

## RESEARCH PAPER

# Effect of $\Delta^9$ -tetrahydrocannabinol, a cannabinoid receptor agonist, on the triggering of transient lower oesophageal sphincter relaxations in dogs and humans

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**Background and purpose:** Transient lower oesophageal sphincter relaxations (TLESRs) are the main mechanism underlying gastro-oesophageal reflux and are a potential pharmacological treatment target. We evaluated the effect of the CB<sub>1</sub>/CB<sub>2</sub> receptor agonist  $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC) on TLESRs in dogs. Based on these findings, the effect of  $\Delta^9$ -THC was studied in healthy volunteers.

**Experimental approach:** In dogs, manometry was used to evaluate the effect of  $\Delta^9$ -THC in the presence and absence of the CB<sub>1</sub> receptor antagonist SR141716A on TLESRs induced by gastric distension. Secondly, the effect of 10 and 20 mg  $\Delta^9$ -THC was studied in 18 healthy volunteers in a placebo-controlled study. Manometry was performed before and for 3 h after meal ingestion on three occasions.

**Key results:** In dogs,  $\Delta^9$ -THC dose-dependently inhibited TLESRs and reduced acid reflux rate. SR141716A significantly reversed the effects of  $\Delta^9$ -THC on TLESRs. Similarly, in healthy volunteers,  $\Delta^9$ -THC significantly reduced the number of TLESRs and caused a non-significant reduction of acid reflux episodes in the first postprandial hour. In addition, lower oesophageal sphincter pressure and swallowing were significantly reduced by  $\Delta^9$ -THC. After intake of 20 mg, half of the subjects experienced nausea and vomiting leading to premature termination of the study. Other side-effects were hypotension, tachycardia and central effects.

**Conclusions and implications:**  $\Delta^9$ -THC significantly inhibited the increase in meal-induced TLESRs and reduced spontaneous swallowing in both dogs and humans. In humans,  $\Delta^9$ -THC significantly reduced basal lower oesophageal sphincter pressure. These findings confirm previous observations in dogs and indicate that cannabinoid receptors are also involved in the triggering of TLESRs in humans.

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**Keywords:** gastro-oesophageal reflux disease; cannabinoid; transient lower oesophageal sphincter relaxation;  $\Delta^9$ -tetrahydrocannabinol

**Abbreviations:** GERD, gastro-oesophageal reflux disease; LES, lower oesophageal sphincter; TLESRs, transient lower oesophageal sphincter relaxations; CB, cannabinoid;  $\Delta^9$ -THC,  $\Delta^9$ -tetrahydrocannabinol; DMN, dorsal motor nucleus of the vagus; NTS, nucleus tractus solitarius; PPIs, proton pump inhibitors

## Introduction

Gastro-oesophageal reflux disease (GERD) is a common disorder with symptoms like heartburn, regurgitation and retrosternal pain (Heading, 1999), resulting from prolonged acid exposure of the oesophageal mucosa. The lower oesophageal sphincter (LES) plays a central role in regulating flow across

the gastro-oesophageal junction by generating a tonic pressure to prevent reflux of gastric contents into the oesophagus. Transient lower oesophageal sphincter relaxations (TLESRs), occurring in the absence of a swallow, are the main mechanism underlying reflux in healthy volunteers and in patients with GERD (Dent *et al.*, 1980; Dodds *et al.*, 1982; Holloway, 2000). Therefore, pharmacological inhibition of TLESRs is a potential target to treat GERD.

TLESRs are triggered by gastric distension resulting from an increased activation of gastric mechanoreceptors in the subcardiac region of the stomach. This leads to activation of a reflex pathway involving gastric vagal afferents, brainstem

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integrative circuits and efferent inhibitory pathways to the LES and crural diaphragm (Mittal *et al.*, 1995). Vagal cooling and cervical vagotomy abolish the triggering of TLESRs, indicating that TLESRs are mediated by a vago-vagal pathway (Martin *et al.*, 1986; Abrahams *et al.*, 2002). The insight in the neural pathway and the neurotransmitters involved in this reflex has increased enormously (Hirsch *et al.*, 2002). To date the most potent pharmacological agents to reduce the rate of TLESRs are GABA<sub>B</sub> receptor agonists. The GABA<sub>B</sub> receptor agonist baclofen reduces the rate of TLESRs and the number of acid reflux episodes in normal subjects (Lidums *et al.*, 2000) and in GERD patients (Zhang *et al.*, 2002; Koek *et al.*, 2003; Vela *et al.*, 2003). This effect most likely depends on inhibition of mechanosensitive gastric vagal afferents and their central synaptic connections with brain stem neurons, leading to a raised threshold for action potential firing and reduction of transmitter release respectively (Partosoedarso *et al.*, 2001).

Cannabinoid (CB) receptors, like GABA<sub>B</sub> receptors, belong to the superfamily of G protein-coupled receptors (Pertwee, 1997; 1999). There are two types of CB receptors, the CB<sub>1</sub> and CB<sub>2</sub> receptors. CB<sub>1</sub> receptors are mainly localized in the central nervous system whereas CB<sub>2</sub> receptors are particularly associated with the immune system. Interestingly, CB<sub>1</sub> receptors have been located in brain areas involved in the triggering of TLESRs as well as in the nodose ganglion from which vagal afferents emanate (Van Sickle *et al.*, 2001; Partosoedarso *et al.*, 2003). Moreover a study in dogs showed that the CB receptor agonist WIN 55,212-2 reduced the occurrence of TLESRs in response to gastric distension by 80% (Lehmann *et al.*, 2002). A study in ferrets confirmed the involvement of CB<sub>1</sub> receptors in the central regulation of LES relaxation and showed the presence of CB<sub>1</sub> receptors in the brain centres involved in the triggering of TLESRs (Partosoedarso *et al.*, 2003). These data indicate that CB<sub>1</sub> agonists may be clinically useful to reduce TLESRs in humans and may have the potential to be used as reflux inhibitors.

