

## Sitafloxacin Activity against *Helicobacter pylori* Isolates, Including Those with *gyrA* Mutations<sup>▽</sup>

Kazunari Murakami,<sup>1</sup> Tadayoshi Okimoto,<sup>1</sup> Masaaki Kodama,<sup>1</sup> Jin Tanahashi,<sup>1</sup> Toshio Fujioka,<sup>1</sup> Fumiaki Ikeda,<sup>2</sup> Hiroe Muraoka,<sup>2</sup> Motoko Takigawa,<sup>2</sup> Takeshi Saika,<sup>2</sup> Miyuki Hasegawa,<sup>2</sup> and Intetsu Kobayashi<sup>3\*</sup>

Department of Gastroenterology, Faculty of Medicine, Oita University, 1-1 Idaiga-oka, Hasama-machi, Oita 879-5593, Japan<sup>1</sup>; Chemotherapy Division, Mitsubishi Chemical Medicine Corporation, 3-30-1 Shimura, Itabashi-ku, Tokyo 174-8555, Japan<sup>2</sup>; and Toho University School of Medicine, 4-16-20 Omori-nishi, Ota-ku, Tokyo 143-0015, Japan<sup>3</sup>

Received 20 November 2008/Returned for modification 17 December 2008/Accepted 6 April 2009

**Sitafloxacin showed MICs of less than or equal to 0.5 µg/ml against 105 isolates of *Helicobacter pylori*, including 44 isolates with mutations in the *gyrA* gene. The highest MICs for garenoxacin and levofloxacin were 8 and 64 times, respectively, higher than the highest MICs observed for sitafloxacin.**

The guidelines for the management and treatment of *Helicobacter pylori* infections established by the European Helicobacter Study Group Third Masstricht Consensus Report recommend an eradication antimicrobial chemotherapy consisting of amoxicillin, clarithromycin, and a proton pump inhibitor alone or in combination with metronidazole and clarithromycin (10). On the other hand, a trend toward increased clarithromycin resistance in Japan has been reported (8); furthermore, high metronidazole resistance rates associated with *H. pylori* eradication failure have been seen in the United States, Europe, and Asia with the exception of Japan (11). In the search for alternative eradication treatment regimens, it has been recently reported in the United States and Europe that levofloxacin may be efficacious in *H. pylori* eradication therapy (7, 13). At the same time, the increased use of levofloxacin-based eradication regimens has led to increasing resistance to levofloxacin in *H. pylori* as a result of mutations in the quinolone resistance-determining region (QRDR) of the *gyrA* gene correlating with the decreased effectiveness of levofloxacin in eradication regimens (16). Sitafloxacin is a recently developed fluoroquinolone with wide-spectrum activity, ranging from gram-positive cocci to gram-negative bacilli (1, 15). We studied the effect of mutations in the *gyrA* gene and its impact on the antimicrobial activity of sitafloxacin in *H. pylori*.

(This study was presented at the 48th Interscience Conference on Antimicrobial Agents and Chemotherapy, Washington, DC, 25 to 28 October 2008.)

A total of 105 *H. pylori* isolates were recovered from the gastric mucosa of patients presenting with gastroduodenal diseases in health care facilities in Japan between 2004 and 2005. Of the 105 patients, 57 (54.3%) were males and 48 (45.7%) were females, and the average age was 57.9 years (range in age from 21 to 84). None of the patients had previously undergone eradication therapy. The spectrum of peptic ulcers included 39 (37.1%) cases of chronic gastritis, 21 (20.0%) gastric ulcers, 18

(17.1%) duodenal ulcers, 9 (8.6%) gastric cancers, 8 (7.6%) gastroduodenal ulcers, and 10 (9.5%) cases with other causes or an unspecified diagnosis. Only one isolate per patient was included among the 105 isolates.

Susceptibilities to sitafloxacin (Daiichi Sankyo, Japan), garenoxacin [a novel des-fluoro(6)quinolone (Astellas, Japan) (6)], and levofloxacin (Daiichi Sankyo, Japan) were determined by agar dilution method according to CLSI guidelines by using drugs with known potency (4, 5). The agar dilution method was performed by serial twofold dilution on Mueller-Hinton agar (Becton Dickinson, MD) with 5% sheep blood using 1 to 3 µl of a McFarland 2.0-adjusted inoculum and

