

Synergistic effects of Miconazole and Polymyxin B on microbial pathogens

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Abstract The therapeutic value of antibiotics depends on the susceptibility of the infecting microorganism and the pharmacological profile of the drugs. To assess the value of an antibiotic combination of polymyxin B and miconazole this study examined the *in vitro* synergistic potential of the two drugs on Gram-negative and Gram-positive bacteria and yeast. Antifungal and antibacterial activity was tested by minimum inhibitory concentration (MIC) of broth macrodilution and urea broth microdilution, by fluorescence microscopy and flow cytometry. Synergism was calculated using the fractional inhibitory concentration index (FIC_i). With *Staphylococcus intermedius* as target we found up to an eightfold reduction of the individual MICs when both drugs were combined. However, the FIC_i was 0.63 suggesting no real interaction between the two drugs. With *Escherichia coli*, *Pseudomonas aeruginosa*, and *Malassezia pachydermatis* as targets the antimicrobial drug combination reduced the MICs of polymyxin B and miconazole from fourfold to hundredfold resulting in FIC_i between 0.06 and 0.5 which defines a synergistic action. Thus, if polymyxin B and miconazole are combined their effect is greater than the sum of the effects observed with polymyxin B and miconazole independently, revealing bactericidal and fungicidal synergism. Our results indicate a strong therapeutic value for the combination of these antimicrobial agents against Gram-negative bacteria and yeast and a weaker value against Gram positive bacteria for clinical situations where these pathogens are involved.

Keywords Miconazole · Polymyxin · Antibiotic synergism · Otitis externa

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Introduction

Polymyxin B is an amphipatic cyclic decapeptide and cationic detergent with a broad antibiotic spectrum. It effects against Gram-negative bacteria such as *Enterobacter*, *Escherichia coli*, *Pseudomonas* spp., *Salmonella* spp. and *Shigella* spp and to a lesser extent, against Gram-positive bacteria as *Staphylococcus* spp. and *Streptococcus* spp. The bactericidal effect of polymyxin B relies on a dual mechanism of action. The sublethal action occurs as polymyxin B binds to the outer membrane of bacteria where it complexes avidly with lipopolysaccharides (Newton 1954) and disorganizes the whole outer membrane (Hancock 1984; Koike et al. 1968; Nikaido and Vaara 1985). This allows polymyxin B to enter the cytoplasmic membrane where it causes leakage of cytoplasmic components entailing the cells to die. It is noteworthy that the membrane permeability changes immediately on contact with the drug. The activity of polymyxin B is antagonized by Mg^{2+} and Ca^{2+} (Newton 1953), thus part of the action of this antibiotic is assumed as competitively displacing Mg^{2+} or Ca^{2+} from the negatively charged phosphate groups of membrane lipids. On parenteral administration polymyxin B is extreme nephrotoxic. However, as it completely lacks absorption from intact skin or mucous membrane and thus produces no systemic reaction, it is applied hitherto almost exclusively in topical therapy. Yet the polymyxins gain increasingly importance as reserve antibiotic. As the emergence of multiresistent Gram-negative bacteria pose serious clinical problems the polymyxins are recommended for severe systemic infections caused by bacteria that are resistant to other antibiotics (Ouderkerk et al. 2003; Pasquale and Tan 2005).

Miconazole, another antimicrobial active substance also aims at interacting with the cytoplasmic membrane of microbiological pathogens. Miconazole belongs to the chemical class of imidazoles with a broad antifungal spectrum against most of the pathogenic fungi and *Candida albicans*. *In vitro* it shows an effect against some Gram-positive germs and *acanthamoeba*, too (Janssen and Van Bever 1979; McEvoy 1992). The drug permeates the chitin of the fungal cell wall and increases the membrane permeability to various intracellular substances (Sreedhara Swamy et al. 1974). Miconazole is known to interfere with the synthesis of fungal and bacterial lipid membranes as it restrains the synthesis of ergosterol which results in accumulation of toxic methylated sterol intermediates in membranes and subsequently in fungal cell growth arrests (Vanden Bossche et al. 1990). Another mode of miconazole antifungal action is the induction of reactive oxygen species in susceptible fungi leading to fungal cell death (Francois et al. 2006; Kobayashi et al. 2002). Side effects are rare if miconazole is applied topically (Sawyer et al. 1975) and to date no resistances are known in animals to miconazole (McEvoy 1992).

The application of both polymyxin B as an antibiotic and miconazole as a fungicide and Gram-positive antibacterial agent is common for those infections, where fungal and bacterial pathogens are aetiological. Both antimicrobial agents are proposed for the treatment of systemic mycosis and have been combined so far in topical preparations to provide a complementary antimicrobial spectrum for the prevention of minor wound infections and the treatment of dermatitis or otitis externa in veterinary medicine (Kiss et al. 1997; Rougier et al. 2005; Studdert and Hughes 1991). Organisms frequently isolated from these lesions include *E. coli*, *P. aeruginosa*, *S. intermedius* and the lipophilic yeast *M. pachydermatis*. The latter is a common inhabitant of the outer ear duct in dogs and cats but can also become an opportunistic pathogen, where an infectious state usually parallels a primary established illness.

The combination of antibiotics may reveal positive drug interactions such as an enhanced activity of the compounds or an increased antimicrobial spectrum. Several studies

demonstrate an either synergistic or additive activity between the outer membrane perturbing agent polymyxin B and specific antibiotics (Rosenthal and Storm 1977), (Richards and Xing 1993), (Booth et al. 1994). Many studies have documented that polymyxin B nonapeptide which lacks bactericidal activity but has an outer membrane-permeabilization action preserved induces an increase in sensitivity to different antibiotics and other drugs (for review see (Vaara 1992)). The potentiation of the antifungal activity of tetracycline by polymyxin B through selective alteration of cell membrane permeability was reported, too (Schwartz et al. 1972), however there are contradictory results on this interaction. Also the lysis of *Neisseria catarrhalis* and other Gram-negative bacteria by lysozyme has been demonstrated in cells pretreated with polymyxin B sulfate. Since neither polymyxin nor lysozyme acting independently on the cells resulted in lysis, these results suggested that pre-treatment with the antibiotic increased the permeability of the cells or disorganized the cell wall and hence rendered the mucopolysaccharide substrate accessible to the action of the enzyme (Warren et al. 1957). Furthermore the antibacterial activity of a combination of miconazole and/or bacitracin with polymyxin B against *Staphylococcus aureus* and *E. coli* was higher than the sum of the effects observed with the two drugs independently (Cornelissen and Van den Bossche 1983). A synergy in antifungal activity against *Candida albicans* was reported as well when polymyxin B was combined with amphotericin B, ketoconazole and miconazole (Moneib 1995) and miconazole applied together with benzoylperoxide markedly increased the bactericidal activity against *Staphylococcus spp.* and *Propionibacterium acnes* (Vanden Bossche et al. 1982).

