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## Long-lasting dysregulation of gene expression in corticostriatal circuits after repeated cocaine treatment in adult rats: Effects on *zif 268* and *homer 1a*

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### Abstract

Human imaging studies show that psychostimulants such as cocaine produce functional changes in several areas of cortex and striatum. These may reflect neuronal changes related to addiction. We employed gene markers (*zif 268*, *homer 1a*) that offer a high anatomical resolution to map cocaine-induced changes in 22 cortical areas and 23 functionally related striatal sectors, in order to determine the corticostriatal circuits altered by repeated cocaine exposure (25 mg/kg, 5 days). Effects were investigated 1 day and 21 days after repeated treatment to assess their longevity. Repeated cocaine treatment increased basal expression of *zif 268* predominantly in sensorimotor areas of the cortex. This effect endured for 3 weeks in some areas. These changes were accompanied by attenuated gene induction by a cocaine challenge. In the insular cortex, the cocaine challenge produced a decrease in *zif 268* expression after the 21-day, but not 1-day, withdrawal period. In the striatum, cocaine also affected mostly sensorimotor sectors. Repeated cocaine resulted in blunted inducibility of both *zif 268* and *homer 1a*, changes that were still very robust 3 weeks later. Thus, our findings demonstrate that cocaine produces robust and long-lasting changes in gene regulation predominantly in sensorimotor corticostriatal circuits. These neuronal changes were associated with behavioral stereotypies, which are thought to reflect dysfunction in sensorimotor corticostriatal circuits. Future studies will have to elucidate the role of such neuronal changes in psychostimulant addiction.

### Keywords

cortex; striatum; psychostimulant; immediate-early gene; dopamine

### INTRODUCTION

Drug addiction is associated with neuronal changes in specific parts of the brain. Imaging studies demonstrated that repeated exposure to psychostimulants produces functional changes in brain regions such as the cerebral cortex and the basal ganglia (e.g., London *et al.*, 1990; Breiter *et al.*, 1997; Beveridge *et al.*, 2006; Porrino *et al.*, 2007). Interactions between the cortex and the basal ganglia are critical for the organization of normal motivated behavior (Albin *et al.*, 1989; DeLong, 1990; Robbins *et al.*, 1998). These interactions are mediated by distinct anatomical loops that arise in all parts of the cortex, project in a topographical manner to

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specific functional domains of the striatum, and from there, via basal ganglia output nuclei and thalamus, back to the cortex (Alexander *et al.*, 1986; 1990; Groenewegen *et al.*, 1990). Identification of the functional loops and their neuronal processes that are altered by psychostimulants will further our understanding of the addiction process and guide treatment approaches.

Various studies identified psychostimulant-induced molecular changes in the basal ganglia, especially the striatum (e.g., Harlan & Garcia, 1998; Berke & Hyman, 2000; Yano & Steiner, 2007). Most previous work focused on limbic areas, which mediate motivational processes (Pierce & Kalivas, 1997) and are thus considered of central importance in early addiction stages. However, imaging studies indicate that, as the disease progresses, associational and sensorimotor domains of the striatum are increasingly affected (Porrino *et al.*, 2007). These domains are implicated in habitual and compulsive aspects of drug taking (Berke & Hyman, 2000; Everitt & Robbins, 2005) and indeed display particularly robust psychostimulant-induced molecular changes (e.g., Steiner & Gerfen, 1993; Badiani *et al.*, 1998; Willuhn *et al.*, 2003; Yano & Steiner, 2005b).

Much less is known regarding molecular changes in the cortex after psychostimulant exposure. Several studies documented effects in limbic-related prefrontal areas (e.g., Freeman *et al.*, 2002; Black *et al.*, 2006), but there is evidence for changes in several other cortical areas as well. For example, we recently showed that the psychostimulant methylphenidate induces immediate-early genes (IEGs) such as *zif 268* and *homer 1a* in a widespread but regionally selective manner, including many sensory and motor cortical areas (Yano & Steiner, 2005a; Cotterly *et al.*, 2007). Cocaine and amphetamine have also been found to induce IEGs in the sensorimotor cortex (e.g., Curran *et al.*, 1996; Badiani *et al.*, 1998), but the regional specificity of these effects remains unclear.

In the present study, we mapped changes in IEG expression induced by acute and repeated cocaine treatment throughout the cortex (22 cortical areas on 4 rostrocaudal levels) and compared these effects with the distribution of striatal gene regulation (23 striatal sectors) to determine the corticostriatal circuits affected. For comparison with our previous studies (Yano & Steiner, 2005a; Cotterly *et al.*, 2007), we assessed the IEGs *zif 268* and *homer 1a*. These genes serve as functional markers (Sharp *et al.*, 1993; Chaudhuri, 1997), but are also of interest because of their direct involvement in neuroplasticity; *zif 268* encodes a transcription factor (Knapska & Kaczmarek, 2004) and *homer 1a* a synaptic plasticity regulator (Xiao *et al.*, 2000).

## MATERIALS AND METHODS

### Subjects

Male Sprague–Dawley rats (175–200 g at the beginning of the experiment; Harlan, Madison, WI, USA) were housed 2 per cage under standard laboratory conditions (12:12-hr light/dark cycle; lights on at 0700 h) with food and water available *ad libitum*. The experiments were performed between 1300 and 1700 h. All procedures met the NIH guidelines for the care and use of laboratory animals and were approved by the Rosalind Franklin University Animal Care and Use Committee.

### Drug treatments

Before the start of the pharmacological treatment, rats were repeatedly handled on three days. The animals then received an injection of vehicle or cocaine (cocaine hydrochloride; Sigma, St. Louis, MO, USA; 25 mg/kg, in 0.02% ascorbic acid, i. p.) once daily for 5 days, in their home cage. On days 6 (withdrawal day 1) or 26 (withdrawal day 21), the animals were

transferred in their home cage to an adjacent room and, 3 hours later, received a challenge injection of cocaine (25 mg/kg) or vehicle (groups VV, CV, VC, CC; n=7–9). After the injection, the rat was placed in the arena of an activity monitoring system (43 × 43 cm; Truscan, Coulbourn Instruments, Allentown, PA, USA), and locomotion (ambulatory distance) and stereotypy (“stereotypy 2”) counts were measured for 30 min. These “stereotypy” counts reflect local, repetitive movements (e.g., head bobbing, focused sniffing). Animals of the 21-day withdrawal groups were handled every third day between repeated drug treatment and challenge injection.

### Tissue preparation and in situ hybridization histochemistry

The rats were killed with CO<sub>2</sub> 30 min after the challenge injection. The brain was rapidly removed, frozen in isopentane cooled on dry ice and then stored at –30 °C until cryostat sectioning. Coronal sections (12 μm) were thaw-mounted onto glass slides (Superfrost/Plus, Daigger, Wheeling, IL, USA), dried on a slide warmer and stored at –30 °C. In preparation for the in situ hybridization histochemistry, the sections were fixed in 4% paraformaldehyde/0.9% saline for 10 min at room temperature, incubated in a fresh solution of 0.25% acetic anhydride in 0.1 M triethanolamine/0.9% saline (pH 8.0) for 10 min, dehydrated, defatted for 2 × 5 min in chloroform, rehydrated, and air-dried. The slides were then stored at –30 °C until hybridization.

Oligonucleotide probes (48-mers; Invitrogen, Rockville, MD, USA) were labeled with [<sup>35</sup>S]-dATP as described earlier (Steiner & Kitai, 2000). The probes had the following sequence: *zif* 268, complementary to bases 352–399, GenBank accession number M18416; *homer 1a*, bases 163–210, NM\_031707. The latter targets the beginning of the *homer 1* transcript, which yields a more robust signal at this timepoint (Bottai *et al.*, 2002). Previous findings indicate that the cocaine-induced signal measured with this probe reflects *homer 1a* expression (Brakeman *et al.*, 1997; Bottai *et al.*, 2002; Zhang *et al.*, 2007).

One hundred μl of hybridization buffer containing labeled probe (~3 × 10<sup>6</sup> cpm) was added to each slide. The sections were coverslipped and incubated at 37 °C overnight. After incubation, the slides were first rinsed in four washes of 1X saline citrate (150 mM sodium chloride, 15 mM sodium citrate), and then washed 3 times 20 min each in 2X saline citrate/50% formamide at 40 °C, followed by 2 washes of 30 min each in 1X saline citrate at room temperature. After a brief water rinse, the sections were air-dried and then apposed to X-ray film (BioMax MR-2, Kodak) for 4–10 days.

### Analysis of autoradiograms

Gene expression in the cortex was assessed in sections from 4 rostrocaudal levels (see Fig. 4): frontal, approximately at +2.7 mm relative to bregma (Paxinos & Watson, 1998); rostral, +1.6; middle, +0.4; caudal, –0.8 (Yano & Steiner, 2005a). Levels of mRNA were measured in a total of 22 cortical regions (from medial to lateral; Paxinos & Watson, 1998): cingulate, medial agranular (M2), motor (M1), somatosensory and insular cortex on frontal to caudal levels, and infralimbic, prelimbic and lateral orbital cortex on the frontal level. Striatal gene expression was determined on rostral, middle and caudal levels in a total of 23 sectors mostly defined by their predominant cortical inputs (Fig. 8, Table 1; Willuhn *et al.*, 2003). Eighteen of these sectors represented the caudate-putamen and 5 the nucleus accumbens (medial and lateral core, medial, ventral and lateral shell) (Yano & Steiner, 2005a).

Hybridization signals on film autoradiograms were measured by densitometry (NIH Image; Wayne Rasband, NIMH, Bethesda, MD, USA). The films were captured using a light table (Northern Light, Imaging Research, St. Catharines, Ontario, Canada) and a Sony CCD camera (Imaging Research). The “mean density” value of a region of interest was measured by placing

a template over the captured image. Mean densities were corrected for background by subtracting mean density values measured over white matter (corpus callosum). Values from corresponding regions in the two hemispheres were then averaged.

Treatment effects were determined by two-factor ANOVA, followed by Newman-Keuls *post hoc* tests to describe differences between individual groups (Statistica, StatSoft, Tulsa, OK, USA). For illustrations of topographies (maps) and correlation analyses, the change in gene expression in a given region was expressed as the percentage of the maximal change (% of max.) observed for a particular probe. The illustrations of film autoradiograms displayed in Figure 2 are computer-generated images, and are contrast-enhanced where necessary. Maximal hybridization signal is black.

## RESULTS

### Behavioral effects

Figure 1 depicts behavioral effects of the cocaine challenge injection (or vehicle) given 1 or 21 days after repeated cocaine or vehicle treatment. Administration of cocaine increased ambulation counts in vehicle-pretreated (VC vs. VV,  $P < 0.001$ ) and in cocaine-pretreated animals (CC vs. CV,  $P < 0.01$ ), an effect that was similar in groups tested on withdrawal days 1 or 21 (Fig. 1A). The cocaine challenge produced significant increases in stereotypy counts in cocaine-pretreated animals (withdrawal day 1: CC vs. CV,  $P < 0.01$ ; CC vs. VC;  $P < 0.05$ ), but this effect was less robust (CC vs. CV,  $P < 0.05$ ) 21 days after repeated cocaine treatment.

### Cocaine effects on gene expression in the cortex

**Zif 268 expression**—Gene expression in the cortex was assessed in a total of 22 areas on four rostrocaudal levels. Absolute density values and challenge-induced values expressed in percentage of basal expression (i.e., in respective vehicle controls) are presented.

