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Meningitis in Preterm Neonates: Importance of Cerebrospinal Fluid Parameters

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Abstract

Objective: Cerebrospinal fluid parameters are of great importance in diagnosing meningitis, but normal values for preterm neonates are based on small, single-center studies. We sought to determine current values for preterm neonate cerebrospinal fluid parameters and assess the association of cerebrospinal fluid parameters with culture proven meningitis.

Study Design: Cohort study of the first lumbar puncture from 4,632 neonates <34 weeks gestation performed in the years 1997-2004 at 150 neonatal intensive care units managed by the Pediatrics Medical Group.

Results: We identified 95 cases of meningitis from the 4,632 lumbar punctures. The area under the receiver operating characteristic curves for white blood cell count, glucose, and protein were 0.80, 0.63, and 0.72 respectively for prediction of culture proven meningitis.

Conclusion: Cerebrospinal fluid parameters used to diagnose meningitis in the absence of dependable cerebrospinal fluid cultures are unreliable. Caution should be employed when interpreting cerebrospinal fluid parameters in the premature neonate.

Keywords

nosocomial infections; central nervous system; diagnosis; preterm

INTRODUCTION

The cumulative incidence of meningitis is highest in the first month of life and is higher in preterm neonates than term neonates.¹ For premature infants who develop meningitis, the neurodevelopmental consequences are often profound.² Despite the relatively high incidence and substantial morbidity, there are only seven published studies describing expected cerebrospinal fluid (CSF) from 220 preterm neonates (Table 1),³⁻⁸ and the most recent of these reports³ appeared prior to the widespread use of surfactant.

CSF parameters (white blood cell (WBC) count, glucose, and protein) are often relied upon to diagnose meningitis in premature infants because the physical exam is unreliable due to the

subtle, nonspecific symptoms at presentation.^{1, 9-11} Reliance upon CSF parameters is especially common in the premature infant because the lumbar puncture (LP) is often delayed until after administration of empirical antibiotics and notification of a positive blood culture.^{11, 12} CSF cultures are similarly compromised in newborns of mothers given intrapartum antibiotics for chorioamnionitis or group B streptococcal colonization.^{13, 14}

When faced with the need to make therapeutic decisions on the interpretation of CSF parameters, pediatricians, family practice physicians, and neonatologists often use The Harriet Lane Handbook as their guide. The normal “cutoff” values for CSF parameters in preterm neonates found in this guide are ≤ 25 WBC/mm³, ≥ 24 mg glucose/dL, and ≤ 170 mg protein/dL.¹⁵ Unfortunately, the diagnostic utility of CSF parameters has not been evaluated for the identification of culture proven meningitis in preterm neonates. Our objective was to provide current values for CSF parameters from a large multi-center cohort of preterm neonates and assess the reliability of CSF parameters to diagnose culture proven meningitis.

MATERIALS AND METHODS

Study Population

We evaluated the first lumbar puncture from neonates <34 weeks gestation discharged from 150 neonatal intensive care units (NICUs) managed by the Pediatrix Medical Group from 1997-2004. The data were obtained from an administrative database as described previously.¹⁶ CSF data were triple-checked for accuracy. We excluded 119 neonates with CSF reservoirs and ventriculoperitoneal shunts, 85 neonates with likely contaminated CSF specimens (54 coagulase-negative *Staphylococcus* (CoNS), 8 mixed species, 6 Gram positive rods, 4 *Streptococcus viridans*, 13 other), and 12 neonates with viral meningitis diagnosed by viral culture. Contaminants were defined as CSF cultures positive for organisms generally considered contaminants (coagulase-negative staphylococci, other skin flora [viridans streptococci and diphtheroids], or mixed organisms)

