# Decorrelating source and sink determinism of nitrogen remobilization during grain filling in wheat

# **Pierre Bancal\***

INRA, UMR 1091 INRA/INAPG Environnement et Grandes Cultures, F-78850 Thiverval Grignon, France

Received: 8 December 2008 Returned for revision: 21 January 2009 Accepted: 23 February 2009 Published electronically: 1 April 2009

• *Background and Aims* Nitrogen (N) remobilization is the major source of N for grain filling in wheat, the other being N uptake after anthesis  $(N_{up})$ ; however, variations in remobilization efficiency are not fully understood. It is hard to tell whether the source or the sink effects predominate, because N in the culm at anthesis  $(N_{ant})$  correlates strongly with both N remobilization  $(N_{rem})$  and grain number  $(G_n)$ , respectively the main source and the main sink.

• *Methods* A pot experiment was thus designed to assess the relative contributions of the source and sink to  $N_{\text{rem}}$  regulation. Using two cultivars of winter wheat (*Triticum aestivum*, 'Apache' and 'Autan'), three pre-anthesis and two post-anthesis N fertilization levels were applied in order to vary the N sources, while ear trimming at anthesis reduced sink size.

• Key Results Unlike results observed at a scale of  $m^2$ , the equation binding  $N_{ant}$  to  $N_{rem}$  exhibited a negative intercept, challenging the concept of nitrogen remobilization efficiency. Before ear trimming,  $G_n$  fitted well to  $N_{ant}$ , with a slope dependent on genotype. To obtain a sink variable that was less correlated with  $N_{ant}$ , the difference  $\delta G_n$  was calculated between actual grain number and that which could be predicted from culm N before trimming. A multiple regression then predicted  $N_{rem}$  ( $r^2 = 0.95$ ) from  $N_{ant}$ ,  $N_{up}$  and  $\delta G_n$ , with fitting unbiased by fertilization treatment, trimming or genotype.

• Conclusions In untrimmed culms,  $\delta G_n$  had a negligible effect, so that  $N_{\text{rem}}$  could be fitted to  $N_{\text{ant}}$  and  $N_{\text{up}}$  only: grain N filling appeared to be determined by sources only ( $N_{\text{ant}}$  and  $N_{\text{up}}$ ), not by sink, and the reduction of  $N_{\text{rem}}$  by  $N_{\text{up}}$  was quantified. In these 'normal' cases, the regulation of  $N_{\text{rem}}$  should thus be located within the N sources themselves. In contrast, ear-trimming needs to be considered with caution as it introduced a sink limitation on  $N_{\text{rem}}$ ; moreover one with an important genotype effect.

**Key words:** *Triticum aestivum*, winter wheat, source/sink, grain filling, nitrogen uptake, grain number, nitrogen harvest index, nitrogen remobilization efficiency, genotype × environment.

# INTRODUCTION

Grain protein concentration (GPC) is one of the principal criteria listed in grain specifications for both the processing of wheat flour (*Triticum aestivum*) and the export of grain (Gooding *et al.*, 1994). In a world seeing an ever-growing demand for grain production, increasing grain nitrogen (N) yield is the only way to improve GPC. Thus many genotype and/or management research programs have been undertaken to improve N yield under a range of climatic conditions (Brancourt-Hulmel *et al.*, 2003, 2005). Environmental concerns also require optimized use of the two sources of grain N filling, i.e. (1) the remobilization of nitrogen ( $N_{\rm rem}$ ) already present in vegetative parts at anthesis ( $N_{\rm ant}$ ), and (2) post-anthesis nitrogen uptake ( $N_{\rm up}$ ; see Table 1 for a list of abreviations used in the text).

However, the factors that determine N fluxes during grain filling remain far from clear. According to Martre *et al.* (2003), grain N yield is mostly source-limited, while Barbottin *et al.* (2005) suggested that sink capacity was the main determinant for variations in  $N_{\rm rem}$ . In fact, it is difficult to distinguish between sink and source, as they are so strongly correlated that grain number can be accurately modelled using

Nant (Abbate et al., 1995; Oscarson, 2000; Demotes-Mainard and Jeuffroy, 2001). Some modellers have even suggested discarding the use of grain number, leading to sink-less models (Sinclair and Jamieson, 2006, 2008). As early as 1979, Martinez-Carrasco and Thorne used surgical ear trimming at around anthesis to disconnect the sink and source, and numerous subsequent papers have reported similar treatments. Both GPC and amount of N per grain always increased following trimming, reaching levels far higher than those observed in control plants, while N yield per ear generally declined, with marked genotype and treatment interactions. According to Ma et al. (1996), N yield was only maintained in some cultivars following moderate trimming (25 % of grains removed); however, trimming experiments did not provide clear evidence for a sink limitation of grain N filling. Indeed, several reports have suggested that as well as modifications to the sink, trimming could also induce a fall in  $N_{\rm rem}$  or  $N_{\rm up}$ , and thus cause a decrease in the source (Mi et al., 2000). Data on the quantitative N balance at the whole-plant level would therefore be welcome in trimming studies, but have seldom been reported.

ANNALS OF

BOTAN

The simplest way to modify the source/sink ratio during grain N filling may be to increase  $N_{up}$  by delaying the last application of fertilizer. This type of crop management has been widely employed in France during the past 10 years; nitrogen application rates are split into three parts so that the

© The Author 2009. Published by Oxford University Press on behalf of the Annals of Botany Company. All rights reserved. For Permissions, please email: journals.permissions@oxfordjournals.org

<sup>\*</sup> For correspondence. E-mail bancal@grignon.inra.fr

TABLE 1. Abbreviations used in the text

Nant	N amount in vegetative parts, including roots, at anthesis (after ear trimming, if performed)
$N_{\rm ant,i}$	N amount before ear trimming
$G_{\rm n}$	Ear grain number measured at the end of grain filling
$\delta G_n$	Difference between actual $G_n$ and its predicted value obtained from
	the genotype-dependant correlation to $N_{\text{ant,i}}$
$N_{\rm up}$	Nitrogen uptake by the culm after anthesis
N <sub>rem</sub>	Nitrogen remobilization from vegetative parts during grain filling
GPC	Grain protein concentration at the end of grain filling
NRE	Nitrogen remobilization efficiency (ratio of $N_{\text{rem}}$ to $N_{\text{ant}}$ )

last fertilization is delayed to around heading. However, fertilization cannot be delayed indefinitely, because the efficiency of N uptake rapidly declines after anthesis, possibly because of leaf senescence. The improved N yield of 'stay green' cultivars largely arises from their continued N uptake after anthesis. probably because of delayed leaf senescence (Borrell et al., 2001). Generally speaking, it appears that a higher level of N uptake after anthesis leads to delayed N decline in vegetative tissues (Martre et al., 2006). In an opinion paper, Barneix (2007) even suggested N uptake and remobilization might be mutually incompatible due to reciprocal inhibition: N remobilization would only - and irreversibly - start once N uptake had slowed down. This point of view is difficult to sustain at the whole-plant level, as early leaves have already senesced well before anthesis, at a time when N uptake is very high. Instead, it should be noted that  $N_{\rm rem}$  is actually a balance between N input and output from vegetative tissues, the former probably increasing in line with  $N_{\rm up}$  levels. Regarding the whole period of grain filling, late fertilizer applications have also been shown to reduce the efficiency of N remobilization (Gooding et al., 2007). Conversely, Triboï and Triboï-Blondel (2002) noted that plants with a lower  $N_{\text{ant}}$ , and hence a lower  $N_{\text{rem}}$  capacity, have a greater propensity to take up nitrogen after anthesis.