To date, no specific CB<sub>1</sub> agonists are available for human use. However, marijuana and its main psychoactive component  $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC) are frequently used as anti-emetics (Tortorice and O'Connell, 1990; Tramer *et al.*, 2001) or as an appetite stimulant for example in patients who fail to respond adequately to conventional therapies, in particular cancer patients receiving chemotherapy or radiotherapy, or patients undergoing HIV therapy (Porter and Felder, 2001; Tramer *et al.*, 2001; Massa and Monory, 2006).  $\Delta^9$ -THC has approximately equal affinity for the CB<sub>1</sub> and CB<sub>2</sub> receptors, but its efficacy is less at the CB<sub>2</sub> receptor (Grotenhermen, 2003). In the present study, therefore, we first evaluated whether  $\Delta^9$ -THC had an inhibitory effect in dogs on the triggering of TLESRs, as previously shown for the specific CB<sub>1</sub> agonist WIN 55,212-2. The pharmacodynamic/kinetic relationship in dogs was then used to design a study in healthy human volunteers evaluating its effect on the occurrence of meal-induced TLESRs.

## Methods

### *Dog study*

**Animals** Ten adult male and female Labrador retrievers were used in the experiments. Cervical oesophagostomies were

made, and after recovery from surgery the dogs were accustomed to rest in a Pavlov stand. Before each experiment, the dogs were fasted overnight with free access to water. A washout of at least 3 days was allowed between experiments. All procedures were approved by the Ethical Committee for Animal Experiments of the Göteborg region.

**Protocol** The dogs were intubated with a water-perfused Dentsleeve multilumen catheter to record gastric, LES and oesophageal pressures. An antimony pH electrode was placed 3 cm above the LES to measure acid reflux episodes and a water-perfused catheter was placed in the hypopharynx to measure swallows.

TLESRs were stimulated by infusion into the stomach through the central lumen of the assembly of an acidified liquid nutrient (30 ml kg<sup>-1</sup>; 100 ml min<sup>-1</sup>) followed by air insufflation (500 ml min<sup>-1</sup>) to maintain gastric pressure between 10 ± 1 mm Hg.

The number of TLESRs was measured for a 45 min period starting from the infusion of the liquid. TLESRs were defined as a rapid decrease of LES pressure (>1 mm Hg s<sup>-1</sup>) to a pressure <2 mm Hg above gastric pressure and a duration of >1 s, without any pharyngeal signal <2 s before onset (Lehmann *et al.*, 1999).

$\Delta^9$ -THC was administered intragastrically (2 ml kg<sup>-1</sup>), 30 min before the start of the experiment (T = 0). In experiments with the CB<sub>1</sub> antagonist SR141716A, the compound was administered intragastrically (2 ml kg<sup>-1</sup>) 45 min before start of measurements, followed by  $\Delta^9$ -THC 15 min later. Except for TLESRs and acid reflux episodes, basal LES pressure, latency time from start to first TLESR and swallowing rate were determined. Basal LES pressure was defined as the average pressure during the 45 min experimental period. Reflux episodes were defined as a drop in pH > 1 u within 5 s. The average pH in the 15 s following nadir pH should be <4.

**Plasma sampling and analysis** Blood samples were taken from a foreleg vein from two dogs at each dose. After separation of the plasma the samples were stored at -18°C until analysis. The analysis of the plasma was performed by using MS/MS detection after liquid extraction and chromatographic separation; 100 µl plasma was extracted by shaking with 700 µl methyl-tert-butyl ether/hexane 50/50 v/v (volume in volume) for 30 min. A 450 µl sample of the organic phase was evaporated to dryness and reconstituted in 150 µl 30% acetonitrile, 0.2% formic acid. A 10 µl aliquot was injected onto a reversed phase C18 column, eluted by a fast gradient and detected by electrospray tandem mass spectrometry in positive ion mode.

**Drugs**  $\Delta^9$ -THC was purchased from Sigma Aldrich (Stockholm, Sweden) and was diluted in 5% ethanol, 80% polyethylene glycol and 15% physiological saline (0.9% NaCl). The intragastrically used doses of  $\Delta^9$ -THC were 0.25 µmol kg<sup>-1</sup> (0.079 mg kg<sup>-1</sup>), 0.40 µmol kg<sup>-1</sup> (0.126 mg kg<sup>-1</sup>) and 0.60 µmol kg<sup>-1</sup> (0.189 mg kg<sup>-1</sup>). SR141716A [rimonabant; N-(piperidin-1-yl)-5-(4-chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxamide hydrochloride] was dissolved in 5% ethanol, 60% polyethylene glycol and 35%

physiological saline (0.9% NaCl). The intragastrically used dose of SR141716A was  $0.22 \mu\text{mol kg}^{-1}$  ( $0.1 \text{ mg kg}^{-1}$ ).

**Data analysis and statistics** Each dog served as its own control. All parameters were calculated with regard to the mean of five preceding control experiments for each dog. Data are presented as means  $\pm$  s.e.mean. Statistical analysis was performed by using Student's paired *t*-tests. A *P* value  $<0.05$  was regarded as statistically significant.

#### Human study

**Subjects** Studies were performed in 18 healthy volunteers (male), ranging in age from 19 to 51 years (median 21.5). Subjects were free of any gastrointestinal symptoms, had no history of gastrointestinal surgery and did not take any medication known to influence gastrointestinal motility. They had no history of cannabis use or any other drug of abuse for at least 2 months prior to the study, and frequent cannabis users were excluded. Urine samples were taken to check for any recent drug abuse. Each volunteer gave written informed consent, and the study was approved by the Medical Ethical Committee of the Academic Medical Centre.

**Study design** The effect of  $\Delta^9$ -THC (10 and 20 mg, p.o.) was evaluated in a randomized double-blind placebo-controlled study. On three different study days, all with a minimum interval of 1 week, subjects underwent combined oesophageal manometry and pH measurements. Intake of alcohol and nicotine was not allowed 24 h prior to and during the study. The studies were performed after an overnight fast. A cannula was inserted into a forearm vein for blood sampling. The manometric and pH catheter were introduced through an anaesthetized nostril and positioned so that the sleeve straddled the LES. The pH electrode was located 5 cm above the proximal margin of the LES. All studies were performed in the sitting position. Recording started just after dosing ( $T = 0$ ). After 45 min of basal recording, the subjects received a meal (550 kcal), consisting of 2 pancakes ( $2 \times 100 \text{ g}$ ) with jam ( $2 \times 15 \text{ g}$ ) and orange juice (200 ml) to trigger the occurrence of TLESRs. After the meal ( $T = 60$ ), patients remained upright, and manometric/pH recordings were obtained for three more hours. Blood pressure and heart rate were recorded before dosing, every 15 min for the first 2 h of the study, and every 30 min for the next 2 h. Blood samples were taken before dosing, at  $T = 30, 60, 120, 240$  and 480 min. After the recording period, subjects had to stay for four more hours in the hospital for further monitoring of blood pressure and pulse once every hour. In order to screen for drug side-effects, symptoms of dizziness, nausea and sleepiness were assessed during the total study period.

