

Disproportionation and self-sorting in molecular encapsulation

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Self-assembled capsules are nanoscale structures made up of multiple synthetic subunits held together by weak intermolecular forces. They act as host structures that can completely surround small molecule guests of the appropriate size, shape and chemical surface. Like their biological counterparts, multimeric enzymes and receptors, the subunits of the capsules are generally identical, and lead to homomeric assemblies of high symmetry. In both biological and synthetic systems small variations in structures are tolerated and lead to heteromeric assemblies with slightly different recognition properties. The synthetic capsules are dynamic, with lifetimes from milliseconds to hours, and allow the direct spectroscopic observation of smaller molecules inside, under ambient conditions at equilibrium in solution. We report here the assembly of hybrid capsules made up of 2 very different structures, both capable of forming their own homomeric capsules through hydrogen bonding. These hybrids exhibit host properties that differ markedly from the parent capsules, and suggest that other capsules may emerge from seemingly unrelated modules that have curved surfaces and are rich in hydrogen bonding capabilities.

Reversible encapsulation complexes are supramolecular structures in which guest molecules are completely surrounded by a self-assembled host structure (1). The complexes provide access to isolated species that cannot be seen in bulk solution (2); they act as nanometric reaction chambers (3), as means to stabilize reagents (4,5), and as spaces where new forms of stereochemistry can emerge (6). The forces holding the assembly together can be hydrophobic effects (7), salt bridges (8), metal/ligand interactions (9,10), and hydrogen bonds. The capsules form when, and only when, suitable guests are present to fill the space inside. For example, 6 molecules of resorcinarene **1** (Fig. 1) and 8 molecules of water form a hexameric capsule **1₆** in the solid state through a seam of 60 hydrogen bonds (11). The capsule also self-assembles in solution with wet solvents such as chloroform and benzene inside (12,13) or in the presence of large quaternary ammonium guests (14,15). The resorcinarene can be elaborated to the cavitand **2**, which dimerizes in chloroform to capsule **2₂** through a seam of 8 bifurcated hydrogen bonds. When dissolved in chloroform, the capsule forms around 3 solvent molecules. Alternatively, a hybrid structure **1.2** forms immediately on mixing the 2 homomeric capsules **1₆** and **2₂** and all 3 capsule assemblies coexist in the same solution (16). The exchange of partners was surprising, because “self-assembly” implies a distinction between self and nonself, with some selection, correction and sorting at work (17,18), but these capsules are related because they share resorcinarene modules that fix their dimensions and symmetries. Here, we report a new example of hybrid formation from 2 unrelated capsules.

Results

The capsule **3₂** is a notional “tennis ball” (19) that self-assembles through hydrogen bonding around small molecules such as methane. At first glance, the formation of a hybrid between **2₂** and **3₂** appears unlikely, given the different sizes, hydrogen bonding patterns, and symmetries of the modules, but modeling gives cause for optimism (Fig. 1). Slight flexing of **3** at its 7-membered rings and **2** at its 9-membered rings brings about a

