uptake of the brain during focal ischemia in rats. *Stroke* 1990;21(0039–2499 8):1199–1204. [PubMed: 2389301]
72. Guo Q, Sayeed I, Baronne LM, Hoffman SW, Guennoun R, Stein DG. Progesterone administration modulates AQP4 expression and edema after traumatic brain injury in male rats. *Exp Neurol* 2006;198(0014–4886 2):469–478. [PubMed: 16445913]
73. Yao XL, Liu J, Lee E, Ling GS, McCabe JT. Progesterone differentially regulates pro- and anti-apoptotic gene expression in cerebral cortex following traumatic brain injury in rats. *J Neurotrauma* 2005;22(0897–7151 6):656–668. [PubMed: 15941375]
74. Djebaili M, Guo Q, Pettus EH, Hoffman SW, Stein DG. The neurosteroids progesterone and allopregnanolone reduce cell death, gliosis, and functional deficits after traumatic brain injury in rats. *J Neurotrauma* 2005;22(0897–7151 1):106–118. [PubMed: 15665606]
75. Cai W, Zhu Y, Furuya K, Li Z, Sokabe M, Chen L. Two different molecular mechanisms underlying progesterone neuroprotection against ischemic brain damage. *Neuropharmacology*. 2008;(0028–3908)
76. Roof RL, Hoffman SW, Stein DG. Progesterone protects against lipid peroxidation following traumatic brain injury in rats. *Mol Chem Neuropathol* 1997;31(1044–7393 1):1–11. [PubMed: 9271001]
77. Miller L, Hunt JS. Regulation of TNF-alpha production in activated mouse macrophages by progesterone. *J Immunol* 1998;160(10):5098–5104. [PubMed: 9590261]
78. Drew PD, Chavis JA. Female sex steroids: Effects upon microglial cell activation. *J Neuroimmunol* 2000;111(1–2):77–85. [PubMed: 11063824]

79. He J, Hoffman SW, Stein DG. Allopregnanolone, a progesterone metabolite, enhances behavioral recovery and decreases neuronal loss after traumatic brain injury. *Restor Neurol Neurosci* 2004;22(0922–6028 1):19–31. [PubMed: 15096691]
80. Pettus EH, Wright DW, Stein DG, Hoffman SW. Progesterone treatment inhibits the inflammatory agents that accompany traumatic brain injury. *Brain Res* 2005;1049(1):112–119. [PubMed: 15932748]10.1016/j.brainres.2005.05.004
81. Chan JR, Phillips LJ 2nd, Glaser M. Glucocorticoids and progestins signal the initiation and enhance the rate of myelin formation. *Proc Natl Acad Sci U S A* 1998;95(18):10459–10464. [PubMed: 9724725]
82. Koenig HL, Schumacher M, Ferzaz B, et al. Progesterone synthesis and myelin formation by schwann cells. *Science* 1995;268(0036–8075 5216):1500–1503. [PubMed: 7770777]
83. Ghomari AM, Ibanez C, El-Etr M, et al. Progesterone and its metabolites increase myelin basic protein expression in organotypic slice cultures of rat cerebellum. *J Neurochem* 2003;86(0022–3042 4):848–859. [PubMed: 12887683]
84. Ghomari AM, Baulieu EE, Schumacher M. Progesterone increases oligodendroglial cell proliferation in rat cerebellar slice cultures. *Neuroscience* 2005;135(0306–4522 1):47–58. [PubMed: 16054770]