**Recording methods** Oesophageal manometry was performed by using a 10 lumen assembly with a sleeve sensor incorporated at its distal end to monitor LES pressure. Side holes monitored pressure in the stomach (1 cm below the distal margin of the sleeve) and at 2, 5, 8, 11, 14, 17 and 20 cm above the LES. A side hole in the pharynx monitored swallows. The side holes and the sleeve were perfused with degassed distilled water at  $0.3 \text{ ml min}^{-1}$  and  $0.6 \text{ ml min}^{-1}$  respectively, by using a pneu-

mohydraulic capillary perfusion pump. The pharyngeal side hole was perfused with air at  $0.6 \text{ ml min}^{-1}$  to reduce pharyngeal triggering of TLESRs. Pressures were sensed by external transducers connected to a polygraph. To measure acid reflux, pH was recorded with an antimony electrode with built-in reference, positioned 5 cm above the proximal margin of the LES. Before and after the study the pH electrode was calibrated at  $37^\circ\text{C}$  by using pH 1.0 and 7.0 buffer solutions. Signals were digitalized, computer-processed, stored and analysed by using commercially available software (Polygram, Syntectics Medical, Stockholm, Sweden).

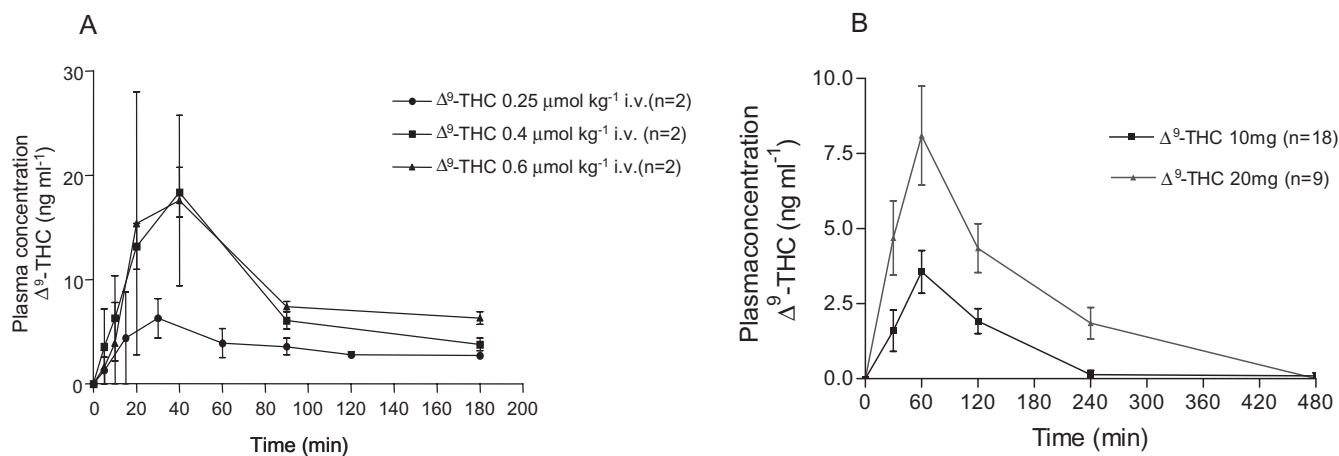
**Materials** The  $\Delta^9$ -THC used was dronabinol (Marinol); the 10 lumen assembly was obtained from Dentsleeve (Adelaide, Australia); the pneumohydraulic capillary perfusion pump from Dentsleeve Pty (Belair, South Australia); the polygraph was from Syntectics Medical (Stockholm, Sweden) and the pH 1.0 and 7.0 buffer solutions were from Medtronic (Skovlunde, Denmark).

**Data analysis** Basal LES pressure was measured at end-expiration relative to intragastric pressure and was determined as visual means of 1 min periods every 15 min.

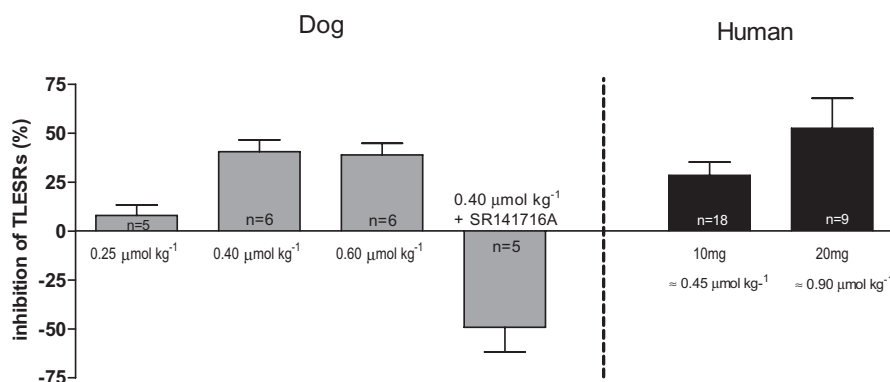
TLESRs were evaluated according to previously published criteria (Holloway *et al.*, 1995): (i) absence of swallowing for 4 s before to 2 s after the onset of LES relaxation; (ii) relaxation rate of  $\geq 1 \text{ mm Hg s}^{-1}$ ; (iii) time from onset to complete relaxation of  $\leq 10 \text{ s}$ ; and (iv) nadir pressure of  $\leq 2 \text{ mm Hg}$ . LES relaxations associated with a swallow that fulfilled the above mentioned criteria ii, iii and iv, and that lasted more than 10 s were included as TLESR. TLESRs were counted for each subject during the preprandial period and the three postprandial hours. In addition, the percentage of TLESRs accompanied by an acid reflux episode was determined, as well as the proportion of acid reflux episodes associated with a TLESR and the total amount of acid reflux episodes and total acid exposure time.

Acid reflux episodes were defined as a decrease in oesophageal pH below 4 for more than 5 s with the pH drop  $> 1 \text{ pH u}$  or, if basal oesophageal pH was already below 4, as a further decrease in pH of at least 1 pH u. Slow drifts of pH below 4 were included in the analysis of the total duration of oesophageal acid exposure but were not scored as acid reflux episodes. The rate of spontaneous swallowing was determined by counting the pharyngeal pressure waves.