TABLE 1. Genetic characteristics of *Helicobacter pylori* isolates in this study

Strain and no. of isolates	Amino acid substitution at position:		
	87	91	Other
Strains with wild-type <i>gyrA</i>			
61	Asn	Asp	
Strains with <i>gyrA</i> mutations at Asn87 ( <i>n</i> = 14)			
11	Lys	Asp	
2	Lys	Asp	Asp143Glu
1	Ile	Asp	
Strains with <i>gyrA</i> mutations at Asp91 ( <i>n</i> = 25)			
9	Asn	Gly	
1	Asn	Gly	Asp145Gly
5	Asn	Asn	
2	Asn	Asn	Asp143Glu
6	Asn	Tyr	
1	Asn	Tyr	Ala97Val
1	Asn	Tyr	Ala129Val
Strains with <i>gyrA</i> mutations in other regions ( <i>n</i> = 5)			
1	Asn	Asp	Thr62Ile
1	Asn	Asp	Asp99Val
1	Asn	Asp	Arg130Lys
1	Asn	Asp	Asp143Glu
1	Asn	Asp	Lys158Arg

\* Corresponding author. Mailing address: Toho University School of Medicine, 4-16-20 Omori-nishi, Ota-ku, Tokyo 143-0015, Japan. Phone and fax: 81-3-3762-9247. E-mail: kobatora@ja2.so-net.ne.jp.

<sup>▽</sup> Published ahead of print on 20 April 2009.

TABLE 2. Correlation of quinolone MICs with *gyrA* mutations for 105 *Helicobacter pylori* isolates

Agent	<i>gyrA</i> mutation	No. of isolates	MIC range	MIC <sub>50</sub>	MIC <sub>90</sub>
Sitafloxacin	Asn87	14	0.03–0.5	0.25	0.5
	Asp91	25	≤0.015–0.25	0.12	0.25
	Other	5	≤0.015–0.12		
	None	61	≤0.015–0.06	0.03	0.03
Garenoxacin	Asn87	14	0.12–4	0.5	1
	Asp91	25	0.12–2	0.5	1
	Other	5	≤0.015–0.5		
	None	61	≤0.015–0.25	0.03	0.06
Levofloxacin	Asn87	14	4–32	8	16
	Asp91	25	2–8	4	8
	Other	5	0.12–4		
	None	61	0.12–2	0.25	0.25

incubation at  $35 \pm 2^\circ\text{C}$  for 72 h under microaerophilic conditions. For quality control, *H. pylori* ATCC 43504 was tested with each run.

PCR amplification and sequence analysis of the QRDR of *gyrA* was performed as previously described by Nishizawa et al. (12). The specific primers used for amplification were GYRA-F (5'-T TTRGCTTATTCMATGAGCGT-3') and GYRA-R (5'-GCAG ACGGCTTGGTARAATA-3'). For sequencing, the ABI Prism 3130xl genetic analyzer (Perkin-Elmer, ABI, CA) was used. Sequences were compared to that of the *H. pylori* wild-type strain (GenBank accession no. L29481).

Forty-four of the 105 *H. pylori* isolates exhibited mutations in the *gyrA* gene (Table 1). Mutations at Asn87 were observed in 14 isolates, while mutations were seen at Asp91 in 25 isolates. The remaining five isolates had mutations in other regions. Table 2 shows the effect of changes in the *gyrA* gene in the QRDR and its effect on the MICs of sitafloxacin, garenoxacin,

and levofloxacin. Mutations involving Asn87 resulted in a shift to higher MIC levels of the drugs than mutations in other regions. Sitafloxacin demonstrated the narrowest MIC distribution with a MIC of  $\leq 0.5$   $\mu\text{g/ml}$  against all isolates. In contrast, the highest MICs for garenoxacin and levofloxacin were 8 and 64 times, respectively, higher than the highest MIC observed for sitafloxacin. A scattergram depicting sitafloxacin MICs versus levofloxacin or garenoxacin MICs for 105 isolates with or without *gyrA* mutations is shown in Fig. 1. From the scattergram, the increase in garenoxacin and levofloxacin MICs relative to the sitafloxacin MICs is observed. With respect to levofloxacin-resistant isolates with MICs ranging from 2 to 32  $\mu\text{g/ml}$ , garenoxacin and sitafloxacin demonstrated MICs ranging from 0.125 to 4  $\mu\text{g/ml}$  and  $\geq 0.015$  to 0.5  $\mu\text{g/ml}$ , respectively.

In recent years, the increased resistance to clarithromycin and metronidazole of *H. pylori* has led to the inclusion of fluoroquinolones in eradication therapy in light of their minimal side effects and improved eradication rates (7, 13). However, resistance to a fluoroquinolone as a result of mutations in *gyrA* has been reported (12). Furthermore, a significant reduction in eradication effectiveness was observed when levofloxacin was used against *H. pylori* isolates exhibiting *gyrA* mutations compared to *H. pylori* isolates with no mutations in *gyrA* (16). Nishizawa et al. found elevated MICs to gatifloxacin along with mutations in *gyrA* in 47.9% of the isolates recovered from patients who had failed *H. pylori* eradication therapy (12).