One day after repeated cocaine treatment, “basal” *zif 268* expression (CV vs. VV) was significantly increased when measured across all cortical areas (total cortex) on rostral ( $P < 0.01$ ), middle ( $P < 0.01$ ) and caudal ( $P < 0.05$ ) levels (Figs. 2A, 3A). Regional analysis (Figs. 4, 6) showed that this effect was most pronounced in the somatosensory cortex on all 4 levels (frontal,  $P < 0.05$ ; rostral,  $P < 0.001$ ; middle,  $P < 0.001$ ; caudal,  $P < 0.01$ ). Weaker increases were seen in the motor (M1) (rostral, caudal,  $P < 0.01$ ), medial agranular (M2) (middle, caudal,  $P < 0.05$ ), and insular (middle,  $P < 0.05$ ) cortex. After the 21-day withdrawal period, a tendency for increased basal *zif 268* expression was still widespread (Fig. 3A), but increased expression was statistically significant only in the somatosensory cortex on the frontal ( $P < 0.01$ ) and middle ( $P < 0.05$ ) levels and in the insular cortex on the middle level ( $P < 0.01$ ) (Figs. 4, 6).

Acute cocaine administration produced increased cortical *zif 268* expression (VC vs. VV), but only when given 1 day after the repeated vehicle treatment. In this group, *zif 268* expression across the total cortex was significantly elevated on the rostral and middle levels (Figs. 2, 3A); the greatest increases were present in the somatosensory cortex (rostral, middle,  $P < 0.01$ ; caudal,  $P < 0.05$ ), followed by the motor cortex (rostral,  $P < 0.001$ ; caudal,  $P < 0.05$ ), the medial agranular cortex (middle,  $P < 0.01$ ) and the cingulate cortex (caudal,  $P < 0.05$ ) (Figs. 5, 6). In contrast, no statistically significant changes in cortical *zif 268* expression were observed when the acute cocaine challenge was administered 21 days after repeated vehicle treatment (Fig. 3A, 5, 6).

The effects of repeated cocaine treatment on cortical *zif 268* expression were also dependent on the withdrawal period. Animals that received the cocaine challenge injection 1 day after repeated cocaine treatment showed no significant changes in any cortical area (CC vs. CV) (Figs. 2A, 3A, 5, 6). When the *zif 268* responses to acute (VC) and repeated (CC) cocaine

treatments were expressed relative to their baseline controls (VV and CV, respectively), repeatedly treated animals (CC) displayed significantly reduced *zif 268* responses compared with acute responses (VC) on all 4 rostrocaudal levels (total cortex, Fig. 3A). Significant decreases were found in 9 of the 22 cortical areas and were again most robust in the somatosensory (rostral, middle,  $P < 0.01$ ) and motor cortex (rostral,  $P < 0.01$ ) (Figs. 5, 6). When the cocaine challenge was given after the 21-day withdrawal period, 21 of the 22 cortical areas showed a tendency for lower *zif 268* mRNA levels than baseline (CC vs. CV) (Fig. 3A), an effect that was statistically significant in the somatosensory cortex on the frontal and rostral levels ( $P < 0.05$ , data not shown) and in the somatosensory cortex ( $P < 0.05$ ) and insular cortex ( $P < 0.01$ ) on the middle level (Fig. 6). When expressed relative to baseline, these animals (CC) displayed a significantly lower *zif 268* response than acute animals (VC) in the middle insular cortex ( $P < 0.05$ ) (Fig. 6).

**Homer 1a expression**—Overall, these cocaine treatments had only modest effects on the expression of *homer 1a* in the cortex (Fig. 2B, 3B). Effects on basal *homer 1a* expression were similar in direction and location to effects on basal *zif 268* expression. One day after repeated cocaine treatment, basal *homer 1a* expression was significantly increased (CV vs. VV) in the rostral somatosensory cortex ( $P < 0.05$ , data not shown), the same area that showed the most robust increase in *zif 268* expression (see above). After the 21-day withdrawal period, significant increases in basal *homer 1a* expression were found in the somatosensory and motor cortex on the middle level ( $P < 0.05$ ) (data not shown). Neither acute cocaine (VC vs. VV) nor a challenge after repeated cocaine treatment (CC vs. CV) produced consistent changes in *homer 1a* expression in these cortical areas, either 1 or 21 days after the repeated treatment (data not shown).

### Cocaine effects on gene expression in the striatum

**Zif 268 expression**—Gene expression in the striatum was measured in a total of 23 sectors on three rostrocaudal levels. While basal *zif 268* expression in the striatum tended to be increased in a majority of sectors, this effect reached statistical significance only in the dorsolateral (sensorimotor) sector on the caudal level (CV vs. VV,  $P < 0.05$ , data not shown), one day after repeated cocaine treatment.

When given 1 day after the repeated vehicle treatment, acute cocaine (VC) induced highly significant ( $P < 0.001$ ) increases in *zif 268* expression (VC vs. VV) in all but 2 of the 18 sectors representing the caudate-putamen (Figs. 2A, 7A, 8A), the exceptions being the ventral sectors on the middle ( $P < 0.01$ ) and caudal ( $P > 0.05$ ) levels (Fig. 8A). While these effects were present on all rostrocaudal levels, they tended to increase from rostral towards caudal and from medial towards lateral, and were most robust in the caudal sensorimotor striatum (Fig. 8A). In the 5 sectors representing the nucleus accumbens (rostral), a significant increase in *zif 268* expression was only seen in the lateral shell ( $P < 0.01$ ) (Fig. 8A). The acute cocaine challenge administered 21 days after repeated vehicle treatment (VC) produced *zif 268* induction with principally the same regional distribution (16/18 sectors,  $P < 0.001$ ; middle ventral,  $P < 0.05$ ; caudal ventral,  $P > 0.05$ ; lateral shell,  $P < 0.05$ ) (Fig. 8A), but the magnitude of the *zif 268* response tended to be somewhat smaller (Figs. 7A, 8A).

After repeated cocaine treatment, significant *zif 268* induction by the cocaine challenge (CC vs. CV,  $P < 0.05$ ) occurred in 14 of the 23 striatal sectors (both 1-day and 21-day withdrawal) (Figs. 2A, 7A, 8A). While the overall topography of induction was similar to that after acute cocaine (CC vs. VC, 1-day withdrawal,  $r = 0.843$ ,  $P < 0.001$ ; 21-day withdrawal,  $r = 0.919$ ,  $P < 0.001$ ), “chronic” *zif 268* induction was attenuated (blunted) throughout the striatum (Figs. 7A, 8A). Thus, a significant decrease in induction (CC vs. VC,  $P < 0.05$ ) was seen in 17 (1-day withdrawal) and 13 sectors (21-day withdrawal), including the nucleus accumbens lateral shell

(1-day withdrawal,  $P < 0.01$ ). This blunting of chronic *zif 268* induction was directly related to acute induction; the more robust the acute induction in a given sector, the more reduced the chronic response in that sector (CC-VC vs. VC, 1-day withdrawal,  $r = -0.896$ ,  $P < 0.001$ ; 21-day withdrawal,  $r = -0.777$ ,  $P < 0.001$ ; Fig. 9A).

**Homer 1a expression**—Overall, the topography of changes in cocaine-induced *homer 1a* expression in the striatum was similar to that for *zif 268* expression. Indeed, regional effects on *homer 1a* and *zif 268* expression were highly correlated for both induction after acute (VC vs. VC, 1-day withdrawal,  $r = 0.907$ ,  $P < 0.001$ ; 21-day withdrawal,  $r = 0.910$ ,  $P < 0.001$ ) and repeated treatments (CC vs. CC, 1-day withdrawal,  $r = 0.937$ ,  $P < 0.001$ ; 21-day withdrawal,  $r = 0.828$ ,  $P < 0.001$ ).

One day after the repeated vehicle treatment, acute cocaine (VC) induced highly significant ( $P < 0.001$ ) increases in *homer 1a* expression (VC vs. VV) in 13/18 sectors of the caudate-putamen (Figs. 2B, 7B, 8B). Induction was less robust or absent in the 3 ventral sectors (rostral,  $P < 0.01$ ; middle, caudal,  $P > 0.05$ ), as well as in the central sectors on the caudal level ( $P < 0.05$ ) (Fig. 8B). Again, these effects tended to increase towards lateral and caudal, and were most pronounced in the caudal sensorimotor sectors. In the nucleus accumbens, the only effect seen was a significant increase in expression in the lateral shell ( $P < 0.01$ ). When given 21 days after repeated vehicle treatment, the acute cocaine challenge (VC) produced *homer 1a* induction with a similar regional distribution (10/18 sectors,  $P < 0.001$ ; rostral medial,  $P > 0.05$ ; middle medial,  $P < 0.01$ ; caudal medial,  $P > 0.05$ ; rostral, middle, caudal ventral,  $P > 0.05$ ; caudal central,  $P < 0.05$ ; no effects in nucleus accumbens) (Fig. 8B), but the magnitude of induction again tended to be somewhat smaller in some sectors (Fig. 8B).

After repeated cocaine treatment, the cocaine challenge induced significant *homer 1a* expression (CC vs. CV,  $P < 0.05$ ) in 9 and 10 of the 23 striatal sectors (Figs. 2B, 7B, 8B). While the overall topography was also similar to that after acute cocaine (CC vs. VC, 1-day withdrawal,  $r = 0.798$ ,  $P < 0.001$ ; 21-day withdrawal,  $r = 0.919$ ,  $P < 0.001$ ), “chronic” *homer 1a* induction was blunted on all 3 rostrocaudal levels (Figs. 7B, 8B). Thus, significantly smaller induction (CC vs. VC,  $P < 0.05$ ) was seen in 13 (1-day withdrawal) and 8 sectors (21-day withdrawal), including the nucleus accumbens lateral shell (1-day withdrawal). Again, this blunting of chronic *homer 1a* induction was related to acute induction; the more robust the acute induction, the more blunted the chronic induction (CC-VC vs. VC, 1-day withdrawal,  $r = -0.811$ ,  $P < 0.001$ ; 21-day withdrawal,  $r = -0.690$ ,  $P < 0.001$ ; Fig. 9B).

## DISCUSSION

In the present study, we used two functional markers, the IEGs *zif 268* and *homer 1a*, to map neuronal changes in cortex and striatum after acute and repeated cocaine treatment. Moreover, by investigating these effects 1 versus 21 days after the repeated treatment, we determined whether such changes endure for a prolonged time period after cocaine exposure. Our findings show that repeated cocaine treatment produces dysregulated gene expression in specific functional domains of the cortex and striatum and that some of these effects last for at least 3 weeks. Our most important findings are summarized as follows. (1) Acute cocaine administration induced *zif 268* expression predominantly in somatosensory and motor areas of the cortex. (2) Repeated cocaine treatment resulted in increased basal expression of *zif 268* in the cortex on all rostrocaudal levels. This effect was most robust in sensorimotor areas one day after the treatment, but was still present 3 weeks later in some areas. (3) These increases in basal expression were accompanied by attenuated *zif 268* inducibility by a cocaine challenge one day after the repeated treatment in all affected cortical regions. (4) After the 3-week withdrawal period, the cocaine challenge produced a decrease in *zif 268* expression in the insular cortex. (5) Similar to the cortical effects, in the striatum, acute induction of *zif 268* was

also most robust in sensorimotor sectors. (6) After repeated cocaine treatment, *zif 268* inducibility was blunted throughout the striatum, an effect that was proportional to the acute induction and lasted for at least 3 weeks. (7) Interestingly, unlike in the striatum, the acute *zif 268* response in the cortex was dependent on the treatment context (handling history); acute cocaine given 1 day, but not 21 days, after repeated vehicle injections significantly enhanced *zif 268* expression. (8) While cocaine had minimal or no effects on *homer 1a* expression in the cortex, cocaine-induced changes in *homer 1a* expression in the striatum were robust and displayed principally identical regional and temporal dynamics as those for *zif 268*. This study is the first to demonstrate long-lasting changes in gene regulation mostly in sensorimotor corticostriatal circuits after repeated cocaine treatment.