Statistical Analysis

The primary outcome variable, meningitis, was defined as a positive CSF culture, a positive CSF Gram stain, or a positive CSF antigen test concordant with the blood culture result. Medians and interquartile ranges (IQR) for CSF parameters were determined for neonates with and without meningitis. Independent variables examined for diagnostic performance in predicting meningitis included: CSF WBC count, CSF glucose, and CSF protein. We evaluated the value of CSF parameters in predicting meningitis by using nonparametric methods to produce receiver operator characteristic (ROC) curves. Unlike the other CSF parameters, lower CSF glucose values are more suggestive of meningitis. In order to produce an ROC curve for CSF glucose, the CSF glucose value was multiplied by (-1). The sensitivity, specificity, positive predictive value (PPV), negative predictive value (NPV), positive likelihood ratio, and negative likelihood ratio were calculated for CSF parameters based on commonly accepted cutoff values for CSF parameters in preterm neonates.^{15, 17-19}

We considered blood and CSF cultures concordant if both cultures were positive with the same organism. We considered CSF and blood cultures discordant if the organism isolated in the CSF culture was different than the organism isolated in the blood culture or if the CSF culture was positive and the blood culture was negative. Mortality was defined as death of the neonate prior to discharge or transfer to another institution.

We conducted the analysis with STATA 8.2 (College Station, Texas) and used nonparametric (Kruskal-Wallis) tests to compare the median CSF parameter values between groups. Reported p-values are two-tailed. In order to examine the median, mean, and range of a sample of our

cohort without meningitis equivalent in size to previous reports, we used a random number generator to indicate a patient in the dataset and collected the CSF WBC for that patient and consecutive patients in the dataset without meningitis. The Duke University Institutional Review Board provided permission to conduct this analysis.

RESULTS

There were 4,632 neonates <34 weeks gestation who underwent lumbar puncture during the study period (Table 2). The majority of lumbar punctures were performed in the first month of life, 79.0% (3659/4632), and the median postnatal day of lumbar puncture was 15. The overall mortality of the cohort was 4.8% (192/4033), and mortality was higher in the neonates with meningitis, 16.1% (13/81), vs. the neonates without meningitis, 4.5% (179/3952), $p < 0.001$. The mortality rate in neonates with *Candida* meningitis was 41.2% (7/17), 14.3% (4/28) in neonates with Gram negative meningitis, and 5.6% (2/36) in neonates with Gram-positive meningitis.

Ninety-five (2.1%) of the CSF specimens were positive. *Staphylococcus aureus* (N=19) and *Candida* (N=19) were the most commonly isolated pathogens in the CSF (Table 3). *E. coli* (N=16) was the most commonly isolated Gram negative species. The blood culture was positive in 794 cases, and 428 (54%) of those were considered likely pathogens (Table 3). *Staphylococcus aureus* (N=100) was the most commonly isolated pathogen in blood. Blood culture results were missing in 281 (6.1%) patients.

Of the neonates with meningitis with available blood culture results, 28/92 (30%) were blood culture negative. Five (5.4%) of the remaining CSF and blood culture pairs were discordant. These included: 2 neonates with *Candida* in the CSF and CoNS in the blood, 1 neonate with Gram positive cocci in the CSF and *Klebsiella* in the blood, 1 neonate with *Staphylococcus aureus* in the CSF and *Candida* in the blood, and 1 neonate with *Staphylococcus aureus* in the CSF and *Enterococcus* in the blood. Fifty-nine neonates (64%) had concordant CSF and blood cultures.

CSF parameters

The median CSF WBC count in neonates with culture proven meningitis was 110 cells/mm³ (IQR, [15, 980], N=55) and 6 cells/mm³ ([2, 16], N=2396) in neonates without meningitis, $p = 0.0001$. The median CSF protein was higher in the neonates with meningitis, 217 mg/dL ([128, 499], N=49), vs. 130 mg/dL ([100, 172], N=2312) in the neonates without meningitis, $p = 0.0001$. The median CSF glucose was lower in neonates with culture proven meningitis, 43 mg/dL ([17, 52], N=50), compared to the neonates without meningitis, 49 mg/dL ([40, 60], N=2329), $p = 0.0005$.