It was the objective of the study to verify the presumed synergistic effect of polymyxin B and miconazole with both bacteria and yeast. We compared the *in vitro* activity of polymyxin B and miconazole as single substances and in combination against *E. coli*, *P. aeruginosa*, *S. intermedius* and *M. pachydermatis*. Bacterial viability was assessed by means of a standard MIC test, fluorescence microscopy and flow cytometry while yeast susceptibility was evaluated using an urea broth microdilution method.

Materials and methods

Antimicrobial agents

Freshly prepared solutions of polymyxin B sulfate (Sigma-Aldrich, Taufkirchen, Germany) in deionized water or miconazole nitrate salt (Sigma-Aldrich) dissolved in 25% (v/v) ethanol and 75% (v/v) polyethylenglycol 400 while heating for 2 h at 60°C were filter sterilized (pore size 0.2 µm) prior use.

Susceptibility testing

For examination of antibiotic susceptibility the tube dilution test was applied. The quality control strains *E. coli* (ATCC 25922), *P. aeruginosa* (ATCC 27853) and *S. intermedius* (ATCC 5739) were used for determination of MICs. For the content of selected tubes from susceptibility testing we determined the minimal bactericidal concentration (MBC) by endpoint colony counting of bacteria on Mueller-Hinton agar. For pre-trial MIC *E. coli* strain XL-1 (Stratagene) was used. *E. coli* and *P. aeruginosa* were cultivated in Mueller-Hinton Broth (Merck/VWR, Darmstadt, Germany; if not mentioned otherwise, all chemicals were obtained from Merck/VWR). *S. intermedius* was propagated either in corynebacteria nutrient (1% casein peptone, tryptic digest, Difco/BD, Heidelberg, Germany,

0.5% yeast extract, Difco/BD, 0.5% glucose and 0.5% NaCl) or in Mueller-Hinton Broth. Bacteria from frozen stocks were subcultured three times on agar plates. Then two single colonies were picked each to inoculate a liquid overnight culture. The bacterial cultures were grown to early log phase at 37°C aerobically by shaking in a reciprocating shaker. Bacterial inoculum was prepared in Mueller-Hinton Broth at 2×10^6 colony forming units (CFU)/ml to give a final concentration of 2×10^6 CFU. To each tube 5.0 ml of bacterial culture, 4.0 ml of growth medium, 0.5 ml of a polymyxin B dilution or 0.5 ml of a dilution of miconazole was added. Each 0.5 ml of dilution of polymyxin B and miconazole was also added in combination to 5.0 ml of bacterial culture and 4.0 ml of growth medium. Samples with single active substance or controls were adjusted to a total volume of 10.0 ml with 0.5 ml sterile deionized water. Solutions of polymyxin B or miconazole consisted of log₁₀ dilution steps between 750 and 0.075 µg/ml and log₂ dilution steps between 750 and 0.046 µg/ml. For all experiments freshly prepared antimicrobial solutions were used.

Bacterial cultures without active substance (C, bacterial inoculum and growth medium) and bacterial cultures with 0.5 ml solvent (L, bacterial inoculum, growth medium and solvent containing 25% ethanol and 75% polyethylenglycol 400, which was also heated to 60°C for 2 h) served as growth controls. Also a sterility control tube (M, growth medium) was included into each MIC test. The cultures were incubated for 18 h at 37°C under aerophilic conditions. After 18 h of incubation the MIC was read as the lowest drug concentration that inhibited bacterial growth.

Each test was performed 5 times. For the first clear tube containing both antimicrobial agents the fractional inhibitory concentration (FIC) was calculated as follows: FIC of drug A (FIC_A) = MIC of drug A in combination/MIC of drug A alone, FIC of drug B (FIC_B) = MIC of drug B in combination/MIC of drug B alone. FIC Index (FIC_i), calculated as the sum of each FIC, was interpreted as follows: FIC_i ≤ 0.5 = synergy, FIC_i > 4.0 = antagonism and FIC_i > 0.5–4 = no interaction (Odds 2003).

Christensen's urea broth microdilution method

The Christensen's urea broth microdilution method (Nakamura et al. 2000) (Rincón et al. 2006) was applied to assay the antifungal activities of polymyxin B and miconazole alone and in combination against *M. pachydermatis*. With this test fungal cell activity is measured. The mechanism is relying upon the release of ammonia during urea hydrolysis to change the pH of the medium, converting the yellow phenol red indicator to pink (Roberts et al. 1978).

Malassezia. pachydermatis reference strain (CBS 1879) was propagated at 35°C on modified Leeming and Notman slants (Leeming and Notman 1987) containing 1% bacteriological peptone (Difco/BD), 1% glucose, 0.2% yeast extract (Difco/BD), 0.8% ox bile (Difco/BD), desiccated, 1% (v/v) glycerol (Sigma-Aldrich, Taufkirchen, Germany), glycerol monostearate (Elbe Fetthandel GmbH, Geesthacht, Germany), 0.5% (v/v) Tween 60 (Sigma-Aldrich), 2% olive oil (Sigma-Aldrich) and 1.5% agar (Sigma-Aldrich). The isolate was subcultured every 2 weeks more than three times before use. The colonies were collected by scraping, suspended with sterile 0.04% Tween 80 in sterile deionized water (pH 7.4) with a glass homogenizer and the suspension was adjusted to an OD at 530 of 0.42 to 0.43 ($\sim 1 \times 10^6$ CFU/ml). Solutions of polymyxin B or miconazole consisted of log₂ dilution steps between 187.5 and 0.01 µg/ml. The urea broth microdilution method was performed in 96-well microtiter plates. Christensen's urea broth contained 0.1% peptone (Difco/BD), 0.0012% phenol red (Sigma-Aldrich), 0.5% sodium chloride, 0.2% monopotassium phosphate, 0.1% glucose, 2% urea, supplemented with 0.1% Tween 80 (Sigma-Aldrich)

and 0.5% Tween 40 (Sigma-Aldrich), pH 5.2. We added to each well 100 μ l Christensen's urea broth, 50 μ l yeast suspension, 25 μ l test drug and 25 μ l sterile deionized water or 50 μ l test drug and no water if both test drugs were used in combination. The final inoculum of *Malassezia pachydermatis* in each well was $\sim 0.25 \times 10^5$ CFU/ml.

The test drugs were arranged in twofold dilution from left to right and in triplicate. The 96-well microtiter plates were sealed with Parafilm® M and incubated at 35°C for 48 h. The results were evaluated visually. The MICs were defined as the lowest drug concentration at which there was complete inhibition of growth and no colour change of the media was observed.