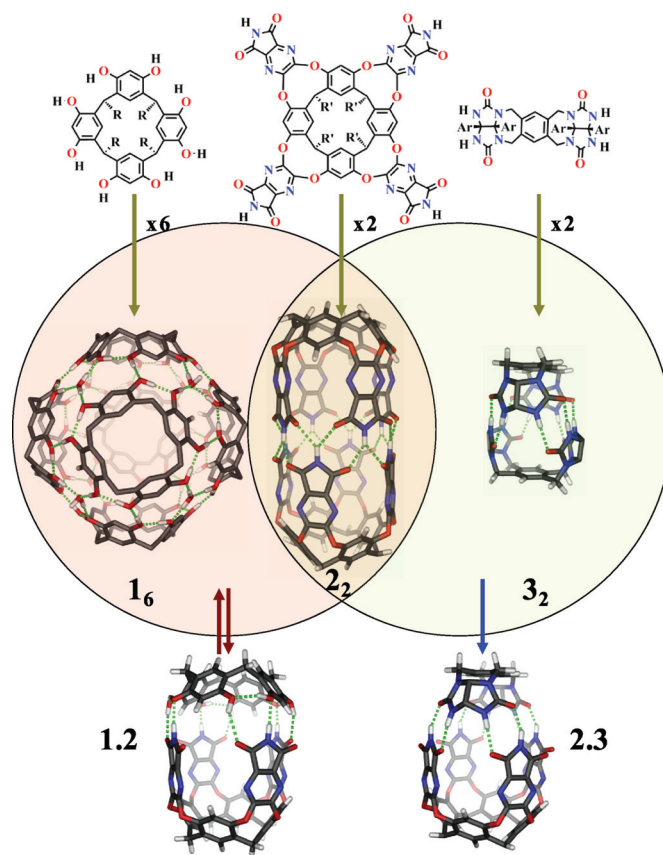


Fig. 1. Line drawings of the subunits and energy minimized structures: The hexameric capsule **1₆**; cylindrical capsule **2₂**; hybrid capsule **1.2**; tennis ball **3₂**; hybrid capsule **2.3**. Peripheral alkyl and aryl groups ($R = R' = C_{11}H_{23}$, $Ar = pC_6H_4-N(Bu)_2$) in the calculated structures have been removed.

match of most of the hydrogen bond donors and acceptors; only 4 imide oxygens are left without hydrogen bond partners. In the experiment, a new assembly emerges immediately and irreversibly upon the addition of **3₂** to **2₂** in $CDCl_3$. The resulting NMR signals indicated complete disappearance of **2₂** (Fig. 2*f*); the homomeric capsules do not coexist in chloroform. The hybrid capsule **2.3** also forms in tetrachloroethane and *p*-xylene (see SI).

We examined a variety of guests expected to be of appropriate size and shape for **2.3**. Ethane is well-accommodated inside **3₂** (Fig. 3*a*), and is coencapsulated with 2,2-paracyclophane in **2₂**

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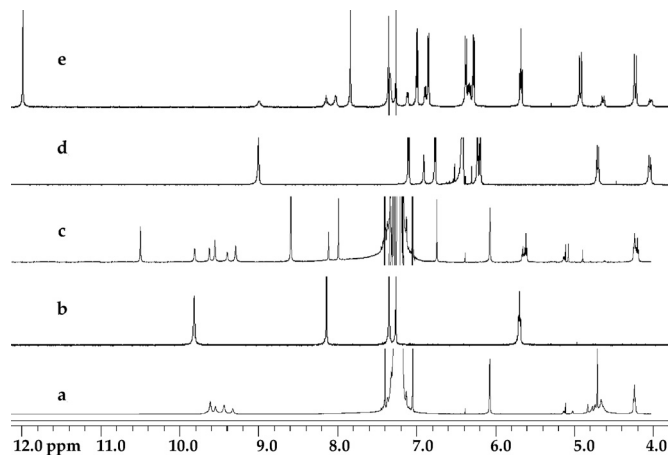


Fig. 2. Partial ^1H NMR spectra of capsular assemblies. (a) Resorcinarene **1** (2 mM) in chloroform; the formation of hexamer **1₆** occurs with 6–8 encapsulated solvent molecules. (b) Two micromolar **2** in chloroform yields the cylindrical capsule **2** with 3 surrounded solvent molecules. (c) Mixing **2** (2 mM) with **1** (2.5 mM) results in the formation of hybrid capsule **1.2**; the original homomeric capsules are also present. (d) Tennis ball **3** (2 mM) in chloroform containing dissolved atmospheric gases. (e) Mixing of **2** (2 mM) and **3** (2 mM) gives the new hybrid capsule **2.3**. Complete NMR peaks assignment of **2.3** are given in the SI.