### **Repeated cocaine treatment produces long-lasting increases in basal *zif 268* expression, but attenuates challenge-induced *zif 268* responses, in the cortex**

IEGs such as *c-fos* and *zif 268* encode transcription factors that regulate other genes and are implicated in various forms of neuronal plasticity (Knapska & Kaczmarek, 2004; Valjent *et al.*, 2006). Psychostimulant-induced changes in the expression of these IEGs thus suggests that repeated drug exposure produces local neuroadaptations or other neuroplastic changes. While imaging studies indeed indicate functional changes in various cortical regions in human or non-human drug users (London *et al.*, 1990; Breiter *et al.*, 1997; Beveridge *et al.*, 2006; Porrino *et al.*, 2007), only relatively few previous molecular studies investigated the effects of repeated psychostimulant exposure on cortical gene regulation, and these typically focused on the prefrontal cortex (e.g., Freeman *et al.*, 2002; Black *et al.*, 2006). Our study is, to our knowledge, the first to quantitatively map gene regulation effects of acute and repeated cocaine treatment throughout the main functional areas of the cerebral cortex.

Our results demonstrate that repeated cocaine treatment produces increased basal *zif 268* expression in the cortex, an effect that occurred predominantly in somatosensory and motor areas, but spread across all rostrocaudal levels examined. Importantly, increased *zif 268* mRNA levels persisted for at least 3 weeks after the treatment in some of the most affected areas. This increased basal *zif 268* expression was accompanied by attenuated inducibility of *zif 268* one day after the treatment. Thus, in these cocaine-pretreated animals, a cocaine challenge failed to further increase *zif 268* expression. Future work will have to determine which neuroadaptations in cellular signaling pathways, or other neuronal changes (see below), produced by repeated cocaine treatment are responsible for these changes in *zif 268* expression.

Regarding the underlying mechanisms, our most recent findings indicate that such psychostimulant-induced molecular changes in the cortex are not specific for cocaine, but may be age-dependent. Thus, similar to cocaine (present study), enhanced basal expression of *zif 268*, as well as attenuated *zif 268* inducibility were also produced by repeated treatment with methylphenidate (Steiner *et al.*, 2008). Cocaine blocks dopamine, norepinephrine and serotonin transporters, while methylphenidate only blocks reuptake of dopamine and norepinephrine, but not serotonin (c. f., Yano & Steiner, 2007). However, both the present findings with cocaine and the above methylphenidate-induced effects (Steiner *et al.*, 2008) were obtained with repeated drug treatment in adult rats. Interestingly, these effects contrast with our prior findings in adolescent rats. Thus, repeated methylphenidate treatment in adolescents produced opposite effects on cortical IEG regulation, namely, reduced basal *zif 268* expression and enhanced *zif 268* and *homer 1a* induction by a (methylphenidate) challenge throughout the cortex (Cotterly *et al.*, 2007). These age-dependent drug effects on cortical gene regulation are likely related to changes in cell-physiological (e.g., Tseng & O'Donnell, 2005; 2007) or other mechanisms (Andersen, 2005) that occur during normal maturation of cortical processes. It will be important to elucidate what the relevant age-dependent mechanisms are and their exact role in cortical gene regulation by psychostimulants.

### Long-lasting blunting of gene induction in the striatum after repeated cocaine treatment

In contrast to the cortex, numerous studies have investigated psychostimulant effects on gene regulation in the striatum and the findings are very consistent (Harlan & Garcia, 1998; Torres & Horowitz, 1999; Yano & Steiner, 2007). For example, we previously described the topography of cocaine-induced gene regulation for *c-fos* and dynorphin in the striatum (Willuhn *et al.*, 2003). Similar to that regional distribution, our present results for *zif 268* and *homer 1a* demonstrate that acute gene induction by cocaine is most pronounced in dorsal and lateral (sensorimotor) sectors of the striatum, with lesser or minimal effects in the medial (associative) or ventral (limbic) striatum. Along the rostrocaudal axis, such gene regulation is moderate rostrally (associative/limbic striatum), increases towards caudal and peaks in the postcommissural striatum (corresponding to the putamen) (present study; Willuhn *et al.*, 2003). While generally similar to gene regulation by amphetamine (e.g., Badiani *et al.*, 1998) and methylphenidate (Yano & Steiner, 2005a; 2005b), differences in the distribution of cocaine effects were also noted. For example, our present study shows that cocaine induces more robust effects in the caudal striatum, compared with methylphenidate (Yano & Steiner, 2005a; 2005b; Cotterly *et al.*, 2007). This difference may indicate a more pronounced role for serotonin in gene regulation in the caudal striatum (see Yano & Steiner, 2007, for discussion).

Repeated cocaine treatment has previously been shown to produce blunting of IEG induction in the striatum (Hope *et al.*, 1992; Steiner & Gerfen, 1993; for reviews, see Harlan & Garcia, 1998; Yano & Steiner, 2007). Our present results demonstrate that this blunting is directly related to the magnitude of acute gene induction. This finding is consistent with the view that such blunting reflects drug-induced compensatory neuroadaptations, possibly both at the epigenetic level (Renthall & Nestler, 2008) and at the systems level (Steiner & Gerfen, 1998). Again, these adaptations appear to be long-lasting; our present results show for the first time that blunting of IEG induction is still very robust 3 weeks after repeated cocaine treatment.

While the regulation of IEGs such as *c-fos* and *zif 268* is fairly well-known, ours is one of the first studies that assessed effects of repeated psychostimulant treatment on the regulation of *homer 1a* in cortex and striatum. A few previous studies showed induction of *homer 1* isoforms (*homer 1a*, *ania-3*) by acute cocaine in the striatum (Brakeman *et al.*, 1997; Berke *et al.*, 1998; Zhang *et al.*, 2007), but little is known on the effects of repeated psychostimulant treatment on *homer 1a* expression (Szumlinski *et al.*, 2008). In our previous study in adolescent rats, we found a marked dissociation between *zif 268* and *homer 1a* regulation by repeated methylphenidate treatment (Cotterly *et al.*, 2007). While the induction of *zif 268* was significantly blunted in the striatum, that of *homer 1a* was modestly decreased in some areas but increased in others, after repeated treatment. The present study demonstrates that, in contrast, repeated cocaine treatment (in adults) produces identical blunting for both *zif 268* and *homer 1a*, regionally and temporally. These findings extend our observations indicating that cocaine produces to some degree more severe (or other) neuroadaptations than methylphenidate (see Yano & Steiner, 2007, for discussion).

Homer/Ves1 proteins are scaffolding proteins that anchor type I metabotropic glutamate receptors to the postsynaptic density and link them to IP3 receptors in the endoplasmic reticulum (Brakeman *et al.*, 1997; Kato *et al.*, 1997; for reviews, see Xiao *et al.*, 2000; Thomas, 2002). These proteins are implicated in calcium signaling, glutamate receptor clustering and trafficking, spine morphogenesis and other processes of synapse structuring (Xiao *et al.*, 2000; Thomas, 2002). These and other findings suggest a role for *homer 1a* in activity-dependent synaptic plasticity (e.g., regulation of the signaling complex and synapse turnover; Xiao *et al.*, 2000; Thomas, 2002). Our findings thus suggest that repeated cocaine treatment affects restructuring of synapses in the striatum. Interestingly, our results also indicate that this effect is much more prominent in striatal neurons than in the cortical neurons.



### Context-dependent effects of acute cocaine on gene regulation in the cortex

A somewhat surprising finding of our study is that *zif 268* induction in the cortex after acute cocaine administration was dependent on the handling history. Other studies have shown IEG induction in the cortex by acute psychostimulants (e.g., Graybiel *et al.*, 1990; Johansson *et al.*, 1994; Curran *et al.*, 1996; Badiani *et al.*, 1998; Yano & Steiner, 2005a). However, most of these previous studies apparently administered the psychostimulant to animals with limited preceding handling/habituation. Our present study underlines the importance of the handling context for cortical, but not striatal, IEG induction by acute cocaine. Thus, in contrast to the robust *zif 268* response when acute cocaine was given 1 day after repeated vehicle treatment, rats that received acute cocaine 3 weeks after the repeated vehicle injections (plus further intermittent handling) did not show significantly increased *zif 268* expression in the cortex. These results thus corroborate previous observations indicating a role of handling/arousal-related activities in cortical IEG induction by psychostimulants (Badiani *et al.*, 1998; Yano & Steiner, 2005a; Conversi *et al.*, 2006). Insofar as these genes are markers for neuroadaptations (see above), these findings highlight the importance of contextual variables for drug-induced neuroplasticity in the cortex, but not in the striatum.

### Functional considerations

Our present results show extensive and long-lasting changes in gene regulation in cortex and striatum after repeated cocaine treatment. Most of these changes tended to be reduced after the 3-week withdrawal period, indicating that these are transient alterations produced by the drug exposure. Others, however, such as the decreased gene expression in the insular cortex, only emerged after 3 weeks. These latter changes are reminiscent of neurobehavioral changes that develop during drug withdrawal (Grimm *et al.*, 2003; Conrad *et al.*, 2008). These effects may thus reflect cortical alterations related to drug withdrawal-induced behavioral changes such as craving (Naqvi *et al.*, 2007).

The finding that repeated cocaine treatment predominantly affects gene regulation in sensorimotor cortical and striatal domains suggests that especially sensorimotor functions are changed by exposure to such drugs, perhaps more so than is currently appreciated. The exact functional consequences of these molecular changes, however, remain to be determined. It has been shown that acute cocaine increases neuronal firing in cortex and striatum (e.g., Pederson *et al.*, 1997; White *et al.*, 1998; Drouin & Waterhouse, 2004; Devonshire *et al.*, 2007), and such changes in neuronal activity are accompanied by changes in the expression of IEGs such as *zif 268* (Chaudhuri *et al.*, 1995; Melzer & Steiner, 1997). The following paragraphs speculate how neuronal functioning may be altered in these corticostriatal circuits after such treatments.

In the cortex, both (excitatory) projection neurons and (inhibitory) interneurons can upregulate IEG expression upon activation (e.g., Chaudhuri *et al.*, 1995; Bertini *et al.*, 2002; Staiger *et al.*, 2002; Van der Gucht *et al.*, 2002). However, by virtue of the relatively low interneuron numbers (<15–30%, depending on area and layer; Ren *et al.*, 1992; Beaulieu, 1993), gene expression area measures as used here should mostly reflect activity changes in projection neurons. Thus, increased basal *zif 268* expression may indicate chronically enhanced basal activity in (some of) these neurons (Drouin & Waterhouse, 2004). Alternatively, as *zif 268* expression was measured following the open-field test, enhanced “basal” *zif 268* expression in these cocaine-pretreated, vehicle-challenged animals (compared with the vehicle-pretreated, vehicle-challenged group) may reflect increased cortical responsiveness (Dong *et al.*, 2005; Nasif *et al.*, 2005) to arousal-enhancing situations such as the behavioral test. Either way, chronic overactivation of these neurons would be expected to cause adaptations in cellular signaling pathways (Hyman & Nestler, 1996), changes that could contribute to the here observed loss of (IEG) responsiveness after the cocaine challenge. It remains to be seen what

the net effects of such complex cellular and molecular adaptations for behaviorally relevant cortical outputs are in such animals.