Fluorescence microscopy

To differentiate live bacteria from dead ones, bacteria were stained in suspension with the Live/Dead BacLight Bacterial Viability Kit (Molecular Probes, Mobitek, Germany). The SYTO 9 nucleic acid stain solution (excitation/emission maxima 480/500) is cell-permeable and stains RNA and DNA in both live and dead bacterial cells green. Propidium iodide (PI, excitation/emission maxima 490/635) penetrates only cells with damaged membranes, thus stains the cells red. Logarithmically growing bacteria cultures were incubated with dilutions of polymyxin B, miconazole or with both substances in combination for 18 h at 37°C while agitation. Each 1.5 ml bacterial sample was concentrated by centrifugation at $10,000 \times g$ for 5 minutes at room temperature and washed once with 0.9% NaCl. The supernatant was removed and the pellet resuspended in 150 μ l 0.9% NaCl. SYTO9 dye (3.34 mM in anhydrous dimethylsulfoxide, DMSO) and PI dye (20 mM in anhydrous DMSO) were combined 1:1 and mixed thoroughly. To each 150 μ l of concentrated bacterial suspension 0.8 μ l of the dye mixture was added, mixed and incubated in the dark at room temperature for 15 minutes. Five μ l of the stained bacterial suspension was transferred onto a 24 \times 46 mm coverslip and visualized directly using a confocal laser scanning microscope Olympus FluoView™ FV1000 (Olympus, Hamburg, Germany). Confocal images were obtained with an argon ion laser as excitation source (excitation wavelength of 488 nm; 1.3% laser power), which was reflected by a dichroic mirror (DM 351/488/543) and focused onto the sample through an oil immersion objective (Olympus Planapochromat 60 x, numerical aperture NA=1.35). Detection channels were set to fluorescein isothiocyanate (FITC, maximum emission 515 nm) and Alexa 594 (maximum emission 618 nm). The live (green fluorescent) and dead (red fluorescent) cells are shown separately and as an overlay with FITC and Alexa 594 bandpass filters. For virtual magnification of bacteria a 3-fold digital zoom was applied.

Bacteria were also visualized with a Zeiss Axiovert 10 microscope (Zeiss, Oberkochen, Germany) equipped with filter set 00 (BP530-585, FT600, LP615) and filter set 09 (BP450-490, FT 510, LP515).

Flow cytometry

Data were acquired with a FC 500 flow cytometer (Beckman Coulter) with an argon ion laser at 488 nm and 20 mW output. Data acquisition and analysis were controlled using Cytomics software. The emission light path contained a 488 nm blocking filter, a 525 bandpass filter before the photomultiplier 1 (PMT) and a 620 nm bandpass filter before the PMT3. The density of the bacterial suspensions was 1×10^6 CFU/ml. Bacterial suspensions were incubated with dilutions of polymyxin B, miconazole or both substances in combination for 18 h at 37°C at aerobic conditions. The cells were harvested by

centrifugation at $10.000\times g$ for 15 min, washed once in 0.9% NaCl and diluted to an OD at 670 nm of 0.03 with 0.9% NaCl. Then 6 μl of 1:1 SYTO9 and PI dye mixture was added to 2 ml of diluted bacterial suspension, mixed well and incubated in the dark at room temperature for 15 min. Populations of bacteria were discriminated as two regions of the log-integrated red-fluorescence (FL3) versus log-integrated green fluorescence (FL1) plot. The percentage of bacteria found within the region identified as those with the "live" cell population was used to estimate the percentage of viable organisms. Using the known viability (proportion of "live" to "killed" bacteria) and measured viability (measured % live bacteria = [# of bacteria in the "live" region/# of bacteria in the "live"+"dead" region] \times 100) and a least-squares fit extrapolated to the 100% "live" point the percent component of dead bacteria in the "live" cell population was estimated.

Results

Antibiotic susceptibility

MICs of two antimicrobials given as single drug each or in combination were determined. The results of the susceptibility tests are summarized in Table 1. *E. coli* strain XL-1 was susceptible to 7.5 $\mu\text{g/ml}$ polymyxin B, but was susceptible to a concentration of polymyxin B as low as 0.07 $\mu\text{g/ml}$ if combined with 0.07 $\mu\text{g/ml}$ miconazole (FIC_i=0.01). No effect on bacterial growth was observed if miconazole was applied alone. The *E. coli* quality control strain was susceptible to 2.93 $\mu\text{g/ml}$ polymyxin B if given as a single antibiotic and to a concentration of 0.18 $\mu\text{g/ml}$ if applied together with the identical concentration of miconazole (FIC_i=0.06). As well miconazole alone could not inhibit *E. coli* growth. *P. aeruginosa* was susceptible to 0.73 $\mu\text{g/ml}$ polymyxin B and showed no effect to miconazole. If polymyxin B acted together with miconazole each drug was inhibitory at a concentration of 0.18 $\mu\text{g/ml}$ (FIC_i=0.25). Thus, when polymyxin B and miconazole were applied in combination they reproducibly showed a clear synergistic antibacterial effect against the Gram-negative strains tested. The Gram-positive *S. intermedius* was susceptible to 0.73 $\mu\text{g/ml}$ miconazole and to 2.93 $\mu\text{g/ml}$ polymyxin B. If identical concentrations of polymyxin B and miconazole were combined a twofold lower dose of miconazole and an

Table 1 *In vitro* susceptibilities of bacterial and yeast strains

Germ strain	Miconazole MIC* Range (Mode)	Polymyxin B MIC Range (Mode)	Miconazole+ Polymyxin B MIC Range (Mode) FIC _i
<i>E. coli</i> XL-1 ^a	no inhibition	7.50	< 0.07 0.01
<i>E. coli</i> (ATCC 25922) ^b	no inhibition	0.73 – 2.93 (2.93)	0.18 – 1.46 (0.18) 0.06
<i>P. aeruginosa</i> (ATCC 27853) ^b	no inhibition	0.73	0.18 – 0.37 (0.18) 0.25
<i>S. intermedius</i> (ATCC 5739) ^b	0.37 – 7.5 (0.73)	2.93 – 7.50 (2.93)	0.37 – 0.75 (0.37) 0.63
<i>M. pachydermatis</i> (CBS1879) ^c	2.92 – 5.85 (2.92)	2.92 – 23.43 (2.92)	0.37 – 1.46 (0.73) 0.50

*MIC, minimal inhibitory concentration, given in $\mu\text{g/ml}$; FIC_i, fractional inhibitory concentration index, given as described in Material and methods