We produced ROC curves using each CSF parameter separately to predict culture proven meningitis (Figures 1-3). The area under the ROC curve was greatest for CSF WBC count (0.80). Using the commonly accepted normal ranges for CSF parameters,^{15, 18, 19} the sensitivity of a CSF WBC count >25 cells/mm³ in predicting meningitis was 71% (Table 4). The sensitivity decreased to 61% and 32% for CSF protein >170 mg/dL and CSF glucose <24 mg/dL respectively.

CSF contaminants

There were 54 neonates excluded from the study with CoNS meningitis. Although often a pathogen in preterm neonates, the organism is a common contaminant and was excluded in our primary analysis in order to concentrate on those neonates with known true pathogens present in their CSF. Including these 54 neonates decreased the median WBC count from 110

cells/mm³ to 37 cells/mm³, decreased the median CSF protein from 217 mg/dL to 194 mg/dL, and increased the median CSF glucose from 43 mg/dL to 45 mg/dL. Similarly the area under the ROC curves decreased from 0.80 to 0.71 for CSF WBC count, 0.72 to 0.67 for CSF protein, and 0.63 to 0.59 for CSF glucose.

Using the commonly accepted normal ranges for CSF parameters,^{15, 18, 19} the sensitivity of a CSF WBC count >25 cells/mm³ for predicting CoNS meningitis was only 37% (Table 5). Similarly, cutoff values for glucose and protein demonstrated lower sensitivity and specificity for predicting CoNS meningitis when compared to their predictive values for other pathogens.

DISCUSSION

This paper reports CSF data from over 4600 neonates born during the years 1997-2004, and thus represents a major advance in the state of our knowledge. The most recent report of normal CSF parameters in preterm neonates involved 71 CSF specimens from 43 very low birth weight (birth weight <1500 grams) neonates, and the highest CSF WBC count observed was 44 cells/mm³.³ In the group of neonates in our study with negative CSF cultures, 333/2396 (14%) of the neonates had CSF WBC counts >44 cells/mm³ and 437/2396 (18%) of the neonates had CSF WBC counts >25 cells/mm³, the commonly accepted upper limit of normal.^{15, 18, 19} Excluding the neonates with >100 RBC in the CSF (possible traumatic specimens), 37/1209 (3.1%) of the neonates with negative CSF cultures had CSF WBC counts >44 cells/mm³, and 62/1209 (5.1%) had CSF WBC counts >25 cells/mm³. However, the median values for CSF parameters in our study were similar to previously reported mean values (Table 1).

In contrast to prior studies, we were able to examine the diagnostic value of CSF parameters in predicting culture proven meningitis by generating ROC curves for each individual CSF parameter. The area under the ROC curve was greatest for CSF WBC count (0.80, [0.73, 0.86]) followed by CSF protein (0.72 [0.64, 0.80]), and CSF glucose (0.63 [0.54, 0.73]). Diagnostic tests with an area under the ROC curve >0.9 are generally considered excellent, and those with an area under the ROC curve between 0.8 and 0.9 are considered good. Although ROC curves are often used to determine cutoff points for diagnostic tests, the process of developing a cutoff point must take into account the costs of treating the disease (financial costs, patient discomfort, drug side effects) in patients without the disease (false positives) with the resulting morbidity and mortality of not treating the disease in patients who truly have the disease (false negatives). Neonatal meningitis is a disease with substantial mortality and morbidity even among treated neonates.^{1, 20} Costs associated with treating false positive patients include prolonged hospital stay, exposure of the neonate to broad spectrum antibiotics, and insertion of central venous catheters for prolonged antibiotic administration.

Using the normal ranges provided in several reference texts,^{15, 17-19} 22% of preterm neonates with culture proven meningitis had normal CSF WBC count, glucose, and protein levels. The PPV of having an abnormality in the CSF was low for each of the CSF parameters (4-10%). The NPV was high for each CSF parameter (98-99%) due to the low incidence of meningitis in the cohort. The likelihood ratios observed for each of the parameters are modest at best (Table 4). For example, a preterm neonate with a 10% pretest probability of having meningitis based on one's clinical experience that has a CSF WBC count >25 cells/mm³ [positive likelihood ratio of 3.7] would have a post-test probability of culture positive meningitis of 29%.