In the striatum, cocaine-induced IEG expression occurs mostly but not exclusively in neurons of the D1 receptor-regulated direct output pathway (Cenci *et al.*, 1992; Johansson *et al.*, 1994; Kosofsky *et al.*, 1995; Badiani *et al.*, 1999; Uslaner *et al.*, 2001). A reduction in cortical input to the striatum has been shown to attenuate psychostimulant-induced IEG expression in striatal projection neurons (Cenci & Björklund, 1993; Vargo & Marshall, 1995; Ferguson & Robinson, 2004). Blunting of striatal IEG induction after repeated cocaine treatment could thus, at least in part, reflect dampened cortical input and/or output in direct pathway neurons. Activity in the direct pathway facilitates cortical activation (see Steiner, 2007, for review), which is reflected in increased cortical IEG expression (Steiner & Kitai, 2000). Reduced activity in the direct pathway would therefore be expected to result in a loss of basal ganglia-related input to the cortex (and IEG expression). Therefore, this scenario proposes chronically reduced/dampened activity or excitability in neurons of the affected cortico-basal ganglia-cortical circuits after repeated cocaine treatment.

Our results show that the most pronounced behavioral consequence of the repeated cocaine treatment was an increase in behavioral stereotypies (repetitive head bobbing, sniffing). Such stereotypies are typical behavioral correlates of repeated psychostimulant/dopamine agonist treatments and have been linked to drug-induced molecular changes in sensorimotor corticostriatal circuits before (Graybiel *et al.*, 2000). Such motor stereotypies appear to reflect a dysfunction in selection or switching of motor actions, one of the main functions of basal ganglia circuits (c.f. Cotterly *et al.*, 2007), and may be related to motor compulsions (Graybiel & Rauch, 2000). Drug-induced neuronal alterations in these circuits may thus play a role in compulsive drug taking (Berke & Hyman, 2000; Everitt & Robbins, 2005), a defining characteristic of drug addiction.

## Acknowledgements

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## References

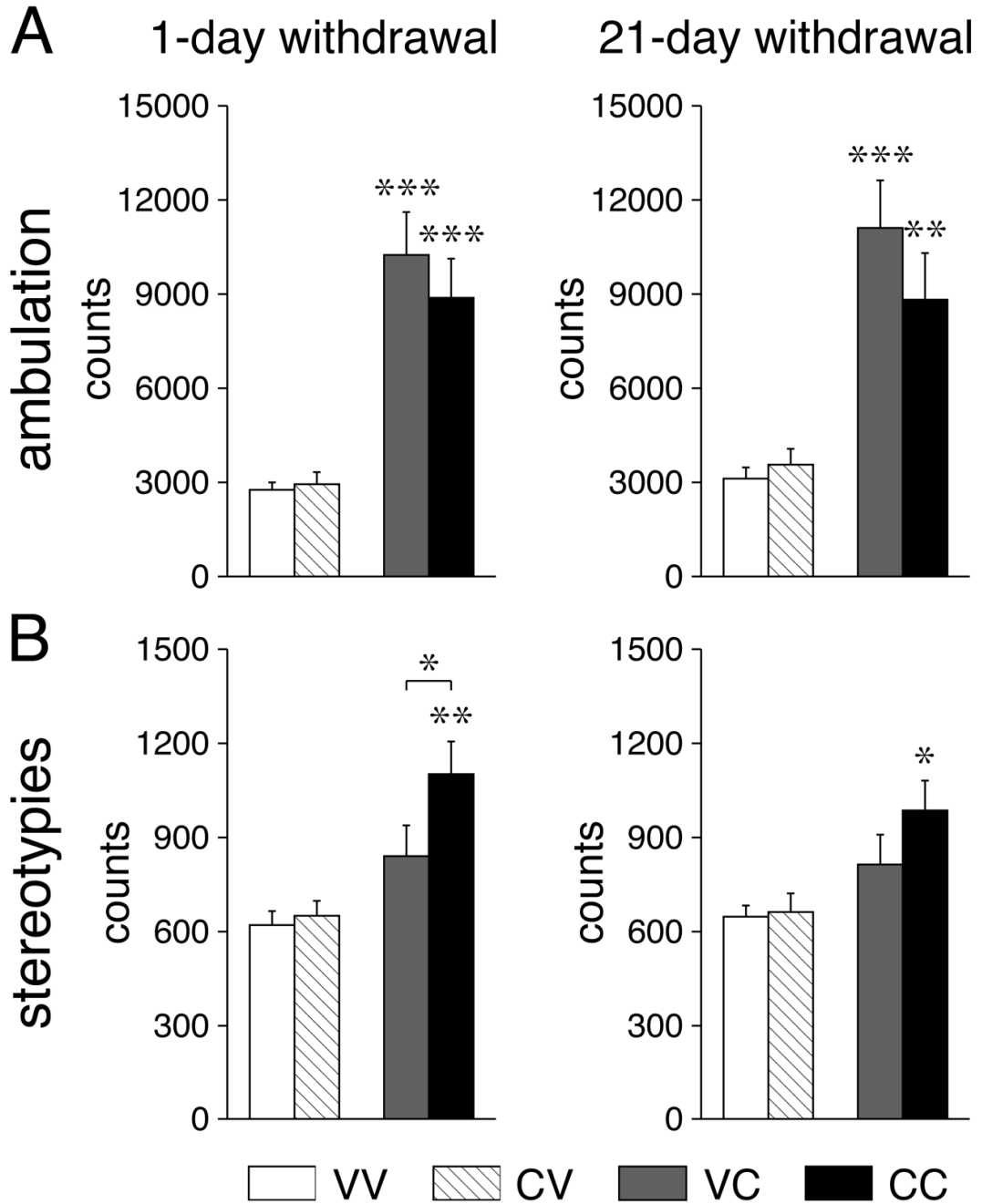
- Albin RL, Young AB, Penney JB. The functional anatomy of basal ganglia disorders. *Trends Neurosci* 1989;12:366–375. [PubMed: 2479133]
- Alexander GE, Crutcher MD, DeLong MR. Basal ganglia-thalamocortical circuits: parallel substrates for motor, oculomotor, “prefrontal” and “limbic” functions. *Prog Brain Res* 1990;85:119–146. [PubMed: 2094891]
- Alexander GE, DeLong MR, Strick PL. Parallel organization of functionally segregated circuits linking basal ganglia and cortex. *Annu Rev Neurosci* 1986;9:357–381. [PubMed: 3085570]
- Andersen SL. Stimulants and the developing brain. *Trends Pharmacol Sci* 2005;26:237–243. [PubMed: 15860370]
- Badiani A, Oates MM, Day HE, Watson SJ, Akil H, Robinson TE. Amphetamine-induced behavior, dopamine release, and c-fos mRNA expression: modulation by environmental novelty. *J Neurosci* 1998;18:10579–10593. [PubMed: 9852594]
- Badiani A, Oates MM, Day HE, Watson SJ, Akil H, Robinson TE. Environmental modulation of amphetamine-induced c-fos expression in D1 versus D2 striatal neurons. *Behav Brain Res* 1999;103:203–209. [PubMed: 10513588]
- Beaulieu C. Numerical data on neocortical neurons in adult rat, with special reference to the GABA population. *Brain Res* 1993;609:284–292. [PubMed: 8508310]
- Berke JD, Hyman SE. Addiction, dopamine, and the molecular mechanisms of memory. *Neuron* 2000;25:515–532. [PubMed: 10774721]

- Berke JD, Paletzki RF, Aronson GJ, Hyman SE, Gerfen CR. A complex program of striatal gene expression induced by dopaminergic stimulation. *J Neurosci* 1998;18:5301–5310. [PubMed: 9651213]
- Bertini G, Peng ZC, Fabene PF, Grassi-Zucconi G, Bentivoglio M. Fos induction in cortical interneurons during spontaneous wakefulness of rats in a familiar or enriched environment. *Brain Res Bull* 2002;57:631–638. [PubMed: 11927366]
- Beveridge TJ, Smith HR, Daunais JB, Nader MA, Porrino LJ. Chronic cocaine self-administration is associated with altered functional activity in the temporal lobes of non human primates. *Eur J Neurosci* 2006;23:3109–3118. [PubMed: 16820001]
- Black YD, Maclaren FR, Naydenov AV, Carlezon WAJ, Baxter MG, Konradi C. Altered attention and prefrontal cortex gene expression in rats after binge-like exposure to cocaine during adolescence. *J Neurosci* 2006;26:9656–9665. [PubMed: 16988036]
- Bottai D, Guzowski JF, Schwarz MK, Kang SH, Xiao B, Lanahan A, Worley PF, Seeburg PH. Synaptic activity-induced conversion of intronic to exonic sequence in Homer 1 immediate early gene expression. *J Neurosci* 2002;22:167–175. [PubMed: 11756499]
- Brakeman PR, Lanahan AA, O'Brien R, Roche K, Barnes CA, Haganir RL, Worley PF. Homer: a protein that selectively binds metabotropic glutamate receptors. *Nature* 1997;386:284–288. [PubMed: 9069287]
- Breiter HC, Gollub RL, Weisskoff RM, Kennedy DN, Makris N, Berke JD, Goodman JM, Kantor HL, Gastfriend DR, Riorden JP, Mathew RT, Rosen BR, Hyman SE. Acute effects of cocaine on human brain activity and emotion. *Neuron* 1997;19:591–611. [PubMed: 9331351]
- Cenci MA, Björklund A. Transection of corticostriatal afferents reduces amphetamine- and apomorphine-induced striatal Fos expression and turning behaviour in unilaterally 6-hydroxydopamine-lesioned rats. *Eur J Neurosci* 1993;5:1062–1070. [PubMed: 8281310]
- Cenci MA, Campbell K, Wictorin K, Björklund A. Striatal *c-fos* induction by cocaine or apomorphine occurs preferentially in output neurons projecting to the substantia nigra in the rat. *Eur J Neurosci* 1992;4:376–380. [PubMed: 12106364]
- Chaudhuri A. Neural activity mapping with inducible transcription factors. *Neuroreport* 1997;8:v-ix.
- Chaudhuri A, Matsubara JA, Cynader MS. Neuronal activity in primate visual cortex assessed by immunostaining for the transcription factor Zif268. *Vis Neurosci* 1995;12:35–50. [PubMed: 7718501]
- Conrad KL, Tseng KY, Uejima JL, Reimers JM, Heng LJ, Shaham Y, Marinelli M, Wolf ME. Formation of accumbens GluR2-lacking AMPA receptors mediates incubation of cocaine craving. *Nature* 2008;454:118–121. [PubMed: 18500330]
- Conversi D, Bonito-Oliva A, Orsini C, Cabib S. Habituation to the test cage influences amphetamine-induced locomotion and Fos expression and increases FosB/DeltaFosB-like immunoreactivity in mice. *Neuroscience* 2006;141:597–605. [PubMed: 16713106]
- Cotterly L, Beverley JA, Yano M, Steiner H. Dysregulation of gene induction in corticostriatal circuits after repeated methylphenidate treatment in adolescent rats: Differential effects on zif 268 and homer 1a. *Eur J Neurosci* 2007;25:3617–3628. [PubMed: 17610581]
- Curran EJ, Akil H, Watson SJ. Psychomotor stimulant- and opiate-induced *c-fos* mRNA expression patterns in the rat forebrain: comparisons between acute drug treatment and a drug challenge in sensitized animals. *Neurochem Res* 1996;21:1425–1435. [PubMed: 8947933]
- DeLong MR. Primate models of movement disorders of basal ganglia origin. *Trends Neurosci* 1990;13:281–285. [PubMed: 1695404]
- Devonshire IM, Mayhew JE, Overton PG. Cocaine preferentially enhances sensory processing in the upper layers of the primary sensory cortex. *Neuroscience* 2007;146:841–851. [PubMed: 17367949]
- Dong Y, Nasif FJ, Tsui JJ, Ju WY, Cooper DC, Hu XT, Malenka RC, White FJ. Cocaine-induced plasticity of intrinsic membrane properties in prefrontal cortex pyramidal neurons: adaptations in potassium currents. *J Neurosci* 2005;25:936–940. [PubMed: 15673674]
- Drouin C, Waterhouse BD. Cocaine-induced vs. behaviour-related alterations of spontaneous and evoked discharge of somatosensory cortical neurons. *Eur J Neurosci* 2004;19:1016–1026. [PubMed: 15009149]
- Everitt BJ, Robbins TW. Neural systems of reinforcement for drug addiction: from actions to habits to compulsion. *Nat Neurosci* 2005;8:1481–1489. [PubMed: 16251991]