^a Results of 2 repeated experiments on different days

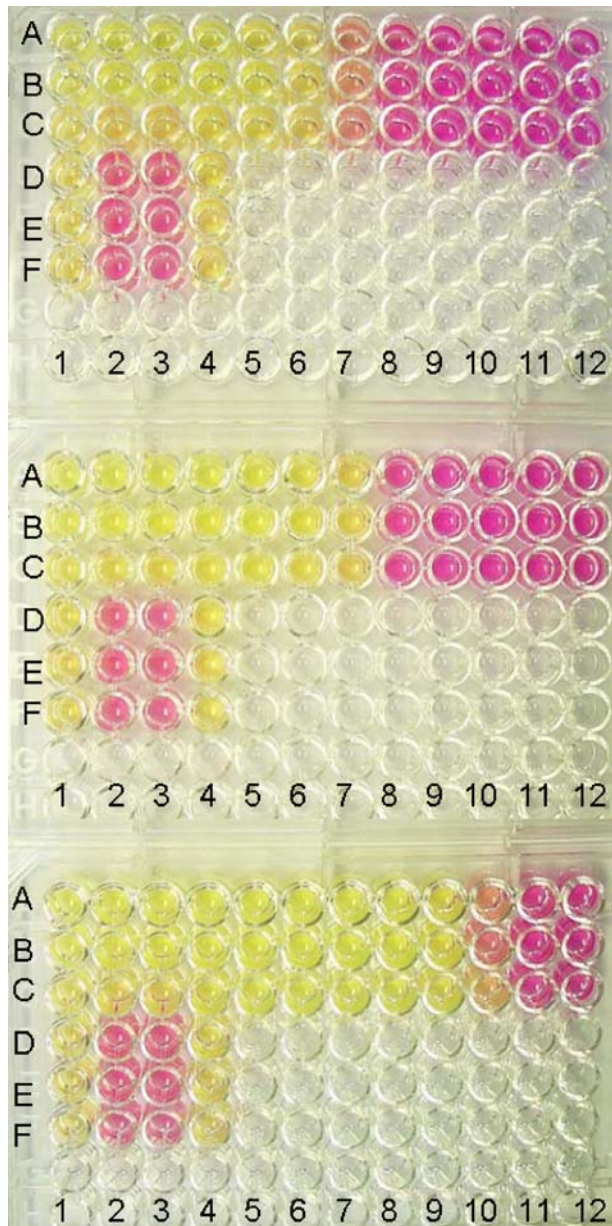
^b Results of 5 repeated experiments on different days

^c Results of 5 repeated experiments on different days with triplicates of each sample

eightfold lower dose of polymyxin B was effective for growth inhibition ($MIC=0.37 \mu\text{g/ml}$, $FIC_i=0.63$). According to a recent revision of the definition of synergy a $FIC_i > 0.5-4.0$ is interpreted as ‘no interaction’ (Odds 2003). Therefore the resulting FIC_i of 0.63 indicates there is no interaction between both drugs if active against *S. intermedicus*.

The mean antifungal activities of miconazole and polymyxin B against *M. pachydermatis* were measured using an urease method, as shown in Fig. 1. If we compared the wells

Fig. 1 Antifungal MIC determination using Christensen’s urea broth microdilution method. Urea hydrolysis by *M. pachydermatis* in microtiter test plates. A positive test is read as a colour change from yellow to pink. The test drugs were arranged in twofold dilution from left to right (row 1–12, 188 $\mu\text{g/ml}$ to 0.09 $\mu\text{g/ml}$) and in triplicate (rows A, B, C). D1, D4, E1, E4, F 1 and F4 represent negative controls (growth medium with distilled water). D2, E2 and F2 are positive controls (all compounds without test drug). D3, E3, F3 contain the solvent of miconazole (polyethylenglycol 400 with ethanol), growth medium and yeast cells as a growth control. From top to bottom: Plate with dilutions of polymyxin B. Plate with dilutions of miconazole. Plate with dilutions of miconazole plus polymyxin B



containing the growth media without *M. pachydermatis* to drug-containing wells with yeast cells we found a MIC of 2.92 µg/ml for miconazole and for polymyxin B as well. If both antimicrobial agents were given in combination susceptibility of *M. pachydermatis* changed to a MIC of 0.73 µg/ml (FIC_i=0,50), which indicates a synergistic response.

Confocal laser scanning microscopy and fluorescence microscopy

Qualitative differentiation of live and dead bacteria was done by confocal laser scanning microscopy (CLSM). CLSM is non-destructive and makes it possible to analyze live biological samples. Living and dead microorganism could be detected *in situ* using an appropriate staining method. Cultures of *S. intermedius* were incubated with different concentrations of polymyxin B and miconazole and stained subsequently with SYTO9 and PI dye. The analysis of bacteria by CLSM allowed the generation of images where the fluorescence of only one dye was visualized. Using narrow bandpass filters the fluorescence of one or the other dye was eliminated respectively. Imaging from the green emission only detects the living green cells and imaging from the red emission detects the dead red cells solely. The left and the right pictures of Fig. 2 represent identical shots of bacterial *S. intermedius* suspensions. The untreated bacteria (2-1 and 2-2) with only a few red (dead) cells are shown in (2-2). After 18 h of incubation with 75 µg/ml miconazole (2-3 and 2-4) more than 50% of the cells became red and are assumed to be dead or in the process to die (2-4). The application of an identical concentration of polymyxin B (2-5 and 2-6) resulted in about 50% red cells (2-6). A tenfold lower concentration of both antimicrobials applied in combination (2-7 and 2-8) revealed strikingly more red cells (2-8) than miconazole or polymyxin B generated when applied alone (2-4 and 2-6, respectively).

To exhibit both dyes within one picture an overlay of two images, each detected with a narrow fluorescence wavelength range was made. Thus the living and dead cells of Fig. 2 are visualized within one image (Fig. 3). Consequently, using CLSM and the Live/Dead BacLight Bacterial Viability Kit the live and the dead organisms can be differentiated and determined even after they had been mixed. Furthermore it was possible to evaluate the antibiotic effect visually.

