

# NIH Public Access

**Author Manuscript**

*Semin Immunol*. Author manuscript; available in PMC 2010 June 1.

#### Published in final edited form as:

*Semin Immunol*. 2009 June ; 21(3): 164–171. doi:10.1016/j.smim.2009.03.001.

## **Barrier immunity and IL-17**

## **Benjamin R. Marks**1 and **Joe Craft**1,2

1*Department of Immunobiology, Yale School of Medicine, New Haven, CT 06520, USA*

2*Section of Rheumatology, Department of Internal Medicine, Yale School of Medicine, New Haven, CT 06520, USA*

## **Abstract**

 $CD4+T_H17$  cells display a featured role in barrier immunity. This effector population of T cells is important for clearance of microorganisms but can also promote autoimmunity at barrier sites. Recent work has indicated that these effector cells share a pathway with  $CD4^+$  regulatory T cells (T<sub>R</sub> cells) that also have a critical function in barrier protection and immune regulation. The development and function of  $T_H$ 17 cells, and their relationship with  $T_R$  cells are discussed.

#### **Keywords**

effector T cells; IL-17; mucosa; regulatory T cells;  $T_H$ 17 cells

## **Introduction**

Mucosal and epithelial surfaces harbor a significant number of immune cells that are necessary to provide host protection at these susceptible sites for pathogen entry. These surfaces, including the gastrointestinal tract, lungs, skin and reproductive tract, present a particularly difficult and precarious scenario for the cells of the immune system. Unlike the sterile environment of the systemic circulation, the blood and the lymphatics, the epithelial layer is bathed in a sea of microorganisms. The evolution of the immune system has occurred so that a symbiotic relationship exists between the commensal bacteria of the gut and human hosts. Teleologically one may predict that the epithelial barriers are constructed so that microorganisms are sequestered outside of a tight barrier so that immune response to resident bacteria is rare. However, more recent data presents the opposite story and indicates that the gut microflora are a critical component to proper immune function and in their absence, the immune system has inadequate development often leading to immune dysregulation. Thus, the immune system that has co-evolved in the setting of diverse microorganisms that are beneficial and potentially harmful to the host has developed a specialized approach to addressing the complex nature of the barrier layer between host and the outside world.

## **The phenotype of T cells at the barrier surfaces**

Given the unique conditions existing at the body's interface with the external environment, the cells of the immune system at these sites operate differently than those in the lymphoid organs.

Contact: Joe Craft @ E-mail: joseph.craft@yale.edu, Telephone: (203) 785-7063, FAX: (203) 785-5415.

**Publisher's Disclaimer:** This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Marks and Craft Page 2

Within the secondary lymphoid organs (SLO), the majority of T cells are naive, identified by the expression of L-selectin (CD62L) and CD45RA (in humans) that upon activation produce IL-2[1]. Naïve cells are primarily recruited to the secondary lymphoid organs such as the lymph node and spleen mediated by L-selectin[2] and the chemokine (C-C motif) receptor 7 (CCR7) [3]. In these locations, naïve cells are continuously surveying available antigen from resident and migrating dendritic cells[4] and awaiting the correct combination of T cell receptor (TCR) signaling in the context of co-stimulation to initiate activation[5]. Following stimulation, T cells differentiate into distinct effector lineages dictated by the activating environment so that the proper immune response occurs[6]. This process requires several days for naïve T cell activation to produce a functional effector T cell exported from the lymph node[7].

In contrast, conventional  $\alpha\beta$  T cells residing at barrier sites have an effector or memory cell phenotype[8]. These T cells express CD44, a molecule important for their non-specific exit from the systemic circulation and residence within peripheral organs[9,10]. They also express distinct chemokine receptors and integrins that allow recruitment to specific sites. Gastrointestinal tract tropism is determined by CCR9 mediated recruitment via the small intestine's expression of CCL25 (TECK) in combination with  $\alpha$ 4 $\beta$ 7 expressed on effector T cells promoting adhesion to mucosal addressin cell adhesion molecule-1 (MAdCAM-1) expressed by the postcapillary endothelial cells in the small intestine[11,12]. For efficient dermal and epidermal homing, T cells express CCR4 and CCR10 that bind to CCL17 and CCL27, respectively, expressed by the skin during resting and inflammatory conditions [13-16]. In addition, T cells necessary for the protection against inhaled pathogens are directed to the lung by the expression of CCR3 and CCR5. While unique adhesion molecules direct effector T cells to specific barrier locations, other molecules such as  $\alpha$ 4 $\beta$ 1 and CCR6 play a more general but important role in recruitment to mucosa and the skin[17,18].

Another important distinction between the  $\alpha\beta$  T cells that reside in the epithelial surfaces compared to the compartment within the lymph node or spleen is the reduced threshold for activation and rapid response to pathogens[8]. At barrier surfaces, effector T cell populations are primed for cytokine secretion. These cells are activated much faster at least partially due to altered co-stimulatory molecule expression. Naïve T cells rely on CD28 as their second signal while memory cells located at epithelial layers make use of ICOS as well as others. The cells at these sites are primed and ready to respond rapidly to any sign of infection to help remove the source before it results in infection[19].

#### **The effector T cell paradigm**

For the past 20 years, the  $T_H$  differentiation paradigm consisted of two mutually exclusive pathways,  $T_H1$  and  $T_H2$ , defined by distinct cytokine production and immune function[20]. The first,  $T_H1$ , is characterized by the production of IFN $\gamma$  and directs cell-mediated immunity. This subset requires the transcription factor T-bet[21], is induced by IFN- $\gamma$  and IL-12 from macrophages or  $DCs[22]$  and requires STAT1 and STAT4 signaling [23,24]. T<sub>H</sub>1 responses are necessary for intracellular pathogen clearance[25] and in mice induce B cells to produce IgG<sub>2a</sub>[26]. The other subset, T<sub>H</sub>2, is characterized by the production of IL-4, IL-5 and IL-13 [27] and in mice, directs B cell secretion of IgE and IgG1[26]. This subset requires the transcription factor GATA3[28], is induced by IL-4 or thymic stromal lymphopoietin from basophils[29], and requires STAT6 signaling[30].  $T_H2$  responses are necessary for clearance of extracellular parasitic infections and cause allergic disease[25]. The two subsets are distinct lineages in that GATA3 and T-bet negatively regulate each other and the presence of IFNγ prevents  $T_H2$  and IL-4 prevents  $T_H1[31]$  differentiation.

More recent work indicates that another subset of effector  $T_H$  cells exists. This lineage is defined by the production of IL-17 and has been given the name  $T_H$ 17. These cells are critical

for protection against extracellular bacteria and fungi and are responsible for several autoimmune conditions[32].

## **The TH17 subset**

#### **Interleukin 17: structure and function**

The  $T_H17$  subset of helper T cells is defined by the production of the IL-17 cytokine. This cytokine was first described 15 years ago and was originally given the name cytotoxic Tlymphocyte-associated antigen 8 (CTLA-8)[33,34], and later renamed IL-17[35]. Subsequent work revealed that it was the first identified in a family of six cytokines, now referred to as IL-17A through F, with IL-17F showing the highest degree of homology with IL-17A followed by B, D, C, and E[36]. The cellular source of IL-17 was originally identified in activated T cells[33,34] but more recently been expanded to include γδ T cells[37], CD8<sup>+</sup> memory T cells [38], neutrophils[38] and monocytes[39]. CD4+ T cells are considered the significant producers of this cytokine.

IL-17 is pro-inflammatory and important for the clearance of extracellular pathogens and multiple autoimmune disorders. Experimental models using mice with defective IL-17 signaling or treated with depleting antibodies show increased susceptibility to lung infection by *Klebsiella pneumonia and Mycoplamsa pneumoniae* and a defect in clearance of *Candida albicans* and *Escherichia coli* [40-43]. This effect has been linked to IL-17-mediated neutrophil recruitment as well as induction of anti-microbial proteins from resident cells. IL-17 stimulates a host of inflammatory cytokines and chemokines, including granulocyte colony-stimulating factor (G-CSF), macrophage inflammatory protein-2 (MIP-2), IL-8, monocyte chemotactic protein-1 (MCP-1), CXCL-8, CXCL-1 and CXCL-10[36,44-47] along with other inflammatory mediators such as prostaglandin E2, nitric oxide, matrix metalloproteases, acute phase proteins and IL-6[45,46,48,49]. Along the same lines, IL-17 can promote unfavorable immune responses indicated by this cytokine's role in multiple autoimmune disorders such as rheumatoid arthritis[50], psoriasis[51], inflammatory bowl disease[52], asthma[53], and multiple sclerosis[54,55].

Given the robust immune response mediated by IL-17, it is not surprising that targets for this cytokine are highly diverse. Studies of mRNA expression indicate that the receptor is present on hematopoetic cells, osteoblasts, fibroblasts, endothelial cells and epithelial cells in the lung, liver, spleen and kidney[35,56]. In fact, this family of receptors is as complex as their ligands. Sequences homology searches have revealed that there are five members, IL-17RA to IL-17RE [36,57]. This group represents a unique family containing domains not observed previously and is structured as a single-pass transmembrane proteins with an extracellular domain and a long intracellular tail [35]. Further analysis indicated that all receptors except IL-17RA have alternative splicing variants that introduce early stop codons allowing for the receptor to be secreted[58,59] and potentially act as a decoy to help reduce IL-17 signaling during an immune response.

