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## The Alzheimer's Disease Mitochondrial Cascade Hypothesis: An Update

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### Abstract

In 2004 we proposed the mitochondrial cascade hypothesis of sporadic Alzheimer's disease (AD). Our hypothesis assumed sporadic and autosomal dominant AD are not etiologically homogeneous, considered evidence that AD pathology is not brain-limited, and incorporated aging theory. The mitochondrial cascade hypothesis asserted: (1) inheritance determines mitochondrial baseline function and durability; (2) mitochondrial durability influences how mitochondria change with age; and (3) when mitochondrial change reaches a threshold, AD histopathology and symptoms ensue. We now review the reasoning used to formulate the hypothesis, discuss pertinent interim data, and update its tenants. Readers are invited to consider the conceptual strengths and weaknesses of this hypothesis.

### Introduction

The amyloid cascade hypothesis has dominated the Alzheimer's disease (AD) research field since 1992 (Hardy and Higgins, 1992). It followed the discovery that amyloid precursor protein (APP) gene mutation could produce dementia with neuritic plaques and neurofibrillary tangles (Goate et al., 1991). The amyloid cascade hypothesis was subsequently bolstered by work showing presenilin (PS) 1 and 2 mutations also cause AD, and that APP, PS1, and PS2 mutation all favor processing of APP to its 42 amino acid beta amyloid (A $\beta$ ) derivative (Levy-Lahad et al., 1995; Sherrington et al., 1995; Scheuner et al., 1996). The hypothesis has evolved over time, with the suspect A $\beta$ 42 bogey shifting from fibrils to protofibrils to oligomers to perhaps dimers (Haass and Steiner, 2001; Hardy and Selkoe, 2002; Lesne et al., 2006; Shankar et al., 2008). The core assumption, though, that A $\beta$ 42 induces tangles, neurodegeneration, and dementia has not changed. The amyloid cascade hypothesis argues forced A $\beta$  production in transgenic animals should faithfully model the disease and A $\beta$  reduction should arrest it. Much of the AD research field has invested heavily in AD transgenic modeling and A $\beta$ -directed therapeutic development and consequently has a vested interest in the amyloid cascade hypothesis.

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While the simplicity of the amyloid cascade hypothesis is in many ways appealing, it does oversimplify some controversial issues. First and foremost, it presumes brain aging and AD are uniformly divergent events. We believe, however, that sporadic AD and brain aging are convergent events (Swerdlow, 2007a; 2007b). To emphasize this we previously proposed the mitochondrial cascade hypothesis, which considers sporadic AD from the perspective of mitochondrial aging theory, our own cytoplasmic hybrid (cybrid) studies of AD mitochondria, basic cell biology principles, and the amyloid cascade hypothesis itself (Swerdlow and Khan, 2004). Since we first proposed this hypothesis in 2004 pertinent data have been published. We now review the conceptual basis of our hypothesis and discuss it from the context of recent AD research.

## What is the Mitochondrial Cascade Hypothesis of Sporadic AD?

The mitochondrial cascade hypothesis asserts inheritance determines mitochondrial baseline function and durability, mitochondrial durability influences how mitochondria change with age, and AD histopathology and symptoms ensue when mitochondrial change reaches a threshold (Swerdlow and Khan, 2004). It presumes autosomal dominant and sporadic AD are not etiologically homogeneous. It is intended to apply only to sporadic AD, but makes predictions about autosomal dominant AD. Mitochondrial dysfunction is seen as a nexus between the sporadic and autosomal dominant forms. In the autosomal dominant forms excessive A $\beta$  impairs mitochondrial function, and it is predicted A $\beta$ -induced mitochondrial dysfunction initiates other AD-characteristic histopathologies. In the sporadic form, age-related mitochondrial changes drive mitochondrial function beyond a functional threshold that induces AD-characteristic histopathologies, including processing of APP to A $\beta$  (Figure 1). Before discussing the specifics of the hypothesis it is necessary to briefly discuss mitochondrial function in AD.

## Mitochondrial Function in AD

Several mitochondrial enzyme activities are altered in AD. Pyruvate dehydrogenase complex, ketoglutarate dehydrogenase complex, and cytochrome oxidase (CO) Vmax activities are all reduced (Swerdlow and Kish, 2002). The CO defect, in particular, is central to the mitochondrial cascade hypothesis. CO constitutes the terminal end of the mitochondrial electron transport chain (ETC). It accepts electrons from cytochrome c, which receives them from more upstream parts of the ETC. CO passes its acquired electrons to oxygen so that H<sub>2</sub>O rather than reactive oxygen species (ROS) is generated. CO is the single greatest site of cell oxygen consumption.

The AD CO Vmax activity reduction was first shown in platelets in 1990 and brain in 1992 (Parker et al., 1990; Kish et al., 1992). Anatomic distribution of the CO defect was initially debated; it is now accepted it occurs at least in platelets, fibroblasts, and large parts of the brain (Swerdlow and Kish, 2002). Reports using different experimental methods suggest CO activity in AD subjects is 10–50% less than in age-matched controls. In general, studies that isolate mitochondria report bigger reductions than studies that do not. CO reduction occurs at all stages of the disease, including mild cognitive impairment (MCI) (Valla et al., 2006).

Because CO activity is reduced outside the brain it presumably is not a neurodegeneration artifact. Whether the Vmax reduction reflects reduced CO enzyme or a structurally altered enzyme is still debated. The Vmax activity rate is reduced in tissue homogenates normalized to total cell protein, in highly purified mitochondrial fractions in which the activity rate is normalized to total mitochondrial protein, and when normalized to activity of the mitochondrial enzyme citrate synthase (Swerdlow and Kish, 2002). Spectral analysis of the enzyme finds it is kinetically altered in AD and lacks one of its two substrate binding sites (Parker and Parks, 1995). Studies report AD brains have increased CO immunochemical staining and CO gene

expression (Hirai et al., 2001; Manczak et al. 2004). All this supports the view CO activity is reduced in AD because the enzyme is structurally different than it is in controls. Other studies, though, report CO protein levels and mRNA levels for several CO genes are reduced in AD and support the possibility CO activity is reduced because there is less CO (Chandrasekaran et al., 1994; Kish et al., 1999). Perhaps both scenarios occur. It is important to note as far as bioenergetics goes this debate does not stop with CO. Some studies suggest mitochondrial DNA (mtDNA) and mitochondrial number increase in AD, while others suggest mtDNA and mitochondrial number decline (de la Monte et al., 2000; Hirai et al., 2001; Baloyannis, 2006).

The CO enzyme complex contains 13 protein subunits. Ten are encoded by nuclear and three by mtDNA genes. Cybrid studies suggest mtDNA is at least partly responsible for reduced CO activity in AD (Swerdlow et al., 1997; Swerdlow, 2007c). Cybrids are cell lines created by placing mitochondria from individual human subjects into cultured cells (King and Attardi, 1989). Cybrids are also useful for assessing the integrity of an individual's mitochondrial genes, because when the individual's mitochondria are transferred the genes inside those mitochondria are also transferred. The mitochondria moved into the cultured cells are grown inside the cells until there are enough to study. Cybrid studies show mitochondria obtained from platelets of persons with AD have reduced CO activity (Swerdlow et al., 2007c). This specific biochemical defect persists over time in the cybrid lines, which supports the view mtDNA differs between AD and control subjects and that these differences can affect CO. Although some studies find specific sequence-level differences between AD and control subject brain and platelet mtDNA, it is not clear such differences also occur in cybrids (Corral-Debrinski et al., 1994; Mecocci et al., 1994; Coskun et al., 2004; Coon et al., 2006). It is still not known how exactly mtDNA from AD subjects differs from that of controls. Regardless, considerable data suggest maternally inherited genes could influence AD phenomena (discussed below).

### **Does Inheritance Influence AD Risk by Influencing Mitochondrial Function?**

Data from epidemiologic, neuropsychological, biomarker, and cell biology studies suggest mitochondrial inheritance could influence AD risk and pathology. One study reported mtDNA haplogroups influence AD risk (van der Walt et al., 2004). Another study found for AD subjects with a demented parent, the demented parent was more often the mother (Edland et al., 1996). This relationship persisted after correcting for greater female longevity, and implies having an AD mother confers a greater risk of AD than having an AD father. A large analysis of Framingham study subject offspring (the Framingham Offspring Study) found neuropsychological test performance in non-demented, middle aged individuals with an AD-affected mother was deficient relative to those with an AD-affected father or no AD-affected parent (Wolf et al., 2005). This may or may not reflect a presymptomatic AD carriage state. Positron emission tomography (PET) studies are relevant to this question. PET quantifies brain glucose utilization and reports this as a "cerebral metabolic rate of glucose" ( $CMR_{glu}$ ). AD subjects have reduced  $CMR_{glu}$  in particular brain regions as do cognitively intact persons with an APOE4 allele (Small et al., 1995; Reiman et al., 1996). Cognitively intact, middle aged individuals with AD mothers but not AD fathers also have AD-like patterns of  $CMR_{glu}$  reduction (Mosconi et al., 2007).

It was also previously reported cybrid cell lines containing mtDNA from individuals with AD mothers had lower CO activity than cybrids containing mtDNA from individuals with AD fathers (Davis et al., 1997). Taken together, epidemiologic (maternal inheritance bias), neuropsychological (differences in cognitive testing between children of AD-affected mothers and fathers), biomarker (PET differences in  $CMR_{glu}$  between children of AD mothers and fathers), and cell biology (perpetuation of AD pathology in cybrid cell lines, as well as a maternal inheritance effect on cybrid CO activity) data suggest mitochondrial inheritance could influence AD risk and pathology.

Nuclear DNA inheritance may also influence AD risk through effects on mitochondrial function. Under in vitro conditions different apolipoprotein E isoforms differentially affect mitochondrial function (Chang et al., 2005). Single nucleotide polymorphisms (SNP) in the gene for TFAM, a protein required for mtDNA replication and expression, and TOMM40, a translocase of the outer mitochondrial membrane protein, may associate with AD (Gunther et al., 2004; Belin et al., 2007; Alvarez et al., 2008; Bekris et al., 2008). We recently concluded a study in which SNP frequencies for the 13 CO genes was determined. Interim data are shown in Figure 2 (Swerdlow et al., 2006). We found non-synonymous nuclear CO gene SNPs in greater than 10% of the general population. CO gene nuclear SNPs occurred in combination with non-synonymous mtDNA CO gene SNPs, which were found in 20% of the general population. Nuclear CO gene untranslated region (UTR) polymorphisms were extremely common. The functional consequences of CO nuclear polymorphisms and CO nuclear DNA-mtDNA compound polymorphisms are currently under study in our laboratory.

