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Effect of dietary conjugated linoleic acid isomers on lipid metabolism in hamsters fed high-carbohydrate and high-fat diets

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Abstract

Dietary conjugated linoleic acids (CLA), have been reported to have a number of isomer-dependent effects on lipid metabolism including reduction in adipose tissue deposition, changes in plasma lipoprotein concentrations and hepatic lipid accumulation. The aim of this study was to compare the effect of individual CLA isomers against lipogenic and high “Western” fat background diets. Golden Syrian hamsters were fed a high-carbohydrate rodent chow or chow supplemented with 17.25% fat formulated to represent the type and amount of fatty acids found in a typical “Western” diet (including 0.2% cholesterol). Diets were further supplemented with 0.25% (w/w) rapeseed oil, *cis*9, *trans*11 (c9,t11)-CLA or *trans*10, *cis*12 (t10,c12)-CLA. Neither isomer had a significant impact on plasma lipid or lipoprotein concentrations. The t10,c12-CLA isomer significantly reduced perirenal adipose tissue depot mass. While adipose tissue acetyl coenzyme A carboxylase and fatty acid synthase mRNA concentrations (as measured by quantitative PCR) were unaffected by CLA, lipoprotein lipase mRNA was specifically reduced by t10,c12-CLA, on both background diets ($p < 0.001$). This was associated with a specific reduction of SREBP1c expression in perirenal adipose tissue ($p = 0.018$). The isomers appear to have divergent effects on liver triacylglycerol content with c9,t11-CLA producing lower concentrations than t10,c12-CLA. We conclude that t10,c12-CLA modestly reduces adipose tissue deposition in the Golden Syrian hamster independently of background diet and this may possibly result from reduced uptake of lipoprotein fatty acids, as a consequence of reduced LPL gene expression.

Keywords

Conjugated linoleic acid; lipoprotein lipase; SREBP; hamster

Introduction

Conjugated Linoleic Acid (CLA) refers to a mix of geometric and positional isomers of linoleic acid where the two double bonds are conjugated. The last decade has seen a plethora of claims, mainly supported by work with animal models that dietary intake of CLA is associated with potential health benefits^{1,2}. These include reduction in fat deposition, protection from atherosclerosis and cancer and enhanced immunity. Translation of these effects into benefits to human health has proven intractable with little evidence to support effects on cardiovascular disease, immunity or cancer²⁻⁴. Effects on fat deposition have also

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been variable, with a recent meta-analysis of 18 human trials concluding that at a dose of 3.2g/day, consumption of mixed CLA isomers (primarily c9,t11-CLA and t10, c12-CLA) produces modest body fat loss in humans⁵. It is of note that mixed isomers of CLA have recently been awarded Generally Regarded as Safe (GRAS) status by the Food and Drug Administration in the USA and can now be added to fluid and flavoured milks, yoghurts, milk-based meal replacements, meal replacement bars, soy milk, and fruit juice*.

The predominant CLA isomer in the diet is the c9,t11 isomer, which is primarily found in milk and meat of ruminant animals, and is produced from desaturation of rumenal *trans*-11 C18:1 through the action of stearoyl CoA desaturase (SCD) in the tissues of the animal⁶. Much smaller amounts of other isomers can also be found in ruminant products. In addition to naturally occurring CLA, dietary supplements are also widely available. Until recently these have tended to be isomeric mixtures (largely equal mixtures of c9,t11-CLA and t10,c12-CLA), however more recently pure c9,t11-CLA and t10,c12-CLA have become available.

Many of the health benefits attributed to CLA have been linked to specific isomers. The t10,c12-CLA isomer is most potent in reducing body fat deposition⁷. The c9,t11-CLA isomer has been suggested to have tumour suppressive actions⁸. Evidence for effects of individual isomers on cardiovascular disease and its associated risk factors has been less conclusive^{2,3}.

Much of the data currently available on the effects of CLA isomers *in vivo* has been gathered from studies using mice. However, it has become increasingly clear that this species shows a much greater reduction in body fat in response to CLA than most other species⁹. In recent years a number of studies have reported the comparative effects of pure CLA isomers on lipid metabolism in the Golden Syrian hamster¹⁰⁻²³. While most of these have reported some degree of reduction in adipose tissue deposition, effects on plasma lipids have been less conclusive. All of these studies have tended to use high doses (0.5% w/w or greater) of CLA against the background of a high fat diet and usually substituting linoleic acid-rich oils with CLA. Such diets have often been supplemented with highly saturated plant oils containing very low concentrations of unsaturated fatty acids. In the present study we feed 0.25% (w/w) pure c9,t11-CLA or t10,c12-CLA against a background diet of either low-fat chow or a high fat diet designed to mimic the fatty acid composition and quantity of a typical "Western" diet.

Materials and Methods

Protocol of animal treatment

All procedures involving hamsters were subject to UK Home Office regulations and animals were housed as previously described¹⁴. Hamsters were anaesthetized using sodium pentobarbitone (Sagatal, 1 mL/kg) and 3-4 ml blood collected by cardiac puncture and placed into EDTA-tubes. While animals were not fasted, they were sacrificed during the light-phase (when food intake would be minimal), between the times of 09.00-12.00h. Plasma was isolated by centrifugation and stored at 4°C until lipoprotein separation which was commenced within 48h of collection. Livers and perirenal and epididymal fat pads were removed, weighed and snap-frozen in liquid nitrogen.