In fact, most of the evidence for antagonism between  $N_{\rm rem}$ and  $N_{\rm up}$  is based on variations in N concentrations in vegetative parts at the end of the grain filling period. When available, quantitative N balances at the whole-plant level tend to suggest that the losses in N yield due to antagonism between  $N_{up}$  and  $N_{\rm rem}$  are quite small. This may be due to the fact that under current crop management methods,  $N_{up}$  is much lower than N<sub>rem</sub> during grain filling (Van Sanford and MacKown, 1987); therefore any  $N_{up}$  antagonism to  $N_{rem}$  would remain moderate and can thus be ignored. This situation is favourable for farmers; however, it could be reversed, because current trends in both plant breeding and crop management are leading to an increase in post-anthesis N uptake. The antagonism between  $N_{\rm rem}$  and  $N_{\rm up}$  thus needs to be quantified under conditions that lead to higher  $N_{\rm up}/N_{\rm rem}$  ratios, because gains in N availability through an increase in  $N_{up}$  may be counteracted by a decrease in  $N_{\rm rem}$ ; in which case, the apparent efficiency of fertilizers would be reduced by increasing the amount of N wasted in straw.

This paper focuses on the regulation of  $N_{\rm rem}$  using two wheat cultivars that differ in terms of their grain N filling. A broad range in  $N_{\rm ant}/N_{\rm up}$  ratios was obtained by combining three levels of N fertilization before anthesis with two levels afterwards. The specific effect of grain number reduction through ear trimming was also studied. Analyses of variance and multiple correlations were used to explore the regulatory relationships between  $N_{\rm rem}$ ,  $N_{\rm up}$  and grain N filling.

# MATERIALS AND METHODS

# Culture and treatments

Two modern cultivars ('Apache' and 'Autan') of winter wheat (*Triticum aestivum* L.) were chosen because of their contrasting patterns of N metabolism. In preliminary field trials, 'Apache' always produced more ears per  $m^2$ , while grain numbers per ear (main culm and tiller mixed) were similar in the two genotypes. N uptake per  $m^2$  before anthesis was greater in 'Apache', but N uptake after anthesis was higher in 'Autan'. The mean grain weight, GPC and amount of N per grain were higher in 'Autan'.

Seeds were sown on 20 October 2004 in an experimental field at the INRA station in Thiverval-Grignon, France  $(48^{\circ}50'\text{N}; 1^{\circ}57'\text{E})$  at a density of 250 seeds m<sup>-2</sup>, under either high or no N fertilization (no-N). Highly fertilized plots were sown in a silt loam, a typical Eutrochrept soil (according to soil taxonomy), while no-N plots were sown in the poorest areas in a sandy embankment. On 25 March, plants in the plots had reached the beginning of stem elongation (growth stage = 31, according to Zadoks *et al.*, 1974). No-N plots exhibited a nitrogen nutrition index of 0.4(NNI; Justes et al., 1994), while the NNI of highly fertilized plots was 0.9. About 560 plants were carefully collected from each of the no-N plots and 280 plants from each of the high-N plots. Their roots were washed extensively before plants were transplanted into perlite-filled pots (two plants per 1.7-L pot) that were then placed outside. The average temperature was 11.5 °C during the next 2 months before anthesis and 17.4 °C during the 2 months after that. During pot culture prior to anthesis, one half of the plants transplanted from the no-N plot received 2.7 mg N per plant on a weekly basis; all other plants received 10.8 mg N per plant. Apart from nitrogen, all plants received the same full fertilization each week, and water when required depending on the weather. Three levels of early fertilization (applied before anthesis) were thus attained: some plants had received low fertilization (low in field, then low in pot), while others had received increasing fertilization (low in field, then high in pot), and others had received high fertilization (high in field, then high in pot).

Ear emergence was recorded for every main culm in order to select 84 pots bearing synchronous plants within each cultivar and early fertilization treatment group. After anthesis, one half of the selected plants received low fertilization, while the other half received high fertilization (1·8 and 7·2 mg N weekly per plant, respectively). Obviously, this latter fertilization could only be absorbed after anthesis; however, because the pots were not washed at anthesis, some of the fertilizer applied before anthesis may have been absorbed afterwards. Thus early and late fertilization could not be regarded as strictly equivalent to N uptake before and after anthesis, respectively. Conversely, N supply and N harvest were balanced over the whole season, suggesting that over this time scale (that of the study) no N leaching occurred from pots.

TABLE 2. Summary of the different genotypes and treatments(crossing of genotype, fertilization before/after anthesis and earreduction) used in the study. Treatment identities are used inFig. 3

Treatment identity	Genotype	Early fertilization	Late fertilization	Ear reduction
A	'Apache'	Low	Low	Control
В	'Apache'	Low	Low	50 % trimming
С	'Apache'	Low	High	Control
D	'Apache'	Low	High	50 % trimming
E	'Apache'	Increasing	Low	Control
F	'Apache'	Increasing	Low	50 % trimming
G	'Apache'	Increasing	High	Control
Н	'Apache'	Increasing	High	50 % trimming
Ι	'Apache'	High	Low	Control
J	'Apache'	High	Low	50 % trimming
Κ	'Apache'	High	High	Control
L	'Apache'	High	High	50 % trimming
М	'Autan'	Low	Low	Control
Ν	'Autan'	Low	Low	50 % trimming
0	'Autan'	Low	High	Control
Р	'Autan'	Low	High	50 % trimming
Q	'Autan'	Increasing	Low	Control
R	'Autan'	Increasing	Low	50 % trimming
S	'Autan'	Increasing	High	Control
Т	'Autan'	Increasing	High	50 % trimming
U	'Autan'	High	Low	Control
V	'Autan'	High	Low	50 % trimming
W	'Autan'	High	High	Control
Х	'Autan'	High	High	50 % trimming

Within 2 d of anthesis, the ears on one half of the plants were trimmed by removing all the odd-numbered spikelets. The experiment as a whole thus consisted 24 treatments with four factors: two genotypes; three early fertilizations; two late fertilizations; and two ear-trimming treatments (Table 2). This provided a fully cross-factor experimental set-up designed to analyse individual effects and interactions. Each treatment was applied to 24 pots, but hereafter only the first and last sampling times are considered, i.e. six pots per treatment, which were harvested as detailed below.

#### Sampling procedure

Three pots containing two plants each were harvested from each treatment at anthesis (17-26 May, depending on the treatment) and at physiological maturity (around 15 July). The roots and above-ground parts were collected, and within the above-ground parts the main culms were separated from tillers. Roots were recovered quantitatively, but they were usually fragmented and therefore could not be directly attributed to either the main culm or the tillers. Instead, the N ratio of the main culm to tillers was calculated for all above-ground parts of each sample, and used to split the corresponding root nitrogen between the main culm and tillers. The samples were dried for 48 h at 80 °C except for the ears, which were freeze-dried. The grains were then separated from the chaff. The spikelets removed from the trimmed ears were also reserved for further analysis. Tiller number and growth varied considerably within a treatment group. Moreover, under some treatments tillers appeared after anthesis and headed without elongation of their stem. The data concerning tillers should therefore be regarded as being of little significance; hereafter in this paper, only the main culm is considered.