85. Ibanez C, Shields SA, El-Etr M, Baulieu EE, Schumacher M, Franklin RJ. Systemic progesterone administration results in a partial reversal of the age-associated decline in CNS remyelination following toxin-induced demyelination in male rats. *Neuropathol Appl Neurobiol* 2004;30(0305–1846 1):80–89. [PubMed: 14720179]
86. Ardeshiri A, Kelley MH, Korner IP, Hurn PD, Herson PS. Mechanism of progesterone neuroprotection of rat cerebellar purkinje cells following oxygen-glucose deprivation. *Eur J Neurosci* 2006;24(0953–816 9):2567–2574. [PubMed: 17100844]
87. Brinton RD, Wang JM. Therapeutic potential of neurogenesis for prevention and recovery from alzheimer's disease: Allopregnanolone as a proof of concept neurogenic agent. *Curr Alzheimer Res* 2006;3(1567–2050 3):185–190. [PubMed: 16842093]
88. Mellon SH, Gong W, Schonemann MD. Endogenous and synthetic neurosteroids in treatment of niemann-pick type C disease. *Brain Res Rev* 2008;57(0165–0173 2):410–420. [PubMed: 17629950]
89. Ciriza I, Azcoitia I, Garcia-Segura LM. Reduced progesterone metabolites protect rat hippocampal neurones from kainic acid excitotoxicity in vivo. *J Neuroendocrinol* 2004;16(0953–8194 1):58–63. [PubMed: 14962077]
90. Djebaili M, Hoffman SW, Stein DG. Allopregnanolone and progesterone decrease cell death and cognitive deficits after a contusion of the rat pre-frontal cortex. *Neuroscience* 2004;123(0306–4522 2):349–359. [PubMed: 14698743]
91. Wang JM, Johnston PB, Ball BG, Brinton RD. The neurosteroid allopregnanolone promotes proliferation of rodent and human neural progenitor cells and regulates cell-cycle gene and protein expression. *J Neurosci* 2005;25(1529–2401 19):4706–4718. [PubMed: 15888646]
92. Xilouri M, Avlonitis N, Calogeropoulou T, Papazafiri P. Neuroprotective effects of steroid analogues on P19-N neurons. *Neurochem Int* 2007;50(0197–0186 4):660–670. [PubMed: 17316905]
93. Charalampopoulos I, Tsatsanis C, Dermizaki E, et al. Dehydroepiandrosterone and allopregnanolone protect sympathoadrenal medulla cells against apoptosis via antiapoptotic bcl-2 proteins. *Proc Natl Acad Sci U S A* 2004;101(21):8209–8214. [PubMed: 15148390]10.1073/pnas.0306631101
94. Charalampopoulos I, Alexaki VI, Tsatsanis C, et al. Neurosteroids as endogenous inhibitors of neuronal cell apoptosis in aging. *Ann N Y Acad Sci* 2006;1088:139–152. [PubMed: 17192562] 10.1196/annals.1366.003
95. Belelli D, Lambert JJ. Neurosteroids: Endogenous regulators of the GABA(A) receptor. *Nat Rev Neurosci* 2005;6(1471–003 7):565–575. [PubMed: 15959466]
96. Carswell HV, Dominiczak AF, Macrae IM. Estrogen status affects sensitivity to focal cerebral ischemia in stroke-prone spontaneously hypertensive rats. *Am J Physiol Heart Circ Physiol* 2000;278(0363–6135 1):H290–H294. [PubMed: 10644611]
97. Hall ED, Pazara KE, Linseman KL. Sex differences in postischemic neuronal necrosis in gerbils. *J Cereb Blood Flow Metab* 1991;11(2):292–298. [PubMed: 1997500]

98. Hurn PD, Macrae IM. Estrogen as a neuroprotectant in stroke. *J Cereb Blood Flow Metab* 2000;20(4):631–652. [PubMed: 10779008]10.1097/00004647-200004000-00001