It is already known dramatic reductions in CO activity are functionally devastating (Prick et al., 1983; Flierl et al., 1997; Cooper and Brown, 2008). The question as far as AD goes is whether CO activity variation within the “normal” range has clinical relevance. We postulate if it does, inheritance-determined function of an individual’s CO could influence AD risk. Both maternal and paternal inheritance would play important roles, but maternal inheritance would play a greater role because of the mtDNA contribution. Interestingly, a secondary analysis of our AD cybrid data found the overall CO activity reduction we observed was mostly driven by cybrids containing mtDNA from younger AD subjects (Figure 3) (Swerdlow, 2007b). This suggests the lower an individual’s inherent CO activity is, the younger the age at which the individual reaches the dementia threshold and is diagnosed with AD. To summarize this section, several lines of evidence support the view inheritance influences mitochondrial function and mitochondria-relevant gene inheritance could influence AD risk.

### **Are Age-Related Mitochondrial Changes Relevant to AD?**

Studies of aging brain mitochondria consistently report reductions of complex I activity, complex IV activity, and increased ROS production (Navarro and Boveris, 2007). Reduced numbers of mitochondria do not appear to account for these changes. Thus, a reasonable explanation is physical changes to the enzyme complexes are responsible. Other age-related mitochondrial changes include reduced membrane potential, increased size, and increased fragility.

Mitochondrial DNA mutations may contribute to age-related mitochondrial changes. In descriptive support of this, mtDNA deletions and oxidative damage accumulate with age in many tissues, especially brain (Corral-Debrinksi et al., 1992; Mecocci et al., 1993). It is easier to find low abundance, heteroplasmic point mutations (microheteroplasmy) in the brains of elderly individuals than in the brains of young individuals (Lin et al., 2002; Simon et al., 2004). To critically assess whether mtDNA mutation accumulation can promote aging, several groups have developed mice with deficient mtDNA polymerase  $\gamma$  (mtPOLG) proofreading capacity (Zhang et al., 2005; Trifunovic et al, 2004.; Kujoth et al., 2005). Characterizations of brain and brain mitochondria are scant, but immunostaining does reveal an increase in CO-negative fibers (Vermulst et al., 2008). Evaluations of multiple tissues, though, certainly show accelerated mtDNA deletion and point mutation accumulation, an age-dependent progressive reduction of ETC enzyme activities, and accelerated aging.

Similar to what is found across the mitochondrial genome, CO genes from aging individuals also accumulate sequence deviations. It was previously shown an age-related increase in CO1 gene microheteroplasmy inversely correlates with CO activity (Lin et al., 2002). If it is correct persons with lower inherent CO activities have an earlier AD onset age and by extension a

greater risk of AD (Figure 3), this finding has direct implications for the mitochondrial cascade hypothesis. It is possible to postulate a scenario in which an individual's CO activity at a given age (inherent CO activity minus the age-related CO activity decline at a particular age) either defines a threshold at which brain aging becomes brain disease, or provides a biomarker that defines when other mitochondrial parameters cross a line demarcating brain aging from brain disease (Figure 4). In this model, the distance between an individual's inherent CO activity and the CO activity disease threshold, in conjunction with how rapidly an individual's CO activity falls with aging, defines the age at which AD presents.

## Is Amyloidosis in Sporadic AD a Primary Event?

The amyloid cascade hypothesis assumes A $\beta$  occupies the apex of a neurodegenerative cascade. Support for this comes from studies of autosomal dominant AD. Research on these variants collectively suggests A $\beta$  more likely mediates disease than other APP derivatives or APP itself (Scheuner., 1996). Proponents extrapolate this paradigm to sporadic AD, which necessarily de-emphasizes the role aging may play by precluding it from the cascade apex.

The mitochondrial cascade hypothesis does not contest the view all pathology in autosomal dominant AD ultimately originates from aberrant A $\beta$  production. We are unconvinced, though, that this is the case in sporadic AD. Unlike autosomal AD, sporadic AD is truly an age-dependent event, and amyloid protein formation is a recognized consequence of aging (Swerdlow, 2007a). In addition to the brain it occurs in the heart and pancreas, and the amyloid protein deposited in each organ is unique. This "senile amyloidosis" has in fact been recognized for decades, and generally is felt to represent a consequence of aging rather than a cause of disease (Ravid et al., 1967; Cornwell and Westermarck, 1980). Indeed, aged individuals with a particular amyloid protein in one organ have an increased likelihood of having a different amyloid protein in a different organ (Schwartz, 1968; Storkel et al., 1983). This classic observation is supported by recent studies showing epidemiologic associations between AD and type II diabetes mellitus (Ott et al., 1999; Haan, 2006).

Cybrid modeling of AD mitochondrial function suggests AD subject mitochondria promote A $\beta$  production. We found increased intra and extracellular A $\beta$ 42 and A $\beta$ 40 levels in AD cybrid cell lines with reduced CO activity (Khan et al., 2000). The extracellular A $\beta$  formed adherent aggregates on flask surfaces. Living cells approached aggregate borders but did not grow on top of the aggregates. This is consistent with reports showing oxidative phosphorylation uncoupling, ATP synthase inhibition, and cytochrome oxidase inhibition converts APP processing towards amyloidogenic derivatives (Gabuzda et al., 1994; Gasparini et al., 1997; Webster et al., 1998).

Exposing cultured cells or isolated mitochondria to A $\beta$  impairs the mitochondrial ETC in general and perhaps CO specifically (Pereira et al., 1998; Canaveri et al., 1999; Crouch et al., 2005). This does not impugn and is actually consistent with the mitochondrial cascade hypothesis. Our hypothesis asserts mitochondrial dysfunction activates a final common pathway that produces all aspects of AD histology. In autosomal dominant AD, A $\beta$  overproduction is the primary cause of the mitochondrial dysfunction. In sporadic AD, mitochondrial function that declines beyond an aging-disease threshold is the primary cause. A $\beta$ , as one downstream product of the mitochondrial functional decline, presumably acts as part of a positive feedback loop that exaggerates mitochondrial dysfunction. We originally proposed cells may have developed this loop to shut down perturbed mitochondria, or else help cells transition from aerobic to anaerobic metabolism (Swerdlow and Khan, 2004). In any case, we do not believe A $\beta$  evolved solely as a toxic time bomb.

The existence of a mitochondria- A $\beta$  nexus is supported by work showing APP, A $\beta$ , PS proteins, and in fact the entire  $\gamma$  secretase complex is in fact found in mitochondria (Yamaguchi

et al., 1992; Anandatheerthavarada et al., 2003; Lustbader et al., 2004; Hansson et al., 2004; Caspersen et al., 2005; Teng and Tang, 2005; Manczak et al., 2006; Devi et al., 2006; Anandatheerthavarada and Devi, 2007; Hansson Petersen et al., 2008). The importance of the mitochondria- A $\beta$  nexus is supported by experiments performed with mtDNA-depleted cells that lack ETC function. NT2 neuronal cells are killed by A $\beta$ , while mtDNA-depleted NT2 cells are not (Cardoso et al., 2001). This implies A $\beta$  toxicity is mediated through effects on mitochondria.

## Are Tau Pathology and Cell Cycle Re-entry Related Events?

Tangles consist of aggregated microtubule associated protein (MAP) tau, tangle tau is excessively phosphorylated at serine and threonine residues, and tau phosphorylation presumably facilitates aggregation (Billingsley and Kincaid, 1997; Lovestone and Reynolds, 1997; Johnson and Hartigan, 1999). Although MAP tau gene mutations drive neurodegeneration, the resultant phenotype is characteristic of frontotemporal dementia rather than AD and altered A $\beta$  production is not characteristic of primary tauopathies (Cairns et al., 2007). Polymorphic variation of the tau gene defines two extended haplotypes, H1 and H2; H1 associates with progressive supranuclear palsy, corticobasal degeneration, Parkinson's disease, but probably not AD (Baker et al., 1999; Pittman et al., 2006; Mukherjee et al., 2007). Data from autosomal dominant AD therefore indicate A $\beta$  perturbations can drive tau phosphorylation and tangle formation, while data showing the converse are lacking. The amyloid cascade hypothesis assumes A $\beta$  drives tangle formation in both autosomal dominant and sporadic AD but in both cases is mechanistically vague on this point.

The mitochondrial cascade hypothesis postulates reduced cell energy promotes tau phosphorylation in both sporadic and autosomal dominant AD (Swerdlow and Khan, 2004). Supporting data come from experiments showing prolonged fasting, complex I inhibition, CO inhibition, mitochondrial uncoupling, and hibernation (a state associated with mitochondrial uncoupling) induce tau phosphorylation (Blass et al., 1990; Yanagisawa et al., 1999; Arendt et al., 2003; Szabados et al., 2004; Escobar-Khondiker et al., 2007; Hartig et al., 2007). In addition to several neurodegenerative disorders, tau phosphorylation also occurs in normal embryogenesis and transformed cells. Repeated cell division in the latter two cases utilizes considerable energy and energy storage is not possible. In AD, we predict energy levels are low due to mitochondrial failure.

Cell growth and cycling are regulated by trophic stimulation and energy status. Trophins bind insulin, insulin-like growth factor 1 (IGF1), and other related receptors belonging to the tyrosine kinase receptor family. In neurons, ligand binding induces insulin receptor substrate 2 (IRS2) tyrosine phosphorylation, phospho-activated IRS2 phosphorylates phosphoinositide 3 kinase (PI3K), and this activating phosphorylation eventually drives phosphorylation and activation of the Akt proto-oncogene protein (Bondy and Cheng, 2004; Manning and Cantely, 2007). We believe cells use this trophin-stimulated pathway to sense whether their external environment will support growth or division. Cell energy status also regulates Akt activity. The mammalian target of rapamycin (mTOR) protein helps cells adapt to fluctuations in their energy supply (Schieke and Finkel, 2006). mTOR complexes with other proteins, and one of these complexes, mTORC2, also phosphorylates Akt (Hresko and Mueckler, 2005; Sarbassov et al., 2005a; Sarbassov et al., 2005b).

