

Human Herpesvirus 6 Inhibits Human Immunodeficiency Virus Type 1 Replication in Cell Culture

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The SF strain of human herpesvirus 6 (HHV-6_{SF}) isolated from the saliva of a human immunodeficiency virus (HIV)-infected individual was shown to inhibit HIV type 1 (HIV-1) replication in both peripheral blood mononuclear cells and purified CD4⁺ lymphocytes. This suppression of HIV-1 replication led to decreased cytopathic effects of HIV-1 and prolonged survival of CD4⁺ cells in culture. Even low levels of HHV-6 added to peripheral blood mononuclear cells showed an inhibitory effect on HIV-1 replication. These results differ from those previously reported showing enhanced HIV-1 production following infection with another strain of HHV-6.

We previously described the high prevalence of human herpesvirus 6 (HHV-6) in the saliva of healthy individuals (5). The prototype virus strain characterized, HHV-6_{SF}, has biologic and molecular features somewhat different from those of other reported HHV-6 strains, but like all other isolates, it preferentially grows in CD4⁺ lymphocytes (5a). Both human immunodeficiency virus type 1 (HIV-1) and HHV-6 can be found in the same individual, but these viruses apparently use a different receptor on CD4⁺ cells for infection (5a, 9). Other investigators have previously reported that coinfection of peripheral blood mononuclear cells (PMC) or established T-cell lines with both viruses leads to an enhanced replication of HIV-1 (8). This observation supports the finding that HHV-6 infection stimulates chloramphenicol acetyltransferase enzyme production when linked to the HIV-1 long terminal repeat (2, 8). We also studied the interaction of these two viruses in PMC and purified CD4⁺ lymphocytes. Our results differ from those previously published, since we found that dual infection of these cultures leads to a marked decrease in HIV replication as well as an inhibitory effect of HIV-1 on HHV-6 expression.

For our studies, the PMC (6) and CD4⁺ lymphocytes (11) were obtained from the blood of adult HIV-1-seronegative individuals whose PMC have not shown the presence of replicating HHV-6. The cells (3×10^6) were cultured as described previously (6) in 2 ml of RPMI 1640 medium containing 5% heated (56°C, 30 min) fetal calf serum, 2 mM glutamine, 1% antibiotics (100 U of penicillin per ml, 100 µg of streptomycin per ml), and 5% interleukin-2 (Electronucleonics, Silver Springs, Md.). In the initial experiments, 0.1 ml of HHV-6_{SF} (with a titer of $10^{6.4}$ infectious particles per ml) and 0.1 ml of HIV-1_{SF33} (reverse transcriptase [RT] activity, 10^6 cpm/ml) were added at the same time to the cultures. Control cultures received the same quantity of either virus alone. After 24 h, the cultures were centrifuged and the cells were suspended in fresh medium without the viruses. Every 3 days, the clarified supernatant was evaluated for HIV-1 by RT assays (1) and the cells were suspended in fresh medium. A portion of the cells was examined for HHV-6 by indirect

immunofluorescence assays (IFA) as described previously (4, 5). In some studies, cells were also evaluated by IFA for HIV antigens with a monoclonal antibody to the viral core (4). Other experiments were conducted with the HIV-1_{SF2} strain by using similar procedures. This virus differs from HIV-1_{SF33} in that it is less cytopathic and does not grow as rapidly in cell culture (6).

The results indicated that HHV-6_{SF} infection inhibits HIV-1 replication. In the first experiments, after 1 week HIV-1_{SF33} had an RT activity of 19,800 cpm/ml in the HHV-6 coinfecting cells compared with over 10^6 cpm/ml in the culture containing the HIV-1 strain alone. During this observation period, HHV-6-staining cells increased from 5 to 100%, with no substantial cell death in the cultures, as measured by trypan blue dye exclusion and viable cell counts. Moreover, by 11 days, HIV-1 was no longer recoverable from the supernatant of the dually infected cultures, as measured by inoculation of the fluid onto fresh PMC cultures. Three other experiments conducted in the same manner gave comparable results.

Similar observations were made as well when 3×10^6 CD4⁺ lymphocytes purified by panning (11) were dually infected with the herpesvirus and the HIV-1_{SF33} or HIV-1_{SF2} strains (Table 1). Inhibition of the more rapidly growing and more cytopathic HIV-1_{SF33} strain was noted somewhat sooner than that of the HIV-1_{SF2} strain. Substantial reduction in replication of both HIV-1 strains in culture occurred within 4 to 7 days after infection. Moreover, HIV-1 infection also decreased the replication of HHV-6. In the dually infected cultures, the herpesvirus antigen-positive cells increased at a slower rate, reaching 100% on day 11, compared with day 7 in the HHV-6_{SF}-infected cultures alone (Table 1). By day 11, the cell viabilities of both infected cultures had decreased to 60 to 79% of the uninfected controls, with the exception of HIV-1_{SF2}-infected cultures, which remained at 93%. These results reflect the known cytopathic properties of HHV-6 and HIV-1_{SF33}.