Experimental diets were fed for 6-wk. 8-12-wk old hamsters were randomly divided into 7 groups of 8 animals. Animals were fed either a chow diet (Rat and Mouse Diet 1, Special Diet Supplies) or the same chow supplemented with 172.5g/kg added fat and 0.2%

*http://www.cognis.com/company/Press+and+Media/Press+Releases/2008/080728_EN_NH.htm

cholesterol (High Fat/High Cholesterol (HF/HC):135g/kg beef tallow, 15g/kg tripalmitin and 22.5g/kg corn oil). Diets were prepared in 2kg batches and frozen at -20°C in 500g aliquots until use. Rapeseed oil/CLA were added directly to ground chow. The high fat diet was prepared by adding cholesterol to melted tallow+ tripalmitin and then mixing this with chow, corn oil and rapeseed oil/CLA. Animals were fed every 2-3 days with un-eaten food being completely removed and discarded. The total fat content of the chow diet was approximately 2.8% and that of the HF/HC was 19.4%. The major fatty acids in the chow diet were 4.38% C14:0, 24.4% C16:0, 10.5% C18:0, 27.4% C18:1 and 27.9% C18:2. Those in the HF/HC diet contained 3.21% C14:0, 31.4% C16:0, 16.7% C18:0, 28.6% C18:1 and 12.8% C18:2. Eight of the diets were further supplemented with 2.5g/kg of either high oleic rapeseed oil (human food grade), c9, t11 CLA (90% c9, t11-CLA, 3.8% t10,c12 -CLA, 3.3% c9-C18:1 and 2.5% other conjugated isomers) or t10, c12 -CLA (94.4% t10, c12-CLA, 2.0% c9, t11-CLA, 0.4% c9-C18:1 and 2.9% other conjugated isomers). The rapeseed supplemented chow and HF/HC diets both contained approximately 0.28% and 0.24% of c9, t11-CLA respectively and no detectable t10, c12-CLA. Supplementation with CLA increased the relative amount of the appropriate isomer to approximately 7% and 1% of the total fatty acids in the chow and HF/HC diets respectively. A further HF/HC diet was supplemented with 1% t10, c12-CLA. CLA isomers were provided by Larodan Fine Chemicals AB, Malmö, Sweden). Animals had free access to food and water, and food was replaced completely every 2-3d. Daily food intake was measured between days 21-27 of the trial.

Fatty acid analysis

The fatty acid composition of perirenal adipose tissue was determined by gas chromatography of fatty acid methyl esters as previously described¹⁷.

Lipoprotein separation and cholesterol and triacylglycerol (TAG) analysis

After removal of triacylglycerol-rich lipoproteins ($d < 1.02$ g/ml), plasma LDL ($d = 1.02$ - 1.06 g/ml) and HDL ($d > 1.06$ g/ml) were isolated from hamster plasma as previously described²⁴. Total plasma cholesterol and TAG concentrations were determined using diagnostic kits from Thermo-Trace (Infinity Cholesterol and Infinity TAG Enzymatic kits, Alpha Laboratories). Hepatic cholesterol ester and TAG concentrations were also determined using Thermo-Trace diagnostic kits.

Determination of mRNA levels

RNA was extracted from liver and perirenal adipose tissue using Trizol (Invitrogen) according to the manufacturer's instructions. Genomic DNA was digested with DNase, RNA purity and yield determined and RT-PCR performed as described previously²⁵. For the relative quantification of cDNA for ATP binding cassette transporter A1 (ABC-A1), acetyl CoA carboxylase (ACC), fatty acid synthase (FAS), lipoprotein lipase (LPL), low density lipoprotein receptor (LDLr), sterol regulatory element binding proteins 1a, 1c and 2 (SREBP1a, SREBP1c and SREBP2) quantitative, real-time PCR was performed using either a PRISM 7700 Sequence Detector (Applied Biosystems) or a Lightcycler 480 (Roche). Primer and probe sequences and real-time PCR methods were as previously described²⁵. Standard curves were used to check assay linearity and to determine sample gene expression in RNA equivalents using the C_T values. In addition to the genes of interest the mRNA concentrations for the housekeeping gene, β -actin, were measured and found not to differ between treatment groups. Relative expression of genes of interest were therefore normalized to β -actin and expressed as arbitrary units.

Statistical Analyses

Data from control animals and those fed either 0.25% c9,t11-CLA or t10,c12-CLA were analysed by two-way ANOVA, with diet as one factor (D), and conjugated linoleic acid (C) as another. Results are expressed as means \pm Standard error of the difference (SED). Treatment effects and differences between means were considered significant when $p < 0.05$. Where an effect of CLA was apparent, but no interaction with background diet, data were further analysed by post-hoc Bonferroni test to determine differential effects of isomers. Selected data for animals fed 0.25% and 1% t10, c12-CLA were analyzed by regression analysis to determine dose-dependent effects.