Cell growth sometimes occurs in conjunction with division and sometimes it does not. Growing to divide and growing not to divide (hypertrophy), though, are fundamentally different processes. Cytoskeletal maintenance is not advantageous in dividing cells and high energy demand precludes energy storage. Growth without division requires a cytoskeleton and energy storage is possible. It is not surprising a single Akt-regulated protein, glycogen synthase kinase

3 $\beta$  (GSK3 $\beta$ ), coordinates cytoskeletal maintenance with energy storage. GSK3 $\beta$  phosphorylates tau and inhibits glycogen synthase (Cohen and Frame, 2001; Grimes and Jope, 2001; Bhat and Budd, 2002). Tau phosphorylation prevents cytoskeleton formation or promotes cytoskeleton disassembly, while glycogen synthase inhibition blocks glucose storage. GSK3 $\beta$  activation is therefore conducive to cell division.

Substantial evidence reveals AD neuron cell cycle re-entry is more common than it is in non-AD brain (Vincent et al., 1996; McShea et al., 1997; Nagy et al., 1997; Arendt et al., 2000; Yang et al., 2001; Herrup and Arendt, 2002; Yang et al., 2003; Mosch et al., 2007). This re-entry proceeds through several parts of the cycle but fails at the G<sub>2</sub>-M interface (Zhu et al., 2004). The mitochondrial cascade hypothesis proposes tau phosphorylation/tangle formation and cell cycle re-entry are related events, and experimental support for this prediction was recently published (McShea et al., 2007). We speculate preserved trophic stimulation in the face of low energy mimics a cycling profile and causes AD neurons to act as dividing cells. If correct, Akt must function differently under conditions of trophic stimulation/high energy than it does under conditions of trophic stimulation/low energy (Figure 5). Trophic stimulation/high energy would presumably cause Akt to inhibit GSK3 $\beta$ , thus facilitating growth without division. Trophic stimulation/low energy would have to activate the Akt proto-oncogene protein in such a way that GSK3 $\beta$  inhibition does not occur. We are currently exploring this possibility. Finally, it is necessary to postulate another factor, perhaps oxidative stress, causes phosphorylated tau to form tangle aggregations.

## Do Mitochondria Contribute Directly to AD Neuron Degeneration?

Published data suggest AD neurons die an apoptotic death (Su et al., 1994; Cotman, 1998; Rohn et al., 2002; Colurso et al., 2003; Blalock et al., 2004; Takuma et al., 2005). These data are sparse and inferential, since it is not possible to directly observe actual dying neurons in AD brain. Mitochondria presumably initiate neurodegeneration-related apoptosis by releasing proteins to the cytoplasm or nucleus that activate programmed cell death (PCD) pathways (Mattson, 2000). Accumulation of the mitochondrial proteins cytochrome c, apoptosis initiating factor (AIF), and Smac/DIABLO beyond their typical mitochondrial confines clearly induces PCD in laboratory models (Saelens et al., 2004). We previously reported cytoplasmic cytochrome c levels are elevated in AD cybrids, and caspase 3 activity is also increased (Khan et al., 2000). Mitochondrial depolarization is associated with cytochrome c egress, and AD cybrid mitochondria are relatively depolarized (Cassarino et al., 1998; Khan et al., 2000; Trimmer et al., 2000; Cardoso et al., 2004).

We believe apoptotic pathways are not maintained in absolute on or off states, but rather reflect mitochondrial integrity, number, or both. Cells with greater mitochondrial surface areas may have higher apoptotic set points, as might cells with failing mitochondria (Khan et al., 2000). One study of AD brain did show increased mitochondrial surface area as well as increased numbers of degrading mitochondria (Hirai et al., 2001). AD cybrids similarly show increased numbers of mitochondria and perturbed mitochondrial infrastructure (Trimmer et al., 2000; Trimmer et al., 2004).

## Considerations and Implications

We believe aging and sporadic AD are fundamentally convergent, not divergent, entities. This reflects clinical and biomarker experience showing cognitive change and A $\beta$  plaque accumulation are common (although not obligatory) consequences of aging and can precede dementia by years if not decades (Swerdlow, 2007a; Aizenstein et al., 2008). The mitochondrial cascade hypothesis asserts declining mitochondrial function initiates these clinical and biomarker changes and therefore represents the primary underlying problem in sporadic AD. We

do not claim, though, that AD pathology simply exaggerates age-related physiologic changes. Whether mitochondrial biogenesis increases or decreases in sporadic AD is pertinent to this point. Studies of brain aging suggest mtDNA copy number and ETC protein levels increase with age; we assume this represents a compensatory response to falling ETC efficiency (Barrientos et al., 1997; Hirai et al., 2001). Data addressing these parameters in AD brain are not uniform, and some investigators conclude mitochondrial biogenesis increases further in AD while others conclude it decreases (de la Monte et al., 2000; Hirai et al., 2001). We await this issue's definitive resolution. If mitochondrial biogenesis increases, it would suggest a physiologic divide between aging and AD arises at the point compensation is no longer possible. If mitochondrial biogenesis decreases, the point at which the cell abandons compensatory efforts could define this divide.

We have not critically questioned whether A $\beta$  is more relevant to AD than APP or other APP derivatives. Most proponents of the amyloid cascade hypothesis believe A $\beta$  is central although this assumption has been challenged (Galvan et al., 2007). While our hypothesis states mitochondrial dysfunction increases A $\beta$  levels and increased A $\beta$  affects mitochondrial function, our A $\beta$  emphasis is partly for heuristic purposes and APP or other APP derivatives could ultimately prove more relevant. A better understanding of how APP, A $\beta$ , or secretase enzymes access mitochondria and whether APP processing occurs in or on mitochondria could help address this.

There are data that argue A $\beta$  is not toxic under physiologic *in vivo* conditions and may even mitigate other sporadic AD-associated pathologies (Nunomura et al., 2001; Aizenstein et al., 2008). A $\beta$  and oxidative stress marker levels inversely correlate in particular brain regions, suggesting A $\beta$  reduces ROS production (Nunomura et al., 2001). Limited A $\beta$  exposure has also been associated with mitochondrial biogenesis in animal and cell models (Reddy et al., 2004; Diana et al., 2008). Our hypothesis is comfortable with both sides of this debate. As we stated in 2004, we believe A $\beta$  production (or other changes in APP physiology) in sporadic AD reflects an adaptive but failed attempt to disable impaired mitochondria, or else an adaptive but failed attempt by aerobically marginal cells to acquire an anaerobic phenotype (Swerdlow and Khan, 2004).

The mitochondrial cascade hypothesis predicts effective autosomal dominant AD treatments could prove partly or wholly ineffective in sporadic AD. Even if A $\beta$  exacerbates mitochondrial failure in sporadic AD, its removal may not adequately restore mitochondrial function to tolerable levels. There have been enormous efforts in recent years to treat sporadic AD by removing fibrillar A $\beta$ , blocking A $\beta$  oligomerization, and preventing formation of A $\beta$ 42. Testing these approaches in small numbers of autosomal dominant AD subjects before attempting them in large numbers of sporadic AD subjects might prove useful. Our hypothesis may also help explain why interventions that benefit APP transgenic mice have thus far not benefited sporadic AD patients (Swerdlow, 2007a; Holmes et al., 2008).

When we first proposed the mitochondrial cascade hypothesis it was acknowledged to at least a small degree by the AD research field and an independent group subsequently advocated a mitochondria-centric AD hypothesis (Swerdlow and Khan, 2005; Roses et al., 2007; Mancuso et al., 2007). A mitochondrial cascade hypothesis has been formulated for sporadic Parkinson's disease (PD), and it would seem reasonable to also formulate a mitochondrial cascade hypothesis for sporadic amyotrophic lateral sclerosis (ALS) (Domingues et al., 2008). Like AD, PD and ALS have rare autosomal forms in which mitochondrial dysfunction may play an intermediary role in neurodegeneration, and common sporadic forms that are associated with mitochondrial dysfunction (Swerdlow, 2007d).



In formulating the mitochondrial cascade hypothesis we considered epidemiologic, clinical, biochemical, and molecular studies of human sporadic AD (Swerdlow and Khan, 2004). We used cybrid cell lines to model sporadic AD mitochondrial function, and data from this model are critical to this hypothesis. Animal data from ETC inhibition models, mtDNA manipulation models, and Mendelian manipulation models also contributed. For example, APP transgenic mice with increased mitochondrial oxidative stress as a consequence of reduced manganese superoxide dismutase show increased plaque deposition, while APP transgenic mice with decreased mitochondrial oxidative stress as a consequence of reduced CO holoenzyme show decreased plaque deposition (Li et al., 2004; Fukui et al., 2007). These studies support our view that mitochondrial physiology influences amyloidosis. How best to model a sporadic disease, though, is a critical question that will impact the popularity of our hypothesis. Without accessible models, interest will likely remain limited regardless of how conceptually reasonable our hypothesis seems.

## References

- Aizenstein HJ, Nebes RD, Saxton JA, Price JC, Mathis CA, Tsopelas ND, Ziolkowski SK, James JA, Snitz BE, Houck PR, Bi W, Cohen AD, Lopresti BJ, DeKosky ST, Halligan EM, Klunk WE. Frequent amyloid deposition without significant cognitive impairment among the elderly. *Arch Neurol* 2008;65:1509–1517. [PubMed: 19001171]
- Alvarez V, Corao AI, Alonso-Montes C, Sanchez-Ferrero E, De Mena L, Morales B, Garcia-Castro M, Coto E. Mitochondrial transcription factor A (TFAM) gene variation and risk of late-onset Alzheimer's disease. *J Alzheimers Dis* 2008;13:275–280. [PubMed: 18430995]
- Anandatheerthavarada HK, Biswas G, Robin MA, Avadhani NG. Mitochondrial targeting and a novel transmembrane arrest of Alzheimer's amyloid precursor protein impairs mitochondrial function in neuronal cells. *J Cell Biol* 2003;161:41–54. [PubMed: 12695498]
- Anandatheerthavarada HK, Devi L. Amyloid precursor protein and mitochondrial dysfunction in Alzheimer's disease. *Neuroscientist* 2007;13:626–638. [PubMed: 17911214]
- Arendt T, Holzer M, Stobe A, Gartner U, Luth HJ, Bruckner MK, Ueberham U. Activated mitogenic signaling induces a process of dedifferentiation in Alzheimer's disease that eventually results in cell death. *Ann N Y Acad Sci* 2000;920:249–255. [PubMed: 11193159]
- Arendt T, Stieler J, Strijkstra AM, Hut RA, Rudiger J, Van der Zee EA, Harkany T, Holzer M, Hartig W. Reversible paired helical filament-like phosphorylation of tau is an adaptive process associated with neuronal plasticity in hibernating animals. *J Neurosci* 2003;23:6972–6981. [PubMed: 12904458]
- Baker M, Litvan I, Houlden H, Adamson J, Dickson D, Perez-Tur J, Hardy J, Lynch T, Bigio E, Hutton M. Association of an extended haplotype in the tau gene with progressive supranuclear palsy. *Hum Mol Genet* 1999;8:711–715. [PubMed: 10072441]
- Baloyannis SJ. Mitochondrial alterations in Alzheimer's disease. *J Alzheimers Dis* 2006;9:119–126. [PubMed: 16873959]
- Barrientos A, Casademont J, Cardellach F, Estivill X, Urbano-Marquez A, Nunes V. Reduced steady-state levels of mitochondrial RNA and increased mitochondrial DNA amount in human brain with aging. *Brain Res Mol Brain Res* 1997;52:284–289. [PubMed: 9495550]
- Bekris LM, Millard SP, Galloway NM, Vuletic S, Albers JJ, Li G, Galasko DR, DeCarli C, Farlow MR, Clark CM, Quinn JF, Kaye JA, Schellenberg GD, Tsuang D, Peskind ER, Yu CE. Multiple SNPs within and surrounding the apolipoprotein E gene influence cerebrospinal fluid apolipoprotein E protein levels. *J Alzheimers Dis* 2008;13:255–266. [PubMed: 18430993]
- Belin AC, Bjork BF, Westerlund M, Galter D, Sydow O, Lind C, Pernold K, Rosvall L, Hakansson A, Winblad B, Nissbrandt H, Graff C, Olson L. Association study of two genetic variants in mitochondrial transcription factor A (TFAM) in Alzheimer's and Parkinson's disease. *Neurosci Lett* 2007;420:257–262. [PubMed: 17537576]
- Bhat RV, Budd SL. GSK3beta signalling: casting a wide net in Alzheimer's disease. *Neurosignals* 2002;11:251–261. [PubMed: 12566926]