#### Assessments of N balances

The dry weight of each sample was measured and then finely ground, and N concentrations were subsequently determined using Dumas' combustion method. N uptake  $(N_{up})$  and net remobilization from vegetative organs  $(N_{rem})$  were derived from Ruske *et al.* (2003) except that, unless specifically indicated, these equations also involved root N:

$$N_{up} = (\text{total N in culm at maturity} - \text{total N in culm at anthesis})$$
 (1)  
 $N_{rem} = (\text{vegetative N at anthesis} - \text{vegetative N at maturity})$  (2)

Hereafter in this paper N at anthesis  $(N_{ant})$  and grain number  $(G_n)$  both refer to the values obtained after ear trimming, in cases where this was done. In some specified cases, estimates prior to trimming  $(N_{i,ant} \text{ and } G_{i,n})$  are also used. The initial grain number  $(G_{i,n})$  in trimmed ears could be estimated as being twice the value of  $G_n$  recorded after trimming. Trimming also removed some chaff N, which was measured in corresponding samples. Nitrogen before trimming  $(N_{i,ant})$  was calculated by correcting  $N_{ant}$  for the nitrogen discarded in the spikelets that were removed.

#### Data analysis

Analyses of variance were performed using Statgraphics Plus (Manugistics, Inc., Rockville, MA) to examine the effects of different genotypes and treatments on various N remobilization parameters. The individual effects of genotype, early fertilization, late fertilization and ear trimming were analysed, as well as their first-level interactions. Statistically significant differences were then determined using the Newman–Keuls test with an overall error rate  $\alpha = 0.01$ .

The contributions of  $N_{ant}$ ,  $G_n$  and  $N_{up}$  to  $N_{rem}$  were analysed using simple and multiple linear regressions. Slopes and intercepts of the regression lines between genotypes were tested for significance and compared using the specific Statgraphics Plus procedure. Finally, analyses of variance were performed on the residuals of the regressions thus described, in order to detect any bias linked to either genotype, early fertilization, late fertilization or ear trimming.

## RESULTS

#### General features of treatments

The various genotypes and treatments resulted in broad variations in main-culm nitrogen at anthesis ( $N_{ant}$ ; Table 3). Analysis of variance indicated no significant difference in  $N_{ant}$  between genotypes under low, early fertilization (17 ± 3 mg culm<sup>-1</sup>; mean ± s.e.), while under increasing or high

TABLE 3. (a) Nitrogen variables in main culms for the different genotypes and treatments, as listed in Table 2. Anthesis N (N	(ant) is
the amount of N in vegetative parts (including roots) at anthesis (after ear trimming, in cases where this was done); ear	grain
number $(G_n)$ was obtained at the end of grain filling; N uptake $(N_{up})$ and remobilization $(N_{rem})$ were calculated from the diffe	rences
between data at anthesis and at the end of grain filling. (b) ANOVA was performed by fully crossing the effects of genotype,	, early
fertilization, late fertilization and ear reduction, as well as first-order interactions between the different factors	

(a) N variables						
	N <sub>ant</sub> (mg)	$G_{n}$	Grain N (mg)	N <sub>up</sub> (mg)	N <sub>rem</sub> (mg)	NRE
Genotype (G)						
'Apache'	30 <sup>a</sup>	38 <sup>b</sup>	$1.4^{\mathrm{a}}$	26 <sup>a</sup>	21 <sup>b</sup>	0.67 <sup>b</sup>
'Autan'	26 <sup>a</sup>	29 <sup>a</sup>	$1.5^{b}$	$28^{\rm a}$	15 <sup>a</sup>	0.56 <sup>a</sup>
Early fertilization	ц (E)					
Low	17 <sup>a</sup>	24 <sup>a</sup>	$1.4^{\mathrm{a}}$	24 <sup>a</sup>	$10^{\mathrm{a}}$	$0.57^{a}$
Increasing	27 <sup>b</sup>	33 <sup>b</sup>	$1.5^{\mathrm{a}}$	$30^{\mathrm{a}}$	17 <sup>b</sup>	0.61 <sup>a</sup>
High	$42^{\circ}$	$42^{c}$	$1.5^{\mathrm{a}}$	$27^{\mathrm{a}}$	$28^{\circ}$	0.66 <sup>b</sup>
Late fertilization	(L)					
Low	28 <sup>a</sup>	34 <sup>a</sup>	$1.3^{\mathrm{a}}$	23 <sup>a</sup>	19 <sup>a</sup>	$0.64^{a}$
High	28 <sup>a</sup>	33 <sup>a</sup>	$1.5^{b}$	31 <sup>b</sup>	17 <sup>a</sup>	0.58 <sup>a</sup>
Ear reduction (R)	1					
Control	30 <sup>a</sup>	44 <sup>b</sup>	$1 \cdot 2^{a}$	30 <sup>a</sup>	21 <sup>b</sup>	$0.68^{a}$
50 % trimming	27 <sup>a</sup>	22 <sup>a</sup>	$1.7^{b}$	24 <sup>a</sup>	16 <sup>a</sup>	0.55ª
(b) ANOVA						
	N <sub>ant</sub> (mg)	G <sub>n</sub>	Grain N (mg)	N <sub>up</sub> (mg)	N <sub>rem</sub> (mg)	NRI
G		***	**	ns	***	***
E	***	***	ns	ns	***	*
L	nd	ns	***	*	ns	*
R	ns	***	***		***	***
$G \times E$		***		ns	**	ns
$G \times L$	nd	ns	ns	ns	ns	ns
$G \times R$	ns	ns	**			**
$E \times L$	nd	ns	ns	ns	ns	ns
$E \times R$	ns	***	*	ns		ns
$L \times R$	nd	ns	ns	ns	ns	ns

In (a) different letters indicate a significant difference between means (P < 0.01; n = 72).

In (b) \* P < 0.01; \*\* P < 0.001; \*\*\* P < 0.0001; ns denotes non-significant effects (P > 0.05); nd indicates that the effect of late fertilization did not apply to  $N_{ant}$ .

fertilization,  $N_{\text{ant}}$  was higher in 'Apache' than in 'Autan' (up to  $48 \pm 5 \text{ vs. } 39 \pm 2 \text{ mg culm}^{-1}$ , respectively). Despite the fact that trimming discarded some chaff N ( $4 \pm 1 \text{ mg culm}^{-1}$ , averaged over all treatments),  $N_{\text{ant}}$  was not significantly lower in trimmed plants (P > 0.05).