Results

Effect on body composition

The average initial starting weight of all animals was 93.0 ± 8.3 g (mean \pm SD) with no significant differences between groups of animals. Over the course of the experiment animals gained an average of 26.2 ± 9 g. As can be seen in Table 1, no significant difference in weight gain (expressed as a percentage of initial weight) was seen between the groups. While there was no effect of either CLA isomer on food intake, animals adapted to the higher energy density of the HF/HC diet by reducing their food intake.

There was a significant reduction in the relative weight of the perirenal adipose tissue depot in animals fed t10, c12-CLA, which was independent of background diet. Further analysis was performed on data from animals fed the HF/HC diet supplemented with 0%, 0.25% and 1% t10, c12-CLA to ascertain whether the effects on perirenal adipose tissue weight were dose dependent (Fig. 1A). Regression analysis failed to demonstrate a statistically significant relationship ($p = 0.094$). By contrast to the perirenal adipose tissue depot, epididymal fat weight was not influenced by CLA feeding, even when t10,c12-CLA was fed at a level of 1% (relative weights of epididymal tissue in animals fed 1% t10,c12-CLA was 1.79 ± 0.35 (mean \pm standard deviation) and not significantly different to values for animals fed 0% CLA or 0.25% t10,c12-CLA).

Fatty acid analysis of perirenal adipose tissue indicated that in animals fed chow, c9,t11-CLA made up $0.24 \pm 0.06\%$ of total fatty acids, which increased to $0.37 \pm 0.05\%$ in those fed the HF/HC diet. Supplementation of each of these diets with 0.25% c9,t11-CLA increased the content of this fatty acid in perirenal adipose tissue to $1.59 \pm 0.18\%$ and $1.24 \pm 0.06\%$, respectively. The t10,c12-CLA isomer was not detectable in the adipose tissue of animals fed the basal chow and high fat diets. Supplementation of the two diets with this isomer increased tissue levels to $0.55 \pm 0.27\%$ and $0.34 \pm 0.04\%$, respectively.

The HF/HC diet caused a 17% increase in liver weight ($P < 0.001$) which was associated with an accumulation of triacylglycerol and, particularly, cholesterol (Table 1). Thin layer chromatography confirmed that most of the cholesterol was in the esterified form (data not shown). Liver mass was also increased in animals fed t10,c12-CLA with the effect being more pronounced on a chow than a high fat diet (18 vs 4% increase). Divergent effects of CLA isomers were seen on liver triacylglycerol levels with animals fed c9,t11-CLA having lower concentrations than either control or t10, c12-CLA fed animals. In animals fed the HF/HC diet there was a highly significant linear relationship between the amount of t10, c12-CLA in the diet and both relative liver weight (Fig 1A) and TAG content (Fig 1B). Hepatic cholesterol content was not affected by either dose of t10, c12-CLA.

Effect on plasma lipids and lipoproteins

The HF/HC diet significantly increased total plasma cholesterol ($p < 0.001$) and triacylglycerol ($p = 0.015$), however there was no further effects of CLA supplementation (table 2). Both LDL and HDL cholesterol were increased by the HF/HC diet but again no effect of CLA supplementation was seen.

Effect of diet on perirenal adipose tissue mRNA concentrations

Feeding the HF/HC diet suppressed lipogenic gene expression in perirenal adipose tissue (table 3) with levels of FAS, ACC and LPL mRNA all significantly reduced ($P < 0.001$). This was associated with reduced expression of each of the SREBP isoforms. While neither CLA isomer had any effect on FAS, or ACC mRNA, the t10,c12-CLA isomer specifically reduced perirenal adipose LPL mRNA by approximately 50% irrespective of background diet. SREBP-1c mRNA levels were also reduced, by approximately 30%, by t10,c12-CLA independently of background diet.

Effect of diet on hepatic mRNA concentrations

FAS and ACC mRNA levels were decreased, but LPL mRNA concentration was increased, by the HF/HC diet (table 4; $P = 0.001$). Consumption of the HF/HC diet was associated with a decrease in expression of hepatic LDL receptor and SREBP-2 and an increase in ABCA1 mRNA expression ($p < 0.001$). However no effect of the HF/HC diet was observed on the levels of either SREBP1a or 1c transcripts in the liver. In contrast with the perirenal tissue, none of the mRNAs measured in the liver were affected by CLA consumption. Within liver ACC, FAS, LDLr mRNAs were all positively correlated to SREBP2 mRNA but were not related to either SREBP1c or 1a mRNA (table 5). LPL mRNA was positively correlated with ABCA1 mRNA which in turn was correlated with SREBP1c mRNA.