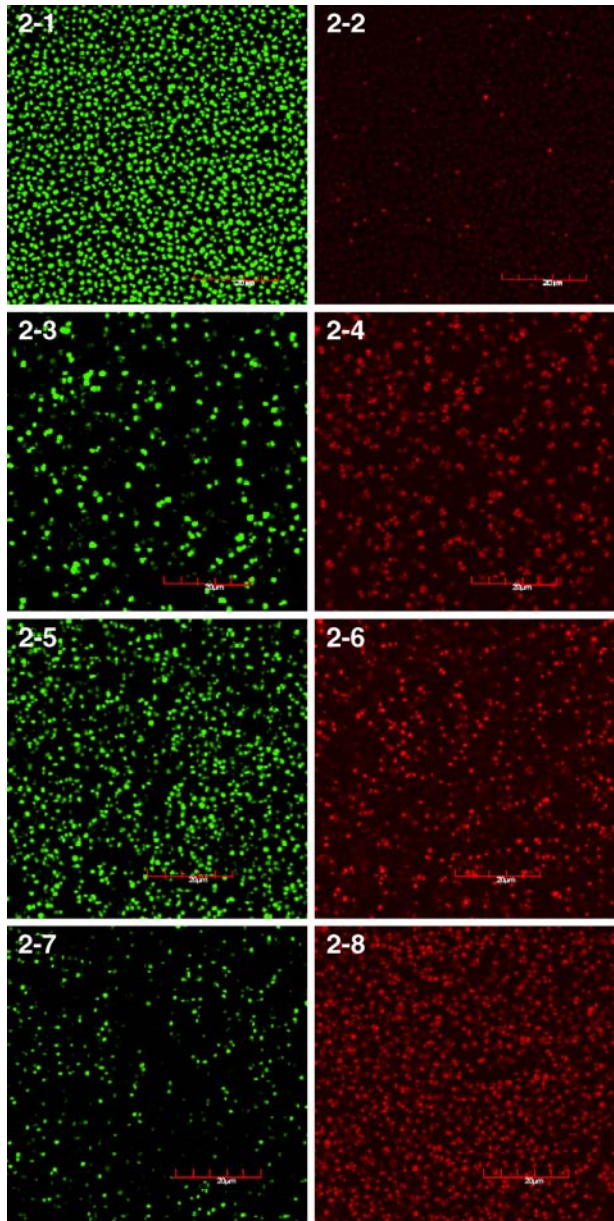
Live and dead bacteria were also visualized with an inverted fluorescence microscope after staining with SYTO9 and PI dye. With this method green and red cells are discriminated using alternately a filter set for FITC and other green fluorochromes or a filter set for red fluorochromes. Owing to the fact that these bandpass filters are not as narrow as in CLSM the pictures representing the green fluorescence also show red cells in the background. If red cells overlay green cells these cells are visualized yellow. Figure 4 demonstrates a qualitative analysis of antimicrobial action against *E. coli* and *P. aeruginosa* after 18 h of antibiotic action. Cells without treatment show very few red bacteria (a, e); following application of 0.73 µg/ml miconazole alone about 10% cells are scored as dead (red) (b, f). An identical concentration of solitary polymyxin B gave rise to about 30% dead (red) cells (c, g) whereas a fourfold lower concentration of each drug, if applied in combination, produced a comparable biological effect (d, h). As a result using this method the antibiotic effectiveness can be quickly evaluated qualitatively as a ratio of live and dead cells within a given population. Visual examination of bacterial cells has approved an enhanced bacterial killing if miconazole and polymyxin B are applied together.

Flow cytometry

Exemplified on two bacterial strains we analysed *E. coli* and *P. aeruginosa* by flow cytometry after antibiotic exposure and staining with the Live/Dead BacLight Bacterial

Fig. 2 Confocal laser scanning microscopy. Samples of *S. intermedius* after treatment for 18 h with polymyxin B, miconazole or with both polymyxin B and miconazole in combination. The cells were stained with SYTO 9 and PI. The left (green) pictures are generated using a FITC bandpass filter and visualize only the living cells, whereas the right (red) pictures are generated from identical presentations using an Alexa 594 bandpass filter to detect the dead cells exclusively.

From top to bottom: Untreated bacteria (2-1 and 2-2). Bacteria treated with 75 $\mu\text{g}/\text{ml}$ miconazole (2-3 and 2-4). Bacteria treated with 75 $\mu\text{g}/\text{ml}$ polymyxin B (2-5 and 2-6). Bacteria treated with 7.5 $\mu\text{g}/\text{ml}$ miconazole plus 7.5 $\mu\text{g}/\text{ml}$ polymyxin B (2-7 and 2-8)

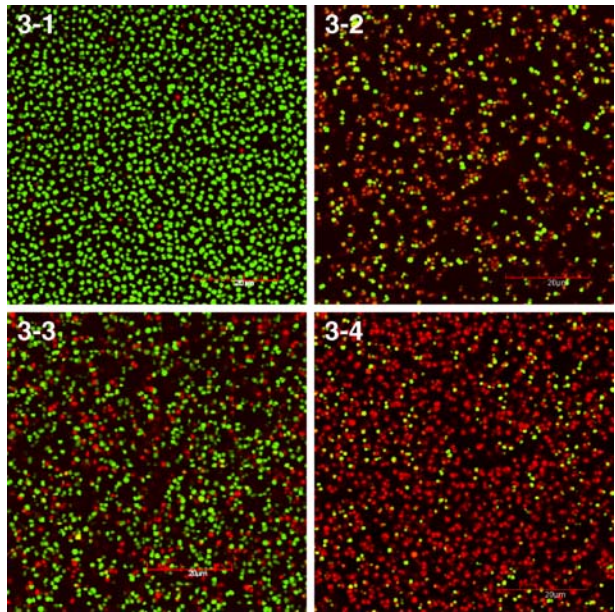


Viability Kit. The percentage of live cells in the population could be calculated by the standard curve shown in Fig. 5. For *E. coli* the rate of live cells was 87% and for *P. aeruginosa* we identified 98% live cells within the population investigated.

When *E. coli* was treated with 2.93 $\mu\text{g}/\text{ml}$ of miconazole, a dead rate of 10.9% was measured which is in the dimension of the normal proportion of dead cells within a population. On the other hand 2.93 $\mu\text{g}/\text{ml}$ of polymyxin B resulted in killing of 45.8% *E. coli* bacteria. Both drugs in combination each at a concentration of 1.46 $\mu\text{g}/\text{ml}$ killed 72.0%

Fig. 3 Overlay pictures of live (green fluorescent) and dead (red fluorescent) *S. intermedius* bacteria are shown using a confocal laser scanning microscope.

Untreated bacteria (3-1). Bacteria treated with 75 $\mu\text{g/ml}$ miconazole (3-2). Bacteria treated with 75 $\mu\text{g/ml}$ polymyxin B (3-3). Bacteria treated with 7.5 $\mu\text{g/ml}$ miconazole plus 7.5 $\mu\text{g/ml}$ polymyxin B (3-4)



of all cells within 18 h (Fig. 6). Following polymyxin B and miconazole application for 18 h flow cytometric analysis of the vitality of *P. aeruginosa* revealed that less cells were affected. After application of 5.86 $\mu\text{g/ml}$ miconazole 8.1% of the bacterial population stained red and therefore was counted dead while 5.86 $\mu\text{g/ml}$ of polymyxin B resulted in only 5.7% dead cells. The combination of 2.93 $\mu\text{g/ml}$ miconazole with 2.93 $\mu\text{g/ml}$ polymyxin B stained 27% of the cells red (results are not shown) which is decidedly a lower rate of dead cells than that found with *E. coli*. Our results show that the survival rate for the determination of bactericidal efficacy of antibacterial agents can be measured by FACS analysis. Against *E. coli* and *P. aeruginosa* we could demonstrate an enhanced effect when both the drugs miconazole and polymyxin B act in combination.