Grain number  $(G_n)$  ranged from  $16.3 \pm 0.9$  in trimmed ears of 'Autan'following low, early fertilization to  $63.2 \pm 2.5$  in control ears of 'Apache' following high, early fertilization. The experimental conditions resulted in very large variations in both  $N_{ant}$  and  $G_n$ , but source and sink always remained correlated, except in the case of the trimming treatment. The overall correlation of  $N_{ant}$  to  $G_n$  was moderate, with  $r^2 =$ 0.46 for n = 72 (Fig.1A), whereas estimates of culm N and grain number before trimming ( $N_{ant,i}$  and  $G_{n,i}$ , respectively) were very strongly correlated (Fig. 1B), with the slope of the regression being significantly affected by genotype (P < 0.01):

$$G_{n,i} = (1.01 \pm 0.06) \times N_{ant,i} + (18 \pm 2);$$
  
 $r^2 = 0.90$  for  $n = 36$  in 'Apache' (3a)

$$G_{n,i} = (0.75 \pm 0.06) \times N_{\text{ant},i} + (17 \pm 2);$$
  
 $r^2 = 0.82$  for  $n = 36$  in 'Autan' (3b)

The amount of N per grain was significantly lower in 'Apache' than in 'Autan' (P < 0.001; Table 3), but a highly significant interaction in genotype × ear reduction (G × R; P < 0.001) was observed because this difference was no longer significant when trimming had been performed. In trimmed ears, the amount of N per grain increased up to  $1.7 \pm 0.1 \text{ mg grain}^{-1}$  in both genotypes. The amount of N per grain was increased by late fertilization (P < 0.0001), but not by early fertilization (P > 0.05) A significant interaction in early fertilization × ear reduction ( $E \times R$ ; P < 0.01), however, suggested that both increasing and high, early fertilization resulted in higher N amounts per grain, but only in trimmed ears.

N uptake after anthesis  $(N_{\rm up})$  ranged from  $12 \pm 2$  to as much as  $45 \pm 3$  mg culm<sup>-1</sup> under the A and W treatments (see Table 2), respectively (Table 3), while its ratio to final N

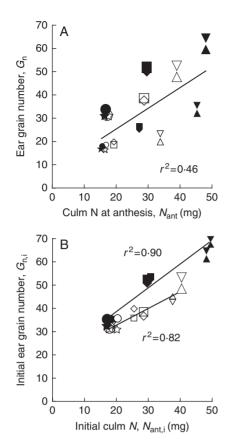


FIG. 1. Relationships between culm N at anthesis (including roots) and grain number. (A) Results after selected plants were trimmed at anthesis by removing one longitudinal half of the ear. (B) Genotype-dependant correlations between initial parameters before trimming. Each point is the mean of three pots containing two main culms. Closed symbols represent the genotype 'Apache' and open symbols the genotype 'Autan'; large symbols represent culms with whole ears and small symbols culms with trimmed ears. The various symbols refer to level of N fertilization before/after anthesis: low/ low, circles; low/high, stars; medium/low, squares; medium/high, diamonds; high/low, triangles; high/high, inverted triangles. In (A) the regression line and correlation coefficient apply to the whole data set (d.f. = 71); in (B) two correlations are presented for the genotypes 'Apache' (closed symbols) and 'Autan' (open symbols).

yield ranged from  $0.4 \pm 0.1$  to  $0.9 \pm 0.3$ . No significant effect of early fertilization on  $N_{\rm up}$  was observed (P > 0.05), while late fertilization clearly enhanced  $N_{\rm up}$  (P < 0.01). N uptake declined following ear trimming in 'Autan' ( $33 \pm 3$  vs.  $22 \pm 3 \text{ mg culm}^{-1}$ ), whereas it was not affected in 'Apache' ( $27 \pm 2 \text{ vs. } 26 \pm 2 \text{ mg culm}^{-1}$ ). N uptake did not correlate with either  $N_{\rm ant}$  or  $G_{\rm n}$  ( $r^2 < 0.12$  for n = 72).

The amount of N remobilized from vegetative parts after anthesis ( $N_{\rm rem}$ ) ranged from 7 to 36 mg culm<sup>-1</sup>, and exhibited highly significant effects of genotype, early fertilization and ear trimming (P < 0.0001); only the effect of late fertilization on  $N_{\rm rem}$  was not significant (P > 0.05). The average decrease in  $N_{\rm rem}$  due to trimming was  $3 \pm 1$  and  $7 \pm 2$  mg culm<sup>-1</sup> in 'Apache' and 'Autan', respectively. In 'Autan' therefore, but not in 'Apache', the decrease in  $N_{\rm rem}$  due to trimming exceeded the amount of N in discarded spikelets. A highly significant G × E interaction (P < 0.001) was observed because the response of  $N_{\rm rem}$  to early fertilization was stronger in 'Apache' than in 'Autan' (after low, increasing and high fertilization before anthesis, respectively:  $10 \pm 1$ ,  $19 \pm 1$  and  $34 \pm 2$  mg culm<sup>-1</sup> in 'Apache' vs.  $9 \pm 1$ ;  $15 \pm 3$  and  $22 \pm 2$ mg culm<sup>-1</sup> in 'Autan').

Lastly, N remobilization efficiency (NRE: ratio of  $N_{\rm rem}$  to  $N_{\rm ant}$ ; Table 3) ranged from  $0.28 \pm 0.13$  to  $0.76 \pm 0.02$ , and varied highly significantly with genotype and ear trimming (P < 0.0001). Indeed, highly significant G × R interactions (P < 0.001) were observed because trimming clearly reduced NRE in 'Autan', but hardly at all in 'Apache'. High, early fertilization increased NRE when compared to low, early fertilization (P < 0.01), whereas high, late fertilization decreased NRE (P < 0.01).

#### Determining N remobilization from vegetative parts

Figure 2A indicates that  $N_{\text{rem}}$  was very strongly correlated to  $N_{\text{ant}}$  ( $r^2 = 0.91$ ; n = 72), with an intercept significantly different from zero. Analysis of variance of the residuals of this regression indicated that it was biased by genotype (P < 0.0001), early (P < 0.01) and late (P < 0.001) fertilization,

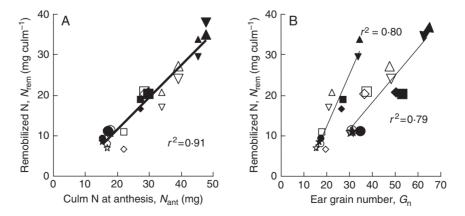


FIG. 2. Relationship between the amount of remobilized nitrogen from vegetative parts, including roots ( $N_{rem}$ ) and (A) nitrogen in vegetative parts at anthesis, after trimming ( $N_{ant}$ ), and (B) actual grain number ( $G_n$ ). Each point is the mean of three pots containing two main culms. Symbols represent the various genotypes and treatments as in Fig.1. In (A) the regression line and correlation coefficient apply to the whole data set (d.f. = 71), while in (B) two correlations are given for whole ears (large symbols) and trimmed ears (small symbols).

as well as by ear trimming (P < 0.0001). Quite clearly,  $N_{\text{ant}}$  is a major determinant of  $N_{\text{rem}}$ , but it is not alone. Grain number was also well correlated with  $N_{\text{rem}}$  (Fig. 2B), but a comparison of the regression lines indicates they were markedly affected by trimming treatment (P < 0.001); the two regression lines are shown in Fig. 2, indicating  $r^2$  at 0.79 and 0.80 (n = 36) for control and trimmed culms, respectively. However, analysis of variance of the residuals indicated that this trimmingdependent correlation was in turn biased by genotype (P < 0.01) and early fertilization (P < 0.001), but not by late fertilization (P > 0.05). Lastly, despite very marked variations in  $N_{\text{up}}$ , within the same range as for  $N_{\text{ant}}$ , no correlation was observed when  $N_{\text{up}}$  was plotted against  $N_{\text{rem}}$  ( $r^2 = 0.03$ , n =72, data not shown).