Discussion

Initial observations that CLA reduced adipose tissue deposition were originally performed in mice fed high carbohydrate/low fat diets^{7,9}. It is now quite clear that under these conditions, in this species, CLA induces a lipotrophic syndrome⁹ in which dramatic adipose tissue loss is associated with severe hyperinsulinemia, insulin resistance and hepatic steatosis. More recently a number of studies have been performed in the Golden Syrian Hamster. In general these studies have explored the impact of CLA isomers against a background of high fat/high cholesterol diets. Thus, one of the aims of the present study was to explore effects on both low and high fat diets. The majority of previous hamster studies have fed highly saturated plant oils as the only source of dietary fat. For example, Portillo and colleagues^{12,13,15,18,21} use semi-synthetic diets supplemented with 10% palm oil. As a result, total food energy from fat was approximately 22% and that from n-6 PUFA only about 3%. This contrasts with typical intakes in many industrialized countries of over 35% energy from total fat and 5-6% from n-6 PUFA²⁶. Hayes *et al.*²⁷ have suggested that the responsiveness of animals to changes in fatty acid composition of the diet is exaggerated at low n-6 PUFA intakes. In many previous studies this may be further confounded by the fact that the effect of CLA isomers is directly compared to diets supplemented with equivalent amounts of linoleic acid. For example, Navarro *et al.*¹² showed that adipose tissue mass (epididymal and perirenal) were not significantly different in hamster fed chow compared to those fed a semi-purified diet containing 0.5% t10,c12-CLA. However when compared to a semi-synthetic diet containing 0.5% high-linoleic acid sunflower oil, adipose tissue weights were significantly lower in both the chow-fed animals and those consuming the semi-synthetic diet supplemented with t10, c12-CLA. It is difficult to ascertain whether the changes seen in such studies are a result of CLA or reduced linoleic acid in the diet. In the present study high fat diets were designed to mimic a "Western" diet in terms of both fatty

acid composition and quantity. The high fat diet used provided approximately 44% of energy from total fat and 5.6% from n-6 PUFA, intakes which would not be uncommon in a human 'Western' diet. Supplementation with CLA isomers has been compared to that with high oleic acid –rape seed oil.

Previous studies of the effects of CLA on lipid metabolism in hamsters have used CLA intakes of 0.5-1% (w/w)¹¹⁻²³. Based on typical food intakes and body weights this represents a daily intake of approximately 30-60mg of CLA/day or 250-500mg/kg body weight/day. This would translate to an intake of 20-40g/day in an 80kg human, which would be clearly unachievable. Alternatively, intake can be expressed as a proportion of total energy intake. Based on an energy content of normal laboratory chow of about 10kJ/g, these levels of supplementation with CLA would represent an approximate intake of 0.5-1mg CLA/kJ. To obtain a similar dose, a human consuming 10MJ/day, would need to consume 5-10g CLA/day. It remains to be established which of these estimations is most appropriate and this probably depends on the extent to which effects are dependent on the accumulation of CLA in the tissue. The amount of CLA used in the present study was approximately equivalent to a human intake of 10g/day (on a body weight basis) or 2.5g/day on an energy intake basis). While still high, this is approaching more achievable human supplementation levels.

As we have previously shown, the high fat/high cholesterol diet increased plasma cholesterol and triacylglycerol^{28,29}. While both LDL and HDL cholesterol were increased, the effect on the former was more significant, resulting in an increase in the ratio of LDL to HDL from 0.1 to 0.25. As previously reported^{28,29} cholesterol-feeding was also associated with increased hepatic cholesterol ester concentration and a more modest elevation in hepatic triacylglycerol. Also, as previously seen, hepatic LDLr mRNA concentrations were reduced by the high fat/high cholesterol diet²⁹. LDLr expression has been shown to be largely regulated by the activity of SREBP2³⁰ and indeed there was a highly significant correlation between SREBP2 and LDLr mRNA concentrations ($p < 0.001$). This diet also had marked effects on adipose tissue and hepatic lipogenic gene expression. ACC and FAS mRNA concentrations were reduced in both tissues while LPL was reduced in adipose tissue but increased in liver. Lipogenic gene expression is regulated by the activity of SREBPs³⁰ and expression of all three isoforms was reduced in adipose tissue. In liver, however, only SREBP2 mRNA was reduced. While it has long been suggested that the expression of lipogenic genes such as ACC and FAS are primarily regulated by SREBP1 isoforms, there is evidence to suggest that SREBP2 may also play a role^{31,32}. In fact, this isoform is a much more potent regulator of gene transcription than SREBP1c (the major SREBP1 isoform in liver)³³. In view of a highly significant correlation between hepatic SREBP2 mRNA and both ACC ($p < 0.001$) and FAS ($p < 0.001$) it is interesting to speculate that the reduction in SREBP2 is responsible for the reduction in expression of these two genes. However, it is also possible that nuclear concentrations of SREBP1c mature protein are decreased in the absence of changes in gene expression. LPL mRNA showed tissue specific responses to the HF/HC diet. In adipose tissue, LPL expression was down-regulated and showed a high degree of correlation with ACC, FAS and each of the SREBPs. However, in liver it was actually up-regulated and showed negative associations with ACC, FAS and SREBP2. LPL has been shown to contain a sterol regulatory element within its promoter and therefore might have been expected to change in the same direction as ACC and FAS³⁴. However, it has also been reported to contain response elements for the liver X receptor (LXR)³⁵. LXR α is a nuclear receptor that is activated by oxygenated derivatives of cholesterol and is known to regulate the expression of a range of genes for enzymes and other proteins involved in cholesterol and lipid metabolism³⁶. It is possible that the increased cholesterol content of the diets has led to the generation of increased LXR ligand(s) and this may, in turn lead to the up-regulation of LPL gene expression. The highly significant correlation

between ABC-A1 mRNA and LPL mRNA supports this hypothesis. ABC-A1 is well established as a target gene for LXR37. However, it is of note that ACC and FAS have both been reported to have LXR response elements within their promoters^{38,39}, and we observed negative correlations in expression of these genes relative to ABC-A1. How high fat/high cholesterol diets mediate such divergent effects on these genes warrant further investigation.