- Billingsley ML, Kincaid RL. Regulated phosphorylation and dephosphorylation of tau protein: effects on microtubule interaction, intracellular trafficking and neurodegeneration. *Biochem J* 1997;323 (Pt 3):577–591. [PubMed: 9169588]
- Blalock EM, Geddes JW, Chen KC, Porter NM, Markesbery WR, Landfield PW. Incipient Alzheimer's disease: microarray correlation analyses reveal major transcriptional and tumor suppressor responses. *Proc Natl Acad Sci U S A* 2004;101:2173–2178. [PubMed: 14769913]
- Blass JP, Baker AC, Ko L, Black RS. Induction of Alzheimer antigens by an uncoupler of oxidative phosphorylation. *Arch Neurol* 1990;47:864–869. [PubMed: 2375692]
- Bondy CA, Cheng CM. Signaling by insulin-like growth factor 1 in brain. *Eur J Pharmacol* 2004;490:25–31. [PubMed: 15094071]
- Cairns NJ, Bigio EH, Mackenzie IR, Neumann M, Lee VM, Hatanpaa KJ, White CL 3rd, Schneider JA, Grinberg LT, Halliday G, Duyckaerts C, Lowe JS, Holm IE, Tolnay M, Okamoto K, Yokoo H, Murayama S, Woulfe J, Munoz DG, Dickson DW, Ince PG, Trojanowski JQ, Mann DM. Neuropathologic diagnostic and nosologic criteria for frontotemporal lobar degeneration: consensus of the Consortium for Frontotemporal Lobar Degeneration. *Acta Neuropathol* 2007;114:5–22. [PubMed: 17579875]
- Canevari L, Clark JB, Bates TE. beta-Amyloid fragment 25–35 selectively decreases complex IV activity in isolated mitochondria. *FEBS Lett* 1999;457:131–134. [PubMed: 10486579]
- Cardoso SM, Santos S, Swerdlow RH, Oliveira CR. Functional mitochondria are required for amyloid beta-mediated neurotoxicity. *FASEB J* 2001;15:1439–1441. [PubMed: 11387250]
- Cardoso SM, Santana I, Swerdlow RH, Oliveira CR. Mitochondria dysfunction of Alzheimer's disease cybrids enhances Aβ toxicity. *J Neurochem* 2004;89:1417–1426. [PubMed: 15189344]
- Caspersen C, Wang N, Yao J, Sosunov A, Chen X, Lustbader JW, Xu HW, Stern D, McKhann G, Yan SD. Mitochondrial Aβ: a potential focal point for neuronal metabolic dysfunction in Alzheimer's disease. *FASEB J* 2005;19:2040–2041. [PubMed: 16210396]
- Cassarino DS, Swerdlow RH, Parks JK, Parker WD Jr, Bennett JP Jr. Cyclosporin A increases resting mitochondrial membrane potential in SY5Y cells and reverses the depressed mitochondrial membrane potential of Alzheimer's disease cybrids. *Biochem Biophys Res Commun* 1998;248:168–173. [PubMed: 9675105]
- Chandrasekaran K, Giordano T, Brady DR, Stoll J, Martin LJ, Rapoport SI. Impairment in mitochondrial cytochrome oxidase gene expression in Alzheimer disease. *Brain Res Mol Brain Res* 1994;24:336–340. [PubMed: 7968373]
- Chang S, ran Ma T, Miranda RD, Balestra ME, Mahley RW, Huang Y. Lipid- and receptor-binding regions of apolipoprotein E4 fragments act in concert to cause mitochondrial dysfunction and neurotoxicity. *Proc Natl Acad Sci U S A* 2005;102:18694–18699. [PubMed: 16344479]
- Cohen P, Frame S. The renaissance of GSK3. *Nat Rev Mol Cell Biol* 2001;2:769–776. [PubMed: 11584304]
- Colurso GJ, Nilson JE, Vervoort LG. Quantitative assessment of DNA fragmentation and beta-amyloid deposition in insular cortex and midfrontal gyrus from patients with Alzheimer's disease. *Life Sci* 2003;73:1795–1803. [PubMed: 12888118]
- Coon KD, Valla J, Szelinger S, Schneider LE, Niedzielko TL, Brown KM, Pearson JV, Halperin R, Dunckley T, Papassotiropoulos A, Caselli RJ, Reiman EM, Stephan DA. Quantitation of heteroplasmy of mtDNA sequence variants identified in a population of AD patients and controls by array-based resequencing. *Mitochondrion* 2006;6:194–210. [PubMed: 16920408]
- Cooper CE, Brown GC. The inhibition of mitochondrial cytochrome oxidase by the gases carbon monoxide, nitric oxide, hydrogen cyanide and hydrogen sulfide: chemical mechanism and physiological significance. *J Bioenerg Biomembr*. 2008
- Cornwell GG 3rd, Westermarck P. Senile amyloidosis: a protean manifestation of the aging process. *J Clin Pathol* 1980;33:1146–1152. [PubMed: 7005266]
- Corral-Debrinski M, Horton T, Lott MT, Shoffner JM, McKee AC, Beal MF, Graham BH, Wallace DC. Marked changes in mitochondrial DNA deletion levels in Alzheimer brains. *Genomics* 1994;23:471–476. [PubMed: 7835898]

- Coskun PE, Beal MF, Wallace DC. Alzheimer's brains harbor somatic mtDNA control-region mutations that suppress mitochondrial transcription and replication. *Proc Natl Acad Sci U S A* 2004;101:10726–10731. [PubMed: 15247418]
- Cotman CW. Apoptosis decision cascades and neuronal degeneration in Alzheimer's disease. *Neurobiol Aging* 1998;19:S29–32. [PubMed: 9562464]
- Crouch PJ, Blake R, Duce JA, Ciccotosto GD, Li QX, Barnham KJ, Curtain CC, Cherny RA, Cappai R, Dyrks T, Masters CL, Trounce IA. Copper-dependent inhibition of human cytochrome c oxidase by a dimeric conformer of amyloid- $\beta$ 1–42. *J Neurosci* 2005;25:672–679. [PubMed: 15659604]
- Davis RE, Miller S, Herrstadt C, Ghosh SS, Fahy E, Shinobu LA, Galasko D, Thal LJ, Beal MF, Howell N, Parker WD Jr. Mutations in mitochondrial cytochrome c oxidase genes segregate with late-onset Alzheimer disease. *Proc Natl Acad Sci U S A* 1997;94:4526–4531. [PubMed: 9114023]
- de la Monte SM, Luong T, Neely TR, Robinson D, Wands JR. Mitochondrial DNA damage as a mechanism of cell loss in Alzheimer's disease. *Lab Invest* 2000;80:1323–1335. [PubMed: 10950123]
- Devi L, Prabhu BM, Galati DF, Avadhani NG, Anandatheerthavarada HK. Accumulation of amyloid precursor protein in the mitochondrial import channels of human Alzheimer's disease brain is associated with mitochondrial dysfunction. *J Neurosci* 2006;26:9057–9068. [PubMed: 16943564]
- Diana A, Simic G, Sinforiani E, Orru N, Pichiri G, Bono G. Mitochondria morphology and DNA content upon sublethal exposure to beta-amyloid(1–42) peptide. *Coll Antropol* 2008;32(Suppl 1):51–58. [PubMed: 18405058]
- Domingues AF, Arduino DM, Esteves AR, Swerdlow RH, Oliveira CR, Cardoso SM. Mitochondria and ubiquitin-proteasomal system interplay: relevance to Parkinson's disease. *Free Radic Biol Med* 2008;45:820–825. [PubMed: 18619530]
- Edland SD, Silverman JM, Peskind ER, Tsuang D, Wijsman E, Morris JC. Increased risk of dementia in mothers of Alzheimer's disease cases: evidence for maternal inheritance. *Neurology* 1996;47:254–256. [PubMed: 8710088]
- Escobar-Khondiker M, Hollerhage M, Muriel MP, Champy P, Bach A, Depienne C, Respondek G, Yamada ES, Lannuzel A, Yagi T, Hirsch EC, Oertel WH, Jacob R, Michel PP, Ruberg M, Höglinger GU. Annonacin, a natural mitochondrial complex I inhibitor, causes tau pathology in cultured neurons. *J Neurosci* 2007;27:7827–7837. [PubMed: 17634376]
- Flierl A, Reichmann H, Seibel P. Pathophysiology of the MELAS 3243 transition mutation. *J Biol Chem* 1997;272:27189–27196. [PubMed: 9341162]
- Fukui H, Diaz F, Garcia S, Moraes CT. Cytochrome c oxidase deficiency in neurons decreases both oxidative stress and amyloid formation in a mouse model of Alzheimer's disease. *Proc Natl Acad Sci U S A* 2007;104:14163–14168. [PubMed: 17715058]
- Gabuzda D, Busciglio J, Chen LB, Matsudaira P, Yankner BA. Inhibition of energy metabolism alters the processing of amyloid precursor protein and induces a potentially amyloidogenic derivative. *J Biol Chem* 1994;269:13623–13628. [PubMed: 8175797]
- Galvan V, Gorostiza OF, Banwait S, Ataie M, Logvinova AV, Sitaraman S, Carlson E, Sagi SA, Chevallier N, Jin K, Greenberg DA, Bredesen DE. Reversal of Alzheimer's-like pathology and behavior in human APP transgenic mice by mutation of Asp664. *Proc Natl Acad Sci U S A* 2006;103:7130–7135. [PubMed: 16641106]
- Gasparini L, Racchi M, Benussi L, Curti D, Binetti G, Bianchetti A, Trabucchi M, Govoni S. Effect of energy shortage and oxidative stress on amyloid precursor protein metabolism in COS cells. *Neurosci Lett* 1997;231:113–117. [PubMed: 9291153]
- Goate A, Chartier-Harlin MC, Mullan M, Brown J, Crawford F, Fidani L, Giuffra L, Haynes A, Irving N, James L, et al. Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. *Nature* 1991;349:704–706. [PubMed: 1671712]
- Grimes CA, Jope RS. The multifaceted roles of glycogen synthase kinase 3 $\beta$  in cellular signaling. *Prog Neurobiol* 2001;65:391–426. [PubMed: 11527574]
- Gunther C, von Hadeln K, Muller-Thomsen T, Alberici A, Binetti G, Hock C, Nitsch RM, Stoppe G, Reiss J, Gal A, Finckh U. Possible association of mitochondrial transcription factor A (TFAM) genotype with sporadic Alzheimer disease. *Neurosci Lett* 2004;369:219–223. [PubMed: 15464268]
- Haan MN. Therapy Insight: type 2 diabetes mellitus and the risk of late-onset Alzheimer's disease. *Nat Clin Pract Neurol* 2006;2:159–166. [PubMed: 16932542]