(Fig. 3*b*). The 2,2-paracyclophane alone is not bound in **2**—a single molecule fills too little of the space whereas 2 molecules fill too much—but the combination of these 2 guests in their encapsulation complexes in the solvent mesitylene-*d*₁₂ gives only the hybrid capsule **2.3** with the 2,2-paracyclophane inside (Fig. 3*c* and *d*). The snug fit of this guest in **2.3** is also shown; if the guest is not allowed to rotate freely in the cavitated, the symmetry planes are lost. The NMR spectrum shows this to be the case in

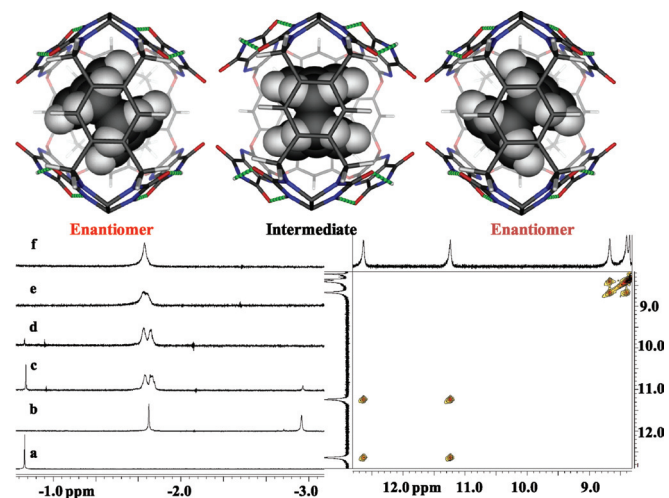


Fig. 3. Rotation of *p*-cyclophane in the hybrid capsule. (Upper) A view into the complex of **2.3** with *p*-cyclophane; rotation of the guest gives an intermediate with planes of symmetry and interconverts the magnetic environments of the cavitated walls. (Lower) (Left) Upfield regions of the ^1H NMR spectra (600 MHz, mesitylene-*d*₁₂) of ethane in **3** (4 mM) (a), coencapsulation of ethane and 2,2-paracyclophane in **2** (4 mM) (b), immediately after mixing a and b (c), c after 15 min (d), d at 310 K (e), d at 320 K (f). The signals at $-1.57 \sim -1.62$ ppm are those of the methylenes of 2,2-paracyclophane located at the tapered end of **2.3**. (Right) 2D ROESY spectrum of encapsulated 2,2-paracyclophane in **2.3**; mixing time 300 ms. Cross-peaks indicate the exchange of environments among the imide and ureido NH signals, as 2,2-paracyclophane slowly rotates inside the capsule at 300 K.

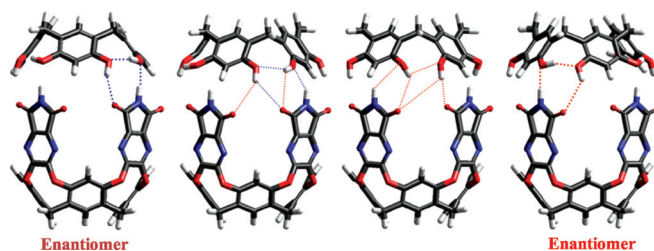


Fig. 4. Racemization of hybrid capsules **1.2**. A slight rotation of the resorcinarene and rearrangement of the hydrogen bond seam causes interconversion of enantiomeric capsules.

the hybrid capsule. Two enantiomeric complexes are present as shown and, with gentle heating, they interconvert ($\Delta G^\ddagger = 15.3$ kcal/mole at coalescence $T = 312$ K); through the rotation of the guest along its long axis. The imide hydrogens show 2 signals in the NMR spectrum with a $\Delta\delta > 1$ ppm; the 2D ROESY spectroscopy showed exchange cross-peaks between them at room temperature and provided $\Delta G^\ddagger = 16.2$ kcal/mol calculated for the guest rotation. Diffusion ordered spectroscopy (DOSY) (20) was performed on the sample of 2,2-paracyclophane in **2.3**. Diffusion coefficients confirmed that all 3 components diffuse as a single species and therefore exist in the same complex (see SI). The packing coefficient of 2,2-paracyclophane in **1.2** is unusually high (70%), and resembles the value for closely packed spheres in the solid state (74%). Along with data from other hybrid capsules and coencapsulation (21) experiments with 2,2-paracyclophane in **2**, the value for solid guests in reversible encapsulation complexes appears to be $\approx 70\%$. The corresponding value for gases in such capsules is $\approx 40\%$ (22).