To date, functional studies of the IL-17 receptor family are still lacking, with most analyses limited to IL-17RA and more recently IL-17RC. While both of these receptors can bind to IL-17 and IL-17F, IL-17RA has a log fold decreased affinity for IL-17F[60] while IL-17RC binds equally to both[61]. IL-17RA exists as a preformed homodimer[62] or can function as a heterodimer pairing with IL-17RC[63]. In fact, IL-17RA may be a generic receptor for all IL-17 family members given the recent report that both IL-17RA and IL-17RB are necessary for IL-17E (IL-25) signaling, a  $T_H2$  inducing pathway, very different from IL-17[64].

Upon ligand binding, the IL-17 receptor undergoes a conformational change facilitating dissociation of the intracellular region. The IL-17RA has a cytoplasmic tail with motifs similar

to the TLR-IL-1 receptor (TIR)[65] superfamily, now termed SEFIR domain (similar expression to FGF receptor, IL-17 receptor, Toll-IL-1R)[66] but does not require the myeloid differentiation factor 88 (MyD88) for signaling. Upon IL-17RA engagement, signaling via Act1[67] promotes TRAF6 ubiquitination of the receptor[68] and activates the NF-κB transcription factor pathway[67,69-71]. However, even with multiple family members and overlap of IL-17RA and IL-17RC in binding, there is minimal redundancy in the function of IL-17RA given that targeted deletion of this receptor causes profound defects in host defense [47].

#### **TH17: a new helper subset**

As mentioned earlier, CD4<sup>+</sup> helper T cells have been divided into two distinct effector lineages,  $T_H1$  and  $T_H2[20]$ . These two subsets develop during the course of an infection and are selected by the inflammatory signals provided by the innate immune system so that a particular type of immune response can be carried out. These two effector lineages have somewhat opposing functions  $[72-74]$  with T<sub>H</sub>1 cells driving cell-mediated immune responses that can cause tissue damage and experimentally characterized as the pathway necessary for delayed type hypersensitivity (DTH)[75], while  $T_H2$  cells promoting antibody-mediated responses and are associated with allergy and the IgE isotype[76].

Although the  $T_H1/T_H2$  paradigm explained many experimental systems and disease models, there were some inconsistencies that provided a framework for the introduction of a new subset [32,77]. These experimental observations related to the IL-12 cytokine family and the discovery of another member, IL-23. IL-12 is a heterodimeric cytokine that is composed of p40 and p35 subunits to make a complete cytokine p70[78,79] that signals via the IL-12 receptor consisting of the IL-12R $\beta$ 1 and IL-12R $\beta$ 2[80,81]. This signaling pathway is necessary for  $T_H$ 1 development and genetic deficiency of this cytokine prevents IFN $\gamma$  production from T cells and mice normally resistant to *Leishmania major* die from the infection[82-84].

However, other experimental models, particular experimental autoimmune encephalitis (EAE) raised questions regarding the simplicity of IL-12 and the IFNγ inducing effect. Initial work using antibodies to block p40 or mice deficient in this subunit showed resistance to EAE, indicating that IL-12 and presumably IFNγ were necessary for disease development[85-87], except that, in the absence of IFNγ, mice were still susceptible to the disease[88]. The data indicated a divergent function between the p40 subunit of IL-12 and IFNγ induction. Support for this notion occurred when p35 deficiency had the same effect on EAE as the IFNγ knockout and opposite effect when p40 was lacking[89]. These observations indicated that the p40 subunit had functions apart from pairing with p35 to induce a  $T_H1$  response.

This issue was resolved when another IL-12 family member, IL-23 was discovered[90] and shown to be critical for the induction of IL-17 from  $CD4^+$  T cells[91]. The IL-23 cytokine shares the p40 subunit but pairs with a unique p19 protein that together bind to a receptor composed of the shared IL-12R $\beta$ 1 and a unique IL-23 receptor[92] (Table 1). Similar to the IL-12 requirement for IFNγ production, it was shown that the related cytokine IL-23 induces T cells to produce IL-17[91]. Soon after, EAE disease induction was proven to be dependent upon IL-23 derived T<sub>H</sub>17 cells and actually protected by the IL-12/IFN $\gamma$  pathway[55,89]. Thus, very similar IL-12 family member cytokines with a common p40 subunit and IL-12Rβ1 induce distinct effector pathways consisting of  $T_H1$  and  $T_H17$ .

#### **TH17 development**

The discovery of IL-23 and the identification of its role in IL-17 mediated disease set the stage for characterization of the new subset,  $T_H17[55,89]$ , and understanding how these effector cells develop. Similar to  $T_H1$  and  $T_H2$ , the  $T_H17$  lineage has a distinct *in vitro* differentiation

pathway. Sorting of naïve T cells and culturing in the presence of TGFβ induces FoxP3 transcription factor and converts the majority of T cells to regulatory T cells  $(T_R)$ ; however, addition of IL-6 to the culture conditions changed the phenotype to IL-17 production[93,94]. In fact, activating T cells with  $T_R$ , producers of TGF $\beta$ , and DCs stimulated with a Toll like receptor agonist, a source of IL-6, also converted the naïve population to  $T_H17[95]$ . In these *in vitro* skewing experiments, the absence of IL-23 had no effect on T<sub>H</sub>17 development leading to the conclusion that TGF $\beta$  and IL-6 mediate the initial T<sub>H</sub>17 differentiation while IL-23 is important for survival and expansion. *In vivo* experiments using genetically altered mice with non-functional TGFβ receptor signaling or impaired T cell production confirmed the role of this cytokine in IL-17 differentiation and subsequent EAE disease development[96,97].

#### **TH17 transcription factors and signaling pathways**

Analogous to T-bet, GATA3 and FoxP3 for  $T_H1$ ,  $T_H2$  and  $T_{REG}$ , the transcription factor retinoic acid receptor-related orphan receptor (ROR)γt directs the differentiation of  $T_H$ 17. Mice that lack RORγt cannot make IL-17 producing T cells and retroviral transduction into naïve cells promotes  $T_H$ 17 development[98]. In addition, another transcription factor of the same family, RORα, plays a synergistic role with RORγt in T<sub>H</sub>17 differentiation[99]. It has recently been shown that Runx1 is an important transcription factor in binding to RORγt and FoxP3 to promote efficient  $T_H$ 17 development[100]. Interferon-regulatory factor (IRF) 4, a mediator of  $T_H$ 2 development is also required but not specific for  $T_H$ 17 induction[101].

The specific sequence of cell signaling events involved in  $T_H17$  development and function has been partially elucidated. IL-23 and IL-6 activate STAT3 signaling, now considered necessary and unique for T<sub>H</sub>17 differentiation[102]. As part of activation, the T<sub>H</sub>17 cells make IL-21 that provides autocrine signaling and can replace the need for IL-6[103-105]. As part of this process, the suppressor of cytokine signaling (Socs) 3 is turned off as it functions as a negative regulator [106].

#### **TH17 regulation**

Analogous to the inhibitory effects that  $T_H1$  cytokines have on  $T_H2$  development and the reverse,  $T_H$ 17 function is also influenced by  $T_H1$  and  $T_H2$  cytokines. *In vitro* activation and differentiation of naïve T cells to the T<sub>H</sub>17 lineage is enhanced with blocking of IFN<sub>Y</sub> and IL-4 [94,107]. Mice lacking the T<sub>H</sub>1 transcription factor T-bet develop exaggerated T<sub>H</sub>17 levels in the setting of autoimmune disease such as myocarditis or during Mycobacterium bacterial infection[108,109]. However, this data does not determine if the presence of these cytokines prevents development of  $T_H17$  cells or regulates the secretion of IL-17 follow lineage commitment. In addition, IL-27, another member of the IL-12 cytokine family, with  $T_H1$ inducing properties, can inhibit  $T_H17$  independent of its  $T_H1$  promoting function[110]. Along the same lines, IL-17E also known as IL-25, has suppressive function and its absence promotes enhanced IL-17 levels that exacerbates  $EAE[110]$  and allows for increased  $T_H17$  cell development in the gut[111]. Thus, the IL-17 cytokine contributes to a specific type of inflammatory response and as appropriate is carefully regulated by other cytokines to promote swift resolution of toxic inflammatory conditions to minimize injury to the host.

#### **TH17: An effector lineage sharing a regulatory T cell pathway**

The T<sub>H</sub>17 subset is often considered a parallel effector lineage to T<sub>H</sub>1 and T<sub>H</sub>2[112] with a distinct role in the pathogenesis of specific autoimmune conditions and a mediator of microbial clearance (Table 2). Early work indicating a dependency on IL-23 presented an analogous developmental pathway to  $T_H1$  induction by IL-12[32] and presented the initial idea that the secretion of related factors IL-12 or IL-23 determined the fate of the developing immune response. However, subsequent data established IL-23 as a survival factor and identified

TGFβ in conjunction with IL-6 as the lineage determining cytokines[93-95]. Thus, the comparison to  $T_H1$  development was diminished and a link to  $T_R$  cells was introduced.