#### Multiple regression analysis of N remobilization

 $N_{\text{rem}}$  was linearly correlated with the main determinants identified above, leading to the following multiple regression ( $r^2 = 0.94$  for n = 72):

$$N_{\rm rem} = (0.70 \pm 0.04) N_{\rm ant} + (0.15 \pm 0.03) G_{\rm n} - (0.07 \pm 0.03) N_{\rm up} - (4.7 \pm 1.0)$$
(4)

The coefficients for  $N_{\text{ant}}$  and  $G_n$  were positive and highly significant (P < 0.0001), but the negative coefficient for  $N_{\text{up}}$  was much less significant (P > 0.01). Lastly, the negative intercept of the relationship was clearly different from zero (P < 0.0001). Such a correlation should be regarded with caution, however, as  $N_{\text{ant}}$  and  $G_n$  were far from being independent from each other. Moreover, the correlation exhibited bias for genotype and late fertilization (P < 0.01), as well as a G × R interaction (P < 0.0001): eqn (4) under-estimated  $N_{\text{rem}}$  in trimmed 'Apache' by  $-1.7 \pm 0.2 \text{ mg culm}^{-1}$  and overestimated  $N_{\text{rem}}$  in trimmed 'Autan' by  $1.8 \pm 0.1 \text{ mg culm}^{-1}$ . In addition, the regression did not discriminate between genotypes despite the fact that their reaction to trimming was clearly different.

To obtain a sink variable that was less correlated to  $N_{\text{ant}}$  than  $G_n$ , the difference  $\delta G_n$  was then calculated between  $G_n$  and grain number, which could be predicted from  $N_{\text{ant,i}}$  using eqn (3). Using  $\delta G_n$  instead of  $G_n$ , a new multiple regression analysis of  $N_{\text{rem}}$  was obtained (eqns 5a, b), which had a high level of significance ( $r^2 = 0.95$  for n = 72) and a constant that did not differ significantly from zero (P > 0.05), provided the slope for  $\delta G_n$  was kept genotype-dependent:

$$N_{\rm rem} = (0.80 \pm 0.02) N_{\rm ant} + (0.05 \pm 0.02) \delta G_{\rm n} - (0.11 \pm 0.02) N_{\rm up} \text{ in 'Apache'}$$
(5a)

$$N_{\rm rem} = (0.80 \pm 0.02) N_{\rm ant} + (0.28 \pm 0.03) \delta G_{\rm n} - (0.11 \pm 0.02) N_{\rm up} \text{ in 'Autan'}$$
(5b)

The coefficients for  $N_{\text{ant}}$  and  $N_{\text{up}}$ , as well as that for  $\delta G_n$  in 'Autan', were highly significant (P < 0.0001), but this was not the case for  $\delta G_n$  in 'Apache' (P > 0.01). The only significant bias (P < 0.01) was observed for trimmed culms: in these plants,  $N_{\text{rem}}$  was under-estimated by  $-1.0 \pm 0.5$  mg culm<sup>-1</sup>

under low, late fertilization and over-estimated by 1.2 + $0.5 \text{ mg culm}^{-1}$  under high, late fertilization. The genotype effect on the coefficient for  $\delta G_n$  suggested that there was almost no sink limitation in 'Apache', in contrast to 'Autan'; this is indicated in Fig. 3, which shows the respective contributions of the terms in eqn (5) to  $N_{\rm rem}$ . As the root-mean-square error (RMSE) of the fit was low  $(2.1 \text{ mg culm}^{-1})$ , the residuals of  $N_{\text{rem}}$  in its estimation by eqns (5a) and (5b) were within 8 % of the actual  $N_{\rm rem}$  in all but one case (exhibiting a very low  $N_{\rm rem}$ ). The major role of  $N_{\text{ant}}$  in determining  $N_{\text{rem}}$  was clear, while  $N_{\text{up}}$  led to a decrease in  $N_{\text{rem}}$  from -1 to -5 mg culm<sup>-1</sup>, generally accounting for between -10 % and -30 % of  $N_{\text{rem}}$ . Lastly, the influence of  $\delta G_n$  was markedly genotype-dependent in trimmed culms: the corresponding loss was  $-8 \pm 1 \% N_{\text{rem}}$  in 'Apache' vs.  $-60 \pm 9\%$  N<sub>rem</sub> in 'Autan'. In contrast, the influence of  $\delta G_{\rm n}$  was very small in untrimmed plants ( $\pm 0.5 \, {\rm mg \, culm^{-1}}$ , less than 3 %  $N_{\rm rem}$ ), and in this case the multiple regression became genotype-independent, as the coefficient of  $\delta G_n$  did not significantly differ from zero for both genotypes (P >0.05), leading to the following, simpler equation  $(r^2 = 0.99)$ for n = 36; RMSE = 1.6 mg culm<sup>-1</sup>):

$$N_{\rm rem} = (0.77 \pm 0.02)N_{\rm ant} - (0.08 \pm 0.02)N_{\rm up}$$
(6)

The coefficients for  $N_{\text{ant}}$  and  $N_{\text{up}}$  were then highly significant (P < 0.0001 and P < 0.001, respectively), without significant bias for either genotype or fertilization treatment, as suggested by analysis of variance for the correlation residuals (data not shown).

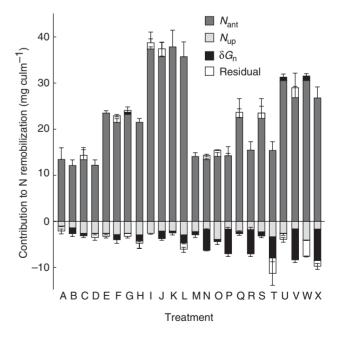


FIG. 3. Contribution to N remobilization in main culm  $(N_{rem})$  for the various treatments as detailed in Table 2. The various terms used in eqn (5) are shown, as well as the residuals of the multiple correlation to  $N_{rem}$  of  $N_{ant}$ ,  $N_{up}$  and  $\delta G_n$ .  $N_{ant}$  is the amount of N in vegetative parts at anthesis (after trimming),  $N_{up}$  is the N uptake after anthesis, and  $\delta G_n$  refers to the difference between  $G_n$  and grain number as predicted from  $N_{ant,i}$  (before trimming) by eqn (3). Bars indicate the s.e.m. for three replicates.

#### Bancal — Nitrogen remobilization in wheat

TABLE 4. (a) Importance of taking roots into account for the assessment of anthesis N, post-anthesis N uptake and N remobilization. Data are expressed as either mg per above-ground parts of the culm or relative to the results obtained using the whole culm. Data in this table can be compared with those in Table 3, which were obtained taking roots into account. (b) ANOVA was performed by fully crossing the effects of genotype, early fertilization, late fertilization and ear reduction, as well as first-order interactions between the different factors