Flow cytometry studies (7) with the Leu3a and Leu2a monoclonal antibodies to CD4 and CD8 cell markers (Becton Dickinson and Co., Mountain View, Calif.) were conducted on CD4⁺ cell populations infected by each virus alone and those dually infected to note any effect on cell subset survival. The relative number of CD4⁺/CD8⁻ cells increased

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TABLE 1. Effect of HHV-6 on HIV-1 replication in CD4⁺ lymphocytes

Virus	Measure of virus replication at ^a :									
	Day 4		Day 7		Day 11					
	HIV RT	HHV-6 Ag	HIV RT	HHV-6 Ag	HIV RT	HHV-6 Ag	Viability	CD4 ⁺	CD8 ⁺	CD4 ⁻ /CD8 ⁻
HHV-6 _{SF}	—	—	—	100	—	100	73	94	5	—
HIV-1 _{SF2}	0.8	—	104.9	—	206	—	93	68	4	27
HIV-1 _{SF2} + HHV-6 _{SF}	1.6	1	3.5	25	1.9	100	79	97	2	1
HIV-1 _{SF33}	32.6	—	183.3	—	148	—	60	57	14	29
HIV-1 _{SF33} + HHV-6 _{SF}	14.5	—	30.5	50	20.2	100	64	87	3	10
None	—	—	—	—	—	—	95	98	2	—

^a Values indicate the following: RT, RT activity (10⁴ cpm/ml); Ag, HHV-6 antigen expression (percentage of cells), as detected by IFA (see text); viability, percentage of cells, as measured by trypan blue dye exclusion. T-cell subsets (CD4⁺, CD8⁺, and CD4⁻/CD8⁻) were assessed by flow cytometry analysis (7) and expressed as the percentage of total cells examined. A representative experiment of three independent studies is presented. —, None detected.

in the HIV-1-infected cultures, most likely reflecting CD4⁺ cell death and/or down modulation of the CD4 antigen in HIV-1-infected cells (3). The absence or low levels of these CD4⁻/CD8⁻ cells in the dually infected cultures and the continued presence of high numbers of CD4⁺ cells supports the lack of substantial HIV-1 replication and cytopathology in these cultures.

We next examined whether low input levels of HHV-6 would have a similar effect on HIV replication. High titers (10⁶ cpm/ml) of HIV-1_{SF2} and HIV-1_{SF33} (0.1 ml) were inoculated onto PMC. At the same time, 0.1 ml of HHV-6_{SF} undiluted or diluted 10- and 100-fold (from a stock of 10^{6.4} infectious particles per ml) was added to separate cultures (Table 2). The initial low level antigen detected at 5 days in the undiluted HHV-6-infected control cultures most likely reflects defective interfering particles (5a). The results again demonstrated that HHV-6 infection had a dramatic inhibiting effect on HIV-1 replication. Even when very limited HHV-6 infection was noted (day 5), with 1% of the cells staining for HHV-6 antigen, the reduction in RT activity was evident (Table 2). The same observation was made after 1 week of

culture. At both time periods, viable cell counts were similar for both infected and uninfected cells (>95% viable). Moreover, as observed with the CD4⁺ cells (Table 1), HIV-1_{SF33} had some inhibitory effect on HHV-6 replication. In three independent studies, this inhibitory effect of the two virus types on each other was noted to varying degrees. The reduction in RT activity was not always linear (Table 2), but the reduction in HIV-1 replication was evident throughout. Moreover, the effect of HIV-1 on HHV-6 expression was not consistently noted with the HIV-1_{SF2} strain.

This inhibition of HIV-1 replication by HHV-6_{SF}, as well as the effect of HIV-1 on HHV-6_{SF} expression, was further examined in timed experiments in which one virus was added to PMC 24 h earlier than the other. In these studies, a dramatic reduction in HIV-1 replication was found in the PMC cultures previously inoculated with HHV-6_{SF}. Similarly, HHV-6_{SF} expression, as demonstrated by antigen production, was substantially inhibited by prior infection of the PMC with HIV-1_{SF33} (Fig. 1). These observations were made when no substantial decrease in cell viability was noted by trypan blue dye exclusion.