Activating naïve T cells *in vitro* in the presence of TGF $\beta$  alone promotes development of T<sub>R</sub> with the addition of IL-6 diverting differentiation to  $T_H$ 17. This was the first indication that this inflammatory subset shared a common lineage with  $T_R[93,94]$ . Further support comes from IL-2, a cytokine necessary for  $T_R$  survival [113]. This cytokine constrains the development of T<sub>H</sub>17 cells so that TGF $\beta$ /IL-6 in the presence of IL-2 had significantly reduced T<sub>H</sub>17 development and expanded FoxP3<sup>+</sup> T<sub>R</sub> cells[114]. However, inflammatory conditions, such as provision of IL-1 with TGFβ/IL-6 in the presence of IL-2 rescued the IL-2 inhibitory effect and restored T<sub>H</sub>17 differentiation[115]. Additional reports find that FoxP3<sup>+</sup> T<sub>R</sub> cells can be converted directly to  $T_H$ 17 producing cells with the correct inflammatory conditions [116]. The vitamin A metabolite, retinoic acid, produced by DCs within the gut, is responsible for preventing inflammation by diverting  $T_H17$  cells into  $T_R[117]$ .

A convincing piece of work proving a common lineage between  $T_H$ 17 and  $T_R$  cells comes from a study using reporter mice to track the expression of FoxP3 and RORγt in T cells. The authors showed that TGFβ signaled in a concentration dependent manner to promote the expression of both FoxP3 and RORγt. FoxP3 directly bound to RORγt preventing  $T_H$ 17 differentiation an effect relieved by IL-6, IL-21 and IL-23[118]. An additional report confirms the suppressive function of FoxP3 on RORγt and adds that Runx1 is critical in binding both transcriptions factors to promote  $T_H17$  development[100].

Additional support for a  $T_H17/T_R$  shared developmental pathway was provided by identification of T cells fated to become  $T_R$  but unable to express FoxP3 due to an insertion of GFP in place of this gene. In so doing, the  $T_R$  fated cells in the absence of FoxP3 converted to RORγt expressing cells and produced IL-17[119]. Thus, in the absence of FoxP3, natural mechanisms selecting for  $T_R$  development default to  $T_H$ 17, suggesting that altering thymic conditions such as IL-6 or IL-1 may select for  $T_H17$  cells from the  $T_R$  compartment.

#### **TH17 associated cytokines**

The  $T_H$ 17 subset is associated with several other cytokines that contribute to this subset's unique function. The IL-17 family member, IL-17F, the closest related cytokine to IL-17 within this family, is also secreted by this lineage[55]. IL-17F can function similar to IL-17 by inducing production of IL-6, IL-8 and CXCL1 from *in vitro* cultured cells and administration of exogenous IL-17F during asthma induction promotes neutrophil recruitment[120,121]. However, despite similarities in protein sequence and function, IL-17F does not have complete redundancy with IL-17. For example, IL-17 knock out mice exhibit reduced arithritis[122], EAE[123] and allergic responses[124]. In fact, recent data indicates a distinct role during gut inflammation given that IL-17 knockout mice have reduced survival during DSS-induced colitis while IL-17F deficient mice are protected[125].

The IL-10 family member, IL-22, is also an established T<sub>H</sub>17 associated cytokine. *In vitro* activation of naïve T cells in the presence of  $T_H17$  skewing conditions, *i.e.* TGFB and IL-6, promotes IL-22 production[126,127]. The source of IL-22 is limited to T cells, NK cells and NK T cells[128]. The receptor for IL-22 consists of the IL-10 receptor (IL-10R) β and IL-22 receptor (IL-22R)[129]. While the IL-10R $\beta$  has broad expression, the IL-22R is limited to the skin, liver, lung and pancreas but not detected in T cells[128,130]. Thus, IL-22 promotes signaling to peripheral organs and does not directly influence T cell responses.

IL-22 is an important factor during inflammation. This cytokine in cooperation with IL-17 induces anti-microbial peptide activation to enhance clearance of bacterial infections[126]. IL-22 knockout mice indicate that this cytokine has an important role in psoriasis and hepatitis.

In the setting of dermal inflammation, IL-22 plays a pathologic role in promoting acanthosis [127]. Conversely, during acute inflammation of the liver, IL-22 is protective and reduces liver enzyme elevation[131].

#### **IL-17: an important cytokine in immune barrier function**

The IL-17 cytokine is a major player in the immune responses at epithelial surfaces. This factor is important for efficient clearance of pathogenic infections and responsible for significant autoimmune pathology.

#### **Lung**

IL-17 is critical for protecting the host from lung-associated pathogens. Studies using IL-17RA and IL-23 knockout mice highlight the importance of this cytokine in the clearance of the pathogen *Klebsiella pneumonia*[41,132]. Other bacterial infections such as *Mycobacterium tuberculosis*[133] and *Mycobacterium bovis*[134] can induce an IL-17 response that is important for preventing lethal disease. A role for IL17 has been suggested in viral infections such as in synergistic recruitment of neutrophils in human rhinovirus infection[135]. This cytokine has also been linked to opportunistic fungal infections such as the HIV related *Pneumocystis carinii*[136] and *Candida albicans*[40].

While IL-17 is considered beneficial for protecting the lungs from the constant exposure to potential pathogens, this cytokine is responsible for directing inflammation during allergic asthma. This cytokine is increased in the airways of people with asthma consistent with its inflammatory role in promoting inflammation[137]. However, its function in allergic lung inflammation is not clear. While IL-17 contributes to the recruitment of neutrophils and eosinophils to the lungs, IL-17RA knockout mice have worse  $T_H2$  disease indicating an inhibitory/protective role in mediating  $T_H2$  type disease[138]. Additional studies in mice indicate that the IL-17 cytokine promotes a distinct type of inflammatory lung disease. Mice that receive  $T_H2$  skewed T cells were responsive to treatment with dexamethasone while  $T_H$ 17 skewed cells induced significant airway inflammation but unresponsive to steroid treatement[139]. Thus, this subset can direct unwanted lung inflammation and may help to explain why some people are resistant to conventional asthma therapy.

### **Gastrointestinal tract**

At this mucosal surface of the gastrointestinal tract, the body is exposed to an abundance of microorganisms, most of which are important for preventing overgrowth of pathogenic bacteria and necessary for immune homeostasis. As such, the immune system has developed mechanisms to distinguish between the harmful and the helpful residents of our gut. One indication that the  $T_H17$  subset plays an important role at this site comes from the study that initially identified RORγt as the lineage specific transcription factor. In this study, the authors found the highest concentration of  $T_H17$  cells were within the lamina propria of the small intestine, almost 10% of  $\alpha\beta$  T cells[98]. This finding is quite striking when considering that it was later shown that mucosal DCs were poor inducers of  $T_H17$  cells, secondarily to the production of retinoic acid, compared to their lymph node counterparts that were much more  $T_H$ 17 permissive[117]. These findings appear to be contradictory unless one considers that the  $T_H17$  cells located in the lamina propria originate somewhere else and the retinoic acid from the mucosal DCs control these imported potentially pathogenic T cells.

There is a dynamic interaction between the commensal bacteria and the immune cells of the gut. Toll-like receptors are responsible for mediating this cross talk and instructing the immune system appropriately[140]. For example, toll-like receptor 9 detects gut flora DNA to regulate the balance between  $T_R$  and  $T_H$ 17 cells of the gut. In the absence of this signaling pathway,

the T cells of the gut are overwhelmed with regulatory cells and prevent productive immune function[141]. Similarly, the native gut flora provides a balance between related IL-17 cytokines, IL-17 and IL-17E (IL-25). Pathogen-free conditions promote  $T_H$ 17 overgrowth and elevated IL-23 while restoration of microorganisms signals for IL-17E to re-establish the proper balance and promote healthy intestinal homeostasis[111].

The IL-17/IL-23 axis is a critical player in the promotion of inflammatory bowl disease. Mouse models of gut inflammation were originally attributed to  $T_H1$  effector subsets based on the observation that antibodies directed at the p40 subunit of IL-12 proved to be an effective treatment[142]. However, the identification of p40 as a shared subunit between IL-12 and IL-23 has prompted a re-examination of gut inflammation. It is now established that the IL-17 pathway is an important cytokine involved in autoimmune disease of the gut[143,144]. In human studies, it was observed that IL-23 and IL-17 are elevated in patients with IBD[145] and that treatment with anti-p40 antibodies is very effective in preventing the disease symptoms [146], most likely due to reduction in IL-23 and subsequently IL-17. The IL-23/IL-17 pathway has also been implicated in the promotion of unwanted gut inflammation with the identification of genetic variations of IL-23, STAT3 and other  $T_H17$  associated genes linked to Crohn's disease and ulcerative colitis[147].

## **Skin**

The dermis/epidermis is another very large barrier organ housing a distinct immune cell population. At this site, similar to the gut, microorganisms are ubiquitous along the outside border. Here too,  $T_H17$  cells are present and help to protect this potential danger zone from pathogen entry. People with an inability to clear the opportunistic fungal infection *Candida* suffer from mucocutaneous candidiasis. A recent report indicates that peripheral blood mononuclear cells from these patients have reduced IL-17 and IL-22 mRNA and that their  $CCR6+ IL-17+ T$  cells are significantly reduced [148]. Mouse studies confirm the role of IL-17 in preventing this cutaneous yeast infection[40].