99. Toung TJ, Traystman RJ, Hurn PD. Estrogen-mediated neuroprotection after experimental stroke in male rats. *Stroke* 1998;29(8):1666–1670. [PubMed: 9707210]
100. Hawk T, Zhang YQ, Rajakumar G, Day AL, Simpkins JW. Testosterone increases and estradiol decreases middle cerebral artery occlusion lesion size in male rats. *Brain Res* 1998;796(1–2):296–298. [PubMed: 9689481]
101. Jover T, Tanaka H, Calderone A, et al. Estrogen protects against global ischemia-induced neuronal death and prevents activation of apoptotic signaling cascades in the hippocampal CA1. *J Neurosci* 2002;22(1529–2401 6):2115–2124. [PubMed: 11896151]
102. Gulinello M, Lebesgue D, Jover-Mengual T, Zukin RS, Etgen AM. Acute and chronic estradiol treatments reduce memory deficits induced by transient global ischemia in female rats. *Horm Behav* 2006;49(2):246–260. [PubMed: 16125703]10.1016/j.yhbeh.2005.07.010
103. Bingham D, Macrae IM, Carswell HV. Detrimental effects of 17beta-oestradiol after permanent middle cerebral artery occlusion. *J Cereb Blood Flow Metab* 2005;25(3):414–420. [PubMed: 15647739]10.1038/sj.jcbfm.9600031
104. Vergouwen MD, Anderson RE, Meyer FB. Gender differences and the effects of synthetic exogenous and non-synthetic estrogens in focal cerebral ischemia. *Brain Res* 2000;878(0006–8993 1–2):88–97. [PubMed: 10996139]
105. Gordon CP, Keller PA. Control of hepatitis C: A medicinal chemistry perspective. *J Med Chem* 2005;48(0022–2623 1):1–20. [PubMed: 15633995]
106. Sohrabji F, Bake S. Age-related changes in neuroprotection: Is estrogen pro-inflammatory for the reproductive senescent brain? *Endocrine* 2006;29(2):191–197. [PubMed: 16785595]10.1385/ENDO:29:2:191
107. Suzuki S, Brown CM, Dela Cruz CD, Yang E, Bridwell DA, Wise PM. Timing of estrogen therapy after ovariectomy dictates the efficacy of its neuroprotective and antiinflammatory actions. *Proc Natl Acad Sci U S A* 2007;104(14):6013–6018. [PubMed: 17389368]
108. Rusa R, Alkayed NJ, Crain BJ, et al. 17beta-estradiol reduces stroke injury in estrogen-deficient female animals. *Stroke* 1999;30(8):1665–1670. [PubMed: 10436119]
109. Dubal DB, Kashon ML, Pettigrew LC, et al. Estradiol protects against ischemic injury. *J Cereb Blood Flow Metab* 1998;18(0271–678 11):1253–1258. [PubMed: 9809515]
110. Liu R, Wen Y, Perez E, et al. 17beta-estradiol attenuates blood-brain barrier disruption induced by cerebral ischemia-reperfusion injury in female rats. *Brain Res* 2005;1060(1–2):55–61. [PubMed: 16212944]10.1016/j.brainres.2005.08.048
111. O'Donnell ME, Lam TI, Tran LQ, Foroutan S, Anderson SE. Estradiol reduces activity of the blood-brain barrier na-K-cl cotransporter and decreases edema formation in permanent middle cerebral artery occlusion. *J Cereb Blood Flow Metab* 2006;26(10):1234–1249. [PubMed: 16421506] 10.1038/sj.jcbfm.9600278