**Plasma sampling and analysis** Blood samples were taken from a forearm vein at six time points during each study day. After separation of plasma, samples were stored at  $-20^\circ\text{C}$  until analysis. Metabolism of  $\Delta^9$ -THC occurs mainly in the liver by hydroxylation and oxidation. Nearly 100 metabolites have been identified for  $\Delta^9$ -THC (Grotenhermen, 2003). The two major metabolites are 11-hydroxy-THC and 11-nor-9-carboxy-THC. Analysis of  $\Delta^9$ -THC, 11-hydroxy-THC and 11-nor-9-carboxy-THC was done by Pharma Bio Research (Assen, The Netherlands) through high performance liquid chromatography with tandem mass spectrometric detection, in compliance with the principles of Good Laboratory Practice. Only the results of  $\Delta^9$ -THC plasma concentration are discussed in this study.



**Figure 1** Mean  $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC) plasma concentration in dogs (A) and humans (B). Peak plasma concentrations occurred 40 and 60 min after drug administration in dogs and humans respectively, and declined rapidly with time. Note the difference in scaling of the axes.



**Figure 2** Percentage of inhibition by different doses of  $\Delta^9$ -tetrahydrocannabinol in dog and human studies. Maximal inhibition of transient lower oesophageal sphincter relaxations (TLESRs) occurred up to 40% and 53% in dogs and humans respectively, with the selective  $\text{CB}_1$  receptor antagonist SR141716A reversing the inhibitory effect of  $\Delta^9$ -tetrahydrocannabinol.

**Statistical analysis** Data are presented as the means  $\pm$  s.e.mean. For statistical analysis, Student's paired *t*-test and a mixed model using pairwise comparisons were used. *P* value  $< 0.05$  was considered to be significant. Based on previous studies and on the assumption that a reduction in TLESRs of 40% would occur between placebo and active drug, a sample size of 18 subjects was estimated to be required for detection of this difference at the 5% significance level with 80% power.

## Results

### Dog study

**Plasma THC levels**  $\Delta^9$ -THC was introduced intragastrically at three different doses: 0.25  $\mu\text{mol kg}^{-1}$  (0.079  $\text{mg kg}^{-1}$ ), 0.40  $\mu\text{mol kg}^{-1}$  (0.126  $\text{mg kg}^{-1}$ ) and 0.60  $\mu\text{mol kg}^{-1}$  (0.189  $\text{mg kg}^{-1}$ ). Maximum  $\Delta^9$ -THC plasma levels were reached after 30–40 min at all doses of  $\Delta^9$ -THC (Fig. 1A). No apparent differences in plasma exposure were seen between 0.4 and 0.6  $\mu\text{mol kg}^{-1}$ .

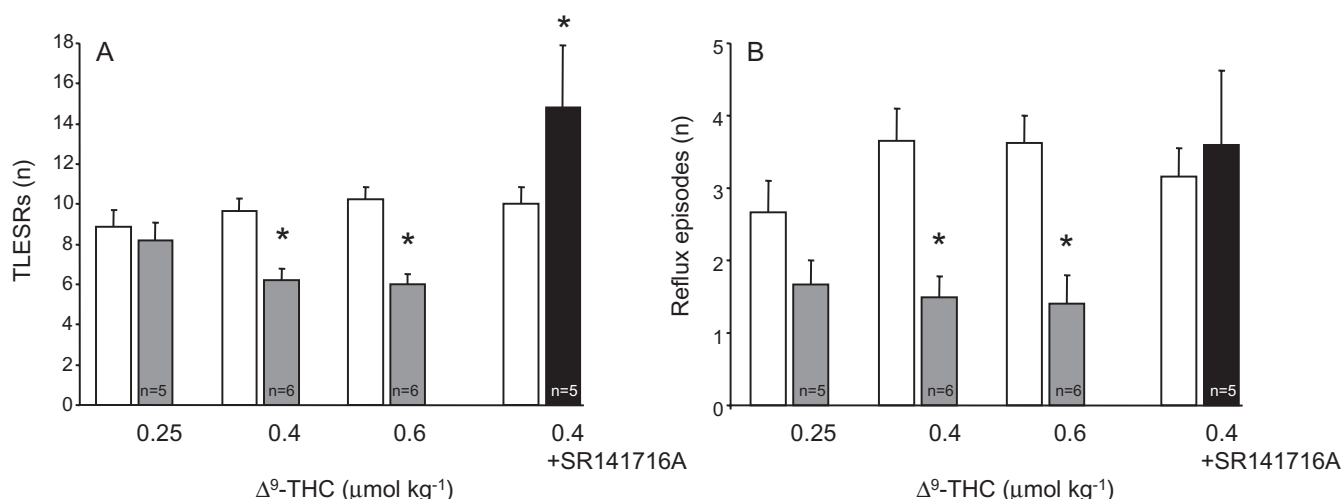
No obvious side-effects were observed at the present doses. A higher dose, 0.8  $\mu\text{mol kg}^{-1}$ , induced emesis in one out

of two dogs, and no further experiments were done at this dose.

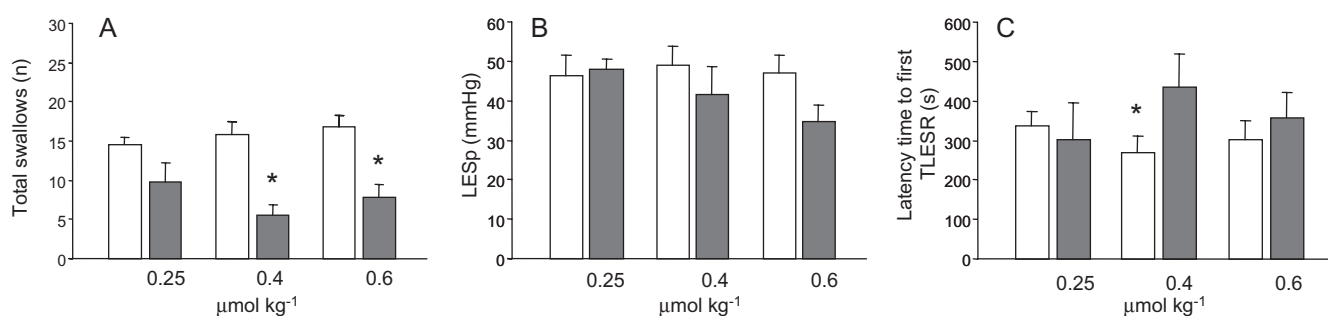
**TLESRs and reflux episodes** The average number of TLESRs in all control experiments was  $9.5 \pm 0.4$  (10 dogs,  $n = 68$ ).  $\Delta^9$ -THC produced a dose-dependent inhibition of TLESRs (Fig. 2). At the highest dose, the rate of TLESRs was significantly reduced during  $\Delta^9$ -THC compared with that in the control experiments (Fig. 3A). Similarly, the number of reflux episodes was significantly reduced by the two highest doses of  $\Delta^9$ -THC (Fig. 3B). The latency time to the occurrence of the first TLESR in the experiments was significantly increased at 0.4  $\mu\text{mol kg}^{-1}$  (Fig. 4C).