	Above-ground anthesis N		Above-ground N increase		Above-ground N remobilization	
	Total (mg)	Ratio to whole culm	Total (mg)	Ratio to whole culm	Total (mg)	Ratio to whole culr
Genotype (G)						
'Apache'	26 <sup>a</sup>	$0.84^{\mathrm{a}}$	30 <sup>a</sup>	1.13 <sup>b</sup>	18 <sup>b</sup>	$0.84^{a}$
'Autan'	23 <sup>a</sup>	$0.86^{\mathrm{a}}$	30 <sup>a</sup>	$1 \cdot 10^{a}$	13 <sup>a</sup>	$0.85^{a}$
Early fertilization (H	E)					
Low	14 <sup>a</sup>	0.83ª	25 <sup>a</sup>	$1.07^{\mathrm{a}}$	8 <sup>a</sup>	0.83 <sup>a</sup>
Increasing	23 <sup>b</sup>	$0.85^{\mathrm{a}}$	32 <sup>a</sup>	$1.08^{a}$	14 <sup>b</sup>	$0.86^{a}$
High	35°	$0.85^{\mathrm{a}}$	32 <sup>a</sup>	1.18 <sup>b</sup>	24 <sup>c</sup>	$0.84^{a}$
Late fertilization (L)	)					
Low	24 <sup>a</sup>	$0.85^{\mathrm{a}}$	26 <sup>a</sup>	1.13 <sup>a</sup>	16 <sup>a</sup>	$0.85^{a}$
High	24 <sup>a</sup>	$0.85^{\mathrm{a}}$	33 <sup>b</sup>	$1.01^{a}$	15 <sup>a</sup>	$0.85^{a}$
Ear reduction (R)						
Control	$26^{a}$	$0.85^{\mathrm{a}}$	33 <sup>a</sup>	$1 \cdot 10^{\mathrm{a}}$	18 <sup>b</sup>	$0.86^{a}$
50 % trimming	23 <sup>a</sup>	$0.84^{\mathrm{a}}$	26 <sup>a</sup>	$1 \cdot 12^{\mathrm{a}}$	13 <sup>a</sup>	0.83 <sup>a</sup>

	Above-ground anthesis N		Above-ground N increase		Above-ground N remobilization	
	Total (mg)	Ratio to whole culm	Total (mg)	Ratio to whole culm	Total (mg)	Ratio to whole culm
G	_	_	ns	*	***	ns
Е	***	ns	_	***	***	ns
L	nd	nd	*	_	ns	ns
R	_	ns	*	ns	***	_
$\mathbf{G} \times \mathbf{E}$	_	ns	ns	ns	**	ns
$G \times L$	nd	nd	ns	ns	ns	ns
$\mathbf{G} \times \mathbf{R}$	ns	ns	_	ns	_	ns
$E \times L$	nd	nd	ns	ns	ns	ns
$\mathbf{E} \times \mathbf{R}$	ns	ns	ns	ns	ns	_
$L \times R$	nd	nd	ns	ns	ns	ns

In (a) different letters indicate a significant difference between means (P < 0.01; n = 72).

In (b) \* P < 0.01; \*\* P < 0.001; \*\*\* P < 0.0001; ns denotes non-significant effects (P > 0.05); nd indicates that the effect of late fertilization did not apply to  $N_{ant}$ .

#### Predictions of remobilization without root measurements

All the data presented so far in this paper have involved the quantification of root N using the experimental design described. However, root sampling is impossible in field assays, and in such cases  $N_{\text{ant}}$ ,  $N_{\text{up}}$  and  $N_{\text{rem}}$  are commonly estimated using  $N_{\text{ant,a}}$ ,  $N_{\text{up,a}}$  and  $N_{\text{rem,a}}$ , obtained by applying eqns (1) and (2) to above-ground parts rather than the whole culm. The values of  $N_{ant,a}$ ,  $N_{up,a}$  and  $N_{rem,a}$  were affected by genotype, fertilization and ear trimming, in a way similar to  $N_{\rm ant}$ ,  $N_{\rm up}$  and  $N_{\rm rem}$ , respectively (Table 4). Both  $N_{\rm ant,a}/N_{\rm ant}$ and  $N_{\rm rem,a}/N_{\rm rem}$  ratios averaged 0.85, with variations that were not significantly linked to genotype, fertilization or ear trimming. NRE and its estimation using the  $N_{\rm rem,a}/N_{\rm ant,a}$  ratio did not differ significantly according to a paired sample comparison (P > 0.05). The value of  $N_{up,a}$  (Table 4) considered remobilization from the roots as an N uptake, and therefore over-estimated  $N_{\rm up}$  by between 1 % and 27 %, depending on genotype (P < 0.01) and early fertilization (P < 0.0001), but not on late fertilization (P > 0.01) or ear trimming (P > 0.01)

0.05). Therefore,  $N_{\rm up}$  could not be precisely evaluated from  $N_{\rm up,a}$  measurements in above-ground parts only. Nevertheless, a direct estimate of  $N_{\rm rem}$  in untrimmed culms was obtained from  $N_{\rm ant,a}$  and  $N_{\rm up,a}$  ( $r^2 = 0.97$  for n = 36; RMSE = 1.8 mg culm<sup>-1</sup>):

$$N_{\rm rem} = (0.91 \pm 0.03) N_{\rm ant,a} - (0.07 \pm 0.02) N_{\rm up,a}$$
(7)

Despite it being less relevant from a physiological point of view, the following regression (eqn 8), also restricted to untrimmed culms, could be more easily tested in a field experiment and even compared with previous studies ( $r^2 = 0.97$  for n = 36; RMSE = 1.3 mg culm<sup>-1</sup>):

$$N_{\rm rem,a} = (0.78 \pm 0.02) N_{\rm ant,a} - (0.06 \pm 0.02) N_{\rm up,a} \quad (8)$$

In both eqns (7) and (8), the coefficients for  $N_{\text{ant,a}}$  and  $N_{\text{up,a}}$  were both highly significant (P < 0.0001 and P < 0.001, respectively), without any significant bias regarding genotype,

fertilization treatment or interactions, as suggested by analysis of variance of the regression residuals. It appeared that eqn (8) was indistinguishable from eqn (6), except that it referred to above-ground values.

## DISCUSSION

#### Representativeness of NRE

This experiment was designed to provide for very large  $N_{up}$  by inducing N deficiencies before anthesis (Triboï and Triboï-Blondel, 2002), so that the ratio of  $N_{\rm up}$  to grain N ranged from 0.4 to 0.9 while the values attained in crops are commonly below 0.5 (Van Sanford and MacKown, 1987). However, despite exploring unusual ranges for the  $N_{\rm up}/N_{\rm rem}$ ratio, the usual values for  $N_{\rm ant}$  and  $N_{\rm rem}$  were also represented in the study. Previous field experiments have reported higher values for NRE (0.4 to 0.9) than those found during the present study (0.3 to 0.8). Kichey et al. (2007) suggested a genotype effect on NRE, but Barbottin et al. (2005) noted that the genotype actually interacted with the year and level of fertilization. For this reason, the low NRE levels obtained in the present study could hardly be explained by the choice of cultivars. Unlike field experiments, this study also took roots into account, which are a major N sink according to Andersson et al. (2004). However, despite the fact that root N varied considerably from 7 % to 27 % of N in vegetative parts (depending on the genotype and treatment), the trends observed in whole culms were essentially maintained when only above-ground parts were examined. The sampling with roots therefore may not provide an explanation for the discrepancy between the results of this study and those in the literature.