Work in mice has shown that t10,c12 CLA dramatically reduces adipose tissue deposition^{7,9}. Recent studies in hamsters have indicated more modest effects on adipose tissue mass^{10,11,18,23,40}. In this study, the t10,c12-CLA isomer also modestly reduced perirenal, but not epididymal adipose tissue mass. It is of note that even when fed at 1%, t10,c12-CLA (as part of a HF/HC diet) failed to reduce epididymal fat mass and no dose-dependent effect was seen on the perirenal depot weight. The effect of t10, c12-CLA on perirenal fat was associated with a reduction in LPL mRNA level, which was decreased by up to almost 50% irrespective of the background diet. The t10,c12-CLA isomer also reduced SREBP-1c mRNA concentrations in the perirenal adipose, and potentially represents the mechanism by which t10,c12-CLA is affecting perirenal LPL mRNA levels. This is supported by a highly significant correlation between LPL and SREBP-1c mRNA levels in perirenal adipose. Zabala *et al.*¹⁸ found that 0.5% and 1% t10,c12 -CLA reduced the mass of epididymal, perirenal and subcutaneous fat depots. The reduction in epididymal fat mass was also shown to be associated with reduced LPL mRNA concentration and enzyme activity. They also demonstrated reduced SREBP1c expression, though in this case SREBP1a was also reduced. It is not clear why no effect on epididymal fat mass was seen in the current experiment. However, it is of note that while Zabala *et al.*¹⁸ were substituting linoleic acid rich oil with CLA, in the current experiment, oleic acid -rich, rapeseed oil was used. Another difference in the findings of these two studies was that while Zabala *et al.*¹⁸ found ACC and FAS expression was reduced in epididymal adipose tissue, in the present experiment these were not affected by t10,c12-CLA in the perirenal depot. The differences in findings between these two studies, depot -specific responses and impact of altering the the linoleic acid content of the diets are all worthy of further study.

In the present study we found no evidence of an effect of either CLA isomer on plasma total, LDL or HDL cholesterol, on either background diet. Previous work in the hamster has produced conflicting results. We have previously reported reduced LDL cholesterol levels in hamsters fed butter enriched in c9,t11-CLA and t11-C18:1¹⁷. While one group has reported reduced LDL in hamsters fed pure c9,t11-CLA²², others have found no effect^{10,12,16}. The results of studies using pure t10,c12-CLA have also produced a range of effects with some authors reporting decreases in LDL cholesterol^{10,12}, one showing an increase¹⁹ and others reporting no effect^{16,22}. The response of HDL cholesterol to pure isomers have been equally varied with some reports of increased HDL with c9,t11-CLA⁴¹ and t10,c12-CLA¹⁶ and others suggesting decreases^{10,20} or no effect^{12,17}. The variability in responses is likely to be a combination of factors including differences in dose of CLA, background diet and age/strain of hamster. It is of note that in humans no significant effect of supplementation with either CLA isomer was seen when comparing to pre-supplementation levels. There was, however, some suggestion of a divergent effect of the two isomers, with c9,t11-CLA decreasing and t10,c12-CLA increasing the ratio of LDL cholesterol to HDL cholesterol⁴². Taken together these data suggests that any effects of CLA using dosages achievable in humans will be small.

The t10,c12-CLA isomer increased liver weight in a dose dependent manner. This is consistent with a number of other hamster studies using concentrations of CLA of 0.5-1%^{10,11,12,15,20}. Previous workers¹⁵ have suggested that the increase in liver weight is associated with an increase in the number of hepatocytes within the livers of animals fed the t10,c12 isomer. At low concentrations we also found divergent effects of the isomers on

liver TAG, with c9,t11-CLA tending to reduce and t10,c12-CLA increase, concentrations. This concurs with previous findings⁴³ which demonstrated divergent effects of these CLA isomers on the level of liver steatosis in apoE knockout mice. One possible theory offered by those authors for this effect is that the c9,t11-CLA may preferentially promote lipolysis in the liver by PPAR α activation. Hepatic TAG increased linearly with dose of t10,c12-CLA, with a doubling of the amount in livers from animals fed 1% t10, c12-CLA. It remains to be established whether feeding a mixture of c9, t11-CLA and t10, c12-CLA will offset the steatosis associated with intake of the latter.