Administration of the selective  $\text{CB}_1$  antagonist SR141716A (0.22  $\mu\text{mol kg}^{-1}$ ) prevented the inhibitory response on TLESRs produced by  $\Delta^9$ -THC, 0.4  $\mu\text{mol kg}^{-1}$  ( $n = 5$ ). The overall response was a significant increase in the number of TLESRs (Fig. 2).

**Basal LES pressure, swallowing rate** Basal LES pressure in control conditions was  $45.4 \pm 1.6$  mm Hg. As shown in Figure 4B,  $\Delta^9$ -THC slightly lowered basal LES pressure, but this effect did not reach statistical significance. In contrast, the



**Figure 3** (A) transient lower oesophageal sphincter relaxations (TLESRs) in dogs.  $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC) produced a dose-dependent inhibition of TLESRs. At the highest dose, the rate of TLESRs was significantly reduced from  $10.2 \pm 0.6$  in control experiments to  $6.0 \pm 0.6$  during  $\Delta^9$ -THC. SR141716A reversed the inhibitory effect of  $\Delta^9$ -THC. (B) The number of reflux episodes was significantly reduced by  $\Delta^9$ -THC at the two highest doses. \* $P < 0.05$  compared with controls (Student's paired *t*-test).



**Figure 4** (A) Swallowing rate in dogs was significantly reduced by  $\Delta^9$ -THC at 0.4 and 0.6  $\mu\text{mol kg}^{-1}$ . (B) No significant effect was seen on basal LES pressure in dogs. (C) Latency time to the first transient lower oesophageal sphincter relaxation (TLESR) was significantly increased at the dose of 0.4  $\mu\text{mol kg}^{-1}$ . \* $P < 0.05$  compared with controls (Student's paired *t*-test). LESp, lower oesophageal sphincter pressure.

rate of swallowing was significantly reduced by  $\Delta^9$ -THC at 0.4 and 0.6  $\mu\text{mol kg}^{-1}$  (Fig. 4A). SR141716A blocked the reduction of reflux episodes and swallows induced by  $\Delta^9$ -THC 0.4  $\mu\text{mol kg}^{-1}$ .

#### Human study

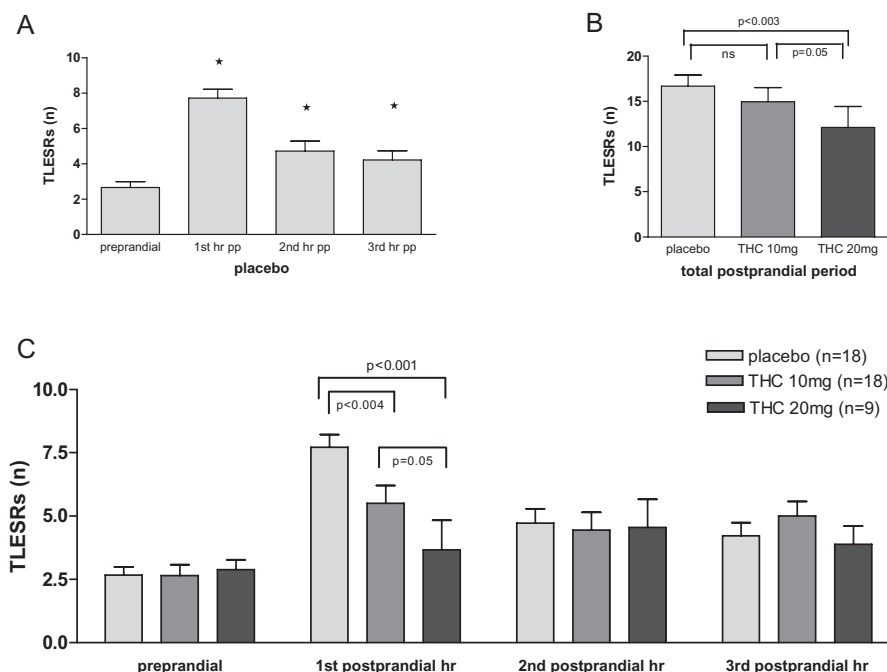
**Plasma THC levels** Maximum  $\Delta^9$ -THC plasma levels were reached at  $T = 60$  min during both doses of  $\Delta^9$ -THC and declined rapidly with time (Fig. 1B). Compared with  $\Delta^9$ -THC 10 mg,  $\Delta^9$ -THC 20 mg reached higher plasma levels and was detected in the plasma for a longer period of time (Fig. 1B). There was a large inter-individual variation in plasma concentrations, with a range in peak concentrations of  $11.8 \text{ ng ml}^{-1}$  during  $\Delta^9$ -THC 10 mg and  $13.6 \text{ ng ml}^{-1}$  during  $\Delta^9$ -THC 20 mg.

**Side-effects** All 18 subjects tolerated the lowest  $\Delta^9$ -THC dose. However, due to nausea and vomiting in the first hour after meal intake, only nine subjects completed the  $\Delta^9$ -THC 20 mg session. Other reported side-effects were dizziness, loss of concentration, sleepiness and confusion. Less frequently reported were a dry mouth and headache. Manifestation of side-effects

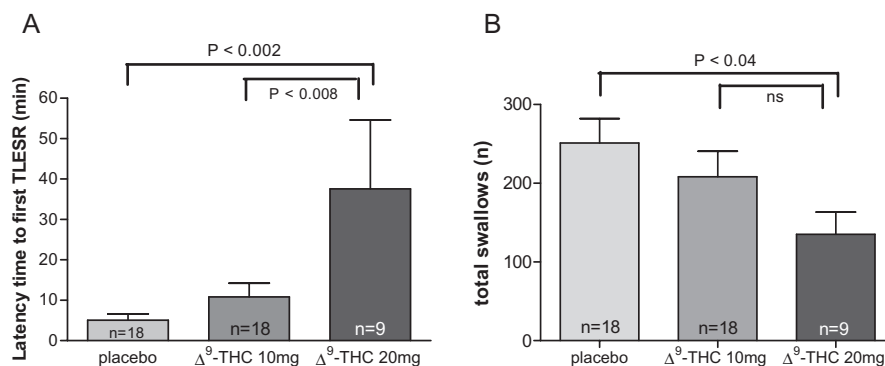
started at approximately  $T = 60$  min, that is at the start of the first postprandial hour and coincided with peak plasma levels. The side-effects were maximal around  $T = 120$  min and lasted for 3–4 h.