These results in cell culture contrast with those previously reported for HHV-6 (8). Our data indicate that HHV-6_{SF} infection inhibits HIV-1 replication in PMC and purified CD4⁺ lymphocytes before any evidence of viral cytotoxicity. The reason for the discrepancy between our results and those of others in which enhanced HIV-1 replication was noted is not clear. It may be related to the HIV-1 or HHV-6 strains used. HHV-6_{SF} does differ biologically from the other HHV-6 strains described elsewhere (5a). Our results suggest, however, that in contrast to observations made with previous biologic and molecular studies (2, 8), infection with this herpesvirus in vivo can lead to an inhibition of HIV-1 replication. Moreover, our findings indicate a difference in the interaction of this herpesvirus with HIV-1 compared with those of other herpesviruses, such as Epstein-Barr virus, in which enhancement of HIV-1 replication appears to take place (10).

The mechanism for this viral inhibition is not known. Because HHV-6 antigen can be found in more than 80% of the cells at the time that some HIV-1 replication can be detected, we assume the inhibition occurs intracellularly. In addition, in some experiments before HIV-1 replication was completely suppressed, cells could be detected that were dually infected, although, as expected, the expression of HIV-1 antigens was reduced. Furthermore, these two viruses use different cellular receptors (5a, 9). Finally, the

TABLE 2. Effect of varying quantities of HHV-6_{SF} on HIV-1 replication

Virus	Virus replication at ^a :			
	Day 5		Day 7	
	Ag ^b	RT ^c	Ag	RT
HHV-6 _{SF}				
10 ⁰	8	—	80	—
10 ⁻¹	20	—	30	—
HIV-1 _{SF2}	—	88	—	436.9
+HHV-6 _{SF} (10 ⁰)	8	34	75	74.5
+HHV-6 _{SF} (10 ⁻¹)	1	12.8	30	83.1
+HHV-6 _{SF} (10 ⁻²)	1	13.4	10	234.2
HIV-1 _{SF33}	—	311.4	—	474.6
+HHV-6 _{SF} (10 ⁰)	5	99.0	80	56.2
+HHV-6 _{SF} (10 ⁻¹)	1	166.4	<1	276.3
+HHV-6 _{SF} (10 ⁻²)	1	161.1	1	126.4

^a Procedures are described in text. —, Not detected. A representative experiment of three independent studies is presented.

^b Values indicate the percentage of PMC staining for HHV-6 antigen by IFA (4).

^c Values indicate HIV-1 RT activity (10³ cpm/ml) in the culture supernatant (1).

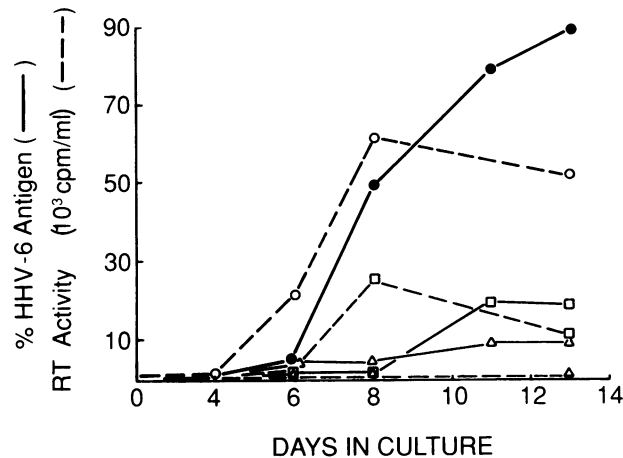


FIG. 1. Human PMC (6×10^6 cells in 2 ml of medium) were inoculated with HHV-6_{SF} (10^5 infectious particles) or HIV-1_{SF33} (10^5 cpm). After 24 h, the cells were washed and split into duplicate cultures. One HHV-6_{SF} culture was superinfected with HIV-1_{SF33}; one HIV-1_{SF33} culture was superinfected with HHV-6. Every 2 to 3 days, the four cultures were centrifuged and a portion of the cells was examined for HHV-6 antigens by IFA (5); the culture fluid was assayed for particle-associated RT activity (1). The data from the single- and dually infected cultures are given. The superinfecting virus, when inoculated onto fresh PMC, gave curves similar to those shown for the cultures receiving the viruses at time zero. Symbols: ●, HHV-6_{SF} alone; ○, HIV-1_{SF33} alone; △, HHV-6_{SF} followed by HIV-1_{SF33} 24 h later; □, HIV-1_{SF33} followed by HHV-6_{SF} 24 h later. A representative experiment from two independent studies is presented.

timed experiments indicate that infection with one virus can inhibit replication of the other virus. It is conceivable, however, that dual infection is not needed and an inhibitory cytokine or some cell surface phenomenon blocks the virus spread and replication. In any case, the results demonstrate that the presence of both viruses is inhibitory in culture and this observation should be considered when culturing fluids or cells for HIV-1. The sensitivity for HIV detection could be decreased if HHV-6 is present in the sample.

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