Many clinicians often use the CSF parameters to "rule out" meningitis in the neonate. According to table 4, the CSF parameter most useful for this purpose is CSF protein. Ninety-six percent of neonates with meningitis have a CSF protein level >90 mg/dL. The negative likelihood ratio for culture proven meningitis of a neonate having a protein level ≤90 mg/dL is 0.2. CSF glucose appears to be more useful to "rule in" meningitis. The positive likelihood

ratio of a glucose level <24 mg/dL is 8.0 and increases to 23.0 for a level <10 mg/dL (Table 4). However, the sensitivity of a glucose level of <24 mg/dL is only 32% and falls to 18% for a level <10 mg/dL. A combination of all 3 parameters provides for a more robust way of “ruling in” meningitis. A neonate undergoing a lumbar puncture with >25 WBC cells/mm³, a glucose of <10 mg/dL, and >250 mg/dL of CSF protein has a 164 fold increase in odds of having a positive CSF culture. However, only 18% of neonates with positive CSF cultures were identified using these cutoff values.

LPs are often delayed because of the clinical instability of the neonate or the low perceived risk of meningitis, and thus, clinicians often defer the LP until after blood culture results are known to be positive and the neonate has been exposed to empirical antibiotics. Although only 2.1% (95/4632) of the neonates in our cohort had meningitis, 30% (28/92) of these neonates had negative blood cultures. These findings are consistent with previous multicenter reports in term and preterm neonates.^{11, 13} In contrast to previous studies describing the causative organisms of sepsis in preterm neonates, we found *Staphylococcus aureus* and *Candida* to be the most commonly isolated species in the CSF.^{21, 22}

Although the administrative database used for this study was collected prospectively, the analysis was retrospective. CSF parameters were missing in the dataset in 45% of neonates. Our analysis of the diagnostic utility of CSF glucose was limited as we did not have temporally related serum glucose levels. We acknowledge that the major weakness of this study was the lack of information regarding type and timing of empirical antibiotics to which the neonates were exposed. It is acknowledged that a number of the neonates with negative CSF cultures likely had a central nervous system infection; however, this acknowledgement does not conflict with our primary hypothesis. Chiefly, that these data suggest that the clinician should employ caution when interpreting “negative” CSF parameter results.

This is the largest collection of CSF parameters in preterm neonates presented to date. These data suggest that CSF parameters relied upon to diagnose meningitis in preterm neonates in the absence of dependable cultures are of marginal diagnostic use, and CSF parameters, either alone or in combination, cannot be used to exclude meningitis. In order to avoid diagnosing this potentially devastating infection on marginally useful CSF parameters, our findings support the argument that the LP should be performed during the initial evaluation of the clinically stable neonate with a suspected systemic infection. A prospective study using mortality and neurodevelopmental follow up will better define the utility of CSF parameters in the premature neonate. Until such study is completed, caution should be employed when interpreting CSF parameters.

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Abbreviations

CSF, cerebrospinal fluid; WBC, white blood cell; LP, lumbar puncture; NICU, neonatal intensive care unit; CoNS, coagulase negative *Staphylococcus*; IQR, interquartile range; ROC, receiver operating characteristic; PPV, positive predictive value; NPV, negative predictive value.