**TLESRs and reflux** Before meal intake, the basal rate of TLESRs was comparable in the three different groups (Fig. 5C). During placebo, meal intake resulted in an increase of TLESRs. This increase was most pronounced in the first postprandial hour, whereas in the second and third hours, the number of TLESRs was only marginally increased (Fig. 5A). Treatment with  $\Delta^9$ -THC dose-dependently reduced meal-induced increase of TLESRs during the first postprandial hour (Figs. 5B and 5C). This corresponds to an inhibition by  $\Delta^9$ -THC 10 and 20 mg of  $28.5 \pm 6.7\%$  and  $52.5 \pm 15.3\%$  respectively (Fig. 2). In line with this, latency time to the first TLESR was significantly increased by  $\Delta^9$ -THC 20 mg (Fig. 6A). During the second and third postprandial hours, however, TLESR rate almost returned to preprandial level and did not differ between the three groups (Fig. 5C).

During basal recordings, acid reflux episodes did not occur in either of the study groups. Similar to the number of



**Figure 5** TLESRs in humans. (A) Meal induced increase in transient lower oesophageal sphincter relaxations (TLESRs) was most pronounced during the first postprandial hour. (B)  $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC) 20 mg, but not  $\Delta^9$ -THC 10 mg, significantly reduced the rate of TLESRs during the total three postprandial hours (mixed model using pairwise comparisons). (C)  $\Delta^9$ -THC dose-dependently decreased the rate of TLESRs during the first postprandial hour (placebo  $7.7 \pm 0.5$ ;  $\Delta^9$ -THC 10 mg  $5.5 \pm 0.7$ ;  $\Delta^9$ -THC 20 mg  $3.7 \pm 1.2$ ) (mixed model using pairwise comparisons). No reduction in TLESRs was seen during the second and third postprandial hours \* $P < 0.05$  compared with the preprandial period during placebo (Student's paired *t*-test).



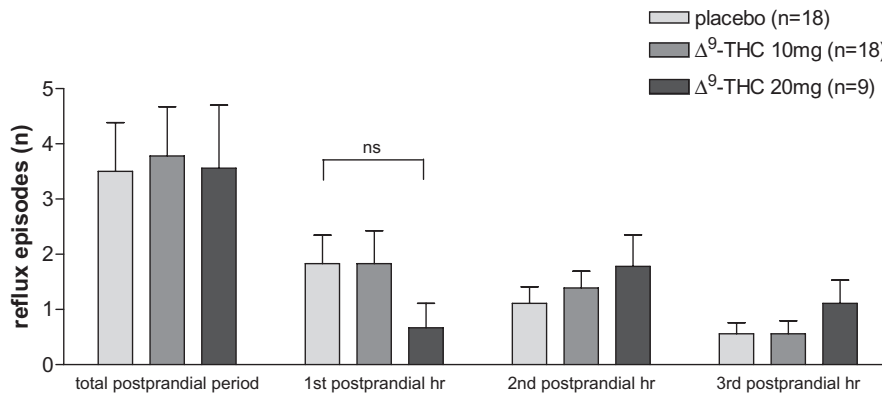
**Figure 6** (A) Latency time in humans was increased significantly from  $5.1 \pm 1.5$  min during placebo to  $37.7 \pm 17.0$  min during  $\Delta^9$ -THC 20 mg ( $P < 0.002$ ). No significant effect was seen with  $\Delta^9$ -THC 10 mg compared with placebo. (B) Mean swallowing rate in humans.  $\Delta^9$ -THC 20 mg induced a significant inhibition of swallowing rate [placebo  $251 \pm 31.1$ ;  $\Delta^9$ -THC 10 mg  $208 \pm 32.6$  (ns);  $\Delta^9$ -THC 20 mg  $135 \pm 28.4$  ( $P < 0.04$ )] (mixed model using pairwise comparisons).  $\Delta^9$ -THC,  $\Delta^9$ -tetrahydrocannabinol; TLESRs, transient lower oesophageal sphincter relaxations.

TLESRs, meal ingestion resulted in an increase in acid reflux episodes in the 3 h postprandial period and in acid exposure time ( $2.9 \pm 1.2\%$ ) during placebo.  $\Delta^9$ -THC did not reduce either the total number of postprandial acid reflux episodes (Fig. 7), or the total acid exposure time [ $\Delta^9$ -THC 10 mg  $3.2 \pm 0.8\%$  (ns);  $\Delta^9$ -THC 20 mg  $4.4 \pm 1.5\%$  (ns)]. During the first postprandial hour, however, the reflux episode rate was decreased with  $\Delta^9$ -THC 20 mg compared with placebo (Fig. 7) but, due to the very low number of episodes involved, this did not reach statistical significance.

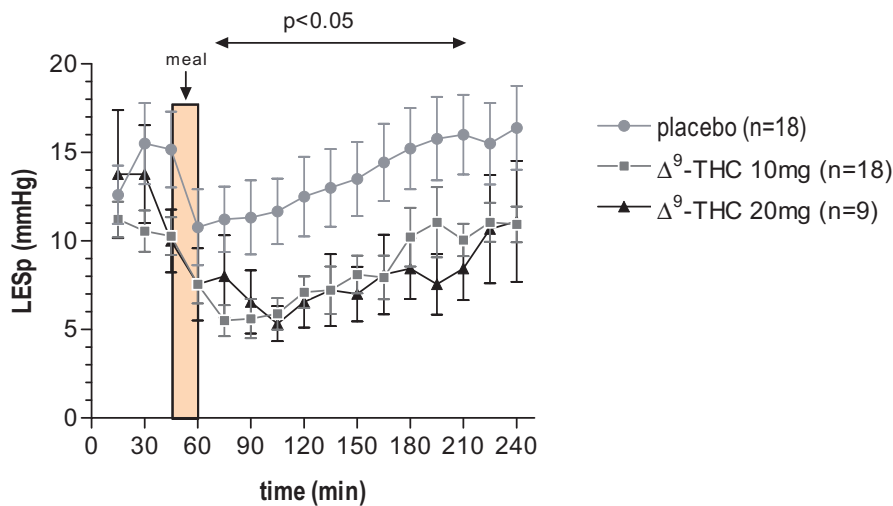
The percentage of TLESRs accompanied by gastro-oesophageal reflux did not significantly differ between the

groups (placebo 17%;  $\Delta^9$ -THC 10 mg 20%;  $\Delta^9$ -THC 20 mg 23%). Acid reflux occurred during a TLESR in 84% of all reflux episodes in the placebo group, compared with 79% in the  $\Delta^9$ -THC 10 mg group (ns) and 87% in the  $\Delta^9$ -THC 20 mg (ns).