Bulky guests such as 2,2-paracyclophane are also taken up by the other hybrid **1.2**. Surprisingly, the number and location of the upfield NMR signals revealed rapid rotation of this guest within that capsule at ambient temperature. It appears that hybrid **1.2** features a wider space inside, forced apart by the diverging OH groups of the resorcinarene “lid.” Hybrid capsule **1.2** is also chiral, because the head-to-tail arrangement of these OH groups can be either clockwise or counterclockwise. The 2 enantiomers are shown in Fig. 4, but they interconvert rapidly even at low temperatures. A mechanism that makes new hydrogen bonds as old ones are broken is proposed and features a slight rotation of the lid. The alternate process involving dissociation and reassembly to interconvert the enantiomers would be energetically much more costly.

The dynamics of hybrid assembly are not accessible by NMR studies; they assemble instantly and completely at millimolar concentrations. They can be studied in real time at the micro- to nanomolar concentration ranges using fluorescence resonance energy transfer (FRET) (23). We recently reported the synthesis of labeled versions of resorcinarenes and cavitand monomers (24,25). Perylene labeled resorcinarene **1_A** (acceptor fluorophore) was added to pyrene labeled cavitand **2_D** (donor fluorophore) in mesitylene with both components at 250 nM. No FRET signal was observed, indicating that the hybrid capsule **1.2** had not formed (Fig. 5). This was expected as there was no suitable guest present to template the formation of any capsule. When 2,2-paracyclophane was added as the guest, a large FRET signal developed after a few days (Fig. 5). The large increase in the acceptor fluorescence was accompanied by a commensurate decrease in the donor fluorescence as **1.2** assembles.

The aromatic rings of the hosts provide strong magnetic shielding for encapsulated guest molecules and the guest proton resonances can be shifted upfield as much as 5 ppm (26). We examined guest species that were not well accommodated by either of **2** or **3** but were of appropriate size and shape for the hybrid

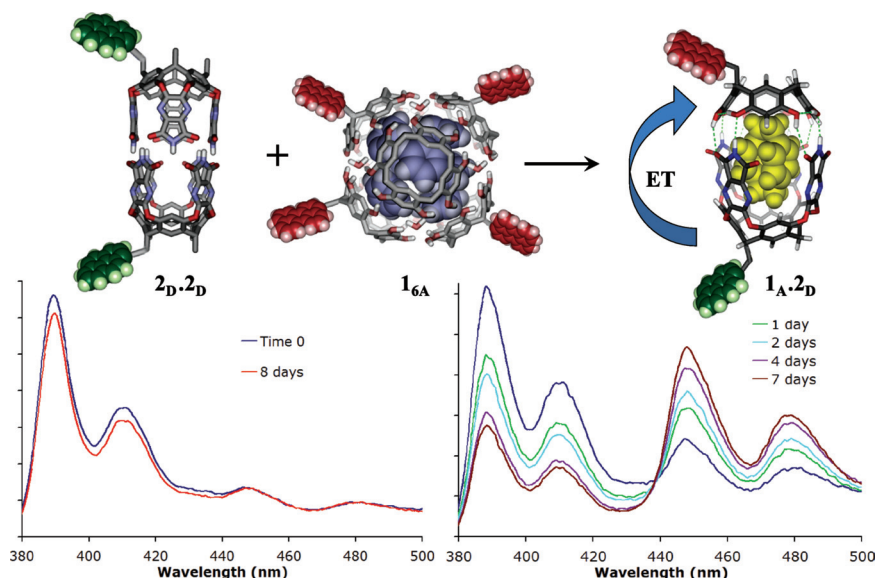


Fig. 5. (Upper) Representation of the donor dye-labeled cylindrical capsule 2_{D2} and the acceptor dye-labeled hexamer 1_{A6} and the hybrid capsule 1_{A-2D} . FRET occurs only upon formation of the hybrid assembly. (Lower Left) Fluorescence spectrum of a mixture of pyrene labeled monomer 2_D and perylene labeled resorcinarene 1_A initially, and after 8 days showing no FRET increase. (Lower Right) Development of FRET with time upon addition of 2,2-paracyclophane as the nucleating guest.