Overall our data suggests low dose supplements with c9, t11-CLA has little impact on lipid metabolism in the hamster irrespective of whether this is fed against the background of a high carbohydrate, lipogenic diet or a high fat/high cholesterol diet. The apparent effect of this isomer in reducing liver TAG is worthy of further investigation as it may potentially offset the steatosis induced by the t10, c12 isomer when mixed isomer preparations are fed. The t10,c12 -CLA isomer reduced the size of the perirenal adipose tissue depot, and this is associated with reduced adipose tissue LPL expression which may be the result of reduced SREBP1c expression. However this was also associated with hepatomegaly and increased hepatic TAG accumulation. The study also highlights important differences in the expression of lipogenic genes in response to high fat/high cholesterol diets and tissue specific differences in these responses.

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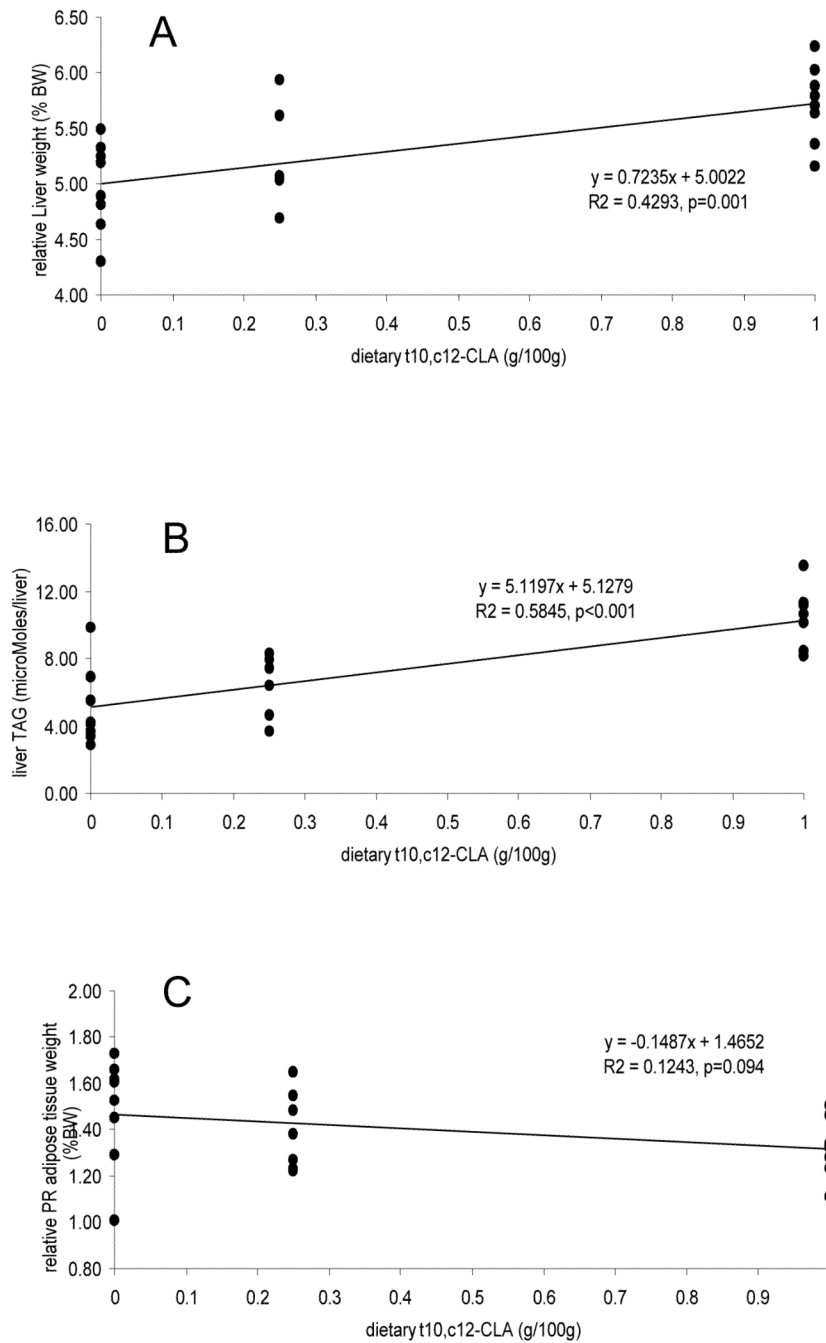


Figure 1.

Groups of 8 hamsters were fed HF/HC supplemented with 0.25% rapeseed oil, 0.25% or 1% t10,c12-CLA. After 6 weeks animals were sacrificed and the weight of perirenal adipose tissue (A) and liver (B) were recorded and expressed relative to total body weight (BW). Liver lipids were extracted and triacylglycerol (TAG) content determined (C).