In addition, the  $T_H$ 17 associated cytokine IL-22 has been at the forefront of autoimmune pathology of the dermis. Mouse models of psoriasis indicate that IL-22 production promotes keratinocyte survival and drives acanthosis[127]. Furthermore, it has been reported that human psoriatic lesions have increased IL-23 mRNA compared to healthy skin[149]. As one might expect, IL-17 expression from psoriatic plaques correlated with disease severity and cytokine levels normalized following treatment with cyclosporine[150].

#### **γδ T cells as a source of IL-17 at epithelial surfaces**

The  $\gamma\delta$  T cell subset makes up a small fraction of the total T cell compartment but serves a distinct conserved function. These T cells develop in the thymus, require random recombination events similar to  $\alpha\beta$  T cells but are produced in waves of subsets defined by the individual  $\gamma$  and  $\delta$  receptors that they express. The different subsets of  $\gamma\delta$  T cells vary in function and location[151], [152]. Thus, this group of immune cells has the potential to generate great diversity in antigen recognition but somehow targets specific T cell subsets to reside in distinct locations to provide a protective function unique to the individual site. γδ T cells are not restricted to classical MHC class I or II molecules like their CD8+ or CD4<sup>+</sup> αβ counterparts. A small population of γδ T cells has been shown to recognize MHC class IB antigens T10 and T22 inmice[153] and other "stress markers" such as MICA in humans through their TCR [154] or by expression of NKG2D[155,156]. Thus,  $\gamma \delta$  T cells do not require foreign antigen to induce activation and promote inflammation but rather respond to endogenous signals that indicate pathogen entry into protected sites. It may be this unconventional activation mechanism that has selected for their niche within the immune system.

γδ T cells have several mechanisms in which they contribute to immune responses. These T cells have been noted to provide common T cell cytokines such as IFNγ and IL-17 during the innate phase of inflammation.  $\gamma \delta$  T cells produce IL-17 and depending on the timing, can represent a majority of cells producing this cytokine[157]. This subset is unique in that these T cells do not require priming to allow effector function causing a delay in IL-17 production but rather can secrete this cytokine immediately upon activation. This ability to produce IL-17 is part of the thymic developmental pathway that selects the individual  $\gamma\delta$  subsets given the observation that Vγ4 thymocytes in mice can produce IL-17 while the Vγ1 cells have minimal production[158] which may be related to the affinity for ligand binding taking place within the thymus[159]. As such,  $\gamma\delta$  T cells may be an important source of the IL-17 cytokine. Given the role of IL-17 in neutrophil recruitment and other early inflammatory responses, this population of T cells being instructed in the thymus to populate host organs, especially the barrier surfaces, and having an ability to produce IL-17 immediately upon activation presents a unique model for T cell production of IL-17 and helps fine tune the immune response.

#### **Conclusions**

T cells are critical in the complex regulation of barrier immunity. These unique sites require dynamic interactions between the cells of the immune system and the surrounding environment. The immune system has evolved not only to prevent unwanted activation in response to microorganisms residing at these sites but makes use of these species to shape the mature compartment guarding the epithelial lining. A newly described helper T cell subset,  $T_H$ 17, has proven to be a major player in protecting the host barrier surfaces. IL-17 has been implicated in clearance of bacterial, viral and fungal infections occurring in the lung, gut and skin as well as the pathogenic mediator of multiple mucosal and cutaneous autoimmune diseases.

The T<sub>H</sub>17 subset, while often described as a third effector lineage, parallel to the T<sub>H</sub>1 and  $T_H2$  subsets, has certain characteristics that may place these cells in a class of their own. One of the most striking findings is the direct lineage relationship with  $T_R$  cells. These two subsets require a common cytokine TFGβ for lineage commitment and recent work has observed direct interactions between the lineage specific transcription factor FoxP3 and RORγt. While  $T_H1$ and  $T_H2$  cells function as late contributors to pathogen clearance, the  $T_H17$  subset is implicated in neutrophil recruitment, a function that is necessary during early inflammation, before the conventional adaptive phase of the immune response. Thus, one may speculate that this subset is already pre-formed waiting for the correct signal to promote inflammation and direct efficient immune clearance and host protection. We are still adding pieces to the puzzle defining the  $T_H$ 17 subset to reveal the true role of this unique and critical helper T cell subset.

## **Acknowledgments**

This work was supported in part by NIH Grants AR40072, AR44076, and P30 AR053495, and by support from Rheuminations, Inc., the Arthritis Foundation and the Connecticut Chapter of the Lupus Foundation of America.

## **References**

- 1. Truneh A, Albert F, Golstein P, Schmitt-Verhulst AM. Early steps of lymphocyte activation bypassed by synergy between calcium ionophores and phorbol ester. Nature 1985;313:318–20. [PubMed: 3918270]
- 2. Gallatin WM, Weissman IL, Butcher EC. A cell-surface molecule involved in organ-specific homing of lymphocytes. Nature 1983;304:30–4. [PubMed: 6866086]
- 3. Gunn MD, Kyuwa S, Tam C, Kukiuchi T, Matsuzawa A, Williams LT, et al. Mice lacking expression of secondary lymphoid organ chemokine have defects in lymphocyte homing and dendritic cell localization. J Exp Med 1999;189:451–60. [PubMed: 9927507]

- 4. Mempel TR, Henrickson SE, Von Andrian UH. T-cell priming by dendritic cells in lymph nodes occurs in three distinct phases. Nature 2004;427:154–9. [PubMed: 14712275]
- 5. June CH, Ledbetter JA, Linsley PS, Thompson CB. Role of the CD28 receptor in T-cell activation. Immunol Today 1990;11:211–6. [PubMed: 2162180]
- 6. Dong C, Juedes AE, Temann UA, Shresta S, Allison JP, Ruddle NH, et al. ICOS co-stimulatory receptor is essential for T-cell activation and function. Nature 2001;409:97–101. [PubMed: 11343121]
- 7. Campbell DJ, Butcher EC. Rapid acquisition of tissue-specific homing phenotypes by CD4(+) T cells activated in cutaneous or mucosal lymphoid tissues. J Exp Med 2002;195:135–41. [PubMed: 11781372]
- 8. Masopust D, Vezys V, Marzo AL, Lefrancois L. Preferential localization of effector memory cells in nonlymphoid tissue. Science 2001;291:2413–7. [PubMed: 11264538]
- 9. St John T, Meyer J, Idzerda R, Gallatin WM. Expression of CD44 confers a new adhesive phenotype on transfected cells. Cell 1990;60:45–52. [PubMed: 2403843]
- 10. Jalkanen S, Reichert RA, Gallatin WM, Bargatze RF, Weissman IL, Butcher EC. Homing receptors and the control of lymphocyte migration. Immunol Rev 1986;91:39–60. [PubMed: 2426181]
- 11. Berlin C, Berg EL, Briskin MJ, Andrew DP, Kilshaw PJ, Holzmann B, et al. Alpha 4 beta 7 integrin mediates lymphocyte binding to the mucosal vascular addressin MAdCAM-1. Cell 1993;74:185–95. [PubMed: 7687523]
- 12. Hamann A, Andrew DP, Jablonski-Westrich D, Holzmann B, Butcher EC. Role of alpha 4-integrins in lymphocyte homing to mucosal tissues in vivo. J Immunol 1994;152:3282–93. [PubMed: 7511642]
- 13. Campbell JJ, Haraldsen G, Pan J, Rottman J, Qin S, Ponath P, et al. The chemokine receptor CCR4 in vascular recognition by cutaneous but not intestinal memory T cells. Nature 1999;400:776–80. [PubMed: 10466728]
- 14. Homey B, Alenius H, Muller A, Soto H, Bowman EP, Yuan W, et al. CCL27-CCR10 interactions regulate T cell-mediated skin inflammation. Nat Med 2002;8:157–65. [PubMed: 11821900]
- 15. Morales J, Homey B, Vicari AP, Hudak S, Oldham E, Hedrick J, et al. CTACK, a skin-associated chemokine that preferentially attracts skin-homing memory T cells. Proc Natl Acad Sci U S A 1999;96:14470–5. [PubMed: 10588729]
- 16. Reiss Y, Proudfoot AE, Power CA, Campbell JJ, Butcher EC. CC chemokine receptor (CCR)4 and the CCR10 ligand cutaneous T cell-attracting chemokine (CTACK) in lymphocyte trafficking to inflamed skin. J Exp Med 2001;194:1541–7. [PubMed: 11714760]
- 17. Sixt M, Bauer M, Lammermann T, Fassler R. Beta1 integrins: zip codes and signaling relay for blood cells. Curr Opin Cell Biol 2006;18:482–90. [PubMed: 16919433]
- 18. Williams IR. Chemokine receptors and leukocyte trafficking in the mucosal immune system. Immunol Res 2004;29:283–92. [PubMed: 15181289]
- 19. Sallusto F, Geginat J, Lanzavecchia A. Central memory and effector memory T cell subsets: function, generation, and maintenance. Annu Rev Immunol 2004;22:745–63. [PubMed: 15032595]
- 20. Mosmann TR, Cherwinski H, Bond MW, Giedlin MA, Coffman RL. Two types of murine helper T cell clone. I. Definition according to profiles of lymphokine activities and secreted proteins. J Immunol 1986;136:2348–57. [PubMed: 2419430]
- 21. Szabo SJ, Kim ST, Costa GL, Zhang X, Fathman CG, Glimcher LH. A novel transcription factor, Tbet, directs Th1 lineage commitment. Cell 2000;100:655–69. [PubMed: 10761931]
- 22. Hsieh CS, Macatonia SE, Tripp CS, Wolf SF, O'Garra A, Murphy KM. Development of TH1 CD4+ T cells through IL-12 produced by Listeria-induced macrophages. Science 1993;260:547–9. [PubMed: 8097338]
- 23. Afkarian M, Sedy JR, Yang J, Jacobson NG, Cereb N, Yang SY, et al. T-bet is a STAT1-induced regulator of IL-12R expression in naive CD4+ T cells. Nat Immunol 2002;3:549–57. [PubMed: 12006974]
- 24. Jacobson NG, Szabo SJ, Weber-Nordt RM, Zhong Z, Schreiber RD, Darnell JE, et al. Interleukin 12 signaling in T helper type 1 (Th1) cells involves tyrosine phosphorylation of signal transducer and activator of transcription (Stat)3 and Stat4. J Exp Med 1995;181:1755–62. [PubMed: 7722452]
- 25. Abbas AK, Murphy KM, Sher A. Functional diversity of helper T lymphocytes. Nature 1996;383:787–93. [PubMed: 8893001]