**Basal LES pressure** Basal LES pressure was comparable in all three groups. Meal ingestion resulted in a reduction of LES pressure. This effect was maximal directly after the meal at  $T = 60$  min, and returned to baseline around  $T = 150$  min.  $\Delta^9$ -THC induced a further decrease in LES pressure after meal intake compared with placebo. This further decrease started at



**Figure 7** Acid reflux episodes in humans. If the total postprandial period is taken into account there is no difference in reflux episode rate. During the first postprandial hour the amount of reflux episodes was decreased from  $1.83 \pm 0.5$  (placebo) to  $0.67 \pm 0.4$  [ $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC) 20 mg]. This difference, however, was not significant, most likely due to the very few episodes detected.



**Figure 8** Effect of  $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC) on basal LES pressure in humans. Both  $\Delta^9$ -THC 10 mg and 20 mg significantly decreased basal LES pressure starting around T = 60 min and this recovered slowly with time ( $P < 0.05$ ) (mixed model using pairwise comparisons). LESp, lower oesophageal sphincter pressure.

T = 45 min, was maximal around T = 100 min, and recovered slowly with time (Fig. 8). No difference in LES pressure between  $\Delta^9$ -THC 10 mg and 20 mg was observed.

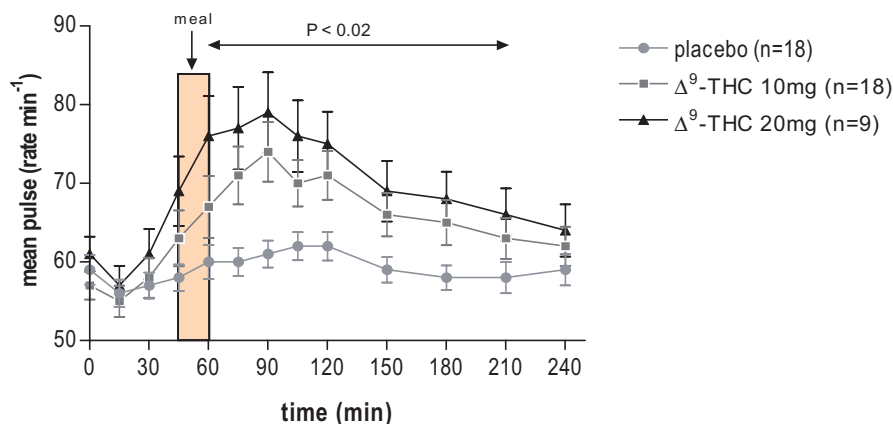
**Swallowing rate** Total swallowing rate was significantly decreased by  $\Delta^9$ -THC 20 mg compared with placebo (Fig. 6B). This decrease was most pronounced during the second and third postprandial hours.

**Blood pressure and pulse** Diastolic and systolic blood pressure were decreased significantly only at T = 45 min and T = 60 min (T = 60; placebo: 130 (111–143)/79 (63–94) mm Hg;  $\Delta^9$ -THC 10 mg: 124 (112–140)/73 (59–89) mm Hg;  $\Delta^9$ -THC 20 mg: 125 (114–140)/71 (66–77) mm Hg ( $P = 0.003$ )).

Tachycardia occurred in both  $\Delta^9$ -THC groups, but was more pronounced after  $\Delta^9$ -THC 20 mg (Fig. 9). Tachycardia started at T = 45 min and reached its maximum around T = 90 min. After  $\Delta^9$ -THC 10 mg the tachycardia lasted till 210 min, whereas after  $\Delta^9$ -THC 20 mg, tachycardia lasted until the end of the experiment (Fig. 9).

## Discussion

TLESRs are the main mechanism underlying gastro-oesophageal reflux, and as such may represent an important potential target for pharmacological treatment of GERD (Dent *et al.*, 1980; Dodds *et al.*, 1982). Insight into the neurotransmitters mediating TLESRs is therefore of crucial importance. Recently, Lehmann *et al.* (2002) showed that exogenous and endogenous activation of CB<sub>1</sub> receptors inhibits TLESRs in dogs; the CB<sub>1</sub> agonist WIN 55,212-2 reduced the occurrence of TLESRs by 80%. In line with this, our study showed that a single dose of the mixed CB<sub>1</sub>/CB<sub>2</sub> agonist  $\Delta^9$ -THC caused a 40% reduction in TLESRs in the same dog model, increased the latency time to the first TLESR and reduced the number of acid reflux episodes. Whether there is a true difference in maximal effect between WIN 55,212-2 and  $\Delta^9$ -THC cannot be ascertained from the present study. Based on these findings, the active dosage was estimated for a human study in which we studied the effect of two different doses of  $\Delta^9$ -THC on the occurrence of



**Figure 9** The effect of the various treatments on mean heart rate in humans. During  $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC) there was a significant tachycardia starting at T = 45 min, with its maximum effect at T = 90 min (placebo 61 beats min<sup>-1</sup>,  $\Delta^9$ -THC 10 mg 74 beats min<sup>-1</sup> and THC 20 mg 79 beats min<sup>-1</sup>); heart rate recovered slowly with time after administration of  $\Delta^9$ -THC. The tachycardia was most marked with  $\Delta^9$ -THC 20 mg (mixed model using pairwise comparisons).

meal-induced TLESRs in healthy subjects. Although the peak plasma levels obtained after a single oral dose of 10 and 20 mg  $\Delta^9$ -THC in healthy subjects was only half of that reached in dogs, TLESRs were significantly inhibited by  $\Delta^9$ -THC. Up to 52% of the TLESRs were inhibited by 20 mg  $\Delta^9$ -THC, but this effect was only observed during the first postprandial hour. It should be emphasized although that in our study, meal intake failed to increase the number of TLESRs during the second and third postprandial hours making it difficult to demonstrate a reduction. Alternatively, the lack of effect on TLESRs in the second and third postprandial hours could result from insufficient serum levels of  $\Delta^9$ -THC. The peak  $\Delta^9$ -THC plasma concentration indeed occurred at the start of the first postprandial hour and declined rapidly with time. However, basal LES pressure was significantly reduced during the entire study, arguing against this possibility. Alternatively, basal LES pressure is more sensitive to  $\Delta^9$ -THC than TLESR.