Discussion

This study was conducted to evaluate the net results of the *in vitro* interactions against bacteria and yeast between polymyxin B and miconazole using different methods. While for decades miconazole has been successfully applied as a very potent antifungal drug polymyxin B has not been used over the past years but in veterinary medicine because less toxic antimicrobials are available. Yet bacteria that are resistant to aminoglycosides, beta-lactams and fluoroquinolones are becoming more common and these germs are often susceptible to the polymyxins.

The tube dilution test for the determination of MICs against the two strains of *E. coli*, against *P. aeruginosa* and against *S. intermedius* demonstrated that all the bacteria were susceptible to polymyxin B. Data of our MIC tests are consistent with previous reports where the MIC against *E. coli* was in the range of 0.4 to 37 $\mu\text{g/ml}$ (Kucers 1997). Against *P. aeruginosa* polymyxin B exhibited excellent potency. We found *P. aeruginosa* to be more sensitive to polymyxin B compared to reported MICs which range from 1.2 to

33.3 $\mu\text{g/ml}$ (Kucers 1997). Little information is available about the susceptibility of *S. intermedius* to different antimicrobial agents. Though many authors quote the antimicrobial activity of polymyxin B to be restricted to Gram-negative bacteria (e.g. Chambers 2001), several studies showed the effectiveness of polymyxin B against *S. intermedius*. On the other hand a low susceptibility of *S. intermedius* strains isolated from dogs to polymyxin B was also reported (Junco and Barrasa 2002). In our study polymyxin was quite effective against *S. intermedius*, our tests revealed a MIC of 5.21 $\mu\text{g/ml}$ which is in a magnitude comparable to the MICs of other bacterial strains investigated. It is noteworthy that

Fig. 4 Fluorescence microscopy of *E. coli* (a, b, c, d) and *P. aeruginosa* (e, f, g, h), treated with either polymyxin B, miconazole or with both substances in combination for 18 h. Cells were then stained with SYTO9 and PI. Identical pictures of bacterial suspensions are generated with a green (left pictures) or red (right pictures) filter set. The red stained cells represent bacteria with damaged membranes and the green cells denote those with intact membranes. Untreated bacterial cells (a, e), bacteria treated with 0.73 $\mu\text{g/ml}$ miconazol (b, f), with 0.73 $\mu\text{g/ml}$ polymyxin B (c, g) and with 0.18 $\mu\text{g/ml}$ miconazole plus 0.18 $\mu\text{g/ml}$ polymyxin B (d, h)

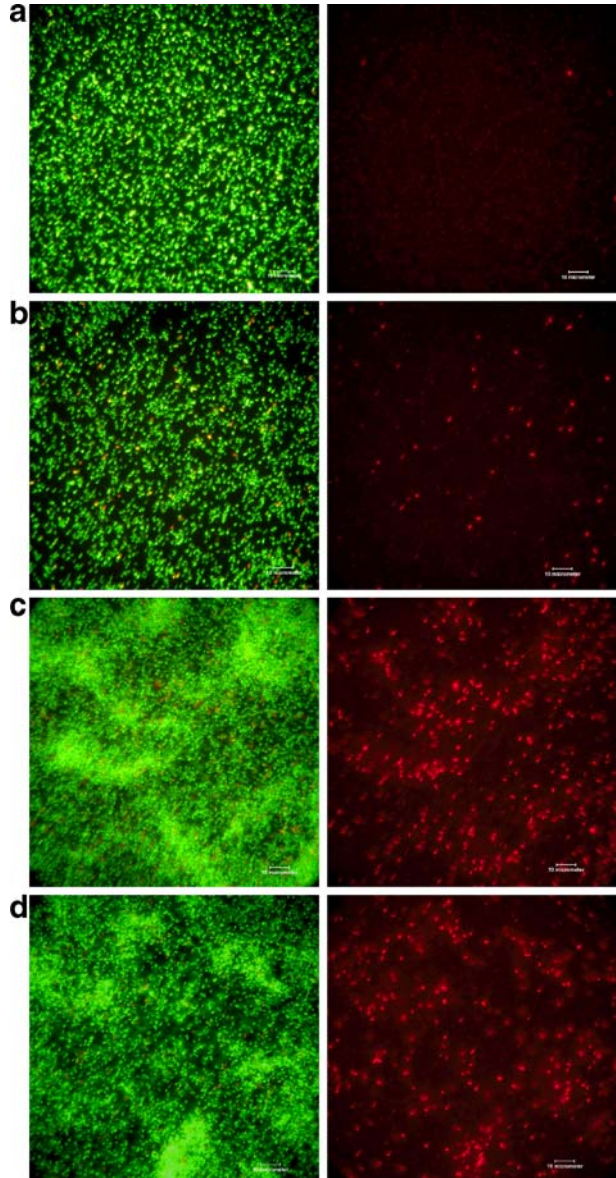
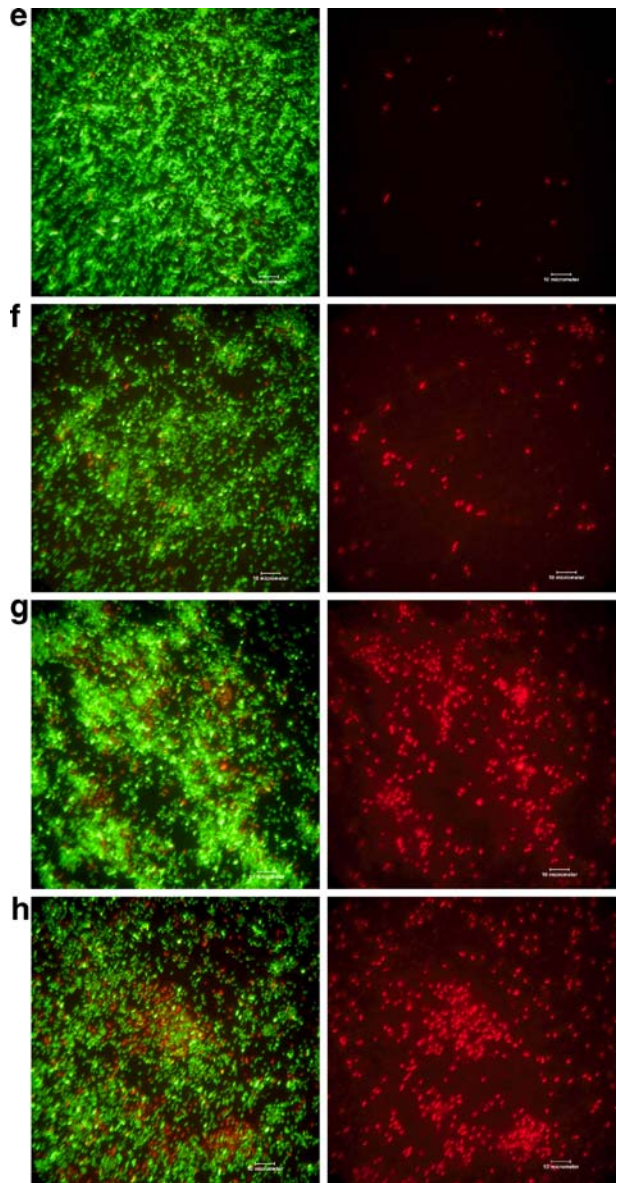


Fig. 4 (continued)



polymyxin B also showed activity against *M. pachydermatis* although the inhibitory concentration of the drug was higher than those against the bacteria tested.