Marks and Craft Page 11

- 26. Snapper CM, Paul WE. Interferon-gamma and B cell stimulatory factor-1 reciprocally regulate Ig isotype production. Science 1987;236:944–7. [PubMed: 3107127]
- 27. Mosmann TR, Coffman RL. TH1 and TH2 cells: different patterns of lymphokine secretion lead to different functional properties. Annu Rev Immunol 1989;7:145–73. [PubMed: 2523712]
- 28. Zheng W, Flavell RA. The transcription factor GATA-3 is necessary and sufficient for Th2 cytokine gene expression in CD4 T cells. Cell 1997;89:587–96. [PubMed: 9160750]
- 29. Sokol CL, Barton GM, Farr AG, Medzhitov R. A mechanism for the initiation of allergen-induced T helper type 2 responses. Nat Immunol 2007;9:310–18. [PubMed: 18300366]
- 30. Kaplan MH, Grusby MJ. Regulation of T helper cell differentiation by STAT molecules. J Leukoc Biol 1998;64:2–5. [PubMed: 9665267]
- 31. Ouyang W, Ranganath SH, Weindel K, et al. Inhibition of Th1 development mediated by GATA-3 through an IL-4-independent mechanism. Immunity 1998;9:745–55. [PubMed: 9846495]
- 32. Harrington LE, Mangan PR, Weaver CT. Expanding the effector CD4 T-cell repertoire: the Th17 lineage. Curr Opin Immunol 2006;18:349–56. [PubMed: 16616472]
- 33. Rouvier E, Luciani MF, Mattei MG, Denizot F, Golstein P. CTLA-8, cloned from an activated T cell, bearing AU-rich messenger RNA instability sequences, and homologous to a herpesvirus saimiri gene. J Immunol 1993;150:5445–56. [PubMed: 8390535]
- 34. Yao Z, Timour M, Painter S, Fanslow W, Spriggs M. Complete nucleotide sequence of the mouse CTLA8 gene. Gene 1996;168:223–5. [PubMed: 8654948]
- 35. Yao Z, Fanslow WC, Seldin MF, Rousseau AM, Painter SL, Comeau MR, et al. Herpesvirus Saimiri encodes a new cytokine, IL-17, which binds to a novel cytokine receptor. Immunity 1995;3:811–21. [PubMed: 8777726]
- 36. Kolls JK, Linden A. Interleukin-17 family members and inflammation. Immunity 2004;21:467–76. [PubMed: 15485625]
- 37. Lockhart E, Green AM, Flynn JL. IL-17 production is dominated by gammadelta T cells rather than CD4 T cells during Mycobacterium tuberculosis infection. J Immunol 2006;177:4662–9. [PubMed: 16982905]
- 38. Ferretti S, Bonneau O, Dubois GR, Jones CE, Trifilieff A. IL-17, produced by lymphocytes and neutrophils, is necessary for lipopolysaccharide-induced airway neutrophilia: IL-15 as a possible trigger. J Immunol 2003;170:2106–12. [PubMed: 12574382]
- 39. Zhou Q, Desta T, Fenton M, Graves DT, Amar S. Cytokine profiling of macrophages exposed to Porphyromonas gingivalis, its lipopolysaccharide, or its FimA protein. Infect Immun 2005;73:935– 43. [PubMed: 15664935]
- 40. Huang W, Na L, Fidel PL, Schwarzenberger P. Requirement of interleukin-17A for systemic anti-Candida albicans host defense in mice. J Infect Dis 2004;190:624–31. [PubMed: 15243941]
- 41. Ye P, Garvey PB, Zhang P, et al. Interleukin-17 and lung host defense against Klebsiella pneumoniae infection. Am J Respir Cell Mol Biol 2001;25:335–40. [PubMed: 11588011]
- 42. Shibata K, Yamada H, Hara H, Kishihara K, Yoshikai Y. Resident Vdelta1+ gammadelta T cells control early infiltration of neutrophils after Escherichia coli infection via IL-17 production. J Immunol 2007;178:4466–72. [PubMed: 17372004]
- 43. Wu Q, Martin RJ, Rino JG, Breed R, Torres RM, Chu HW. IL-23-dependent IL-17 production is essential in neutrophil recruitment and activity in mouse lung defense against respiratory Mycoplasma pneumoniae infection. Microbes Infect 2007;9:78–86. [PubMed: 17198762]
- 44. Awane M, Andres PG, Li DJ, Reinecker HC. NF-kappa B-inducing kinase is a common mediator of IL-17-, TNF-alpha-, and IL-1 beta-induced chemokine promoter activation in intestinal epithelial cells. J Immunol 1999;162:5337–44. [PubMed: 10228009]
- 45. Moseley TA, Haudenschild DR, Rose L, Reddi AH. Interleukin-17 family and IL-17 receptors. Cytokine Growth Factor Rev 2003;14:155–74. [PubMed: 12651226]
- 46. Shen F, Ruddy MJ, Plamondon P, Gaffen SL. Cytokines link osteoblasts and inflammation: microarray analysis of interleukin-17- and TNF-alpha-induced genes in bone cells. J Leukoc Biol 2005;77:388–99. [PubMed: 15591425]
- 47. Ye P, Rodriguez FH, Kanaly S, Stocking KL, Schurr J, Schwarzenberger P, et al. Requirement of interleukin 17 receptor signaling for lung CXC chemokine and granulocyte colony-stimulating factor

expression, neutrophil recruitment, and host defense. J Exp Med 2001;194:519–27. [PubMed: 11514607]