We observed that 41 (39%) of the *H. pylori* isolates recovered from patients prior to undergoing their first eradication regimen already showed resistance to levofloxacin based on a breakpoint of  $>1$   $\mu\text{g/ml}$  (2). The rate of resistance to levofloxacin we observed was considerably higher than the figure found in the *American College of Gastroenterology Guideline*

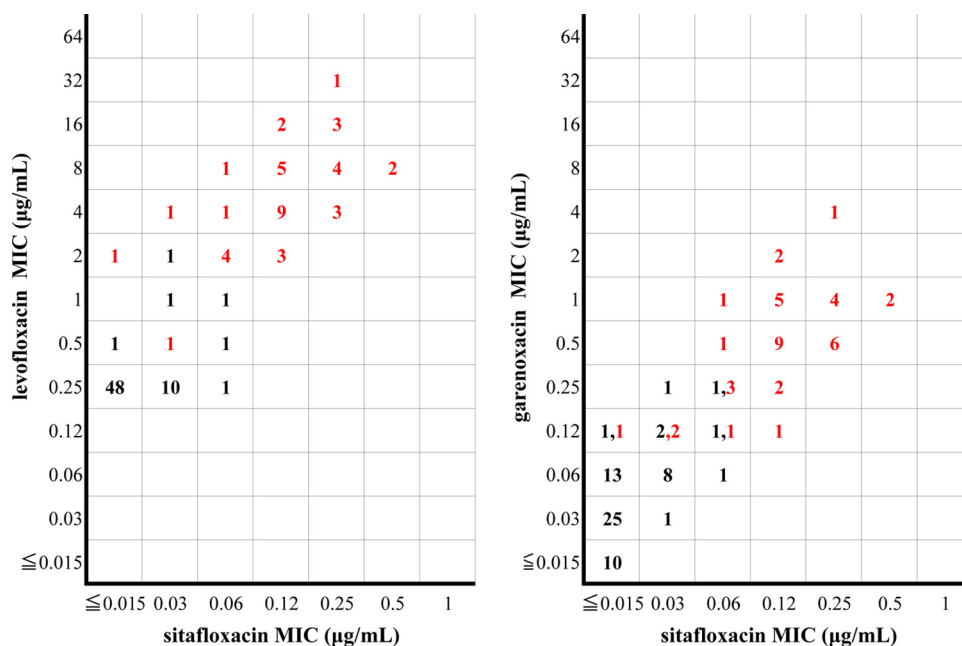


FIG. 1. Scattergram of sitafloxacin MICs versus levofloxacin or garenoxacin MICs for 105 *Helicobacter pylori* isolates. The red and black numbers indicate the frequency of isolates with and without *gyrA* mutations, respectively.

(3). The prevalence of levofloxacin-resistant *H. pylori* may be associated with the increasing use of fluoroquinolones in clinical practice for many indications in Japan since the 1980s. There was little difference observed in the rate of resistance to clarithromycin between levofloxacin-resistant (14/41 [34%]) and -susceptible isolates (20/64 [31%]) in this study (data not shown). Compared to data generated in southeast Asia and Europe (9, 14), the prevalence of metronidazole-resistant isolates observed in our study was lower (5/105 [4.8%]). Two of the five isolates were resistant to levofloxacin.

Bogaerts et al. has previously reported that two amino acid substitutions due to mutations in *gyrA* elevated MICs to levofloxacin, ciprofloxacin, and moxifloxacin (2). While we did not observe *H. pylori* isolates with mutations in both Asn87 and Asp91, there is a need for continued surveillance for the emergence of high resistance in Japan in light of the high use of fluoroquinolones.

In this study, we observed the superior antibacterial activity of sitafloxacin against *H. pylori* compared to that of levofloxacin and garenoxacin even in the presence of mutations in *gyrA*. With respect to other organisms, sitafloxacin has been reported to have high affinity to DNA gyrase and topoisomerase IV along with superior antibacterial activity (1). As *H. pylori* lacks the topoisomerase IV enzyme, the high affinity of sitafloxacin to DNA gyrase may account for its lower MIC against *H. pylori*. Our report is the first to demonstrate the antibacterial activity of sitafloxacin against *H. pylori* isolates with *gyrA* mutations. Based on sitafloxacin's superior antibacterial activity, clinical trials of second- or third-line eradication therapy including sitafloxacin are warranted.