assemblies. All of the guests reported to fit in the hybrid 1.2 fit in the new hybrid assembly as well, but 2.3 accepted a broader range of guests. Two different complexes—carceroisomers (27)—are formed with unsymmetrical guests like *p*-ethyl toluene 4 (Fig. 6). This guest is too long to “tumble” within the hybrid and shows 2 orientations. The favored isomer (67%) positions the tolyl methyl deep in the tapered end for strong C-H/ π interactions (28,29). With *p*-cymene 5 the single methyl in the tapered end of the cavitation was even more favorable (92%), because the isopropyl group does not fit into this space comfortably. Chiral guests such 1,7-dioxaspiro[5.5]undecane 6 or menthol 7 are encapsulated in 2.3 , and they rotate freely inside because no diastereomeric complexes appear in their NMR spectra (see SI).

Midsized alkanes such as 2-methyl heptane 8 or 2,2-dimethyl hexane 9 are also taken up in 2.3 ; in both cases the carceroisomer with ω -methyl group at the tapered end is slightly favored (57%

and 61%, respectively). These alkanes appear too long to tumble freely in the hybrid, but 2D ROESY spectroscopy shows otherwise (Fig. 7). The cross-peaks show the carceroisomers interconvert on the NMR timescale at room temperature. Two different motions are possible: In one of them, 3 completely dissociates from 2 to make more room for the tumbling of the alkane. Evidence to support this mechanism is from 2D spectra that shows exchange of 2.3 with the excess of 3_2 in the solution. The other possible mechanism involves “breathing” along the seam of hydrogen bonds to make enough space for guest rotation. The ΔG^\ddagger s for the tumbling motions of 8 and 9 in 2.3 were calculated as 16.6 kcal/mol and 16.3 kcal/mol, respectively.

The openings and seams of hydrogen bonds at the “upper” part of 2.3 favor bulky, polar guest features, a preference evident in the spectra with $10-15$. The complex with 11 is modeled in Fig. 8, where the hydroxyl groups of the guest have a number of hydrogen-bonding opportunities. This structure resembles the encapsulation complexes of Raymond (30), that feature “dangling arms.”

Discussion

The homomeric capsules have either strong hydrogen bond donors (imide N-H's in 2_2) or strong hydrogen bond acceptors (ureido carbonyls in 3_3), and these complement each other optimally in the hybrid 2.3 . A downfield shift of 2 ppm is observed for the appropriate N-H signal in the NMR spectrum, indicating stronger hydrogen bonding in the hybrid capsule. The entropy changes of the hybridization process is hard to estimate in $CDCl_3$ as the occupancy of 3_2 is unknown. There are 3 solvent molecules in 2_2 and 2 in the hybrid capsule 2.3 . Once formed, the hybrid 2.3 thwarts attempts at self-sorting, unlike hybrid 1.2 . For example, addition of *p,p'*-dimethyl stilbene, one of the best occupants of 2_2 has no effect on the 2,2-paracyclophane complex with 2.3 . In short, the hybrid capsule 1.2 , assembles even in direct competition with its respective homomeric capsules. Heteromeric species that assemble in preference to homomeric ones are known, especially when the modules have close structural relationships (31,32). In contrast, many proteins (33) and the much smaller resorcin[4]arenes and pyrogallol[4]arenes show different behavior. The latter differ marginally yet strict self-sorting of