References

1. Overall JC Jr. Neonatal bacterial meningitis. Analysis of predisposing factors and outcome compared with matched control subjects. *J Pediatr* 1970;76:499–511. [PubMed: 5420788]
2. Stoll BJ, et al. Neurodevelopmental and growth impairment among extremely low-birth-weight infants with neonatal infection. *JAMA* 2004;292:2357–65. [PubMed: 15547163]
3. Rodriguez AF, Kaplan SL, Mason EO Jr. Cerebrospinal fluid values in the very low birth weight infant. *J Pediatr* 1990;116:971–4. [PubMed: 2189977]
4. Sarff LD, Platt LH, McCracken GH Jr. Cerebrospinal fluid evaluation in neonates: comparison of high-risk infants with and without meningitis. *J Pediatr* 1976;88:473–7. [PubMed: 1245961]
5. Otila E. Studies on the cerebrospinal fluid in premature infants. *Acta Paediatrica* 1948;1948:7–99.
6. Gyllensward A, Malmstrom S. The cerebrospinal fluid in immature infants. *Acta Paediatr* 1962:54–62.
7. Wolf H, Hoepffner L. The cerebrospinal fluid in the newborn and premature infant. *World Neurol* 1961;2:871–8. [PubMed: 13786275]
8. Samson K. Die liquordiagnostik im kindesalter. *Ergeb Inn Med Kinderheilkd* 1931;1931:551–778.
9. Feigin RD, McCracken GH Jr, Klein JO. Diagnosis and management of meningitis. *Pediatr Infect Dis J* 1992;11:785–814. [PubMed: 1448332]
10. Schwersenski J, McIntyre L, Bauer CR. Lumbar puncture frequency and cerebrospinal fluid analysis in the neonate. *Am J Dis Child* 1991;145:54–8. [PubMed: 1985430]
11. Stoll BJ, et al. To tap or not to tap: high likelihood of meningitis without sepsis among very low birth weight infants. *Pediatrics* 2004;113:1181–6. [PubMed: 15121927]
12. Fielkow S, Reuter S, Gotoff SP. Cerebrospinal fluid examination in symptom-free infants with risk factors for infection. *J Pediatr* 1991;119:971–3. [PubMed: 1960620]
13. Wiswell TE, et al. No lumbar puncture in the evaluation for early neonatal sepsis: will meningitis be missed? *Pediatrics* 1995;95:803–6. [PubMed: 7761203]
14. Weiss MG, Ionides SP, Anderson CL. Meningitis in premature infants with respiratory distress: role of admission lumbar puncture. *J Pediatr* 1991;119:973–5. [PubMed: 1960621]
15. Robertson, J.; Shilkofski, N. *The Harriet Lane Handbook*. Vol. 17th ed.. Mosby; Philadelphia: 2005. p. 557
16. Garges HP, et al. Neonatal meningitis: what is the correlation among cerebrospinal fluid cultures, blood cultures, and cerebrospinal fluid parameters? *Pediatrics* 2006;117:1094–100. [PubMed: 16585303]
17. Katz, SL.; Gershon, AA.; Hotez, PJ. *Krugman's Infectious Diseases of Children*. Vol. 10th ed.. Mosby; St. Louis: 1998. p. 769-770.
18. McMillan, JA., et al. *Oski's Pediatrics*. Vol. 3rd ed.. Lippincott Williams and Wilkens; Philadelphia: 1999. p. 414-420.
19. Fanaroff, AA.; Martin, RJ. *Neonatal-Perinatal Medicine Diseases of the Fetus and Newborn*. Vol. 7th ed.. Vol. 2. Mosby; St. Louis: 2002. p. 719-720.
20. Harvey D, Holt DE, Bedford H. Bacterial meningitis in the newborn: a prospective study of mortality and morbidity. *Semin Perinatol* 1999;23:218–25. [PubMed: 10405191]
21. Stoll BJ, et al. Changes in pathogens causing early-onset sepsis in very-low-birth-weight infants. *N Engl J Med* 2002;347:240–7. [PubMed: 12140299]
22. Stoll BJ, et al. Late-onset sepsis in very low birth weight neonates: the experience of the NICHD Neonatal Research Network. *Pediatrics* 2002;110:285–91. [PubMed: 12165580]