112. Pelligrino DA, Santizo R, Baughman VL, Wang Q. Cerebral vasodilating capacity during forebrain ischemia: Effects of chronic estrogen depletion and repletion and the role of neuronal nitric oxide synthase. *Neuroreport* 1998;9(0959–4965 14):3285–3291. [PubMed: 9831465]
113. Hurn PD, Littleton-Kearney MT, Kirsch JR, Dharmarajan AM, Traystman RJ. Postischemic cerebral blood flow recovery in the female: Effect of 17 beta-estradiol. *J Cereb Blood Flow Metab* 1995;15(4):666–672. [PubMed: 7790416]
114. Mori M, Tsukahara F, Yoshioka T, Irie K, Ohta H. Suppression by 17beta-estradiol of monocyte adhesion to vascular endothelial cells is mediated by estrogen receptors. *Life Sci* 2004;75(0024–3205 5):599–609. [PubMed: 15158369]
115. Wen Y, Perez EJ, Green PS, Sarkar SN, Simpkins JW. nNOS is involved in estrogen mediated neuroprotection in neuroblastoma cells. *Neuroreport* 2004;15(0959–4965 9):1515–1518. [PubMed: 15194886]
116. Alkayed NJ, Goto S, Sugo N, et al. Estrogen and bcl-2: Gene induction and effect of transgene in experimental stroke. *J Neurosci* 2001;21(19):7543–7550. [PubMed: 11567044]

117. Xu Y, Zhang W, Klaus J, et al. Role of cocaine- and amphetamine-regulated transcript in estradiol-mediated neuroprotection. *Proc Natl Acad Sci U S A* 2006;103(39):14489–14494. [PubMed: 16971488]10.1073/pnas.0602932103
118. Keller JN, Germeyer A, Begley JG, Mattson MP. 17Beta-estradiol attenuates oxidative impairment of synaptic Na⁺/K⁺-ATPase activity, glucose transport, and glutamate transport induced by amyloid beta-peptide and iron. *J Neurosci Res* 1997;50(0360–4012 4):522–530. [PubMed: 9404714]
119. Vedder H, Anthes N, Stumm G, Wurz C, Behl C, Krieg JC. Estrogen hormones reduce lipid peroxidation in cells and tissues of the central nervous system. *J Neurochem* 1999;72(0022–3042 6):2531–2538. [PubMed: 10349864]
120. Behl C, Manthey D. Neuroprotective activities of estrogen: An update. *J Neurocytol* 2000;29(0300–4864 5–6):351–358. [PubMed: 11424951]
121. Weaver CE Jr, Marek P, Park-Chung M, Tam SW, Farb DH. Neuroprotective activity of a new class of steroidal inhibitors of the N-methyl-D-aspartate receptor. *Proc Natl Acad Sci U S A* 1997;94(0027–8424 19):10450–10454. [PubMed: 9294231]
122. Shughrue PJ, Bushnell CD, Dorsa DM. Estrogen receptor messenger ribonucleic acid in female rat brain during the estrous cycle: A comparison with ovariectomized females and intact males. *Endocrinology* 1992;131(1):381–388. [PubMed: 1612018]
123. Mor G, Nilsen J, Horvath T, et al. Estrogen and microglia: A regulatory system that affects the brain. *J Neurobiol* 1999;40(0022–3034 4):484–496. [PubMed: 10453051]
124. Azcoitia I, Sierra A, Garcia-Segura LM. Localization of estrogen receptor beta-immunoreactivity in astrocytes of the adult rat brain. *Glia* 1999;26(0894–1491 3):260–267. [PubMed: 10340766]
125. Sawada M, Alkayed NJ, Goto S, et al. Estrogen receptor antagonist ICI182,780 exacerbates ischemic injury in female mouse. *J Cereb Blood Flow Metab* 2000;20(1):112–118. [PubMed: 10616799] 10.1097/00004647-200001000-00015