Table 1
Changes in body composition of hamsters fed a chow or high fat, high cholesterol diet supplemented with 0.25% (wt/wt) CLA isomers (n= 8 animals per group)

		CLA				ANOVA ¹		
		None	e9,t11	t10,e12	mean		SED	P value
Initial body weight (g)	Chow	95.1	90.2	97.2	94.1	D	2.34	0.355
	HF/HC	89.2	90.6	96.0	91.9	C	2.87	0.096
	mean	92.1	90.4	96.6	93.0	D×C	4.06	0.531
Final body weight (g)	Chow	124.9	110.9	120.2	118.7	D	3.72	0.071
	HF/HC	114.1	118.7	127.2	120.0	C	3.03	0.662
	mean	119.5	114.8	123.7	119.3	D×C	5.26	0.025
Change in body weight (g)	Chow	29.8	20.7	23.0	24.5	D	3.07	0.580
	HF/HC	24.9	28.1	31.1	28.0	C	2.51	0.167
	mean	27.3	24.4	27.1	26.3	D×C	4.35	0.067
Daily feed intake (g)	Chow	7.70	6.51	6.70	6.97	D	0.38	<0.001
	HF/HC	5.75	5.65	6.02	5.80	C	0.31	0.248
	mean	6.72	6.08	6.36	6.39	D×C	0.53	0.192
PR adipose weight (g)	Chow	2.05	1.50	1.49	1.68	D	0.16	0.160
	HF/HC	1.71	1.70	1.79	1.73	C	0.13	0.700
	mean	1.88	1.60	1.64	1.71	D×C	0.22	0.106
PR adipose weight (% final BW) ²	Chow	1.61	1.34	1.24	1.40	D	0.08	0.712
	HF/HC	1.49	1.40	1.40	1.43	C	0.09	0.049
	mean	1.55 ^a	1.37 ^{ab}	1.32 ^b	1.41	D×C	0.13	0.334
Epi adipose weight (g)	Chow	2.35	2.01	2.37	2.24	D	0.17	0.441
	HF/HC	2.12	2.38	2.43	2.31	C	0.14	0.628
	mean	2.24	2.19	2.40	2.28	D×C	0.24	0.231
Epi adipose weight (% final BW) ²	Chow	1.86	1.80	1.97	1.88	D	0.10	0.718
	HF/HC	1.84	1.99	1.90	1.91	C	0.09	0.705
	mean	1.85	1.90	1.93	1.89	D×C	0.15	0.434
Liver weight (g)	Chow	5.25	4.45	5.99	5.23	D	0.26	<0.001
	HF/HC	5.67	5.98	6.65	6.10	C	0.22	<0.001
	mean	5.46 ^a	5.21 ^a	6.32 ^b	5.66	D×C	0.37	0.098
Liver weight (% final BW) ²	Chow	4.19	4.03	4.96	4.39	D	0.11	<0.001
	HF/HC	4.99	4.94	5.21	5.04	C	0.14	<0.001
	mean	4.59	4.49	5.08	4.72	D×C	0.19	0.041
Liver TAG (µmole/liver)	Chow	4.63	2.30	4.11	3.68	D	0.50	0.005
	HF/HC	5.05	3.93	6.52	5.17	C	0.61	0.002
	mean	4.84 ^a	3.12 ^b	5.31 ^a	4.42	D×C	0.86	0.275
Liver Total Cholesterol (µmole/liver)	Chow	1.43	1.87	0.45	1.34	D	0.20	<0.001
	HF/HC	14.35	12.27	13.87	13.57	C	0.25	0.240
	mean	7.99	7.21	7.16	7.45	D×C	0.35	0.738

¹Statistical values were obtained by two-way ANOVA with background diet (D, chow or HF/HC) as one factor and CLA (C, none, c9,t11 or t10,c12) as a second factor. SED: standard error of the difference

²Tissue weight expressed as percent of final body weight (BW)

Table 2
Changes in blood lipid profile and liver lipids in hamsters fed a chow or high fat, high cholesterol diet supplemented with 0.25% (wt/wt) CLA isomers (n=8 animals per group)

		CLA			ANOVA ¹		
		None	e9,t11	t10,c12	mean	SED	P value
Total Plasma chol (mM)	Chow	3.53	3.54	3.09	3.39	0.18	<0.001
	HF/HC mean	6.58 5.06	6.39 4.97	6.73 4.91	6.57 4.98	0.22 0.31	0.794 0.187
LDL chol (mM)	Chow	0.23	0.24	0.16	0.21	0.06	<0.001
	HF/HC mean	0.80 0.52	0.83 0.53	0.83 0.50	0.82 0.52	0.08 0.11	0.882 0.759
HDL chol	Chow	2.27	1.97	2.17	2.14	0.12	<0.001
	HF/HC mean	3.08 2.67	3.08 2.52	3.34 2.76	3.16 2.65	0.15 0.21	0.284 0.430
Plasma TAG (mM)	Chow	1.26	1.30	1.44	1.33	0.13	0.015
	HF/HC mean	1.95 1.61	1.55 1.42	1.46 1.45	1.65 1.49	0.15 0.22	0.424 0.094

¹ Statistical values were obtained by two-way ANOVA with background diet (D, chow or HF/HC) as one factor and CLA (C, none, e9,t11 or t10,c12) as a second factor. SED: standard error of the difference

Table 3
Changes in gene expression in perirenal adipose as a result of feeding either a chow or a high fat, high cholesterol diet supplemented with 0.25% (wt/wt) CLA isomers (n=8 animals per group). Data are expressed in arbitrary unit relative to β -actin mRNA