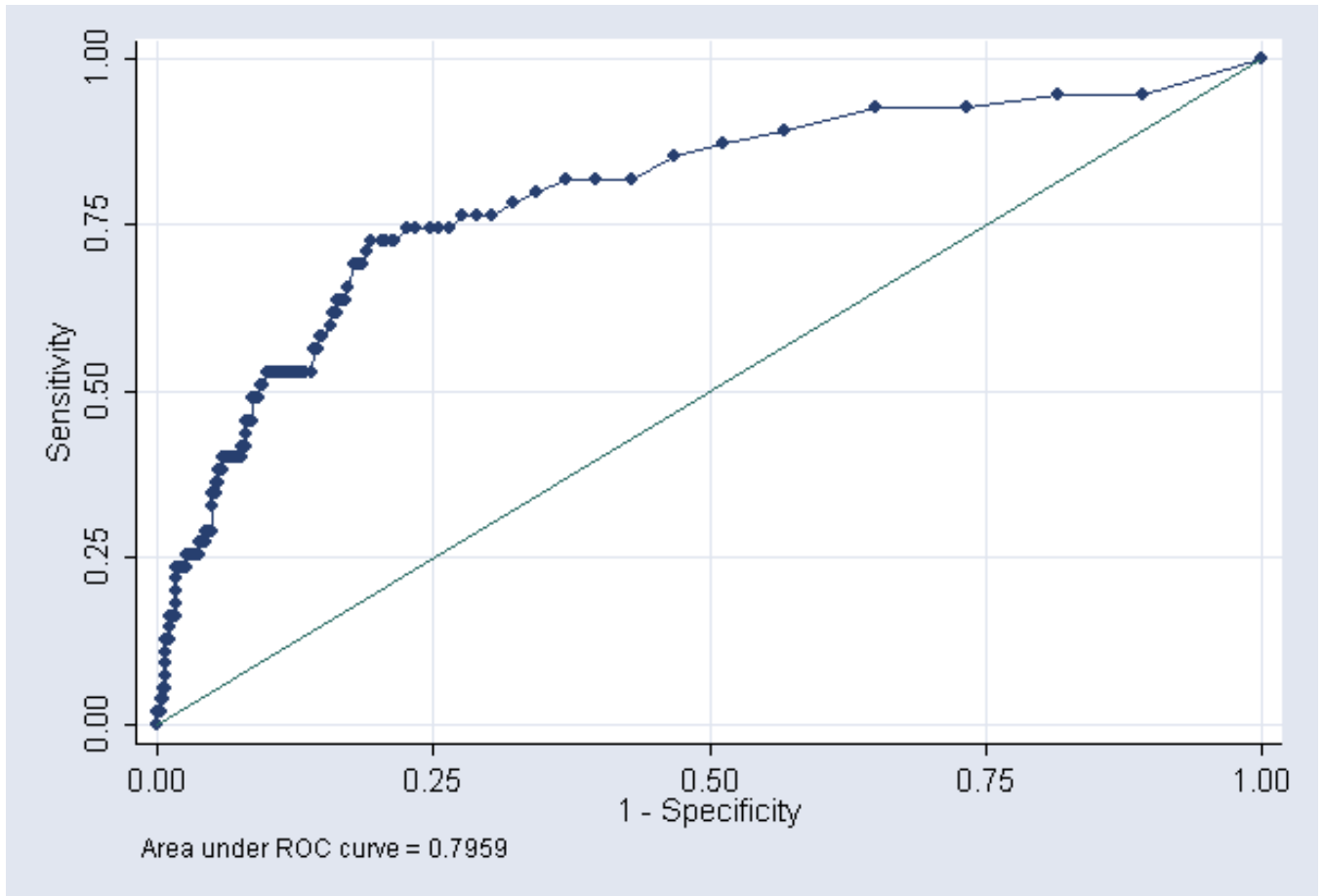


Figure 1. CSF WBC count - ROC curve showing performance of CSF parameters as a diagnostic tool for predicting meningitis