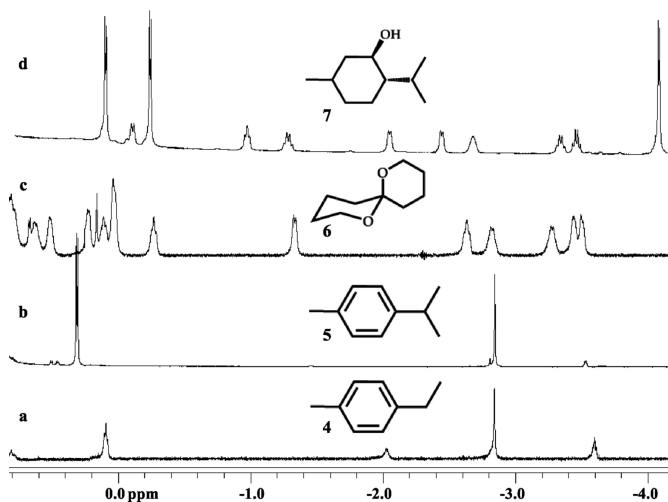


Fig. 6. Upfield regions of 1H NMR spectra show signals for encapsulation of *p*-ethyl toluene (a), *p*-cymene (b), racemic 1,7-dioxaspiro[5.5]undecane (c), (\pm) menthol in the hybrid capsule 3.2 (mesitylene-*d*12, T = 300 K, 600 MHz) (d).

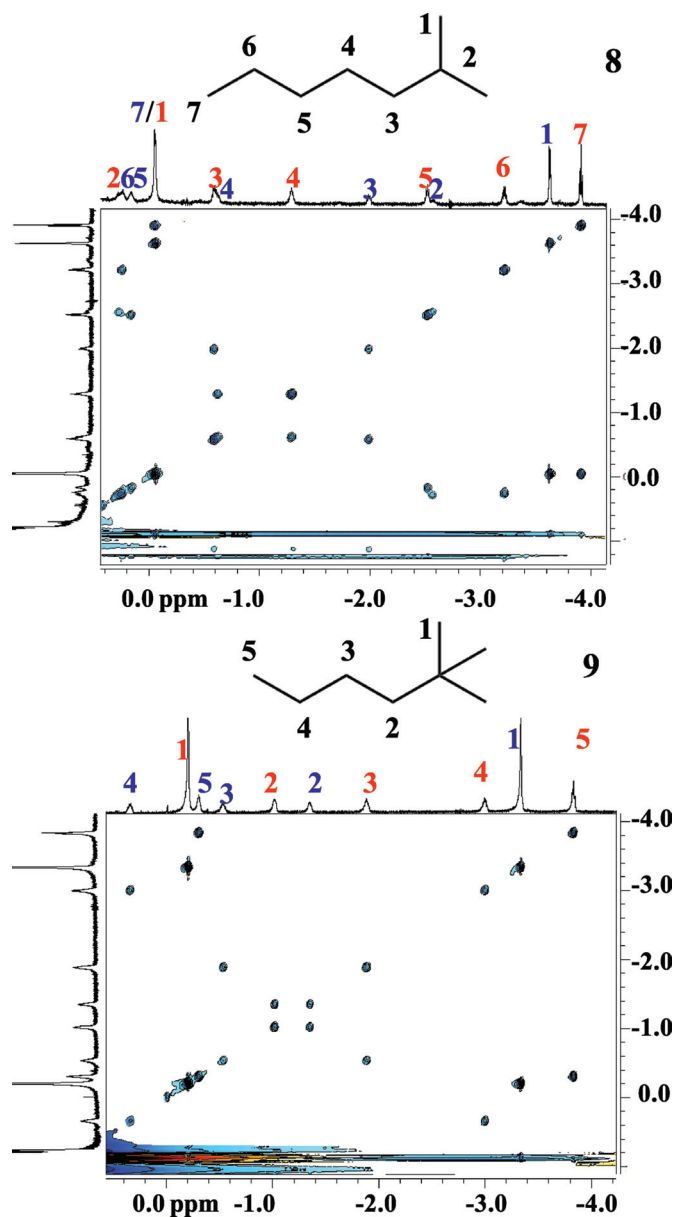


Fig. 7. Upfield regions of 2D ROESY spectra (mesitylene- d_12 , T = 300 K, 600 MHz). Cross-peaks show exchange between 2 carceroisomers (shown in a different color) of **8** in **3.2** (Upper) and **9** (Lower).