		CLA			ANOVA ¹		
		None	e9,t11	t10,c12	mean	SED	P value
ACC	Chow	1.70	1.43	1.87	1.67	0.16	<0.001
	HF/HC	0.81	0.62	0.58	0.69	0.19	0.533
	mean	1.25	1.05	1.23	1.18	0.27	0.371
FAS	Chow	1.79	1.93	1.55	1.76	0.18	0.001
	HF/HC	0.72	0.83	0.45	0.66	0.22	0.227
	mean	1.25	1.38	1.00	1.21	0.31	0.995
LPL	Chow	1.78	1.51	0.96	1.41	0.14	<0.001
	HF/HC	1.00	1.14	0.53	0.89	0.17	<0.001
	mean	1.39 ^a	1.32 ^a	0.74 ^b	1.15	0.24	0.427
SREBP-1a	Chow	1.63	1.25	1.43	1.44	0.14	<0.001
	HF/HC	0.88	1.23	0.69	0.94	0.17	0.429
	mean	1.26	1.24	1.06	1.19	0.24	0.056
SREBP-1c	Chow	1.66	1.55	1.27	1.49	0.14	0.003
	HF/HC	1.14	1.32	0.64	1.03	0.18	0.018
	mean	1.40 ^a	1.44 ^a	0.96 ^b	1.26	0.25	0.513
SREBP-2	Chow	1.44	1.27	1.19	1.30	0.15	<0.001
	HF/HC	0.97	0.77	0.50	0.75	0.18	0.154
	mean	1.21	1.02	0.84	1.02	0.26	0.803

¹ Statistical values were obtained by two-way ANOVA with background diet (D, chow or HF/HC) as one factor and CLA (C, none, e9,t11 or t10,c12) as a second factor. SED: standard error of the difference

Table 4
Changes in gene expression in the liver as a result of feeding either a chow or a high fat, high cholesterol diet supplemented with 0.25% (wt/wt) CLA isomers (n=8 animals per group). Data are expressed in arbitrary unit relative to β -actin mRNA

		CLA				ANOVA ¹		
		None	e9,t11	t10,c12	mean	SED	P value	
LDLr	Chow	1.81	1.49	1.17	1.49	D	0.13	<0.001
	HF/HC	0.42	0.54	0.34	0.43	C	0.16	0.082
	mean	1.11	1.01	0.75	0.96	D×C	0.23	0.199
ACC	Chow	1.56	1.27	1.35	1.40	D	0.13	0.001
	HF/HC	0.49	0.77	0.38	0.54	C	0.16	0.515
	mean	1.02	1.02	0.87	0.97	D×C	0.22	0.159
FAS	Chow	1.77	1.53	1.55	1.61	D	0.15	<0.001
	HF/HC	0.37	0.70	0.36	0.47	C	0.18	0.647
	mean	1.06	1.12	0.95	1.04	D×C	0.25	0.280
LPL	Chow	0.56	0.54	0.46	0.52	D	0.17	<0.001
	HF/HC	1.81	1.52	1.89	1.74	C	0.21	0.695
	mean	1.19	1.03	1.18	1.13	D×C	0.30	0.569
SREBP-1a	Chow	1.14	1.62	1.24	1.34	D	0.14	0.154
	HF/HC	1.24	1.21	1.05	1.17	C	0.17	0.203
	mean	1.19	1.41	1.15	1.25	D×C	0.24	0.493
SREBP-1c	Chow	1.14	1.19	1.30	1.21	D	0.11	0.587
	HF/HC	1.07	1.30	1.07	1.15	C	0.14	0.569
	mean	1.10	1.25	1.18	1.18	D×C	0.19	0.458
SREBP-2	Chow	2.32	2.06	1.57	1.98	D	0.21	<0.001
	HF/HC	0.72	1.37	0.83	0.97	C	0.25	0.133
	mean	1.52	1.71	1.20	1.48	D×C	0.35	0.137
ABC-AI	Chow	0.81	1.00	0.80	0.87	D	0.12	<0.001
	HF/HC	1.39	1.49	1.36	1.41	C	0.14	0.447
	mean	1.10	1.24	1.08	1.14	D×C	0.20	0.940

¹ Statistical values were obtained by two-way ANOVA with background diet (D, chow or HF/HC) as one factor and CLA (C, none, e9,t11 or t10,c12) as a second factor. SED: standard error of the difference

Table 5

Correlation matrices for mRNA concentrations in liver

FAS	0.962***								
LPL	-0.542***	-0.618***							
LDLr	0.672***	0.698***	-0.615***						
ABCA1	-0.339*	-0.419**	0.824***	-0.382**					
SREBP1a	0.271	0.253	-0.168	0.418**	0.036				
SREBP1c	0.379*	0.283	0.053	0.203	0.450**	0.385**			
SREBP2	0.626***	0.694***	-0.453**	0.691***	-0.317*	0.151	0.002		
	ACC	FAS	LPL	LDLr	ABCA1	SREBP1a	SREBP1c		

* Figures represent correlation coefficients, p<0.05

** Figures represent correlation coefficients, p<0.01

*** Figures represent correlation coefficients, p<0.001