- 48. Fossiez F, Djossou O, Chomarat P, Flores-Romo L, Ait-Yahia S, Maat C, et al. T cell interleukin-17 induces stromal cells to produce proinflammatory and hematopoietic cytokines. J Exp Med 1996;183:2593–603. [PubMed: 8676080]
- 49. LeGrand A, Fermor B, Fink C, Pisetsky DS, Weinberg JB, Vail TP, et al. Interleukin-1, tumor necrosis factor alpha, and interleukin-17 synergistically up-regulate nitric oxide and prostaglandin E2 production in explants of human osteoarthritic knee menisci. Arthritis Rheum 2001;44:2078–83. [PubMed: 11592370]
- 50. Koenders MI, Lubberts E, Oppers-Walgreen B, van den Bersselaar L, Helsen MM, Di Padova FE, et al. Blocking of interleukin-17 during reactivation of experimental arthritis prevents joint inflammation and bone erosion by decreasing RANKL and interleukin-1. Am J Pathol 2005;167:141– 9. [PubMed: 15972960]
- 51. van Beelen AJ, Teunissen MB, Kapsenberg ML, de Jong EC. Interleukin-17 in inflammatory skin disorders. Curr Opin Allergy Clin Immunol 2007;7:374–81. [PubMed: 17873575]
- 52. Fujino S, Andoh A, Bamba S, Ogawa A, Hata K, Araki Y, et al. Increased expression of interleukin 17 in inflammatory bowel disease. Gut 2003;52:65–70. [PubMed: 12477762]
- 53. Hellings PW, Kasran A, Liu Z, Vandekerckhove P, Wuyts A, Overbergh L, et al. Interleukin-17 orchestrates the granulocyte influx into airways after allergen inhalation in a mouse model of allergic asthma. Am J Respir Cell Mol Biol 2003;28:42–50. [PubMed: 12495931]
- 54. Lock C, Hermans G, Pedotti R, Pedotti R, Brendolan A, Schadt E, et al. Gene-microarray analysis of multiple sclerosis lesions yields new targets validated in autoimmune encephalomyelitis. Nat Med 2002;8:500–8. [PubMed: 11984595]
- 55. Langrish CL, Chen Y, Blumenschein WM, Mattson J, Basham B, Sedgwick JD, et al. IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. J Exp Med 2005;201:233– 40. [PubMed: 15657292]
- 56. Yao Z, Spriggs MK, Derry JM, Strockbine L, Park LS, VandenBos T, et al. Molecular characterization of the human interleukin (IL)-17 receptor. Cytokine 1997;9:794–800. [PubMed: 9367539]
- 57. Weaver CT, Hatton RD, Mangan PR, Harrington LE. IL-17 family cytokines and the expanding diversity of effector T cell lineages. Annu Rev Immunol 2007;25:821–52. [PubMed: 17201677]
- 58. Haudenschild D, Moseley T, Rose L, Reddi AH. Soluble and transmembrane isoforms of novel interleukin-17 receptor-like protein by RNA splicing and expression in prostate cancer. J Biol Chem 2002;277:4309–16. [PubMed: 11706037]
- 59. Tian E, Sawyer JR, Largaespada DA, Jenkins NA, Copeland NG, Shaughnessy JD Jr. Evi27 encodes a novel membrane protein with homology to the IL17 receptor. Oncogene 2000;19:2098–109. [PubMed: 10815801]
- 60. Hymowitz SG, Filvaroff EH, Yin JP, et al. IL-17s adopt a cystine knot fold: structure and activity of a novel cytokine, IL-17F, and implications for receptor binding. Embo J 2001;20:5332–41. [PubMed: 11574464]
- 61. Kuestner RE, Taft DW, Haran A, Brandt CS, Brender T, Lum K, et al. Identification of the IL-17 receptor related molecule IL-17RC as the receptor for IL-17F. J Immunol 2007;179:5462–73. [PubMed: 17911633]
- 62. Kramer JM, Yi L, Shen F, Maitra A, Jiao X, Jin T, et al. Evidence for ligand-independent multimerization of the IL-17 receptor. J Immunol 2006;176:711–5. [PubMed: 16393951]
- 63. Toy D, Kugler D, Wolfson M, Vanden Bos T, Gurgel J, Derry J, et al. Cutting edge: interleukin 17 signals through a heteromeric receptor complex. J Immunol 2006;177:36–9. [PubMed: 16785495]
- 64. Rickel EA, Siegel LA, Yoon BR, Rottman JB, Kugler DG, Swart DA, et al. Identification of functional roles for both IL-17RB and IL-17RA in mediating IL-25-induced activities. J Immunol 2008;181:4299–310. [PubMed: 18768888]
- 65. Rock FL, Hardiman G, Timans JC, Kastelein RA, Bazan JF. A family of human receptors structurally related to Drosophila Toll. Proc Natl Acad Sci U S A 1998;95:588–93. [PubMed: 9435236]
- 66. Novatchkova M, Leibbrandt A, Werzowa J, Neubuser A, Eisenhaber F. The STIR-domain superfamily in signal transduction, development and immunity. Trends Biochem Sci 2003;28:226– 9. [PubMed: 12765832]

- 67. Qian Y, Liu C, Hartupee J, Altuntas CZ, Gulen MF, Jane-Wit D, et al. The adaptor Act1 is required for interleukin 17-dependent signaling associated with autoimmune and inflammatory disease. Nat Immunol 2007;8:247–56. [PubMed: 17277779]
- 68. Rong Z, Cheng L, Ren Y, Li Z, Li Y, Li H, et al. Interleukin-17F signaling requires ubiquitination of interleukin-17 receptor via TRAF6. Cell Signal 2007;19:1514–20. [PubMed: 17346928]
- 69. Chang SH, Park H, Dong C. Act1 adaptor protein is an immediate and essential signaling component of interleukin-17 receptor. J Biol Chem 2006;281:35603–7. [PubMed: 17035243]
- 70. Maitra A, Shen F, Hanel W, Mossman K, Tocker J, Swart D, et al. Distinct functional motifs within the IL-17 receptor regulate signal transduction and target gene expression. Proc Natl Acad Sci U S A 2007;104:7506–11. [PubMed: 17456598]
- 71. Schwandner R, Yamaguchi K, Cao Z. Requirement of tumor necrosis factor receptor-associated factor (TRAF)6 in interleukin 17 signal transduction. J Exp Med 2000;191:1233–40. [PubMed: 10748240]
- 72. Fernandez-Botran R, Sanders VM, Mosmann TR, Vitetta ES. Lymphokine-mediated regulation of the proliferative response of clones of T helper 1 and T helper 2 cells. J Exp Med 1988;168:543–58. [PubMed: 2970518]
- 73. Gajewski TF, Fitch FW. Anti-proliferative effect of IFN-gamma in immune regulation. I. IFNgamma inhibits the proliferation of Th2 but not Th1 murine helper T lymphocyte clones. J Immunol 1988;140:4245–52. [PubMed: 2967332]
- 74. Gajewski TF, Goldwasser E, Fitch FW. Anti-proliferative effect of IFN-gamma in immune regulation. II. IFN-gamma inhibits the proliferation of murine bone marrow cells stimulated with IL-3, IL-4, or granulocyte-macrophage colony-stimulating factor. J Immunol 1988;141:2635–42. [PubMed: 2971726]
- 75. Cher DJ, Mosmann TR. Two types of murine helper T cell clone. II. Delayed-type hypersensitivity is mediated by TH1 clones. J Immunol 1987;138:3688–94. [PubMed: 2953788]
- 76. Coffman RL, Carty J. A T cell activity that enhances polyclonal IgE production and its inhibition by interferon-gamma. J Immunol 1986;136:949–54. [PubMed: 2934482]
- 77. McKenzie BS, Kastelein RA, Cua DJ. Understanding the IL-23-IL-17 immune pathway. Trends Immunol 2006;27:17–23. [PubMed: 16290228]
- 78. Kobayashi M, Fitz L, Ryan M, Ryan M, Hewick RM, Clark SC, et al. Identification and purification of natural killer cell stimulatory factor (NKSF), a cytokine with multiple biologic effects on human lymphocytes. J Exp Med 1989;170:827–45. [PubMed: 2504877]
- 79. Stern AS, Podlaski FJ, Hulmes JD, Pan YC, Quinn PM, Wolitzky AG, et al. Purification to homogeneity and partial characterization of cytotoxic lymphocyte maturation factor from human Blymphoblastoid cells. Proc Natl Acad Sci U S A 1990;87:6808–12. [PubMed: 2204066]
- 80. Presky DH, Yang H, Minetti LJ, Chua AO, Nabavi N, Wu CY, et al. A functional interleukin 12 receptor complex is composed of two beta-type cytokine receptor subunits. Proc Natl Acad Sci U S A 1996;93:14002–7. [PubMed: 8943050]
- 81. Wu C, Wang X, Gadina M, O'Shea JJ, Presky DH, Magram J. IL-12 receptor beta 2 (IL-12R beta 2) deficient mice are defective in IL-12-mediated signaling despite the presence of high affinity IL-12 binding sites. J Immunol 2000;165:6221–8. [PubMed: 11086056]
- 82. Magram J, Connaughton SE, Warrier RR, Carvajal DM, Wu CY, Ferrante J, et al. IL-12-deficient mice are defective in IFN gamma production and type 1 cytokine responses. Immunity 1996;4:471– 81. [PubMed: 8630732]
- 83. Mattner F, Magram J, Ferrante J, Launois P, Di Padova K, Behin R, et al. Genetically resistant mice lacking interleukin-12 are susceptible to infection with Leishmania major and mount a polarized Th2 cell response. Eur J Immunol 1996;26:1553–9. [PubMed: 8766560]
- 84. Park AY, Hondowicz BD, Scott P. IL-12 is required to maintain a Th1 response during Leishmania major infection. J Immunol 2000;165:896–902. [PubMed: 10878364]
- 85. Leonard JP, Waldburger KE, Goldman SJ. Prevention of experimental autoimmune encephalomyelitis by antibodies against interleukin 12. J Exp Med 1995;181:381–6. [PubMed: 7528773]
- 86. Constantinescu CS, Wysocka M, Hilliard B, Ventura ES, Lavi E, Trinchieri G, et al. Antibodies against IL-12 prevent superantigen-induced and spontaneous relapses of experimental autoimmune encephalomyelitis. J Immunol 1998;161:5097–104. [PubMed: 9794448]