- Haass C, Steiner H. Protofibrils, the unifying toxic molecule of neurodegenerative disorders? *Nat Neurosci* 2001;4:859–860. [PubMed: 11528409]
- Hansson CA, Frykman S, Farmery MR, Tjernberg LO, Nilsberth C, Pursglove SE, Ito A, Winblad B, Cowburn RF, Thyberg J, Ankarcrona M. Nicastrin, presenilin, APH-1, and PEN-2 form active gamma-secretase complexes in mitochondria. *J Biol Chem* 2004;279:51654–51660. [PubMed: 15456764]
- Hansson Petersen CA, Alikhani N, Behbahani H, Wiehager B, Pavlov PF, Alafuzoff I, Leinonen V, Ito A, Winblad B, Glaser E, Ankarcrona M. The amyloid beta-peptide is imported into mitochondria via the TOM import machinery and localized to mitochondrial cristae. *Proc Natl Acad Sci U S A* 2008;105:13145–13150. [PubMed: 18757748]
- Hardy JA, Higgins GA. Alzheimer's disease: the amyloid cascade hypothesis. *Science* 1992;256:184–185. [PubMed: 1566067]
- Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science* 2002;297:353–356. [PubMed: 12130773]
- Hartig W, Stieler J, Boerema AS, Wolf J, Schmidt U, Weissfuss J, Bullmann T, Strijkstra AM, Arendt T. Hibernation model of tau phosphorylation in hamsters: selective vulnerability of cholinergic basal forebrain neurons- implications for Alzheimer's disease. *Eur J Neurosci* 2007;25:69–80. [PubMed: 17241268]
- Herrup K, Arendt T. Re-expression of cell cycle proteins induces neuronal cell death during Alzheimer's disease. *J Alzheimers Dis* 2002;4:243–247. [PubMed: 12226544]
- Hirai K, Aliev G, Nunomura A, Fujioka H, Russell RL, Atwood CS, Johnson AB, Kress Y, Vinters HV, Tabaton M, Shimohama S, Cash AD, Siedlak SL, Harris PL, Jones PK, Petersen RB, Perry G, Smith MA. Mitochondrial abnormalities in Alzheimer's disease. *J Neurosci* 2001;21:3017–3023. [PubMed: 11312286]
- Holmes C, Boche D, Wilkinson D, Yadegarfar G, Hopkins V, Bayer A, Jones RW, Bullock R, Love S, Neal JW, Zotova E, Nicoll JA. Long-term effects of Abeta42 immunisation in Alzheimer's disease: follow-up of a randomised, placebo- controlled phase I trial. *Lancet* 2008;372:216–223. [PubMed: 18640458]
- Hresko RC, Mueckler M. mTOR. RICTOR is the Ser473 kinase for Akt/protein kinase B in 3T3-L1 adipocytes. *J Biol Chem* 2005;280:40406–40416. [PubMed: 16221682]
- Johnson GV, Hartigan JA. Tau protein in normal and Alzheimer's disease brain: an update. *J Alzheimers Dis* 1999;1:329–351. [PubMed: 12214129]
- Khan SM, Cassarino DS, Abramova NN, Keeney PM, Borland MK, Trimmer PA, Krebs CT, Bennett JC, Parks JK, Swerdlow RH, Parker WD Jr, Bennett JP Jr. Alzheimer's disease cybrids replicate beta-amyloid abnormalities through cell death pathways. *Ann Neurol* 2000;48:148–155. [PubMed: 10939564]
- King MP, Attardi G. Human cells lacking mtDNA: repopulation with exogenous mitochondria by complementation. *Science* 1989;246:500–503. [PubMed: 2814477]
- Kish SJ, Bergeron C, Rajput A, Dozic S, Mastrogiacomo F, Chang LJ, Wilson JM, DiStefano LM, Nobrega JN. Brain cytochrome oxidase in Alzheimer's disease. *J Neurochem* 1992;59:776–779. [PubMed: 1321237]
- Kish SJ, Mastrogiacomo F, Guttman M, Furukawa Y, Taanman JW, Dozic S, Pandolfo M, Lamarche J, DiStefano L, Chang LJ. Decreased brain protein levels of cytochrome oxidase subunits in Alzheimer's disease and in hereditary spinocerebellar ataxia disorders: a nonspecific change? *J Neurochem* 1999;72:700–707. [PubMed: 9930743]
- Kujoth GC, Hiona A, Pugh TD, Someya S, Panzer K, Wohlgemuth SE, Hofer T, Seo AY, Sullivan R, Jobling WA, Morrow JD, Van Remmen H, Sedivy JM, Yamasoba T, Tanokura M, Weindruch R, Leeuwenburgh C, Prolla TA. Mitochondrial DNA mutations, oxidative stress, and apoptosis in mammalian aging. *Science* 2005;309:481–484. [PubMed: 16020738]
- Lesne S, Koh MT, Kotilinek L, Kaye R, Glabe CG, Yang A, Gallagher M, Ashe KH. A specific amyloid-beta protein assembly in the brain impairs memory. *Nature* 2006;440:352–357. [PubMed: 16541076]
- Levy-Lahad E, Wasco W, Poorkaj P, Romano DM, Oshima J, Pettingell WH, Yu CE, Jondro PD, Schmidt SD, Wang K, et al. Candidate gene for the chromosome 1 familial Alzheimer's disease locus. *Science* 1995;269:973–977. [PubMed: 7638622]