As expected from the known antimicrobial spectrum of miconazole the fungicide was not inhibitory against the Gram-negative bacteria tested. Yet *S. intermedius* and the yeast *M. pachydermatis* were both susceptible at a MIC of 0.73 and 2.92 $\mu\text{g/ml}$ respectively. These values are in agreement with corresponding studies who noticed concentrations of 0.1 to 10 $\mu\text{g/ml}$ to be inhibitory against Gram-positive bacteria and many fungi including *M. furfur* (McEvoy 1992). With most of the sensitive fungi a MIC of 0.5 to 2.0 $\mu\text{g/ml}$ was reported. However it should be kept in mind that the exactness of *in vitro* miconazole

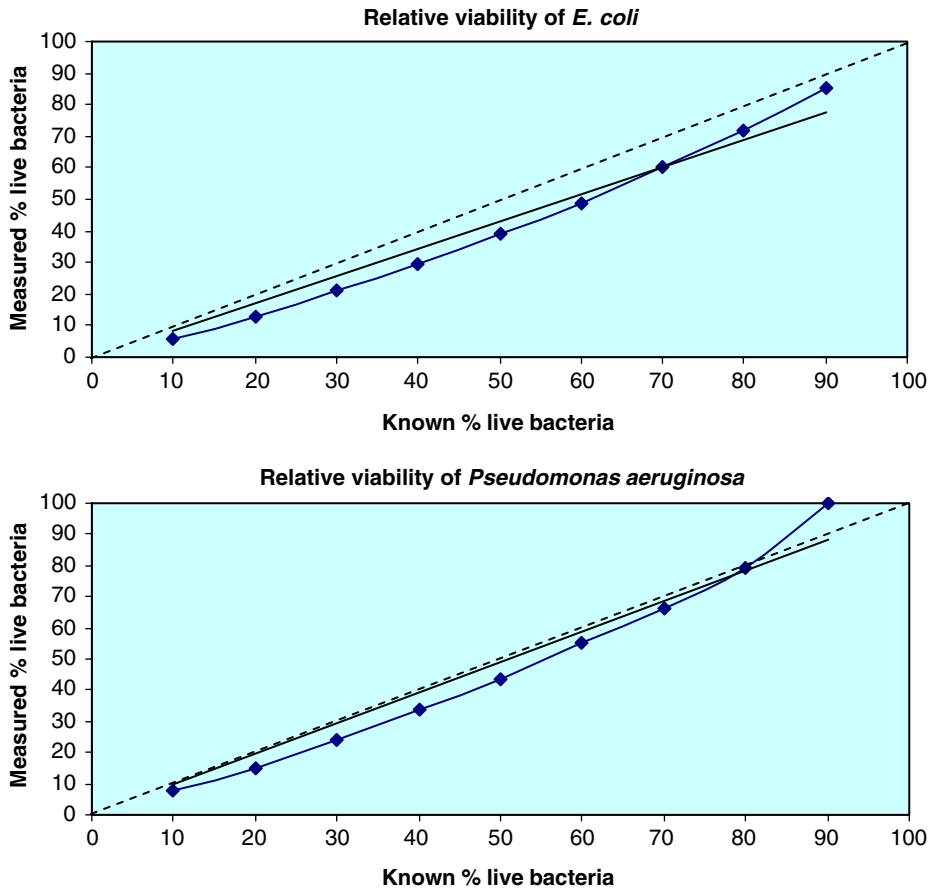
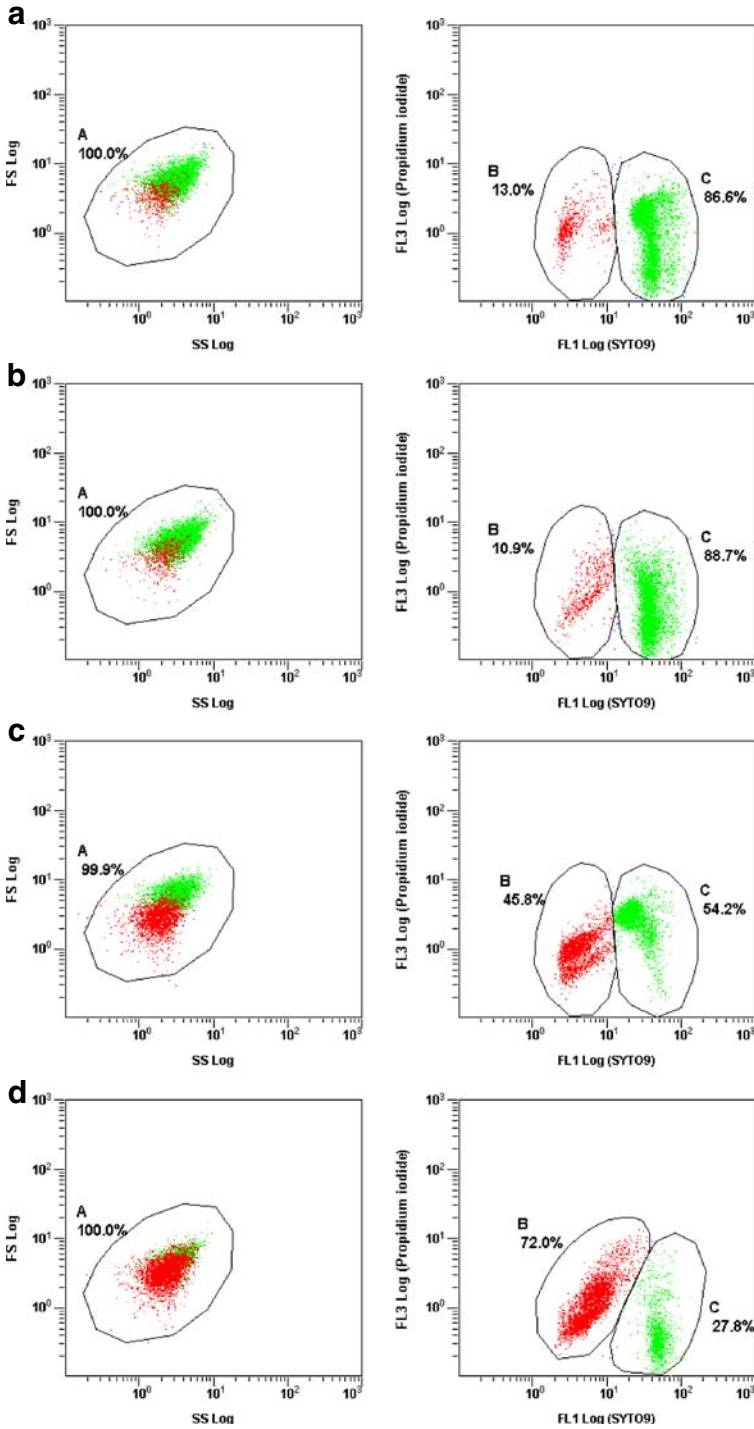