In addition to its effect on TLESRs,  $\Delta^9$ -THC 20 mg also tended to decrease the amount of reflux episodes by 37% during the first postprandial hour. This effect, however, did not reach statistical significance, most likely due to the very low number of reflux episodes recorded, even during placebo. As the percentage of reflux episodes during TLESRs was not affected by  $\Delta^9$ -THC, the reduction in reflux episodes observed in the first postprandial hour most likely results from the inhibition of TLESRs. On the other hand, the total acid exposure time was not affected by  $\Delta^9$ -THC, suggesting a prolonged acid exposure per acid reflux episode. As clearance of refluxate relies on sufficient swallowing, this finding could be explained by the decrease in swallowing rate observed after  $\Delta^9$ -THC. Suppression of spontaneous swallowing is also observed after baclofen (Blackshaw *et al.*, 1999; Lehmann *et al.*, 1999; Lidums *et al.*, 2000; Zhang *et al.*, 2002) and WIN 55,212-2 (Lehmann *et al.*, 2002).

It should be noted although that  $\Delta^9$ -THC is a non-selective CB receptor agonist binding to other receptors such as the recently described CB receptor GPR55 (Pertwee, 2007; Ryberg *et al.*, 2007). Moreover  $\Delta^9$ -THC has 5-HT<sub>3</sub> receptor blocking properties (Fan, 1995), which may be relevant to

our current findings as these receptors have been shown to play a role in the occurrence of TLESRs (Ruth *et al.*, 1998; Holloway, 2001). However, the highly selective CB<sub>1</sub> receptor antagonist SR141716A (Pertwee, 1997; 1999) effectively blocked the inhibitory effect of  $\Delta^9$ -THC in dogs and even stimulated the occurrence of TLESRs. The stimulating effect induced by the combination of  $\Delta^9$ -THC and SR141716A in the present study (40%) was higher compared with the combination of WIN 55,212-2 and SR141716A (10%) but comparable with previous observations observed with a single administration of SR141716A (Lehmann *et al.*, 2002). Although we cannot exclude the possibility that the effects are due to an interaction with other receptors, the data obtained with the selective CB<sub>1</sub> receptor antagonist SR141716A in dogs suggest that, at least in dogs,  $\Delta^9$ -THC interacts with CB<sub>1</sub> receptors. Ideally, these experiments should have also been tested in humans. However, experiments in healthy subjects evaluating the combination of  $\Delta^9$ -THC and a selective CB<sub>1</sub> antagonist were not performed for ethical reasons, that is, in view of the observed side-effects at the highest dose of THC tested.

Although the exact mechanism of action of  $\Delta^9$ -THC cannot be determined from our experiments, we speculate that it acts peripherally on the afferent limb of the neural pathway and/or within the brainstem nuclei controlling TLESRs. In the human brain, CB<sub>1</sub> receptors are present in the dorsal motor nucleus of the vagus, providing the preganglionic parasympathetic motor innervation of the foregut, and in the nucleus tractus solitarius (NTS), which receives sensory afferent fibres from the autonomic nervous system (Glass *et al.*, 1997). In addition, CB<sub>1</sub> receptor expression has been observed in cell bodies of the nodose ganglion (Partosoedarso *et al.*, 2003). No expression, however, has been shown on vagal efferent neurons (Van Sickle *et al.*, 2001). Therefore,  $\Delta^9$ -THC most probably inhibits TLESRs via the CB<sub>1</sub> receptor on central terminals of vagal primary afferents or on interneurons in the NTS synapsing with vagal motor neurons. The latter site of action may be more likely because vagotomy does not affect CB<sub>1</sub>-like immunoreactivity in the ferret NTS (Partosoedarso *et al.*, 2003).



Although the observed inhibitory effect of  $\Delta^9$ -THC on spontaneous swallowing rate might well be a manifestation of diminished salivation, it could also provide further evidence for a central mechanism of action. Clearly, future studies are required to prove this hypothesis.

A drawback of our study is the occurrence of emesis, and subsequent vomiting in half of the subjects at the highest dose tested leading to premature termination. This was rather unexpected as  $\Delta^9$ -THC, in the same dose range, is clinically used as an anti-emetic in patients undergoing chemotherapy (Tramer *et al.*, 2001). Possibly the combination of meal ingestion and oesophageal intubation may have led to these effects. On the other hand, vomiting is a well documented side-effect of  $\Delta^9$ -THC (Grotenhermen, 2003). Other frequent side-effects were tachycardia, hypotension and psychotropic effects, starting approximately 60 min after drug administration and lasting till the end of the study.

Blockade of TLESRs may be a more physiological approach to prevent both acid and non-acid reflux than currently available therapies, a feature especially important in patients who fail to respond or insufficiently respond to proton pump inhibitors. Previous studies with baclofen indeed showed a significant reduction in reflux episodes and improved symptoms in GERD patients. Although our study shows that  $\Delta^9$ -THC reduces the occurrence of TLESRs by more than 50%, there are several reasons why the use of a mixed CB<sub>1</sub>/CB<sub>2</sub> agonist is not an attractive approach to treat GERD. First of all, administration of  $\Delta^9$ -THC resulted in significant central and cardiovascular side-effects. Secondly, the LES pressure and spontaneous swallowing were both significantly reduced by  $\Delta^9$ -THC, facilitating reflux and impairing the clearance of refluxate respectively. To what extent the same applies to a selective CB<sub>1</sub> agonist cannot be concluded from this study, but clearly potential new CB agonists should be devoid of these properties before they can be considered as interesting new drugs to treat reflux.

In conclusion, the present study demonstrates that  $\Delta^9$ -THC significantly inhibits the increase in TLESRs evoked by meal ingestion and reduces spontaneous swallowing in both dogs and humans. Furthermore,  $\Delta^9$ -THC reduces basal LES pressure in humans. These findings confirm previous findings in dogs and indicate that CB receptors are also involved in the triggering of TLESRs in humans.

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## Conflicts of interest

JJ, AC, MR and AL are all full-time employees of AstraZeneca. GB has participated as an investigator in clinical studies sponsored by AstraZeneca.

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