- Ferguson SM, Robinson TE. Amphetamine-evoked gene expression in striatopallidal neurons: regulation by corticostriatal afferents and the ERK/MAPK signaling cascade. *J Neurochem* 2004;91:337–348. [PubMed: 15447667]
- Freeman WM, Brebner K, Lynch WJ, Patel KM, Robertson DJ, Roberts DC, Vrana KE. Changes in rat frontal cortex gene expression following chronic cocaine. *Mol Brain Res* 2002;104:11–20. [PubMed: 12117546]
- Graybiel AM, Canales JJ, Capper-Loup C. Levodopa-induced dyskinesias and dopamine-dependent stereotypies: a new hypothesis. *Trends Neurosci* 2000;23:S71–S77. [PubMed: 11052223]
- Graybiel AM, Moratalla R, Robertson HA. Amphetamine and cocaine induce drug-specific activation of the *c-fos* gene in striosome-matrix compartments and limbic subdivisions of the striatum. *Proc Natl Acad Sci USA* 1990;87:6912–6916. [PubMed: 2118661]
- Graybiel AM, Rauch SL. Toward a neurobiology of obsessive-compulsive disorder. *Neuron* 2000;28:343–347. [PubMed: 11144344]
- Grimm JW, Lu L, Hayashi T, Hope BT, Su TP, Shaham Y. Time-dependent increases in brain-derived neurotrophic factor protein levels within the mesolimbic dopamine system after withdrawal from cocaine: implications for incubation of cocaine craving. *J Neurosci* 2003;23:742–747. [PubMed: 12574402]
- Groenewegen HJ, Berendse HW, Wolters JG, Lohman AH. The anatomical relationship of the prefrontal cortex with the striatopallidal system, the thalamus and the amygdala: evidence for a parallel organization. *Prog Brain Res* 1990;85:95–116. [PubMed: 2094917]
- Harlan RE, Garcia MM. Drugs of abuse and immediate-early genes in the forebrain. *Mol Neurobiol* 1998;16:221–267. [PubMed: 9626665]
- Hope B, Kosofsky B, Hyman SE, Nestler EJ. Regulation of immediate early gene expression and AP-1 binding in the rat nucleus accumbens by chronic cocaine. *Proc Natl Acad Sci USA* 1992;89:5764–5768. [PubMed: 1631058]
- Hyman SE, Nestler EJ. Initiation and adaptation: a paradigm for understanding psychotropic drug action. *Am J Psychiatry* 1996;153:151–162. [PubMed: 8561194]
- Johansson B, Lindström K, Fredholm BB. Differences in the regional and cellular localization of *c-fos* messenger RNA induced by amphetamine, cocaine and caffeine in the rat. *Neuroscience* 1994;59:837–849. [PubMed: 7520134]
- Kato A, Ozawa F, Saitoh Y, Hirai K, Inokuchi K. *ves1*, a gene encoding VASP/Ena family related protein, is upregulated during seizure, long-term potentiation and synaptogenesis. *FEBS Lett* 1997;412:183–189. [PubMed: 9257717]
- Knapska E, Kaczmarek L. A gene for neuronal plasticity in the mammalian brain: *Zif268/Egr-1/NGFI-A/Krox-24/TIS8/ZENK?* *Prog Neurobiol* 2004;74:183–211. [PubMed: 15556287]
- Kosofsky BE, Genova LM, Hyman SE. Substance P phenotype defines specificity of *c-fos* induction by cocaine in developing rat striatum. *J Comp Neurol* 1995;351:41–50. [PubMed: 7534774]
- London ED, Cascella NG, Wong DF, Phillips RL, Dannals RF, Links JM, Herning R, Grayson R, Jaffe JH, Wagner HN. Cocaine-induced reduction of glucose utilization in human brain. A study using positron emission tomography and [fluorine 18]-fluorodeoxyglucose. *Arch Gen Psychiatry* 1990;47:567–574. [PubMed: 2350209]
- Melzer P, Steiner H. Stimulus-dependent expression of immediate-early genes in rat somatosensory cortex. *J Comp Neurol* 1997;380:145–153. [PubMed: 9073089]
- Naqvi NH, Rudrauf D, Damasio H, Bechara A. Damage to the insula disrupts addiction to cigarette smoking. *Science* 2007;315:531–534. [PubMed: 17255515]
- Nasif FJ, Hu XT, White FJ. Repeated cocaine administration increases voltage-sensitive calcium currents in response to membrane depolarization in medial prefrontal cortex pyramidal neurons. *J Neurosci* 2005;25:3674–3679. [PubMed: 15814798]
- Paxinos, G.; Watson, C. *The Rat Brain in Stereotaxic Coordinates*. Academic Press; New York: 1998.
- Pederson CL, Wolske M, Peoples LL, West MO. Firing rate dependent effect of cocaine on single neurons of the rat lateral striatum. *Brain Res* 1997;760:261–265. [PubMed: 9237544]
- Pierce RC, Kalivas PW. A circuitry model of the expression of behavioral sensitization to amphetamine-like psychostimulants. *Brain Res Rev* 1997;25:192–216. [PubMed: 9403138]

- Porrino LJ, Smith HR, Nader MA, Beveridge TJ. The effects of cocaine: a shifting target over the course of addiction. *Prog Neuropsychopharmacol Biol Psychiatry* 2007;31:1593–1600. [PubMed: 17900777]
- Ren JQ, Aika Y, Heizmann CW, Kosaka T. Quantitative analysis of neurons and glial cells in the rat somatosensory cortex, with special reference to GABAergic neurons and parvalbumin-containing neurons. *Exp Brain Res* 1992;92:1–14. [PubMed: 1486945]
- Renthal W, Nestler EJ. Epigenetic mechanisms in drug addiction. *Trends Mol Med* 2008;14:341–350. [PubMed: 18635399]
- Robbins TW, Granon S, Muir JL, Durantou F, Harrison A, Everitt BJ. Neural systems underlying arousal and attention. Implications for drug abuse. *Ann N Y Acad Sci* 1998;846:222–237. [PubMed: 9668410]
- Sharp FR, Sagar SM, Swanson RA. Metabolic mapping with cellular resolution: c-fos vs. 2-deoxyglucose. *Crit Rev Neurobiol* 1993;7:205–228. [PubMed: 8221912]
- Staiger JF, Masanek C, Bisler S, Schleicher A, Zuschratter W, Zilles K. Excitatory and inhibitory neurons express c-Fos in barrel-related columns after exploration of a novel environment. *Neuroscience* 2002;109:687–699. [PubMed: 11927151]
- Steiner, H. Basal ganglia – cortex interactions: Regulation of cortical function by D1 dopamine receptors in the striatum. In: Tseng, KY.; Atzori, M., editors. *Monoaminergic Modulation of Cortical Excitability*. Springer; Berlin: 2007. p. 265–285.
- Steiner H, Gerfen CR. Cocaine-induced *c-fos* messenger RNA is inversely related to dynorphin expression in striatum. *J Neurosci* 1993;13:5066–5081. [PubMed: 7504719]
- Steiner H, Gerfen CR. Role of dynorphin and enkephalin in the regulation of striatal output pathways and behavior. *Exp Brain Res* 1998;123:60–76. [PubMed: 9835393]
- Steiner H, Kitai ST. Regulation of rat cortex function by D1 dopamine receptors in the striatum. *J Neurosci* 2000;20:5449–5460. [PubMed: 10884328]
- Steiner H, Lim SAO, Beverley JA. Repeated methylphenidate treatment: Age-dependent effects on gene regulation in the cortex. *Soc Neurosci Abstr* 2008;34:59.29.
- Szumilinski KK, Ary AW, Lominac KD. Homers regulate drug-induced neuroplasticity: implications for addiction. *Biochem Pharmacol* 2008;75:112–133. [PubMed: 17765204]
- Thomas U. Modulation of synaptic signalling complexes by Homer proteins. *J Neurochem* 2002;81:407–413. [PubMed: 12065649]
- Torres G, Horowitz JM. Drugs of abuse and brain gene expression. *Psychosom Med* 1999;61:630–650. [PubMed: 10511013]
- Tseng KY, O'Donnell P. Post-pubertal emergence of prefrontal cortical up states induced by D1-NMDA co-activation. *Cereb Cortex* 2005;15:49–57. [PubMed: 15217899]
- Tseng KY, O'Donnell P. Dopamine modulation of prefrontal cortical interneurons changes during adolescence. *Cereb Cortex* 2007;17:1235–1240. [PubMed: 16818475]
- Uslaner J, Badiani A, Norton CS, Day HE, Watson SJ, Akil H, Robinson TE. Amphetamine and cocaine induce different patterns of c-fos mRNA expression in the striatum and subthalamic nucleus depending on environmental context. *Eur J Neurosci* 2001;13:1977–1983. [PubMed: 11403691]
- Valjent E, Aubier B, Corbillé AG, Brami-Cherrier K, Caboche J, Topilko P, Girault JA, Hervé D. Plasticity-associated gene *Krox24/Zif268* is required for long-lasting behavioral effects of cocaine. *J Neurosci* 2006;26:4956–4960. [PubMed: 16672671]
- Van der Gucht E, Clerens S, Cromphout K, Vandesande F, Arckens L. Differential expression of c-fos in subtypes of GABAergic cells following sensory stimulation in the cat primary visual cortex. *Eur J Neurosci* 2002;16:1620–1626. [PubMed: 12405976]
- Vargo JM, Marshall JF. Time-dependent changes in dopamine agonist-induced striatal Fos immunoreactivity are related to sensory neglect and its recovery after unilateral prefrontal cortex injury. *Synapse* 1995;20:305–315. [PubMed: 7482290]
- White IM, Doubles L, Rebec GV. Cocaine-induced activation of striatal neurons during focused stereotypy in rats. *Brain Res* 1998;810:146–152. [PubMed: 9813293]
- Willuhn I, Sun W, Steiner H. Topography of cocaine-induced gene regulation in the rat striatum: Relationship to cortical inputs and role of behavioural context. *Eur J Neurosci* 2003;17:1053–1066. [PubMed: 12653981]