- 87. Segal BM, Dwyer BK, Shevach EM. An interleukin (IL)-10/IL-12 immunoregulatory circuit controls susceptibility to autoimmune disease. J Exp Med 1998;187:537–46. [PubMed: 9463404]
- 88. Ferber IA, Brocke S, Taylor-Edwards C, et al. Mice with a disrupted IFN-gamma gene are susceptible to the induction of experimental autoimmune encephalomyelitis (EAE). J Immunol 1996;156:5–7. [PubMed: 8598493]
- 89. Cua DJ, Sherlock J, Chen Y, Murphy CA, Joyce B, Seymour B, et al. Interleukin-23 rather than interleukin-12 is the critical cytokine for autoimmune inflammation of the brain. Nature 2003;421:744–8. [PubMed: 12610626]
- 90. Oppmann B, Lesley R, Blom B, Timans JC, Xu Y, Hunte B, et al. Novel p19 protein engages IL-12p40 to form a cytokine, IL-23, with biological activities similar as well as distinct from IL-12. Immunity 2000;13:715–25. [PubMed: 11114383]
- 91. Aggarwal S, Ghilardi N, Xie MH, de Sauvage FJ, Gurney AL. Interleukin-23 promotes a distinct CD4 T cell activation state characterized by the production of interleukin-17. J Biol Chem 2003;278:1910–4. [PubMed: 12417590]
- 92. Parham C, Chirica M, Timans J, Vaisberg E, Travis M, Cheung J, et al. A receptor for the heterodimeric cytokine IL-23 is composed of IL-12Rbeta1 and a novel cytokine receptor subunit, IL-23R. J Immunol 2002;168:5699–708. [PubMed: 12023369]
- 93. Bettelli E, Carrier Y, Gao W, Korn T, Strom TB, Oukka M, et al. Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. Nature 2006;441:235–8. [PubMed: 16648838]
- 94. Mangan PR, Harrington LE, O'Quinn DB, Helms WS, Bullard DC, Elson CO, et al. Transforming growth factor-beta induces development of the T(H)17 lineage. Nature 2006;441:231–4. [PubMed: 16648837]
- 95. Veldhoen M, Hocking RJ, Atkins CJ, Locksley RM, Stockinger B. TGFbeta in the context of an inflammatory cytokine milieu supports de novo differentiation of IL-17-producing T cells. Immunity 2006;24:179–89. [PubMed: 16473830]
- 96. Li MO, Wan YY, Flavell RA. T cell-produced transforming growth factor-beta1 controls T cell tolerance and regulates Th1- and Th17-cell differentiation. Immunity 2007;26:579–91. [PubMed: 17481928]
- 97. Veldhoen M, Hocking RJ, Flavell RA, Stockinger B. Signals mediated by transforming growth factorbeta initiate autoimmune encephalomyelitis, but chronic inflammation is needed to sustain disease. Nat Immunol 2006;7:1151–6. [PubMed: 16998492]
- 98. Ivanov II, McKenzie BS, Zhou L, Tadokoro CE, Lepelley A, Lafaille JJ, et al. The orphan nuclear receptor RORgammat directs the differentiation program of proinflammatory IL-17+ T helper cells. Cell 2006;126:1121–33. [PubMed: 16990136]
- 99. Yang XO, Pappu BP, Nurieva R, Akimzhanov A, Kang HS, Chung Y, et al. T Helper 17 Lineage Differentiation Is Programmed by Orphan Nuclear Receptors RORalpha and RORgamma. Immunity. 2007
- 100. Zhang F, Meng G, Strober W. Interactions among the transcription factors Runx1, RORgammat and Foxp3 regulate the differentiation of interleukin 17-producing T cells. Nat Immunol. 2008
- 101. Brustle A, Heink S, Huber M, Rosenplanter C, Stadelmann C, Yu P, et al. The development of inflammatory T(H)-17 cells requires interferon-regulatory factor 4. Nat Immunol 2007;8:958–66. [PubMed: 17676043]
- 102. Yang XO, Panopoulos AD, Nurieva R, Chang SH, Wang D, Watowich SS, et al. STAT3 regulates cytokine-mediated generation of inflammatory helper T cells. J Biol Chem 2007;282:9358–63. [PubMed: 17277312]
- 103. Korn T, Bettelli E, Gao W, Awasthi A, Jager A, Strom TB, et al. IL-21 initiates an alternative pathway to induce proinflammatory T(H)17 cells. Nature 2007;448:484–7. [PubMed: 17581588]
- 104. Nurieva R, Yang XO, Martinez G, Zhang Y, Panopoulos AD, Ma L, et al. Essential autocrine regulation by IL-21 in the generation of inflammatory T cells. Nature 2007;448:480–3. [PubMed: 17581589]
- 105. Zhou L, Ivanov II, Spolski R, Min R, Shenderov K, Egawa T, et al. IL-6 programs T(H)-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways. Nat Immunol 2007;8:967–74. [PubMed: 17581537]

- 106. Chen Z, Laurence A, Kanno Y, Pacher-Zavisin M, Zhu BM, Tato C, et al. Selective regulatory function of Socs3 in the formation of IL-17-secreting T cells. Proc Natl Acad Sci U S A 2006;103:8137–42. [PubMed: 16698929]
- 107. Park H, Li Z, Yang XO, Chang SH, Nurieva R, Wang YH, et al. A distinct lineage of CD4 T cells regulates tissue inflammation by producing interleukin 17. Nat Immunol 2005;6:1133–41. [PubMed: 16200068]
- 108. Cruz A, Khader SA, Torrado E, Fraga A, Pearl JE, Pedrosa J, et al. Cutting edge: IFN-gamma regulates the induction and expansion of IL-17-producing CD4 T cells during mycobacterial infection. J Immunol 2006;177:1416–20. [PubMed: 16849446]
- 109. Rangachari M, Mauermann N, Marty RR, Dirnhofer S, Kurrer MO, Komnenovic V, et al. T-bet negatively regulates autoimmune myocarditis by suppressing local production of interleukin 17. J Exp Med 2006;203:2009–19. [PubMed: 16880257]
- 110. Batten M, Li J, Yi S, Kljavin NM, Danilenko DM, Lucas S, et al. Interleukin 27 limits autoimmune encephalomyelitis by suppressing the development of interleukin 17-producing T cells. Nat Immunol 2006;7:929–36. [PubMed: 16906167]
- 111. Zaph C, Du Y, Saenz SA, Nair MG, Perrigoue JG, Taylor BC, et al. Commensal-dependent expression of IL-25 regulates the IL-23-IL-17 axis in the intestine. J Exp Med 2008;205:2191–8. [PubMed: 18762568]
- 112. Weaver CT, Harrington LE, Mangan PR, Gavrieli M, Murphy KM. Th17: an effector CD4 T cell lineage with regulatory T cell ties. Immunity 2006;24:677–88. [PubMed: 16782025]
- 113. Setoguchi R, Hori S, Takahashi T, Sakaguchi S. Homeostatic maintenance of natural Foxp3(+) CD25 (+) CD4(+) regulatory T cells by interleukin (IL)-2 and induction of autoimmune disease by IL-2 neutralization. J Exp Med 2005;201:723–35. [PubMed: 15753206]
- 114. Laurence A, Tato CM, Davidson TS, Kanno Y, Chen Z, Yao Z, et al. Interleukin-2 signaling via STAT5 constrains T helper 17 cell generation. Immunity 2007;26:371–81. [PubMed: 17363300]
- 115. Kryczek I, Wei S, Vatan L, Escara-Wilke J, Szeliga W, Keller ET, et al. Cutting edge: opposite effects of IL-1 and IL-2 on the regulation of IL-17+ T cell pool IL-1 subverts IL-2-mediated suppression. J Immunol 2007;179:1423–6. [PubMed: 17641006]
- 116. Xu L, Kitani A, Fuss I, Strober W. Cutting edge: regulatory T cells induce CD4+CD25-Foxp3- T cells or are self-induced to become Th17 cells in the absence of exogenous TGF-beta. J Immunol 2007;178:6725–9. [PubMed: 17513718]
- 117. Mucida D, Park Y, Kim G, Turovskaya O, Scott I, Kronenberg M, et al. Reciprocal TH17 and regulatory T cell differentiation mediated by retinoic acid. Science 2007;317:256–60. [PubMed: 17569825]
- 118. Zhou L, Lopes JE, Chong MM, Ivanov II, Min R, Victora GD, et al. TGF-beta-induced Foxp3 inhibits T(H)17 cell differentiation by antagonizing RORgammat function. Nature 2008;453:236– 40. [PubMed: 18368049]
- 119. Gavin MA, Rasmussen JP, Fontenot JD, Vasta V, Manganiello VC, Beavo JA, et al. Foxp3 dependent programme of regulatory T-cell differentiation. Nature 2007;445:771–5. [PubMed: 17220874]
- 120. Hizawa N, Kawaguchi M, Huang SK, Nishimura M. Role of interleukin-17F in chronic inflammatory and allergic lung disease. Clin Exp Allergy 2006;36:1109–14. [PubMed: 16961709]
- 121. Oda N, Canelos PB, Essayan DM, Plunkett BA, Myers AC, Huang SK. Interleukin-17F induces pulmonary neutrophilia and amplifies antigen-induced allergic response. Am J Respir Crit Care Med 2005;171:12–8. [PubMed: 15477493]
- 122. Nakae S, Nambu A, Sudo K, Iwakura Y. Suppression of immune induction of collagen-induced arthritis in IL-17-deficient mice. J Immunol 2003;171:6173–7. [PubMed: 14634133]
- 123. Komiyama Y, Nakae S, Matsuki T, Nambu A, Ishigame H, Kakuta S, et al. IL-17 plays an important role in the development of experimental autoimmune encephalomyelitis. J Immunol 2006;177:566– 73. [PubMed: 16785554]
- 124. Nakae S, Komiyama Y, Nambu A, Sudo K, Iwase M, Homma I, et al. Antigen-specific T cell sensitization is impaired in IL-17-deficient mice, causing suppression of allergic cellular and humoral responses. Immunity 2002;17:375–87. [PubMed: 12354389]