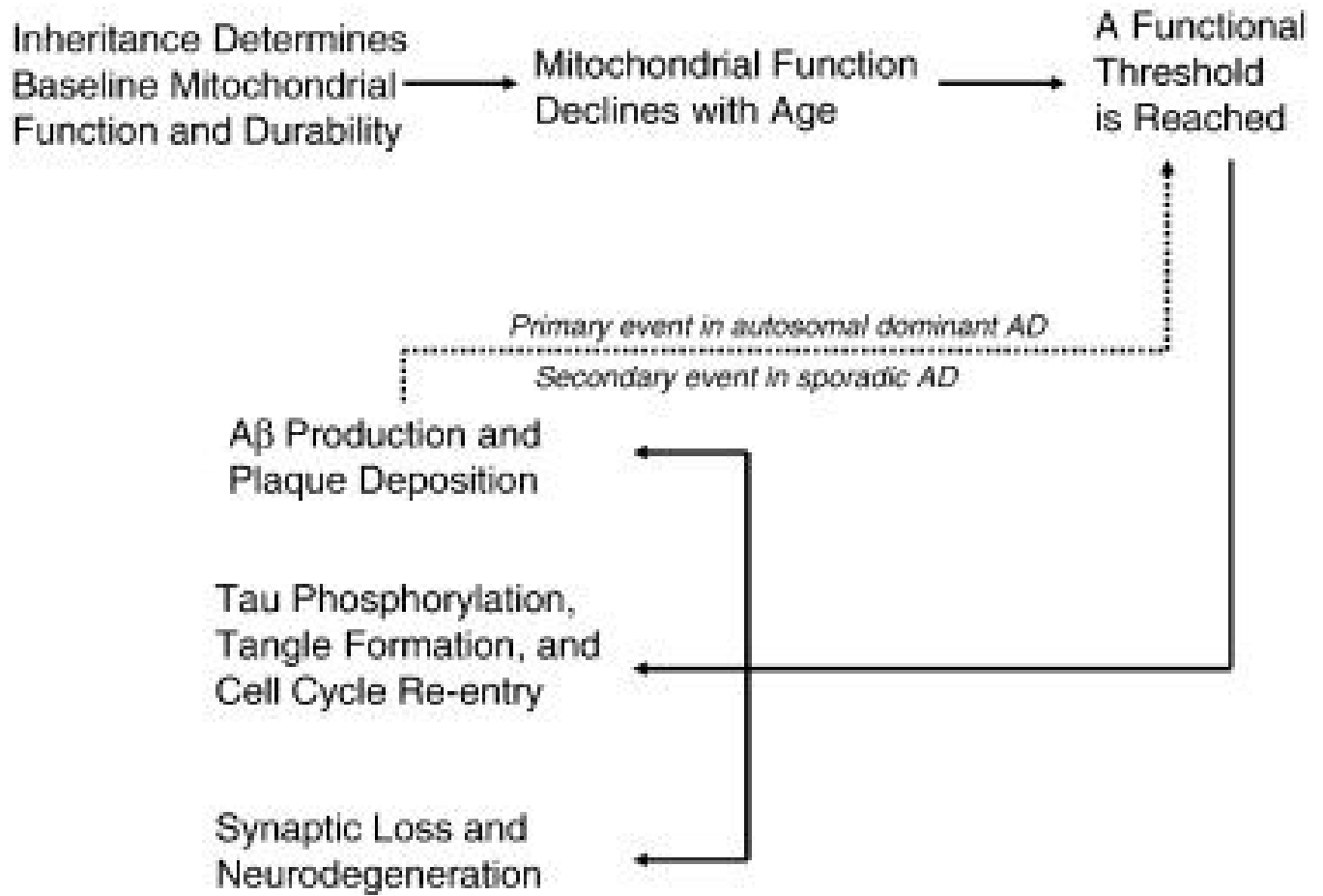
- Li F, Calingasan NY, Yu F, Mauck WM, Toidze M, Almeida CG, Takahashi RH, Carlson GA, Flint Beal M, Lin MT, Gouras GK. Increased plaque burden in brains of APP mutant MnSOD heterozygous knockout mice. *J Neurochem* 2004;89:1308–1312. [PubMed: 15147524]
- Lin MT, Simon DK, Ahn CH, Kim LM, Beal MF. High aggregate burden of somatic mtDNA point mutations in aging and Alzheimer's disease brain. *Hum Mol Genet* 2002;11:133–145. [PubMed: 11809722]
- Lovestone S, Reynolds CH. The phosphorylation of tau: a critical stage in neurodevelopment and neurodegenerative processes. *Neuroscience* 1997;78:309–324. [PubMed: 9145789]
- Lustbader JW, Cirilli M, Lin C, Xu HW, Takuma K, Wang N, Caspersen C, Chen X, Pollak S, Chaney M, Trinchese F, Liu S, Gunn-Moore F, Lue LF, Walker DG, Kuppusamy P, Zewier ZL, Arancio O, Stern D, Yan SS, Wu H. ABAD directly links Abeta to mitochondrial toxicity in Alzheimer's disease. *Science* 2004;304:448–452. [PubMed: 15087549]
- Mancuso M, Coppede F, Murri L, Siciliano G. Mitochondrial cascade hypothesis of Alzheimer's disease: myth or reality? *Antioxid Redox Signal* 2007;9:1631–1646. [PubMed: 17887917]
- Manczak M, Park BS, Jung Y, Reddy PH. Differential expression of oxidative phosphorylation genes in patients with Alzheimer's disease: implications for early mitochondrial dysfunction and oxidative damage. *Neuromolecular Med* 2004;5:147–162. [PubMed: 15075441]
- Manczak M, Anekonda TS, Henson E, Park BS, Quinn J, Reddy PH. Mitochondria are a direct site of A beta accumulation in Alzheimer's disease neurons: implications for free radical generation and oxidative damage in disease progression. *Hum Mol Genet* 2006;15:1437–1449. [PubMed: 16551656]
- Manning BD, Cantley LC. AKT/PKB signaling: navigating downstream. *Cell* 2007;129:1261–1274. [PubMed: 17604717]
- Mattson MP. Apoptosis in neurodegenerative disorders. *Nat Rev Mol Cell Biol* 2000;1:120–129. [PubMed: 11253364]
- McShea A, Harris PL, Webster KR, Wahl AF, Smith MA. Abnormal expression of the cell cycle regulators P16 and CDK4 in Alzheimer's disease. *Am J Pathol* 1997;150:1933–1939. [PubMed: 9176387]
- McShea A, Lee HG, Petersen RB, Casadesus G, Vincent I, Linford NJ, Funk JO, Shapiro RA, Smith MA. Neuronal cell cycle re-entry mediates Alzheimer disease-type changes. *Biochim Biophys Acta* 2007;1772:467–472. [PubMed: 17095196]
- Mecocci P, MacGarvey U, Kaufman AE, Koontz D, Shoffner JM, Wallace DC, Beal MF. Oxidative damage to mitochondrial DNA shows marked age-dependent increases in human brain. *Ann Neurol* 1993;34:609–616. [PubMed: 8215249]
- Mecocci P, MacGarvey U, Beal MF. Oxidative damage to mitochondrial DNA is increased in Alzheimer's disease. *Ann Neurol* 1994;36:747–751. [PubMed: 7979220]
- Mosch B, Morawski M, Mittag A, Lenz D, Tarnok A, Arendt T. Aneuploidy and DNA replication in the normal human brain and Alzheimer's disease. *J Neurosci* 2007;27:6859–6867. [PubMed: 17596434]
- Mosconi L, Brys M, Switalski R, Mistur R, Glodzik L, Pirraglia E, Tsui W, De Santi S, de Leon MJ. Maternal family history of Alzheimer's disease predisposes to reduced brain glucose metabolism. *Proc Natl Acad Sci U S A* 2007;104:19067–19072. [PubMed: 18003925]
- Mukherjee O, Kauwe JS, Mayo K, Morris JC, Goate AM. Haplotype-based association analysis of the MAPT locus in late onset Alzheimer's disease. *BMC Genet* 2007;8:3. [PubMed: 17266761]
- Nagy Z, Esiri MM, Cato AM, Smith AD. Cell cycle markers in the hippocampus in Alzheimer's disease. *Acta Neuropathol* 1997;94:6–15. [PubMed: 9224524]
- Navarro A, Boveris A. The mitochondrial energy transduction system and the aging process. *Am J Physiol Cell Physiol* 2007;292:C670–686. [PubMed: 17020935]
- Nunomura A, Perry G, Aliev G, Hirai K, Takeda A, Balraj EK, Jones PK, Ghanbari H, Wataya T, Shimohama S, Chiba S, Atwood CS, Petersen RB, Smith MA. Oxidative damage is the earliest event in Alzheimer disease. *J Neuropathol Exp Neurol* 2001;60:759–767. [PubMed: 11487050]
- Ott A, Stolk RP, van Harskamp F, Pols HA, Hofman A, Breteler MM. Diabetes mellitus and the risk of dementia: The Rotterdam Study. *Neurology* 1999;53:1937–1942. [PubMed: 10599761]
- Parker WD Jr, Filley CM, Parks JK. Cytochrome oxidase deficiency in Alzheimer's disease. *Neurology* 1990;40:1302–1303. [PubMed: 2166249]

- Parker WD Jr, Parks JK. Cytochrome c oxidase in Alzheimer's disease brain: purification and characterization. *Neurology* 1995;45:482–486. [PubMed: 7898701]
- Pereira C, Santos MS, Oliveira C. Mitochondrial function impairment induced by amyloid beta-peptide on PC12 cells. *Neuroreport* 1998;9:1749–1755. [PubMed: 9665595]
- Pittman AM, Fung HC, de Silva R. Untangling the tau gene association with neurodegenerative disorders. *Hum Mol Genet* 2006;15(Spec No 2):R188–195. [PubMed: 16987883]
- Prick MJ, Gabreels FJ, Trijbels JM, Janssen AJ, le Coultre R, van Dam K, Jaspar HH, Ebels EJ, Op de Coul AA. Progressive poliodystrophy (Alpers' disease) with a defect in cytochrome aa3 in muscle: a report of two unrelated patients. *Clin Neurol Neurosurg* 1983;85:57–70. [PubMed: 6303665]
- Ravid M, Gafni J, Sohar E, Missmahl HP. Incidence and origin of non-systemic microdeposits of amyloid. *J Clin Pathol* 1967;20:15–20. [PubMed: 5334538]
- Reddy PH, McWeeney S, Park BS, Manczak M, Gutala RV, Partovi D, Jung Y, Yau V, Searles R, Mori M, Quinn J. Gene expression profiles of transcripts in amyloid precursor protein transgenic mice: up-regulation of mitochondrial metabolism and apoptotic genes is an early cellular change in Alzheimer's disease. *Hum Mol Genet* 2004;13:1225–1240. [PubMed: 15115763]
- Reiman EM, Caselli RJ, Yun LS, Chen K, Bandy D, Minoshima S, Thibodeau SN, Osborne D. Preclinical evidence of Alzheimer's disease in persons homozygous for the epsilon 4 allele for apolipoprotein E. *N Engl J Med* 1996;334:752–758. [PubMed: 8592548]
- Rohn TT, Rissman RA, Davis MC, Kim YE, Cotman CW, Head E. Caspase-9 activation and caspase cleavage of tau in the Alzheimer's disease brain. *Neurobiol Dis* 2002;11:341–354. [PubMed: 12505426]
- Roses AD, Saunders AM, Huang Y, Strum J, Weisgraber KH, Mahley RW. Complex disease-associated pharmacogenetics: drug efficacy, drug safety, and confirmation of a pathogenetic hypothesis (Alzheimer's disease). *Pharmacogenomics J* 2007;7:10–28. [PubMed: 16770341]
- Saelens X, Festjens N, Vande Walle L, van Gorp M, van Loo G, Vandenabeele P. Toxic proteins released from mitochondria in cell death. *Oncogene* 2004;23:2861–2874. [PubMed: 15077149]
- Sarbassov DD, Guertin DA, Ali SM, Sabatini DM. Phosphorylation and regulation of Akt/PKB by the rictor-mTOR complex. *Science* 2005a;307:1098–1101. [PubMed: 15718470]
- Sarbassov DD, Ali SM, Sabatini DM. Growing roles for the mTOR pathway. *Curr Opin Cell Biol* 2005b;17:596–603. [PubMed: 16226444]
- Scheuner D, Eckman C, Jensen M, Song X, Citron M, Suzuki N, Bird TD, Hardy J, Hutton M, Kukull W, Larson E, Levy-Lahad E, Viitanen M, Peskind E, Poorkaj P, Schellenberg G, Tanzi R, Wasco W, Lannfelt L, Selkoe D, Younkin S. Secreted amyloid beta-protein similar to that in the senile plaques of Alzheimer's disease is increased in vivo by the presenilin 1 and 2 and APP mutations linked to familial Alzheimer's disease. *Nat Med* 1996;2:864–870. [PubMed: 8705854]
- Schieke SM, Finkel T. Mitochondrial signaling, TOR, and life span. *Biol Chem* 2006;387:1357–1361. [PubMed: 17081107]
- Schwartz, P. New patho-anatomic observations on amyloidosis in the aged. Fluorescence microscopic investigations. In: Mandema, E.; Ruinen, L.; Scholten, JH.; Cohen, AS., editors. *Amyloidosis*. Amsterdam: Excerpta Medica Foundation; 1968. p. 400-417.
- Shankar GM, Li S, Mehta TH, Garcia-Munoz A, Shepardson NE, Smith I, Brett FM, Farrell MA, Rowan MJ, Lemere CA, Regan CM, Walsh DM, Sabatini BL, Selkoe DJ. Amyloid-beta protein dimers isolated directly from Alzheimer's brains impair synaptic plasticity and memory. *Nat Med* 2008;14:837–842. [PubMed: 18568035]
- Sherrington R, Rogaev EI, Liang Y, Rogaeva EA, Levesque G, Ikeda M, Chi H, Lin C, Li G, Holman K, et al. Cloning of a gene bearing missense mutations in early-onset familial Alzheimer's disease. *Nature* 1995;375:754–760. [PubMed: 7596406]
- Simon DK, Lin MT, Zheng L, Liu GJ, Ahn CH, Kim LM, Mauck WM, Twu F, Beal MF, Johns DR. Somatic mitochondrial DNA mutations in cortex and substantia nigra in aging and Parkinson's disease. *Neurobiol Aging* 2004;25:71–81. [PubMed: 14675733]
- Small GW, Mazziotta JC, Collins MT, Baxter LR, Phelps ME, Mandelkern MA, Kaplan A, La Rue A, Adamson CF, Chang L, et al. Apolipoprotein E type 4 allele and cerebral glucose metabolism in relatives at risk for familial Alzheimer disease. *JAMA* 1995;273:942–947. [PubMed: 7884953]