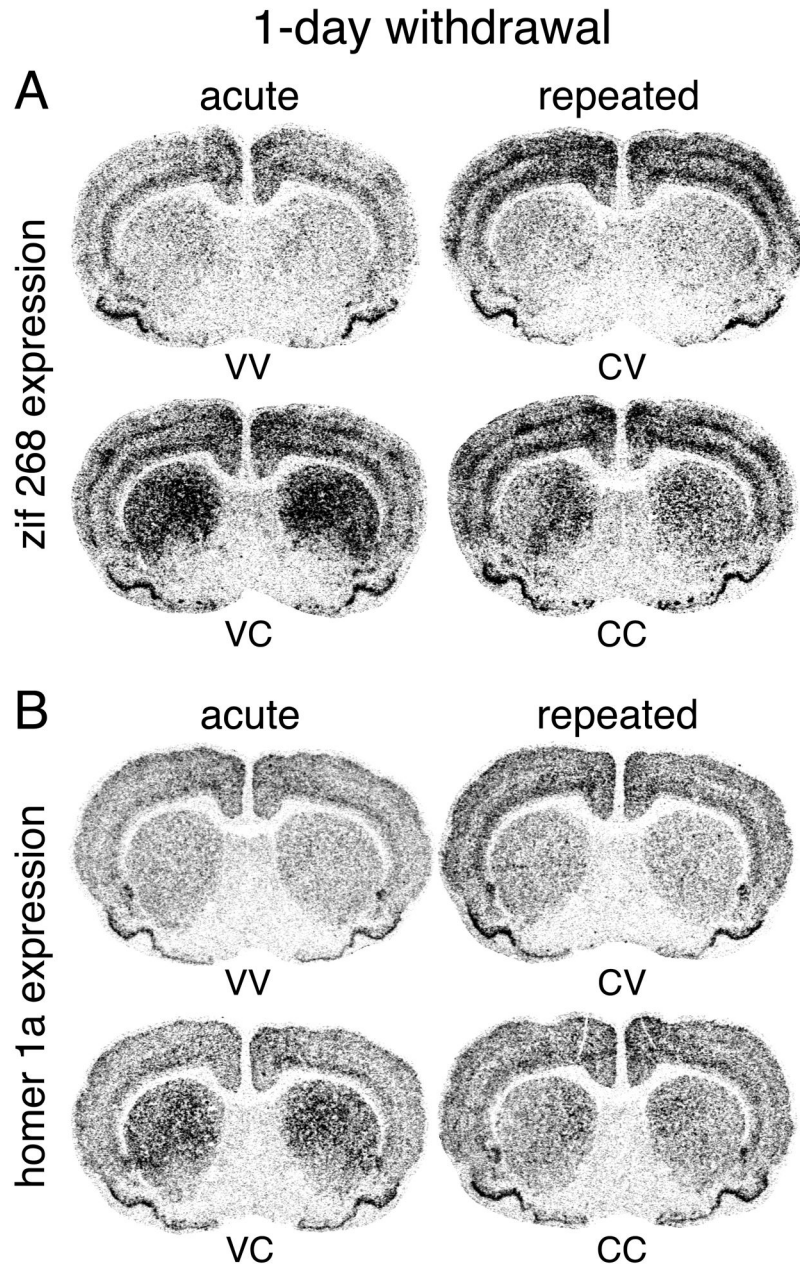
- Xiao B, Tu JC, Worley PF. Homer: a link between neural activity and glutamate receptor function. *Curr Opin Neurobiol* 2000;10:370–374. [PubMed: 10851183]
- Yano M, Steiner H. Methylphenidate (Ritalin) induces Homer 1a and zif 268 expression in specific corticostriatal circuits. *Neuroscience* 2005a;132:855–865. [PubMed: 15837145]
- Yano M, Steiner H. Topography of methylphenidate (Ritalin)-induced gene regulation in the striatum: differential effects on *c-fos*, substance P and opioid peptides. *Neuropsychopharmacology* 2005b; 30:901–915. [PubMed: 15637641]
- Yano M, Steiner H. Methylphenidate and cocaine: the same effects on gene regulation? *Trends Pharmacol Sci* 2007;28:588–596. [PubMed: 17963850]
- Zhang GC, Mao LM, Liu XY, Parelkar NK, Arora A, Yang L, Hains M, Fibuch EE, Wang JQ. In vivo regulation of Homer1a expression in the striatum by cocaine. *Mol Pharmacol* 2007;71:1148–1158. [PubMed: 17234898]



**Fig 1.**

Repeated cocaine treatment increases levels of behavioral stereotypies. Ambulation (A) and stereotypy counts (B) (mean±SEM) are shown for animals that received 5 daily injections of vehicle or cocaine (25 mg/kg, i.p.) followed 1 day (left) or 21 days later (right) by a vehicle injection (groups VV and CV) or a cocaine challenge (25 mg/kg) (groups VC and CC).

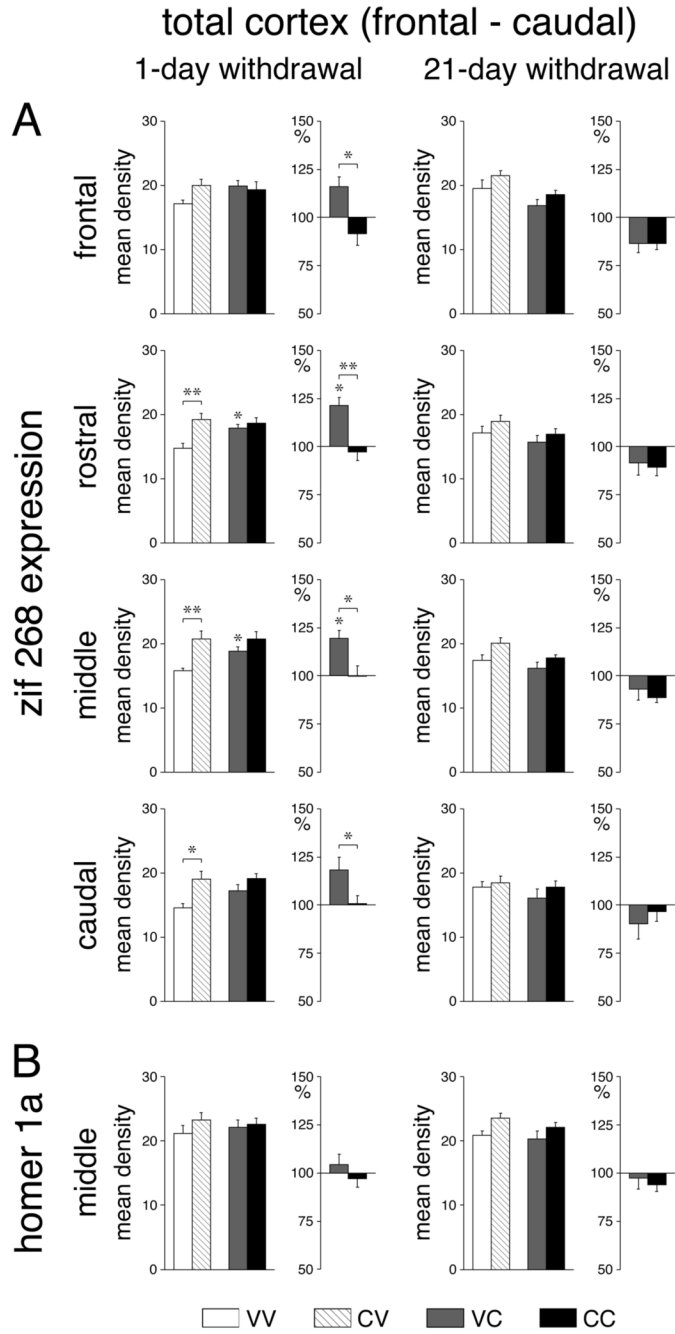
Ambulatory activity and stereotypies were assessed for 30 min in a novel openfield, immediately after the injection (see Materials and Methods). \*  $P < 0.05$ , \*\*  $P < 0.01$ , \*\*\*  $P < 0.001$  vs. respective vehicle control or as indicated.



**Fig. 2.**

Effects of repeated cocaine treatment on IEG expression in the cortex and striatum after 1-day withdrawal. Illustrations of film autoradiograms depict expression of *zif 268* (A) and *homer 1a* (B) in cortex and striatum on the mid-striatal level for rats that received 6 vehicle injections (VV), 5 vehicle injections followed one day later by a cocaine challenge (25 mg/kg) (VC) (acute cocaine), or 5 cocaine injections (25 mg/kg) followed by a vehicle (CV) or a cocaine injection (CC) (repeated cocaine, 1-day withdrawal). Repeated cocaine treatment significantly increased basal *zif 268* expression in the cortex (CV vs. VV) (similar trend for *homer 1a*). In contrast, challenge-induced expression of *zif 268* in the cortex and *zif 268* and *homer 1a* in the striatum were attenuated (CC vs. CV compared with VC vs. VV).



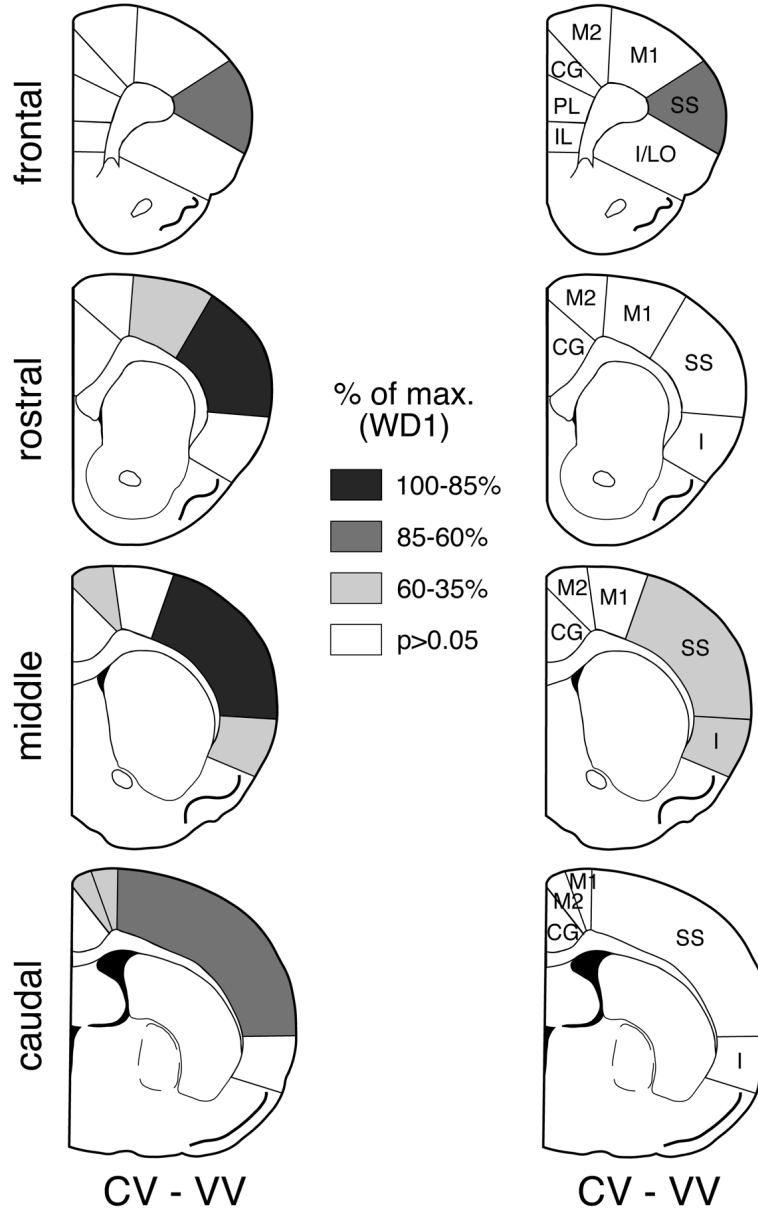


**Fig. 3.** Repeated cocaine treatment increases basal expression and attenuates challenge-induced expression of *zif 268* in the cortex. Effects on *zif 268* (A) and *homer 1a* (B) expression (mean density values; mean±SEM, arbitrary units) in the total cortex on frontal, rostral, middle and caudal levels are shown for rats that were treated with 6 vehicle injections (VV), 5 cocaine injections (25 mg/kg) followed 1 day later (left column) or 21 days later (right column) by a vehicle injection (CV), 5 vehicle injections followed by a cocaine challenge (25 mg/kg) (VC), or 5 cocaine injections followed by a cocaine challenge (CC). Absolute values (left) and relative values (% of vehicle controls, right) are depicted. \* P<0.05, \*\* P<0.01 vs. vehicle controls or as indicated.