#### REFERENCES

1. Akasaka, T., S. Kurosaka, Y. Uchida, M. Tanaka, K. Sato, and I. Hayakawa. 1998. Antibacterial activities and inhibitory effects of sitafloxacin (DU-6859a) and its optical isomers against type II topoisomerases. *Antimicrob. Agents Chemother.* **42**:1284–1287.
2. Bogaerts, P., C. Berhin, H. Nizet, and Y. Glupczynski. 2006. Prevalence and mechanisms of resistance to fluoroquinolones in *Helicobacter pylori* strains from patients living in Belgium. *Helicobacter* **11**:441–445.
3. Chey, W. D., B. C. Wong, and the Practice Parameters Committee of the American College of Gastroenterology. 2007. American College of Gastroenterology guideline on the management of *Helicobacter pylori* infection. *Am. J. Gastroenterol.* **102**:1808–1825.
4. Clinical and Laboratory Standards Institute. 2006. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. Approved standard M7-A7, 7th ed. Clinical and Laboratory Standards Institute, Wayne, PA.
5. Clinical and Laboratory Standards Institute. 2008. Performance standards for antimicrobial susceptibility testing; 18th informational supplement. M100-S18. Clinical and Laboratory Standards Institute, Wayne, PA.
6. Fung-Tomc, J. C., B. Minassian, B. Kolek, E. Huczko, L. Aleksunes, T. Stickle, T. Washo, E. Gradelski, L. Valera, and D. P. Bonner. 2000. Antibacterial spectrum of a novel des-fluoro(6) quinolone, BMS-284756. *Antimicrob. Agents Chemother.* **44**:3351–3356.
7. Gisbert, J. P., and F. Morena. 2006. Systematic review and meta-analysis: levofloxacin-based rescue regimens after *Helicobacter pylori* treatment failure. *Aliment. Pharmacol. Ther.* **23**:35–44.
8. Kobayashi, I., K. Murakami, M. Kato, S. Kato, T. Azuma, S. Takahashi, N. Uemura, T. Katsuyama, Y. Fukuda, K. Haruma, M. Nasu, and T. Fujioka. 2007. Changing antimicrobial susceptibility epidemiology of *Helicobacter pylori* strains in Japan between 2002 and 2005. *J. Clin. Microbiol.* **45**:4006–4010.
9. Kulsunti Wong, P., C. Chomvarin, K. Chaicumpar, W. Namwat, W. Kaewkes, P. Mairlang, and A. Sangchan. 2008. Antimicrobial susceptibility of *Helicobacter pylori* isolated from gastric biopsies in dyspeptic patients. *Southeast Asian J. Trop. Med. Public Health* **39**:1102–1109.
10. Malfertheiner, P., F. Megraud, C. O'Morain, F. Bazzoli, E. El-Omar, D. Graham, R. Hunt, T. Rokkas, N. Vakil, and E. J. Kuipers. 2007. Current concepts in the management of *Helicobacter pylori* infection: the Maastricht III Consensus Report. *Gut* **56**:772–781.
11. Mégraud, F. 2004. *H. pylori* antibiotic resistance: prevalence, importance, and advances in testing. *Gut* **53**:1374–1384.
12. Nishizawa, T., H. Suzuki, K. Kurabayashi, T. Masaoka, H. Muraoka, M. Mori, E. Iwasaki, I. Kobayashi, and T. Hibi. 2006. Gatifloxacin resistance and mutations in *gyrA* after unsuccessful *Helicobacter pylori* eradication in Japan. *Antimicrob. Agents Chemother.* **50**:1538–1540.
13. Nista, E. C., M. Candelli, M. A. Zocco, F. Cremonini, V. Ojetti, R. Finizio, C. Spada, G. Cammarota, G. Gasbarrini, and A. Gasbarrini. 2006. Levofloxacin-based triple therapy in first-line treatment for *Helicobacter pylori* eradication. *Am. J. Gastroenterol.* **101**:1985–1990.
14. Romano, M., M. R. Iovene, M. I. Russo, A. Rocco, R. Salerno, D. Cozzolino, A. P. Pilloni, M. A. Tufana, and G. Nardone. 2008. Failure of first-line eradication treatment significantly increases prevalence of antimicrobial-resistant *Helicobacter pylori* clinical isolates. *J. Clin. Pathol.* **61**:1112–1115.
15. Sato, K., K. Hoshino, M. Tanaka, I. Hayakawa, and Y. Osada. 1992. Antimicrobial activity of DU-6859, a new potent fluoroquinolone, against clinical isolates. *Antimicrob. Agents Chemother.* **36**:1491–1498.
16. Shirasaka, D., N. Aoyama, I. Miki, Y. Matsumoto, H. Miyaji, M. Toyoda, T. Mitani, Y. Morita, T. Tamura, and T. Azuma. 2006. The efficiency and safety of second-eradication therapy for *Helicobacter pylori* with metronidazole and levofloxacin. *J. Gastroenterol.* **103S**:782. (In Japanese.)