- 125. Yang XO, Chang SH, Park H, Nurieva R, Shah B, Acero L, et al. Regulation of inflammatory responses by IL-17F. J Exp Med 2008;205:1063–75. [PubMed: 18411338]
- 126. Liang SC, Tan XY, Luxenberg DP, Karim R, Dunussi-Joannopoulos K, Collins M, et al. Interleukin (IL)-22 and IL-17 are coexpressed by Th17 cells and cooperatively enhance expression of antimicrobial peptides. J Exp Med 2006;203:2271–9. [PubMed: 16982811]
- 127. Zheng Y, Danilenko DM, Valdez P, Kasman I, Eastham-Anderson J, Wu J, et al. Interleukin-22, a T(H)17 cytokine, mediates IL-23-induced dermal inflammation and acanthosis. Nature 2007;445:648–51. [PubMed: 17187052]
- 128. Wolk K, Kunz S, Witte E, Friedrich M, Asadullah K, Sabat R. IL-22 increases the innate immunity of tissues. Immunity 2004;21:241–54. [PubMed: 15308104]
- 129. Kotenko SV, Izotova LS, Mirochnitchenko OV, Esterova E, Dickensheets H, Donnelly RP, et al. Identification, cloning, and characterization of a novel soluble receptor that binds IL-22 and neutralizes its activity. J Immunol 2001;166:7096–103. [PubMed: 11390454]
- 130. Aggarwal S, Xie MH, Maruoka M, Foster J, Gurney AL. Acinar cells of the pancreas are a target of interleukin-22. J Interferon Cytokine Res 2001;21:1047–53. [PubMed: 11798462]
- 131. Zenewicz LA, Yancopoulos GD, Valenzuela DM, Murphy AJ, Karow M, Flavell RA. Interleukin-22 but not interleukin-17 provides protection to hepatocytes during acute liver inflammation. Immunity 2007;27:647–59. [PubMed: 17919941]
- 132. Happel KI, Dubin PJ, Zheng M, Ghilardi N, Lockhart C, Quinton LJ, et al. Divergent roles of IL-23 and IL-12 in host defense against Klebsiella pneumoniae. J Exp Med 2005;202:761–9. [PubMed: 16157683]
- 133. Khader SA, Bell GK, Pearl JE, Fountain JJ, Rangel-Moreno J, Cilley GE, et al. IL-23 and IL-17 in the establishment of protective pulmonary CD4+ T cell responses after vaccination and during Mycobacterium tuberculosis challenge. Nat Immunol 2007;8:369–77. [PubMed: 17351619]
- 134. Umemura M, Yahagi A, Hamada S, Begum MD, Watanabe H, Kawakami K, et al. IL-17-mediated regulation of innate and acquired immune response against pulmonary Mycobacterium bovis bacille Calmette-Guerin infection. J Immunol 2007;178:3786–96. [PubMed: 17339477]
- 135. Wiehler S, Proud D. Interleukin-17A modulates human airway epithelial responses to human rhinovirus infection. Am J Physiol Lung Cell Mol Physiol 2007;293:L505–15. [PubMed: 17545490]
- 136. Rudner XL, Happel KI, Young EA, Shellito JE. Interleukin-23 (IL-23)-IL-17 cytokine axis in murine Pneumocystis carinii infection. Infect Immun 2007;75:3055–61. [PubMed: 17403873]
- 137. Molet S, Hamid Q, Davoine F, Nutku E, Taha R, Page N, et al. IL-17 is increased in asthmatic airways and induces human bronchial fibroblasts to produce cytokines. J Allergy Clin Immunol 2001;108:430–8. [PubMed: 11544464]
- 138. Schnyder-Candrian S, Togbe D, Couillin I, Mercier I, Brombacher F, Quesniaux V, et al. Interleukin-17 is a negative regulator of established allergic asthma. J Exp Med 2006;203:2715– 25. [PubMed: 17101734]
- 139. McKinley L, Alcorn JF, Peterson A, Dupont RB, Kapadia S, Logar A, et al. TH17 cells mediate steroid-resistant airway inflammation and airway hyperresponsiveness in mice. J Immunol 2008;181:4089–97. [PubMed: 18768865]
- 140. Rakoff-Nahoum S, Paglino J, Eslami-Varzaneh F, Edberg S, Medzhitov R. Recognition of commensal microflora by toll-like receptors is required for intestinal homeostasis. Cell 2004;118:229–41. [PubMed: 15260992]
- 141. Hall JA, Bouladoux N, Sun CM, Wohlfert EA, Blank RB, Zhu Q, et al. Commensal DNA Limits Regulatory T Cell Conversion and Is a Natural Adjuvant of Intestinal Immune Responses. Immunity. 2008
- 142. Neurath MF, Fuss I, Kelsall BL, Stuber E, Strober W. Antibodies to interleukin 12 abrogate established experimental colitis in mice. J Exp Med 1995;182:1281–90. [PubMed: 7595199]
- 143. Hue S, Ahern P, Buonocore S, Kullberg MC, Cua DJ, McKenzie BS, et al. Interleukin-23 drives innate and T cell-mediated intestinal inflammation. J Exp Med 2006;203:2473–83. [PubMed: 17030949]
- 144. Kullberg MC, Jankovic D, Feng CG, Hue S, Gorelick PL, McKenzie BS, et al. IL-23 plays a key role in Helicobacter hepaticus-induced T cell-dependent colitis. J Exp Med 2006;203:2485–94. [PubMed: 17030948]
- 145. Schmidt C, Giese T, Ludwig B, Mueller-Molaian I, Marth T, Zeuzem S, et al. Expression of interleukin-12-related cytokine transcripts in inflammatory bowel disease: elevated interleukin-23p19 and interleukin-27p28 in Crohn's disease but not in ulcerative colitis. Inflamm Bowel Dis 2005;11:16–23. [PubMed: 15674109]
- 146. Mannon PJ, Fuss IJ, Mayer L, Elson CO, Sandborn WJ, Present D, et al. Anti-interleukin-12 antibody for active Crohn's disease. N Engl J Med 2004;351:2069–79. [PubMed: 15537905]
- 147. Cho JH. The genetics and immunopathogenesis of inflammatory bowel disease. Nat Rev Immunol 2008;8:458–66. [PubMed: 18500230]
- 148. Eyerich K, Foerster S, Rombold S, Seidl HP, Behrendt H, Hofmann H, et al. Patients with chronic mucocutaneous candidiasis exhibit reduced production of Th17-associated cytokines IL-17 and IL-22. J Invest Dermatol 2008;128:2640–5. [PubMed: 18615114]
- 149. Lee E, Trepicchio WL, Oestreicher JL, Pittman D, Wang F, Chamian F, et al. Increased expression of interleukin 23 p19 and p40 in lesional skin of patients with psoriasis vulgaris. J Exp Med 2004;199:125–30. [PubMed: 14707118]
- 150. Lowes MA, Kikuchi T, Fuentes-Duculan J, Cardinale I, Zaba LC, Haider AS, et al. Psoriasis vulgaris lesions contain discrete populations of Th1 and Th17 T cells. J Invest Dermatol 2008;128:1207– 11. [PubMed: 18200064]
- 151. Itohara S, Farr AG, Lafaille JJ, Bonneville M, Takagaki Y, Haas W, et al. Homing of a gamma delta thymocyte subset with homogeneous T-cell receptors to mucosal epithelia. Nature 1990;343:754– 7. [PubMed: 2154700]
- 152. Scotet E, Nedellec S, Devilder MC, Allain S, Bonneville M. Bridging innate and adaptive immunity through gammadelta T-dendritic cell crosstalk. Front Biosci 2008;13:6872–85. [PubMed: 18508701]
- 153. Crowley MP, Fahrer AM, Baumgarth N, Hampl J, Gutgemann I, Teyton L, et al. A population of murine gammadelta T cells that recognize an inducible MHC class Ib molecule. Science 2000;287:314–6. [PubMed: 10634788]
- 154. Wu J, Groh V, Spies T. T cell antigen receptor engagement and specificity in the recognition of stress-inducible MHC class I-related chains by human epithelial gamma delta T cells. J Immunol 2002;169:1236–40. [PubMed: 12133944]
- 155. Bauer S, Groh V, Wu J, Steinle A, Phillips JH, Lanier LL, et al. Activation of NK cells and T cells by NKG2D, a receptor for stress-inducible MICA. Science 1999;285:727–9. [PubMed: 10426993]
- 156. Pennington DJ, Vermijlen D, Wise EL, Clarke SL, Tigelaar RE, Hayday AC. The integration of conventional and unconventional T cells that characterizes cell-mediated responses. Adv Immunol 2005;87:27–59. [PubMed: 16102571]
- 157. Stark MA, Huo Y, Burcin TL, Morris MA, Olson TS, Ley K. Phagocytosis of apoptotic neutrophils regulates granulopoiesis via IL-23 and IL-17. Immunity 2005;22:285–94. [PubMed: 15780986]
- 158. Roark CL, Simonian PL, Fontenot AP, Born WK, O'Brien RL. gammadelta T cells: an important source of IL-17. Curr Opin Immunol 2008;20:353–7. [PubMed: 18439808]
- 159. Jensen KD, Su X, Shin S, Li L, Youssef S, Yamasaki S, et al. Thymic selection determines gammadelta T cell effector fate: antigen-naive cells make interleukin-17 and antigen-experienced cells make interferon gamma. Immunity 2008;29:90–100. [PubMed: 18585064]





transcription factors