126. Sampei K, Goto S, Alkayed NJ, et al. Stroke in estrogen receptor-alpha-deficient mice. *Stroke* 2000;31(3):738–43. [PubMed: 10700513]discussion 744
127. Dubal DB, Zhu H, Yu J, et al. Estrogen receptor alpha, not beta, is a critical link in estradiol-mediated protection against brain injury. *Proc Natl Acad Sci U S A* 2001;98(0027–8424 4):1952–1957. [PubMed: 11172057]
128. Jeppesen LL, Jorgensen HS, Nakayama H, Raaschou HO, Olsen TS, Winther K. Decreased serum testosterone in men with acute ischemic stroke. *Arterioscler Thromb Vasc Biol* 1996;16(1079–5642 6):749–754. [PubMed: 8640402]
129. Yang SH, Perez E, Cutright J, et al. Testosterone increases neurotoxicity of glutamate in vitro and ischemia-reperfusion injury in an animal model. *J Appl Physiol* 2002;92(8750–7587 1):195–201. [PubMed: 11744660]
130. Cheng J, Alkayed NJ, Hurn PD. Deleterious effects of dihydrotestosterone on cerebral ischemic injury. *J Cereb Blood Flow Metab* 2007;27(9):1553–1562. [PubMed: 17311081]10.1038/sj.jcbfm.9600457
131. Yang SH, Liu R, Wu SS, Simpkins JW. The use of estrogens and related compounds in the treatment of damage from cerebral ischemia. *Ann N Y Acad Sci* 2003;1007(0077–8923):101–107. [PubMed: 14993044]
132. Jones KJ. Gonadal steroids as promoting factors in axonal regeneration. *Brain Res Bull* 1993;30(3–4):491–498. [PubMed: 8457899]
133. Kujawa KA, Jacob JM, Jones KJ. Testosterone regulation of the regenerative properties of injured rat sciatic motor neurons. *J Neurosci Res* 1993;35(3):268–273. [PubMed: 8350388]10.1002/jnr.490350306
134. Tanzer L, Jones KJ. Gonadal steroid regulation of hamster facial nerve regeneration: Effects of dihydrotestosterone and estradiol. *Exp Neurol* 1997;146(1):258–264. [PubMed: 9225759]10.1006/exnr.1997.6529
135. Chen R, Cohen LG, Hallett M. Nervous system reorganization following injury. *Neuroscience* 2002;111(4):761–773. [PubMed: 12031403]

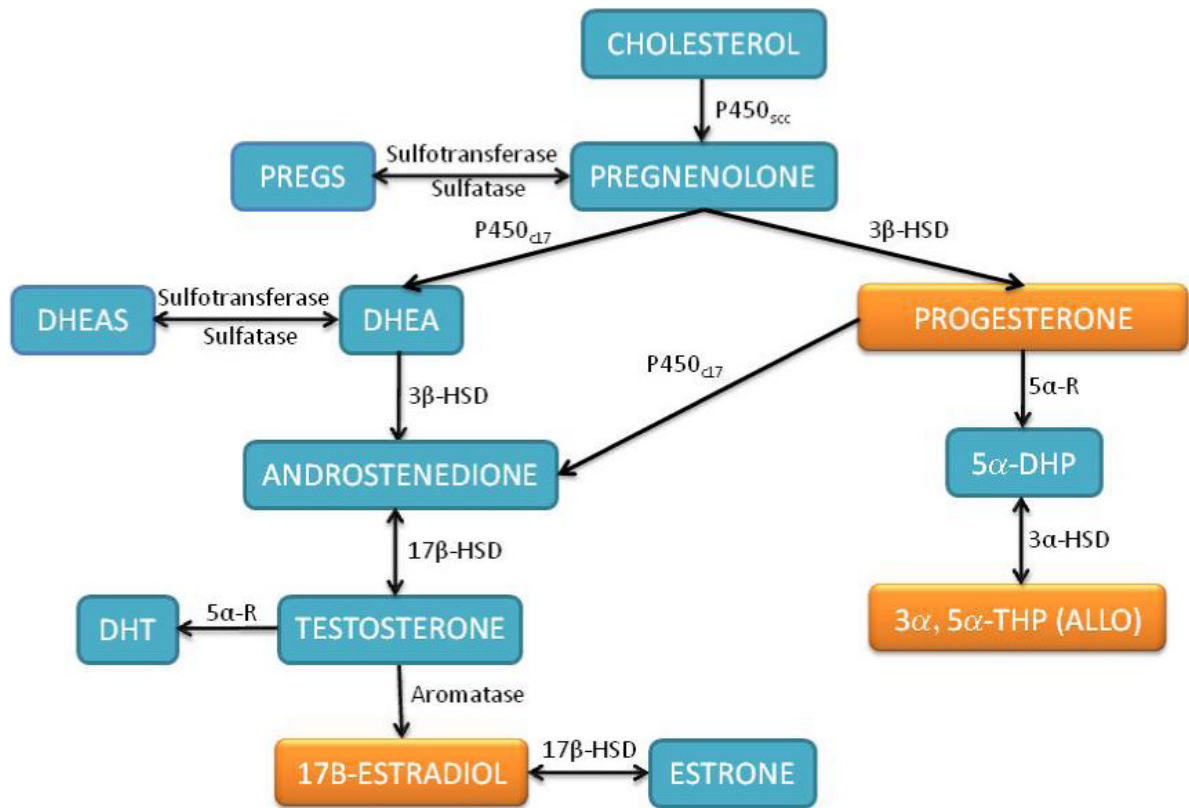


Figure 1. Overview of Neurosteroid Synthesis

Cholesterol is converted into pregnenolone within the mitochondria and then further converted into DHEA via P450_{c17} or progesterone by 3β-HSD. DHEA is converted into testosterone via the intermediate androstenedione. Progesterone can also be converted to testosterone by P450_{c17}. Testosterone is converted to its more potent analog DHT by 5α-R or converted into 17β-estradiol by the enzyme Aromatase. Progesterone is converted to its highly active metabolites 5α-DHP and 3α,5α-THP (ALLO) by consecutive reductions by 5α-R and then 3α-HSD. Both pregnenolone and DHEA can be interconverted to sulfated esters by sulfotransferase and sulfatase enzymes. P450_{scc}, mitochondrial cholesterol side-chain cleavage enzyme; P450_{c17}, microsomal 17 hydroxylase; 3β-HSD, 3β-hydroxysteroid dehydrogenase; 5α-R, 5α-reductase; 5α-HSD, 5α-hydroxysteroid dehydrogenase; 17β-HSD, 17β-hydroxysteroid dehydrogenase; Aromatase, P450-aromatase; DHT, dihydrotestosterone; 3α,5α-THP, Allopregnanolone.