their hexameric capsules occurs in solution (34). In the gas phase the whole range of heteromeric species is observed (35). Attempts to form the remaining possible hybrid combination (**1,3**) showed no evidence of hybridization between the components; instead, self-sorting to **1₆** and **3₂** was observed. However, cavitand **2** exhibits a remarkable affinity for glycolurils beyond the tennis ball (36,37). Perhaps the rich array of hydrogen bond

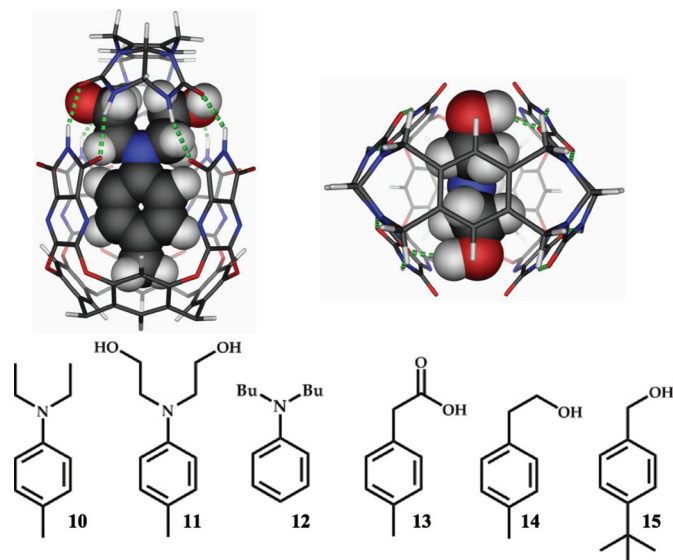


Fig. 8. (Upper) Semiempirical energy-minimized complex of **11** in **2.3** (Peripheral alkyl and aryl groups in the modeled structures have been removed). (Lower) Line drawings of the some suitable guest for hybrid capsule **2.3**.

donors and acceptors of glycolurils (**38**) is responsible for their promiscuity with other modules of the proper scale and curvature. Whatever the reason, the hybrid capsule **2.3** is an exception to self-sorting phenomena and expands the repertoire of new encapsulated species.

Materials and Methods

All reagents were obtained from commercial suppliers and used without further purification. NMR spectra were recorded on a Bruker DRX-600 spectrometer. The NMR samples were prepared as follows. A 2 mM concentration of **1** and/or **2** and/or **3** and 1–20 mM concentrations of the appropriate guest were mixed with 0.6 mL of mesitylene- d_{12} in a NMR tube. The tube was placed in an ultrasonic bath (230 W) and sonicated for 5–10 min. ^1H NMR and 2D spectra were recorded on a Bruker DRX-600 spectrometer with a 5-mm QNP probe. Proton (^1H) chemical shifts are reported in parts per million (δ) with respect to tetramethylsilane (TMS, $\delta = 0$) and referenced internally with respect to the protio solvent impurity. The ROESY spectra were recorded at 300 K at 600 MHz with the phase-sensitive ROESY pulse sequence supplied with the Bruker software. Each of the 512 F1 increments was the accumulation of 40 scans with a 300-ms mixing time.

Fluorescence measurements were obtained using a Fluorolog-3 Model FL3-21 spectrofluorometer. Mesitylene was distilled before use to remove fluorescent impurities. All measurements were conducted at 25.0 ± 0.1 °C. Solutions of each capsule **1₆** (**D**) and **2₂** (**A**) were prepared at 7.7×10^{-6} M and allowed to equilibrate for at least 48 h. Equimolar amounts of each solution were then mixed, and an aliquot of the mixture was diluted into 3 mL of mesitylene so that the concentration of each capsule was 250 nM. The fluorescence spectrum was then recorded with an excitation wavelength of 350 nm.

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