Fig. 5 Analysis of bacterial cells by flow cytometry. The known viability (proportion of ‘live’ to ‘killed’ bacteria) is plotted against the measured viability to identify the percent component of dead bacteria in the ‘live’ cell population

susceptibility testing depends on the measuring method and MIC values do not reflect the activity *in vivo*.

The MIC values derived from our tube dilution tests showed some variation. This may be due to the fact that cations such as Mg^{2+} , Al^{3+} , and Ca^{2+} can adversely affect the activity of antimicrobials. Several investigators have related increased resistance of *P. aeruginosa* to aminoglycosides with increased concentrations of Mg^{2+} and Ca^{2+} in the media and conversely the antibacterial activity of aminoglycoside antibiotics was shown to be enhanced in media with low cationic content (D’Amato et al. 1975). Variations of these ions within lot-to-lot variation in Mueller-Hinton broth may be responsible for varying susceptibilities (Pollock et al. 1978). As well the activity of miconazole is affected by the medium composition as the effect of miconazole was notably reduced even by very low concentrations of Mg^{2+} and Ca^{2+} cations (Sreedhara Swamy et al. 1974).

In CLSM analysis of drug action against *S. intermedius* we found a much higher dose of miconazole and polymyxin B to be necessary for a visual bactericidal effect. Presumably due to the thick cell wall of Gram-positive bacteria this strain reacted much slower to



◀ **Fig. 6** Live/dead discrimination of *E. coli* after incubation with either polymyxin B, miconazole or both substances in combination for 18 h. Cells were analysed by flow cytometry using a double colour staining consisting of SYTO9 and PI. Dot plots on the left show the whole cell populations, where green dots represent live cells and red dots denote dead cells. Dot plots on the right illustrate the bacterial populations separated into two regions B and C with B representing the dead cells (red, FL3), and C corresponding to the living cell population (green, FL1). Cells in B and C are displayed as % of the total population *A. E. coli* untreated (a), *E. coli* treated with 2,93 µg/ml miconazole (b), with 2.93 µg/ml polymyxin B (c), with 1.46 µg/ml miconazole plus 1.46 µg/ml polymyxin B (d)

miconazole and polymyxin B than Gram-negative cells and thus much higher drug doses were necessary to demonstrate a killing effect within 18 h. On the other hand at the concentration, where a dilution of miconazole or polymyxin B caused bacteriostasis we detected only small differences in drug susceptibility between the Gram-positive and the Gram-negative bacteria.

The quantitative measurement of vital and dead cells after antibiotic action by means of flow cytometry was evaluated. When bacteria are treated with drugs that are assumed to change cell permeability this may also affect intracellular granularity and cell morphology and may encompass cell clumping as well. Measurement of antibiotic effects by flow cytometry detects such cellular changes through forward and side scatter. The loss of light scatter may delineate a change in cell size and could also represent a loss of cell wall constituents such as protein or RNA. This is likely to occur with the antibiotics used here. We found that regions are shifted after cells are exposed to polymyxin B and miconazole. Thus, for discrimination between live and dead cells we had to rearrange the regions after antibiotic exposure.

It was also noticed that miconazole at concentrations >11 µg/ml a high percentage of *E. coli* cells stained red with PI (97.4%) and 18.1% of *P. aeruginosa* cells became red after an identical dose of miconazole. Data from the tube dilution tests however demonstrated that this concentration of miconazole could neither inhibit growth of *E. coli* nor of *P. aeruginosa*. MBC assaying of those miconazole-exposed bacteria approved that the PI-stained cells were all able to recover and to reproduce. Obviously under the condition of solitary miconazole application bacterial membranes are compromised allowing PI to penetrate into the cells and thus bacteria are falsely scored as dead in this assay. However, the lethal action is solely attained if miconazole is applied together with polymyxin B. In accordance to the data above miconazole is known to change the membrane permeability at fungistatic concentrations (Sawyer et al. 1975, McEvoy 1992) while at fungicide concentrations the cell wall remains intact but intracellular compounds are destroyed (McEvoy 1992). As a result, the analysis of antimicrobial activity by flow cytometry or fluorescence microscopy using PI staining is possible although there are some restrictions concerning specific drugs such as miconazole, which may produce false results at certain concentrations. Consequently each drug to be analysed using this method should be pretested conducting precedent MBC testing on representative samples.

The combination of miconazole and polymyxin B produced synergistic inhibition of several germs at various *in vitro* conditions in our hands. On the basis of our data the antimicrobial activity of miconazole and polymyxin B was enhanced in all strains tested when both drugs are combined. Synergy refers to the phenomenon where the combination of drugs generates an effect that is greater than the sum of the effects produced by each of the components alone (Sabath 1968). This synergistic activity is related predominantly to the ability of polymyxin B to increase the penetration of hydrophobic antibiotics such as miconazole thus enabling miconazole to gain access to the intracellular space were it

displays its lethal action. The results we obtained with the combination of miconazole and polymyxin B in all strains tested consistently showed a synergism of both drugs. Our results are in agreement with previous *in vitro* studies where a synergism of miconazole and polymyxin B was found for *S. aureus*, *E. coli* (Cornelissen and Van den Bossche 1983) and *C. albicans*. Provided the *in vitro* antibacterial susceptibility predicts therapeutic response these data verify the potential for the use of combinations of the two antimicrobial agents in the topical treatment of infections caused by the prominent pathogenic microorganisms causative for otitis externa in dogs. The clear synergistic tendencies displayed by the two-drug combination allows for the reduction of the antibiotic concentrations, which minimizes toxicity and the probability of formation of resistances to these antibiotics. This may be of special interest in the case of *S. intermedius* which, ensuing antimicrobial therapy, is increasingly reported to be resistant to many antibiotics and failures in treatments cause problems in small animal practice. As a consequence the combination of both antimicrobials substantiates their clinical role in current formulations and offers a very effective therapeutic treatment.

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