## changes in basal *zif* 268 expression

1-day withdrawal

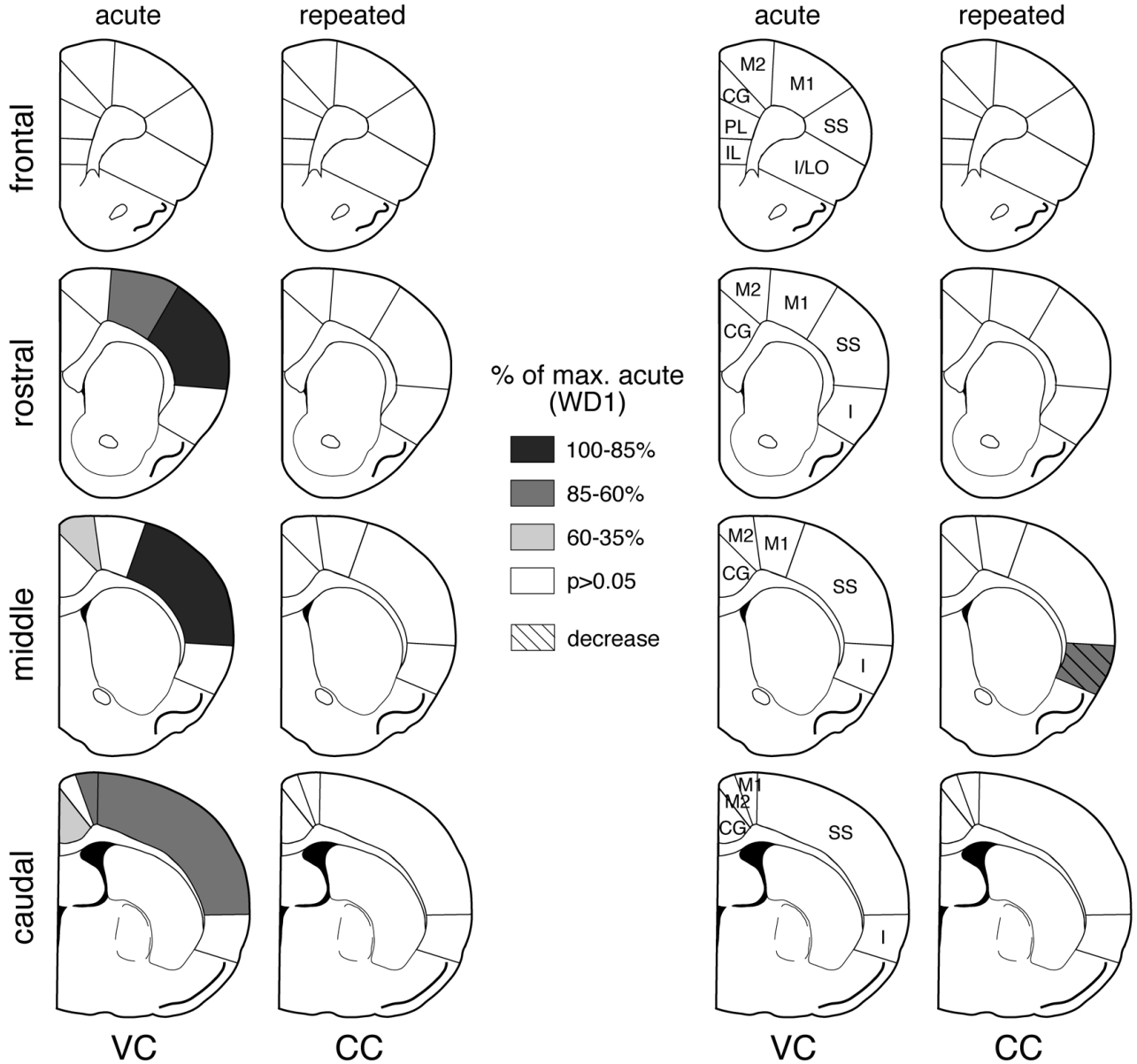
21-day withdrawal



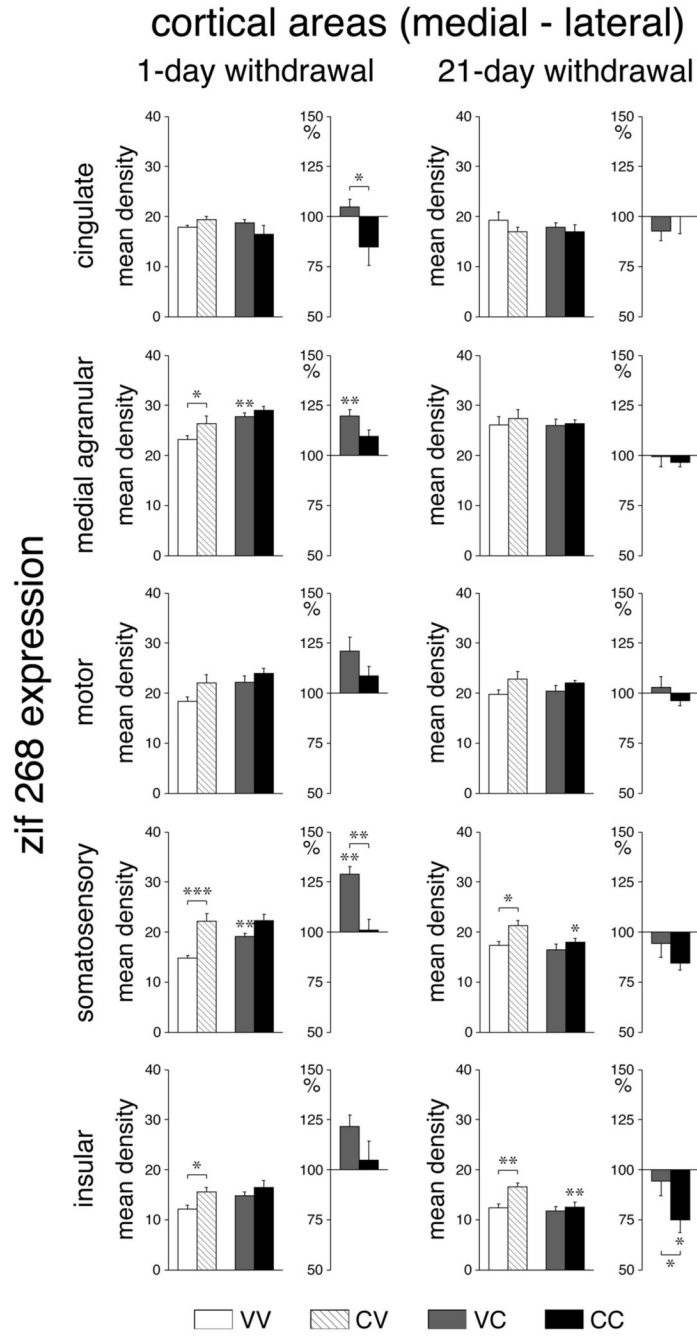
**Fig. 4.**

Distribution of increases in basal *zif* 268 expression in the cortex after repeated cocaine treatment: 1-day vs. 21-day withdrawal. The maps illustrate the differences in *zif* 268 expression between animals that received 5 cocaine injections followed by a vehicle injection (CV) and controls that received 6 vehicle injection (VV) on frontal, rostral, middle and caudal levels, 1 day (left column) and 21 days (right column) after repeated cocaine administration. The values are expressed relative to the maximal change on withdrawal day 1 (% of max. WD1). Areas with significant differences ( $P < 0.05$ ) are coded as indicated. Areas without significant effects are in white. IL, infralimbic; PL, prelimbic; CG, cingulate; M2, medial agranular; M1, motor; SS, somatosensory; I, insular; I/LO, insular/lateral orbital.

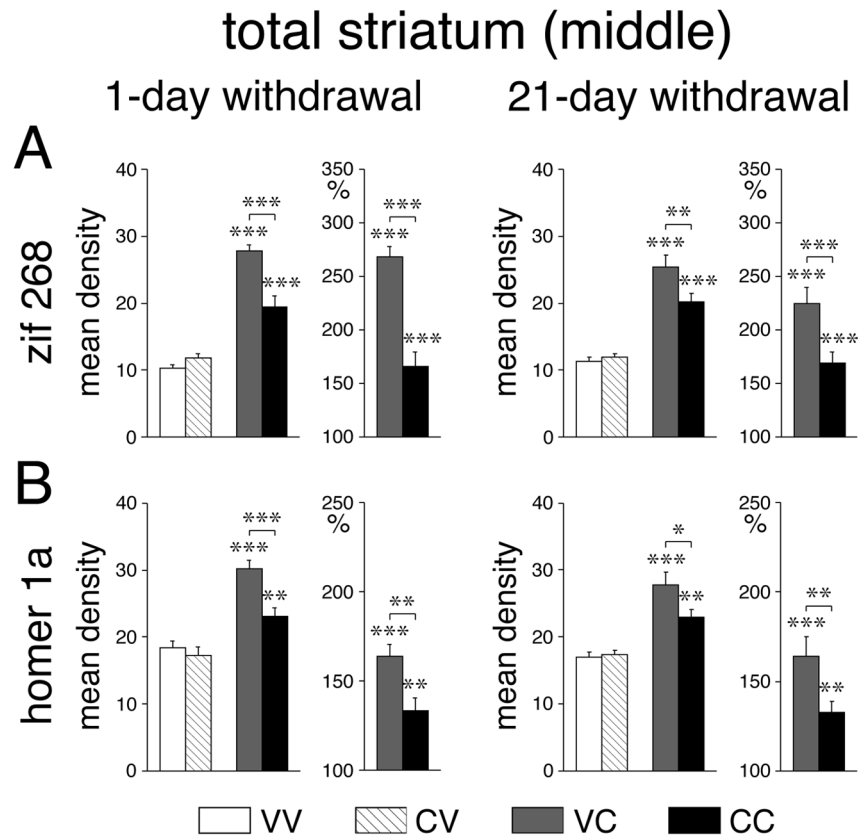
## changes in challenge-induced *zif* 268 expression 1-day withdrawal                      21-day withdrawal



**Fig. 5.** Distribution of changes in cocaine challenge-induced *zif* 268 expression in the cortex after repeated cocaine treatment: 1-day vs. 21-day withdrawal. The maps depict changes in *zif* 268 expression on frontal, rostral, middle and caudal levels after acute (VC) and repeated (CC) cocaine administration, 1 day (left column) and 21 days (right column) after repeated treatment. The value in a given region was first expressed as the percentage of that in vehicle controls, and was then normalized relative to the maximal change in the acute group on withdrawal day 1 (% of max. acute, WD1). Areas with significant differences ( $P < 0.05$ ) are coded as indicated. Areas without significant effects are in white. IL, infralimbic; PL, prelimbic; CG, cingulate; M2, medial agranular; M1, motor; SS, somatosensory; I, insular; I/LO, insular/lateral orbital.