- Storkel S, Bohl J, Schneider HM. Senile amyloidosis: principles of localization in a heterogeneous form of amyloidosis. *Virchows Arch B Cell Pathol Incl Mol Pathol* 1983;44:145–161. [PubMed: 6139907]
- Su JH, Anderson AJ, Cummings BJ, Cotman CW. Immunohistochemical evidence for apoptosis in Alzheimer's disease. *Neuroreport* 1994;5:2529–2533. [PubMed: 7696596]
- Swerdlow RH, Parks JK, Cassarino DS, Maguire DJ, Maguire RS, Bennett JP Jr, Davis RE, Parker WD Jr. Cybrids in Alzheimer's disease: a cellular model of the disease? *Neurology* 1997;49:918–925. [PubMed: 9339668]
- Swerdlow RH, Kish SJ. Mitochondria in Alzheimer's disease. *Int Rev Neurobiol* 2002;53:341–385. [PubMed: 12512346]
- Swerdlow RH, Khan SM. A “mitochondrial cascade hypothesis” for sporadic Alzheimer's disease. *Med Hypotheses* 2004;63:8–20. [PubMed: 15193340]
- Swerdlow RH, Khan S. Alzheimer research forum live discussion: A “Mitochondrial Cascade Hypothesis” for sporadic Alzheimer's disease. *Journal of Alzheimer's Disease* 2005;8:311–315.
- Swerdlow RH, Ghosh S, Wang KX. Polymorphism-determined variation of the cytochrome oxidase enzyme. *Soc Neurosci Abstr* 2006;32:79.1.
- Swerdlow RH. Is aging part of Alzheimer's disease, or is Alzheimer's disease part of aging? *Neurobiol Aging* 2007a;28:1465–1480. [PubMed: 16876913]
- Swerdlow RH. Pathogenesis of Alzheimer's disease. *Clin Interv Aging* 2007b;2:347–359. [PubMed: 18044185]
- Swerdlow RH. Mitochondria in cybrids containing mtDNA from persons with mitochondrialriopathies. *J Neurosci Res* 2007c;85:3416–3428. [PubMed: 17243174]
- Swerdlow RH. Treating neurodegeneration by modifying mitochondria: potential solutions to a “complex” problem. *Antioxid Redox Signal* 2007d;9:1591–1603. [PubMed: 17663643]
- Szabados T, Dul C, Majtenyi K, Hargitai J, Penzes Z, Urbanics R. A chronic Alzheimer's model evoked by mitochondrial poison sodium azide for pharmacological investigations. *Behav Brain Res* 2004;154:31–40. [PubMed: 15302108]
- Takuma K, Yan SS, Stern DM, Yamada K. Mitochondrial dysfunction, endoplasmic reticulum stress, and apoptosis in Alzheimer's disease. *J Pharmacol Sci* 2005;97:312–316. [PubMed: 15750290]
- Teng FY, Tang BL. Widespread gamma-secretase activity in the cell, but do we need it at the mitochondria? *Biochem Biophys Res Commun* 2005;328:1–5. [PubMed: 15670741]
- Trifunovic A, Wredenberg A, Falkenberg M, Spelbrink JN, Rovio AT, Bruder CE, Bohlooly YM, Gidlof S, Oldfors A, Wibom R, Tornell J, Jacobs HT, Larsson NG. Premature ageing in mice expressing defective mitochondrial DNA polymerase. *Nature* 2004;429:417–423. [PubMed: 15164064]
- Trimmer PA, Swerdlow RH, Parks JK, Keeney P, Bennett JP Jr, Miller SW, Davis RE, Parker WD Jr. Abnormal mitochondrial morphology in sporadic Parkinson's and Alzheimer's disease cybrid cell lines. *Exp Neurol* 2000;162:37–50. [PubMed: 10716887]
- Trimmer PA, Keeney PM, Borland MK, Simon FA, Almeida J, Swerdlow RH, Parks JP, Parker WD Jr, Bennett JP Jr. Mitochondrial abnormalities in cybrid cell models of sporadic Alzheimer's disease worsen with passage in culture. *Neurobiol Dis* 2004;15:29–39. [PubMed: 14751768]
- Valla J, Schneider L, Niedzielko T, Coon KD, Caselli R, Sabbagh MN, Ahern GL, Baxter L, Alexander G, Walker DG, Reiman EM. Impaired platelet mitochondrial activity in Alzheimer's disease and mild cognitive impairment. *Mitochondrion* 2006;6:323–330. [PubMed: 17123871]
- van der Walt JM, Dementieva YA, Martin ER, Scott WK, Nicodemus KK, Kroner CC, Welsh-Bohmer KA, Saunders AM, Roses AD, Small GW, Schmechel DE, Murali Doraiswamy P, Gilbert JR, Haines JL, Vance JM, Pericak-Vance MA. Analysis of European mitochondrial haplogroups with Alzheimer disease risk. *Neurosci Lett* 2004;365:28–32. [PubMed: 15234467]
- Vermulst M, Wanagat J, Kujoth GC, Bielas JH, Rabinovitch PS, Prolla TA, Loeb LA. DNA deletions and clonal mutations drive premature aging in mitochondrial mutator mice. *Nat Genet* 2008;40:392–394. [PubMed: 18311139]
- Vincent I, Rosado M, Davies P. Mitotic mechanisms in Alzheimer's disease? *J Cell Biol* 1996;132:413–425. [PubMed: 8636218]

- Webster MT, Pearce BR, Bowen DM, Francis PT. The effects of perturbed energy metabolism on the processing of amyloid precursor protein in PC12 cells. *J Neural Transm* 1998;105:839–853. [PubMed: 9869322]
- Wolf PA, Beiser A, Au R, Auerbach S, DeCarli C. Parental Occurrence of Dementia Linked to Lower Cognitive Function in the Framingham Offspring Study. *Neurology* 2005;64(Suppl 1):A267–A268.
- Yamaguchi H, Yamazaki T, Ishiguro K, Shoji M, Nakazato Y, Hirai S. Ultrastructural localization of Alzheimer amyloid beta/A4 protein precursor in the cytoplasm of neurons and senile plaque-associated astrocytes. *Acta Neuropathol* 1992;85:15–22. [PubMed: 1363016]
- Yanagisawa M, Planel E, Ishiguro K, Fujita SC. Starvation induces tau hyperphosphorylation in mouse brain: implications for Alzheimer's disease. *FEBS Lett* 1999;461:329–333. [PubMed: 10567721]
- Yang Y, Geldmacher DS, Herrup K. DNA replication precedes neuronal cell death in Alzheimer's disease. *J Neurosci* 2001;21:2661–2668. [PubMed: 11306619]
- Zhang D, Mott JL, Chang SW, Stevens M, Mikolajczak P, Zassenhaus HP. Mitochondrial DNA mutations activate programmed cell survival in the mouse heart. *Am J Physiol Heart Circ Physiol* 2005;288:H2476–2483. [PubMed: 15840907]
- Zhu X, McShea A, Harris PL, Raina AK, Castellani RJ, Funk JO, Shah S, Atwood C, Bowen R, Bowser R, Morelli L, Perry G, Smith MA. Elevated expression of a regulator of the G2/M phase of the cell cycle, neuronal CIP-1-associated regulator of cyclin B, in Alzheimer's disease. *J Neurosci Res* 2004;75:698–703. [PubMed: 14991845]