The absence of high NRE values may have originated from the use of measurements on the scale of the culm, rather than the m<sup>2</sup> scale as reported in the literature. The relationships between  $N_{\text{ant}}$  and  $N_{\text{rem}}$  appeared to be affected by the scale considered. At the m<sup>2</sup> scale, the simple linear regression of  $N_{\rm ant}$  to  $N_{\rm rem}$  shows a positive intercept (e.g. in Barbottin et al., 2005). Consequently a higher  $N_{ant}$  thus mathematically led to a higher NRE, which may have occurred when different levels of fertilization prior to anthesis were compared. The higher the fertilization, the higher was  $N_{\text{ant}}$  and the lower was the NRE, as previously noted by Cox et al. (1986). In contrast, the data in this paper suggest that the simple linear regression of  $N_{\text{ant}}$  to  $N_{\text{rem}}$  displayed a negative intercept at the culm scale. Therefore a higher  $N_{\text{ant}}$  thus mathematically led to a lower NRE, leading to an effect of early fertilization in contradiction to findings in the literature (Table 3). In fact, Fig. 2A indicates that  $N_{\text{ant}}$  and  $N_{\text{rem}}$  are aligned in the overall correlation; thus the variation in NRE was only based on the negative intercept of the relationship between  $N_{\rm ant}$ and  $N_{\rm rem}$ . It is possible to imagine that a certain level of N will be immobilized in the dead tissues of a single culm by the time that anthesis occurs. This would result in the negative intercept observed at the culm level. At a later stage, grain N filling would lead to N remobilization from senescing (but still alive) plant organs. The present data suggest that the corresponding slope, which could be termed the 'physiological NRE', was unaffected by fertilization. The ratio of  $N_{\rm rem}$  to  $N_{\rm ant}$ , which could be termed the 'culm NRE', was, however, biased by the intercept of the correlation, and increased with fertilization. At the  $m^2$  level, the relationships between  $N_{ant}$  and  $N_{rem}$  become still more complicated because of tillering, resulting in a positive intercept and a 'crop NRE' that decreases with fertilization. Therefore, the use of the NRE could lead to confusion.

## N<sub>rem</sub> was essentially source-determined

The principal determinant for  $N_{\rm rem}$  was by far  $N_{\rm ant}$ , with slopes in different equations of around 0.75-0.80. This result agreed well with that of Barbottin et al. (2005), who reported that, over a very broad range of  $N_{\rm ant}$ , the slope of  $N_{\rm rem}$  vs.  $N_{\rm ant}$  was 0.76 under simple regressions regardless of genotype, provided that neither important fertilization was applied at anthesis nor that stresses occurred thereafter. The data also indicated a weaker correlation between  $N_{\rm rem}$  and  $G_{\rm n}$ , which could be associated with the link between  $N_{\rm ant}$ and  $G_{n}$ . Studies in the literature have reported a correlation between  $N_{\text{ant}}$  and  $G_{\text{n}}$  at the m<sup>2</sup> level, although this was mostly linked to the degree of tillering. This correlation was also observed at the main-culm level, where it was genotypedependent: 'Autan' produced less grain than 'Apache', even for the same  $N_{\text{ant}}$ . This suggests that the plants themselves somehow regulate their sink capacity  $(G_n)$  to their source level  $(N_{ant})$  around anthesis (Sinclair and Jamieson, 2006). The use of the  $\delta G_n$  difference rather than  $G_n$  to characterize the sink led to a reduction in the sensitivity of  $N_{\rm rem}$  determinism to both sink and genotype; in untrimmed culms their effects became negligible. The absence of a constant in eqns (6)-(8), which could therefore probably be extended to the m<sup>2</sup> level, indicated that  $N_{\text{rem}}$  was fully predicted using source data only ( $N_{\text{ant}}$  and  $N_{\text{up}}$ ). However, it did not mean that sink and genotype had no influence on grain N filling, but rather that the sink influence was taken into account by the genotype-dependent relationship through  $G_n$  and  $N_{ant}$ . Any event disturbing this relationship, either before or after anthesis, could lead to reintroduction of both the genotype and sink effects, for instance when disease or water stress occurr during grain filling, as was observed by Barbottin et al. (2005). Sink and genotype effects also appeared in the response to ear halving, but multiple regressions suggested that the source remained the principal determinant of  $N_{\rm rem}$ , although modulated by sink and genotype. On the other hand, in many cases explicitly accounting for the sink may be redundant, as Jamieson and Semenov (2000) showed that it is simpler to fit grain N filling to sources than to sinks. If regulation somehow occurs, it should be located in sources themselves; according to Hörtensteiner and Feller (2002) and Gregersen and Holm (2007) vegetative parts seem to regulate  $N_{\rm rem}$  themselves by means of a generic senescence program. Nevertheless, despite numerous models in the literature that have described grain N filling regulation by source/sink interactions, to my knowledge no groups have yet described how this regulation could be attained otherwise.

## Sink effects in trimmed ears

Ear trimming always resulted in an increase in the amount of N per grain, even though it reduced N yield per ear in most cases. As early as 1991, Jenner et al. suggested that the relationship between source availability and sink activity shifts gradually as a function of the range of the source/sink ratio. If this ratio is low, grain filling would be proportional to source activity, approaching a constant-rate, saturated pattern with higher source availability. These authors also suggested that the dry-matter filling of grain is similar to a saturated pattern, while the grain N filling of whole ears tends to be within the range of a proportional pattern. Ear trimming would thus shift grain N filling to the range of the source/ sink ratio leading to a saturated pattern. Bancal and Soltani (2002) modelled this type of mixed source and sink determinism, but only in the case of partitioning between several sinks. In fact, grain N filling is too frequently considered to be a single-sink task, whereas <sup>15</sup>N labelling studies have indicated that N absorbed after anthesis may actually be incorporated, and not just temporarily stored, in vegetative parts (Oscarson, 1996; Kichey et al., 2007), thus suggesting that vegetative parts also behave as a sink for newly absorbed N. The model described by Bancal and Soltani (2002) used at least two parameters per sink that mostly affected sink activity at either low or high source/sink ratios, respectively. These parameters might vary according to genotype, which could also explain the genotype effect on trimming response suggested by the genotype-dependent coefficient for  $\delta G_n$ . According to Martre et al. (2003), genotypes exhibiting a larger amount of N per grain in untrimmed ears reached their saturation level earlier following trimming, which was also observed during this study in 'Autan' vs. 'Apache'. These hypotheses would not preclude alternative models explaining a trimming effect due to carbon (Dingkuhn et al., 2007) or hormones (Yang, et al., 2003). These puzzling results obtained for trimmed ears should be extended to control plants only with caution.

## Negative effect of N<sub>up</sub> on N<sub>rem</sub>

The negative effect of late fertilization on NRE has long been recognised (Gooding *et al.*, 2007); however, labelling studies have suggested that remobilization is not actually inhibited but counterbalanced by the incorporation of newly absorbed N in vegetative tissues. The harvest index of N absorbed after anthesis is lower than 100 %. Using <sup>15</sup>N fertilizers, Kichey *et al.* (2007) reported that 89.7 % to 93.4 % of this N was translocated to grains, while under eqn (6) of this study the difference from 1 for the slope of  $N_{\rm rem}$  to  $N_{\rm up}$ suggested that  $92 \pm 2$  % of N absorbed after anthesis was translocated to grains in untrimmed ears. Therefore, the unusually high ratios of  $N_{\rm up}$  to grain N did not diminish translocation efficiency. Current trends towards late fertilization or plant breeding to ensure continued N uptake would therefore not result in high levels of N waste in straw.