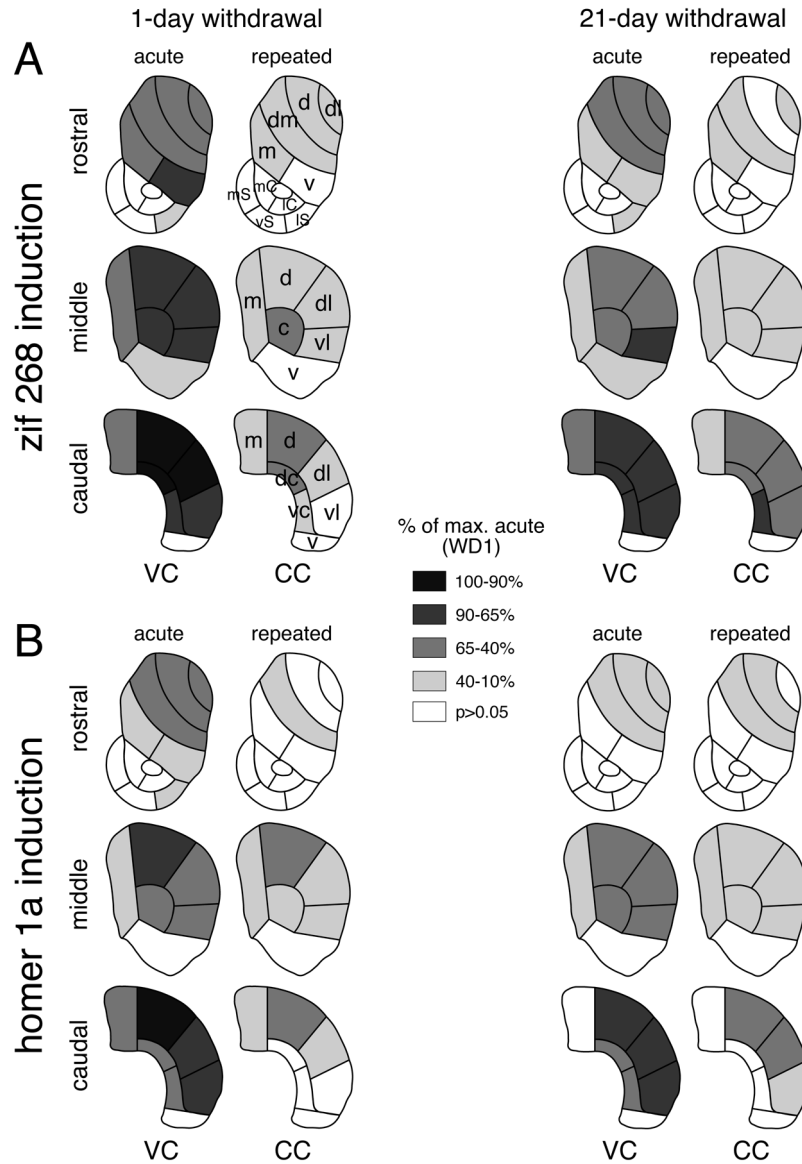


**Fig. 6.** Regional specificity of repeated cocaine-induced changes in cortical *zif268* expression: 1-day vs. 21-day withdrawal. Effects on *zif268* expression (mean density values; mean±SEM) in the different cortical areas on the middle level are given for VV, CV, VC and CC groups, 1 day (left column) or 21 days (right column) after repeated treatment. Absolute values (left) and relative values (% of vehicle controls, right) are depicted. \* P<0.05, \*\* P<0.01, \*\*\* P<0.001 vs. vehicle controls or as indicated.

**Fig. 7.**

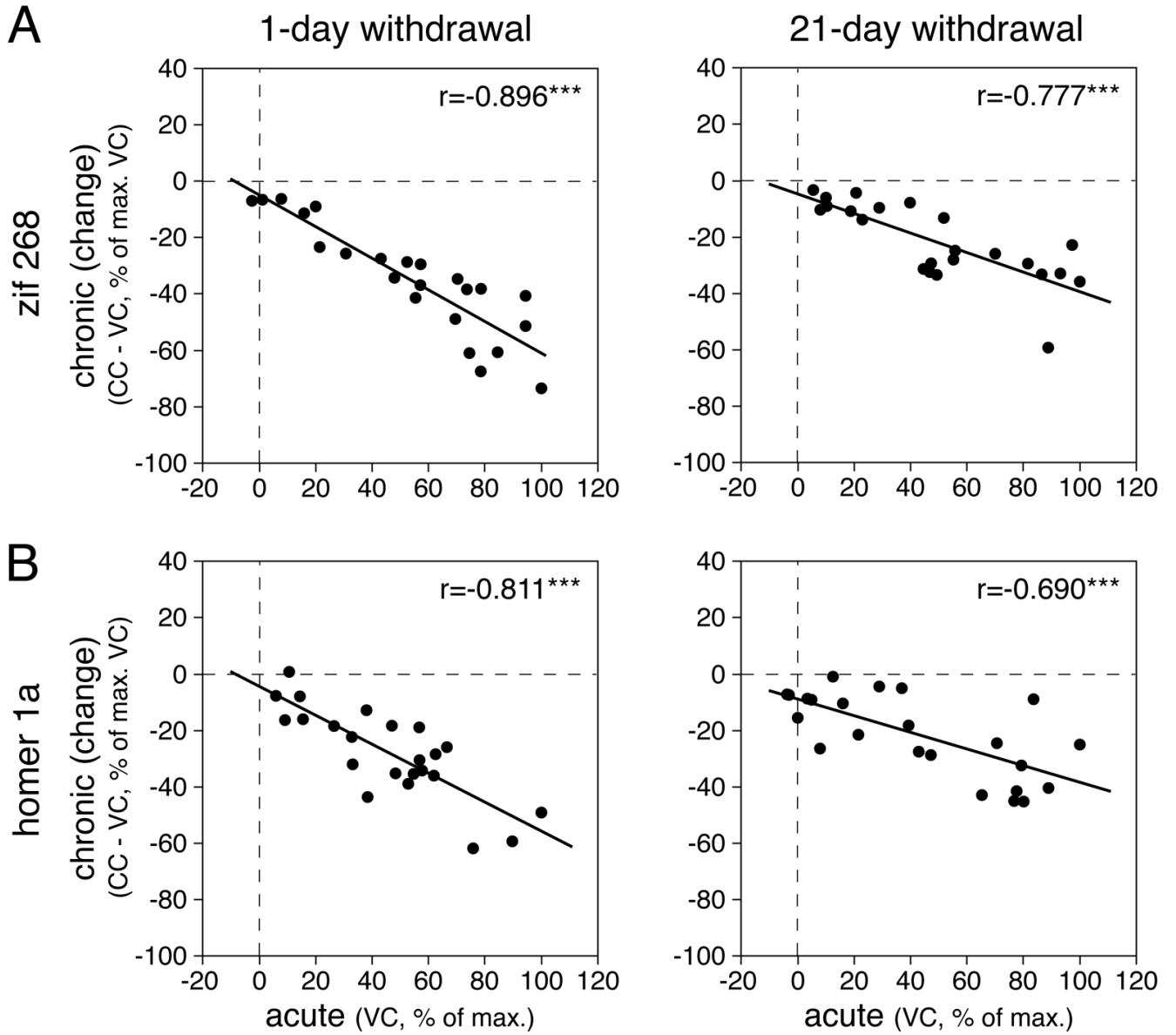
Repeated cocaine treatment blunts inducibility of *zif 268* and *homer 1a* in the striatum. Effects on *zif 268* (A) and *homer 1a* (B) expression (mean density values; mean±SEM) in the total striatum on the middle level are shown for VV, CV, VC and CC groups, 1 day (left column) or 21 days (right column) after repeated treatment. Absolute values (left) and relative values (% of vehicle controls, right) are depicted. \* P<0.05, \*\* P<0.01, \*\*\* P<0.001 vs. vehicle controls or as indicated.

changes in challenge-induced gene expression



**Fig. 8.** Distribution of changes in cocaine challenge-induced *zif 268* and *homer 1a* expression in the striatum after repeated cocaine treatment: 1-day vs. 21-day withdrawal. The maps illustrate changes in *zif 268* (A) and *homer 1a* (B) expression in the rostral, middle and caudal striatum after acute (VC) and repeated (CC) cocaine administration, 1 day (left column) and 21 days (right column) after repeated treatment. The value in a given sector was first expressed as the percentage of that in vehicle controls, and was then normalized relative to the maximal change in the acute group on withdrawal day 1 (% of max. acute, WD1). Sectors with significant differences ( $P < 0.05$ ) are coded as indicated. Sectors without significant effects are in white. Caudate-putamen: m, medial; dm, dorsomedial; d, dorsal; dl, dorsolateral; vl, ventrolateral; v, ventral; c, central; vc, ventral central; dc, dorsal central; nucleus accumbens: mC, medial core; IC, lateral core; mS, medial shell; vS, ventral shell; lS, lateral shell.

striatum: acute vs. chronic gene induction



**Fig. 9.** Relationship between acute IEG induction and IEG induction after repeated (chronic) cocaine treatment in the striatum. Scatterplots show correlations between acute induction (VC, in % of max. VC) and changes in chronic induction (CC-VC, in % of max. VC) for *zif 268* (A) and *homer 1a* expression (B) in the 23 striatal sectors. \*\*\*  $P < 0.001$ .

**Table 1**

Functional domains: striatal sectors and their cortical inputs

level	striatal sector	main cortical input regions
rostral	dorsolateral	motor (M1), somatosensory
	dorsal	medial agranular (M2), motor (M1), somatosensory
	dorsomedial	cingulate
	medial	cingulate, prelimbic
	ventral	insular, lateral orbital
	NAC medial core	prelimbic
	NAC lateral core	insular
	NAC medial shell	prelimbic
	NAC ventral shell	infralimbic
	NAC lateral shell	insular, lateral orbital
middle	medial	cingulate, prelimbic
	dorsal	medial agranular (M2), motor (M1), somatosensory
	dorsolateral	motor (M1), somatosensory
	ventrolateral	motor (M1), somatosensory
	ventral	insular
	central	insular, lateral orbital
caudal	medial	cingulate, prelimbic
	dorsal	medial agranular (M2), motor (M1), somatosensory
	dorsolateral	motor (M1), somatosensory
	ventrolateral	motor (M1), somatosensory
	ventral	insular
	ventral central	insular
	dorsal central	insular, lateral orbital

The function of the 23 striatal sectors (left) is mostly determined by their cortical inputs (simplified, right). These inputs are described and discussed in detail in Willuhn *et al.* (2003). NAC, nucleus accumbens.