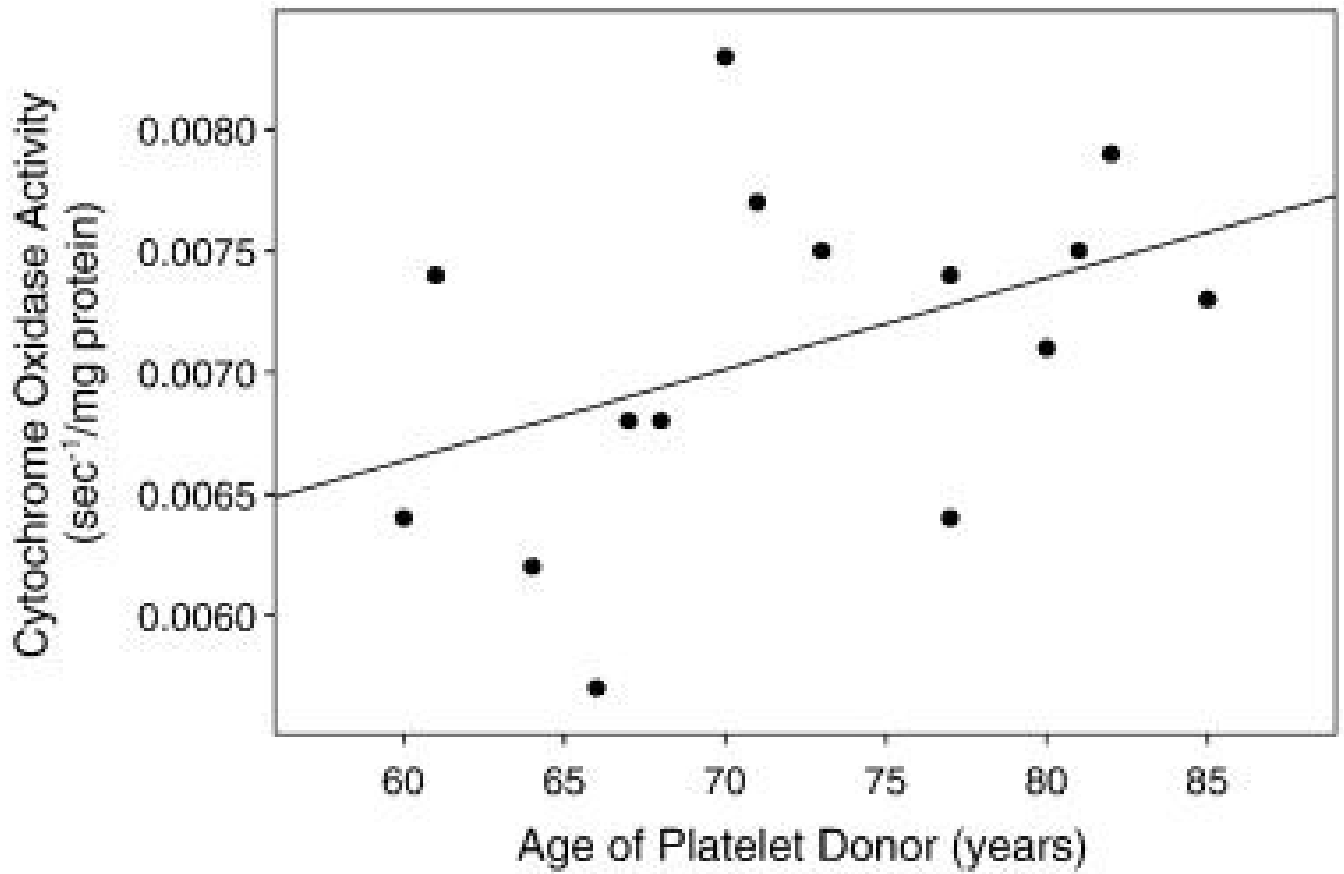


**Figure 1.**  
The mitochondrial cascade hypothesis.

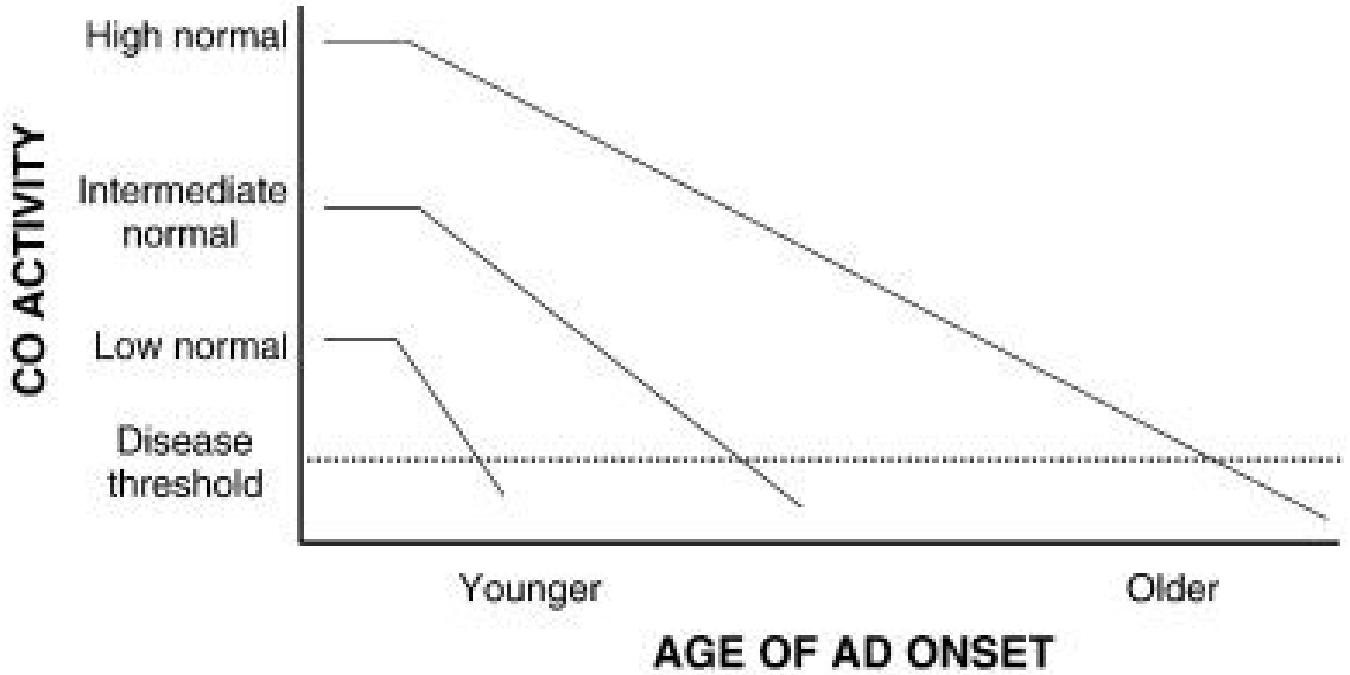
GENE(S)	POLYMORPHISM(S)	AA CHANGE(S)	FREQUENCY
CO1,2,3,4	None	None	37/50
CO4	G77A, GCU→ACU	Ala→Thr	3/50
CO3	G9477A, GUU→AUU	Val→Ile	2/50
CO3	A9667G, AAC→AGC	Asn→Ser	1/50
CO1	G6261A, GCC→AAC	Ala→Thr	1/50
CO4	G77A, GCU→ACU	Ala→Thr	
CO2	G7859A, GAU→AAU	Asp→Asn	1/50
CO1	G6366A, GUC→AUC	Val→Ile	1/50
CO3	G9966A, GUC→AUC	Val→Ile	1/50
CO1	G6261A, GUU→AUU	Val→Ile	1/50
CO3	G9966A, GUC→AUC	Val→Ile	1/50
CO4	G77A, GCU→ACU	Ala→Thr	
CO1	T6253C, AUA→ACA	Met→Thr	1/50

**Figure 2.**

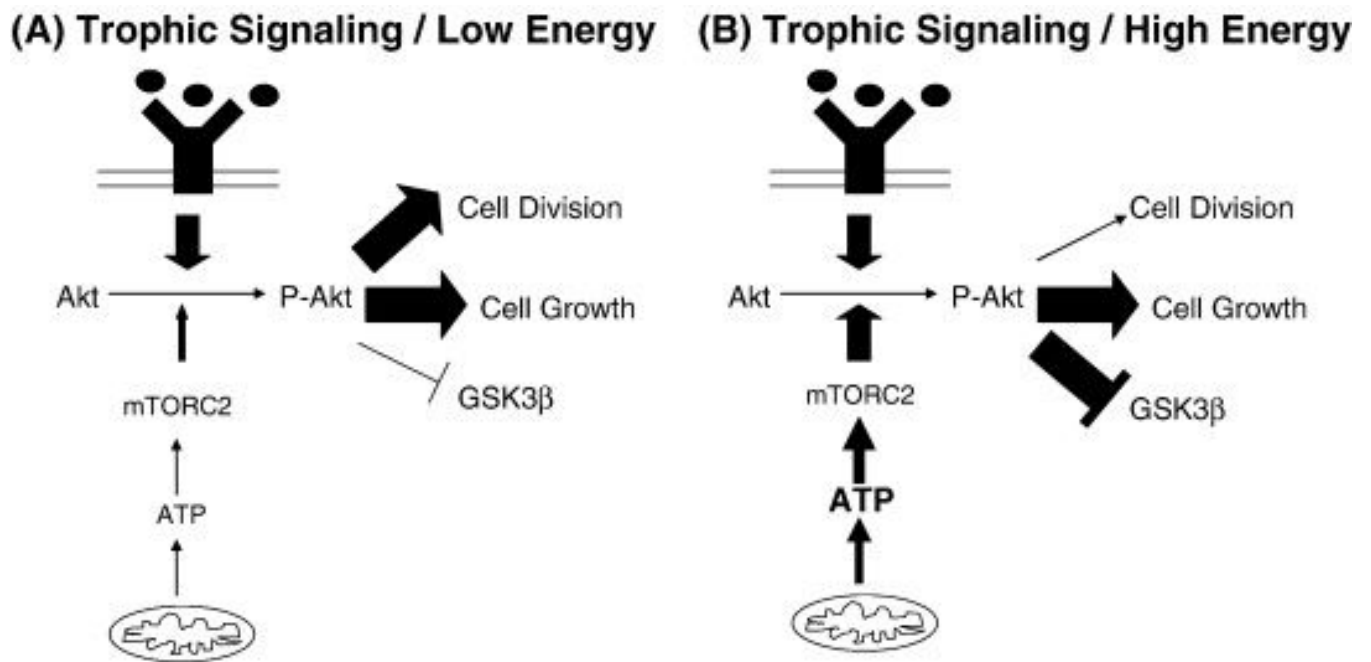
Non-synonymous open reading frame polymorphisms in 50 individuals who underwent CO1, CO2, CO3, and CO4 gene sequencing. The listed polymorphisms from the four genes analyzed define 11 distinct CO enzyme complexes among the 50 individuals. Synonymous polymorphisms, untranslated region (UTR) polymorphisms, and variation in the other 9 CO genes are not listed. The nucleotide numbers for CO1, CO2, and CO3 are from the Cambridge mtDNA sequence. The CO4 polymorphism occurs at nucleotide 77 of the mRNA. AA=amino acid.



**Figure 3.** Cytochrome oxidase activities from 15 cybrid cell lines prepared by transferring AD subject platelet mitochondria to NT2 cells depleted of endogenous mtDNA. Enzyme activities are plotted as a function of the platelet donor's age. The ages of the subjects roughly estimates the ages at which their AD developed, and suggests those with lower cytochrome oxidase activities had a younger age of onset.



**Figure 4.** An individual’s cytochrome oxidase activity at a given age reflects their baseline activity minus an age-dependent activity decline. Activity baselines and rates of decline vary between individuals. For heuristic purposes we have shown a cytochrome oxidase activity threshold below which AD symptoms manifest. Those with high baseline activities and slow decline rates would remain above the disease threshold until a very advanced age, while those with lower baseline activities and more rapid decline rates would cross the threshold at a much younger age. CO=cytochrome oxidase.



**Figure 5.**

Hypothetical scheme in which Akt drives cell growth and division under conditions of trophic stimulation and low cell energy levels, and drives cell growth without division under conditions of trophic stimulation and higher energy levels. GSK3 $\beta$ =glycogen synthase kinase 3 beta; mTORC2=mammalian target of rapamycin complex 2; P=phospho.