#### Conclusions

In untrimmed ears, N yield could be predicted accurately without data for grain number. Sink and genotype effects were only modulations of the major regulation by both sources  $N_{\text{ant}}$  and  $N_{\text{up}}$ , and moreover they only appeared in response to severe stresses (such as ear halving). The results

obtained on trimmed ears thus differed qualitatively from the others and should therefore be extended to control plants with caution.  $N_{\rm rem}$  was positively correlated with  $N_{\rm ant}$  and negatively with  $N_{\rm up}$ , but even with a very high  $N_{\rm up}$  the negative impact of  $N_{\rm up}$  on  $N_{\rm rem}$  remained small. It will therefore continue to be useful to investigate how the uptake of late nitrogen fertilization can be increased.

# ACKNOWLEDGEMENTS

This study was financed by Génoplante [B04 project]. My thanks go to M. Chapon for the management of plant cultures, sampling and measurements.

## LITERATURE CITED

- Abbate PE, Andrade FH, Culot JP. 1995. The effects of radiation and nitrogen on number of grains in wheat. *Journal of Agricultural Science* 124: 351–360.
- Andersson A, Johanson E, Oscarson P. 2004. Nitrogen redistribution from the roots in post-anthesis plants of spring wheat. *Plant and Soil* 264: 321–332.
- Bancal P, Soltani F. 2002. Source-sink partitioning. Do we need Münch? Journal of Experimental Botany 53: 1919–1928.
- Barbottin A, Lecompte C, Bouchard C, Jeuffroy MH. 2005. Nitrogen remobilization during grain filling in wheat: genotypic and environmental effects. *Crop Science* **45**: 1141–1150.
- Barneix AJ. 2007. Physiology and biochemistry of source-regulated protein accumulation in the wheat grain. *Journal of Plant Physiology* 164: 581–590.
- Brancourt-Hulmel M, Doussinault G, Lecomte C, Bérard P, LeBuanec B, Trottet M. 2003. Genetic improvement of agronomic traits of winter wheat cultivars released in France from 1946 to 1992. *Crop Science* 43: 37–45.
- Brancourt-Hulmel M, Heumez E, Pluchard P, et al. 2005. Indirect versus direct selection of winter wheat for low input or high input levels. *Crop Science* **45**: 1427–1431.
- Borrell AK, Hammer GL, van Oosterom EJ. 2001. Stay-green: a consequence of the balance between supply and demand for nitrogen during grain filling ? Annals of Applied Biology 138: 81–95.
- Cox MC, Qualset CO, Rains DW. 1986. Genetic variation for nitrogen assimilation and translocation in wheat. III. Nitrogen translocation in relation to grain-yield and protein. *Crop Science* 26: 737–740.
- **Demotes-Mainard S, Jeuffroy MH. 2001.** Incorporating radiation and nitrogen nutrition into a model of kernel number in wheat. *Crop Science* **41**: 415–423.
- Dingkuhn M, Luquet D, Clément-Vidal A, Tambour L, Kim HK, Song YH. 2007. Is plant growth driven by sink regulation? Implications for crop models, phenotyping approaches and ideotypes. *Wageningen U.R. Frontis* 21: 155–168.
- Gooding MJ, Smith SP, Davies WP, Kettlewell PS. 1994. Effects of lateseason applications of propiconazole and tridemorph on disease, senescence, grain development and the breadmaking quality of winter wheat. *Crop Protection* 13: 362–370.
- Gooding MJ, Gregory PJ, Ford KE, Ruske RE. 2007. Recovery of nitrogen from different sources following applications to winter wheat at and after anthesis. *Field Crops Research* 100: 143–154.
- Gregersen PL, Holm PB. 2007. Transcriptome analysis of senescence in the flag leaf of wheat (*Triticum aestivum* L.). *Plant Biotechnology Journal* 5: 192–206.
- Hörtensteiner S, Feller U. 2002. Nitrogen metabolism and remobilization during senescence. *Journal of Experimental Botany* 53: 927–937.
- Jamieson PD, Semenov MA. 2000. Modelling nitrogen uptake and redistribution in wheat. *Field Crops Research* 68: 21–29.
- Jenner CF, Ugalde TD, Aspinall D. 1991. The physiology of starch and protein deposition in the endosperm of wheat. Australian Journal of Plant Physiology 18: 221–226.

- Justes E, Mary B, Meynard JM, Machet JM, Thelier-Huche L. 1994. Determination of a critical nitrogen dilution curve for winter wheat crops. Annals of Botany 74: 397–407.
- Kichey T, Hirel B, Heumez E, Dubois F, Le Gouis J. 2007. In winter wheat (*Triticum aestivum* L.), post-anthesis nitrogen uptake and remobilization to the grain correlates with agronomic and nitrogen physiological markers. *Field Crops Research* 102: 22–32.
- Ma YZ, MacKown CT, Van Sanford DA. 1996. Differential effects of partial spikelet removal and defoliation on kernel growth and assimilate partitioning among wheat cultivars. *Field Crops Research* 47: 201–209.
- Martinez-Carrasco R, Thorne GN. 1979. Physiological factors limiting grain size in wheat. *Journal of Experimental Botany* 30: 669–679.
- Martre P, Porter JR, Jamieson PD, Triboï E. 2003. Modeling grain nitrogen accumulation and protein composition to understand sink/source regulations of nitrogen remobilization for wheat. *Plant Physiology* 133: 1959–1967.
- Martre P, Jamieson PD, Semenov MA, Zyskowski RF, Porter JR, Triboï E. 2006. Modelling protein content and composition in relation to crop nitrogen dynamics for wheat. *European Journal of Agronomy* 25: 138–154.
- Mi GH, Tang L, Zhang FS, Zhang JH. 2000. Is nitrogen uptake after anthesis in wheat regulated by sink size? *Field Crops Research* 68: 183–190.
- **Oscarson P. 1996.** Transport of recently assimilated <sup>15</sup>N nitrogen to individual spikelets in spring wheat grown in culture solution. *Annals of Botany* **78**: 479–488.

- Oscarson P. 2000. The strategy of the wheat plant in acclimating growth and grain production to nitrogen availability. *Journal of Experimental Botany* 51: 1921–1929.
- Ruske RE, Gooding MJ, Jones SA. 2003. The effects of triazole and strobilurin fungicide on nitrogen uptake, partitioning, remobilization and grain N accumulation in winter wheat cultivars. *Journal of Agricultural Science* 140: 395–407.
- Sinclair TR, Jamieson PD. 2006. Grain number, wheat yield, and bottling beer: an analysis. *Field Crops Research* 98: 60–67.
- Sinclair TR, Jamieson PD. 2008. Yield and grain number of wheat: a correlation or causal relationship? Authors' response to 'The importance of grain or kernel number in wheat: a reply to Sinclair and Jamieson' by R.A. Fischer. *Field Crops Research* 105: 22–26.
- Triboï E, Triboï-Blondel A-M. 2002. Productivity and grain or seed composition: a new approach to an old problem. *European Journal of Agronomy* 16: 163–186.
- Van Sanford DA, MacKown CT. 1987. Cultivar differences in nitrogen remobilization during grain fill in soft red winter wheat. *Crop Science* 27: 295–300.
- Yang JC, Zhang JH, Wang ZQ, Zhu QS, Liu LL. 2003. Involvement of abscisic acid and cytokinins in the senescence of carbon reserves in wheat subjected to water stress during grain filling. *Plant, Cell and Environment* 26: 1621–1631.
- Zadoks JC, Chang TT, Konzak CF. 1974. A decimal code for the growth stages of cereals. Weed Research 44: 415–421.