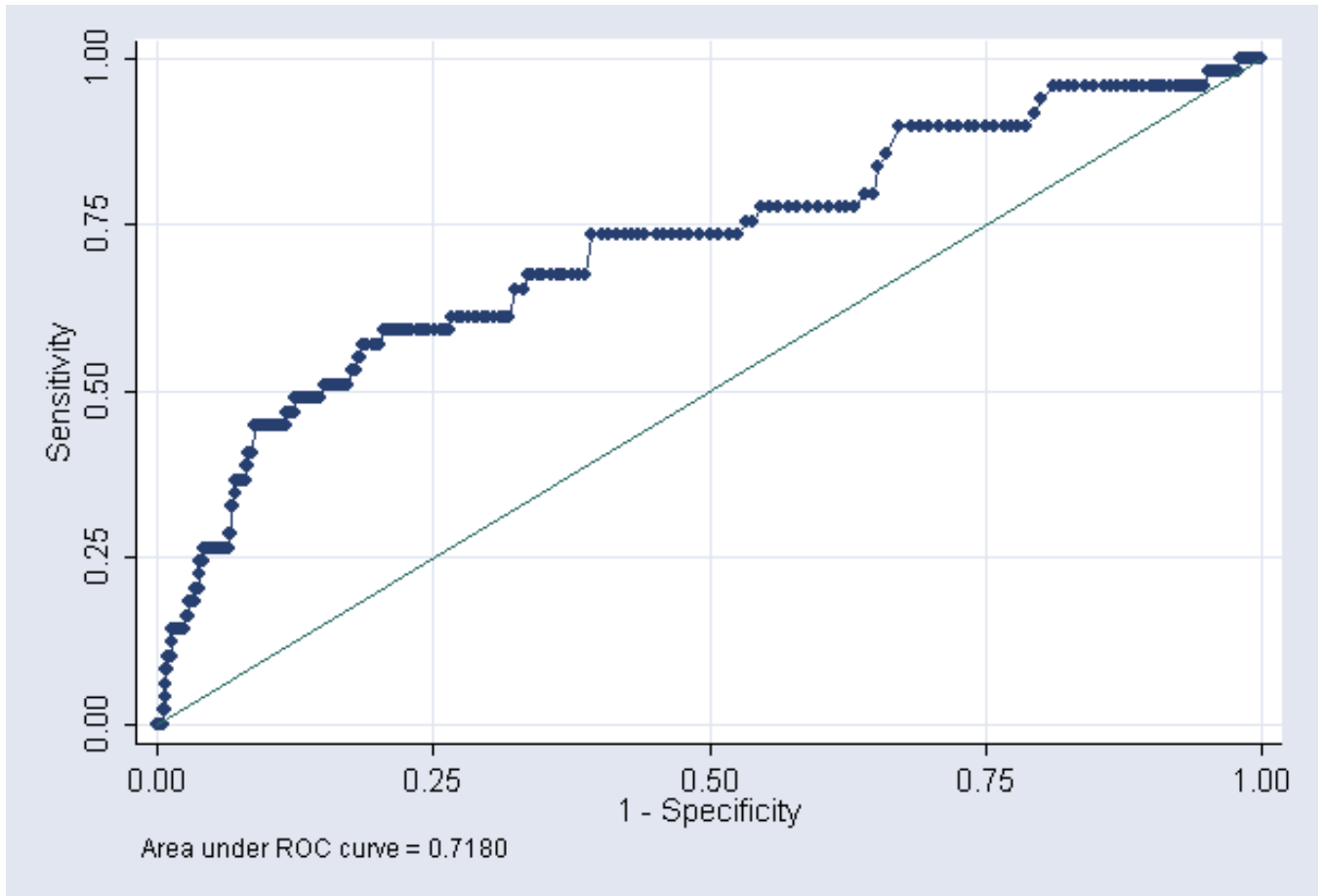


Figure 2.
CSF protein - ROC curve showing performance of CSF parameters as a diagnostic tool for predicting meningitis

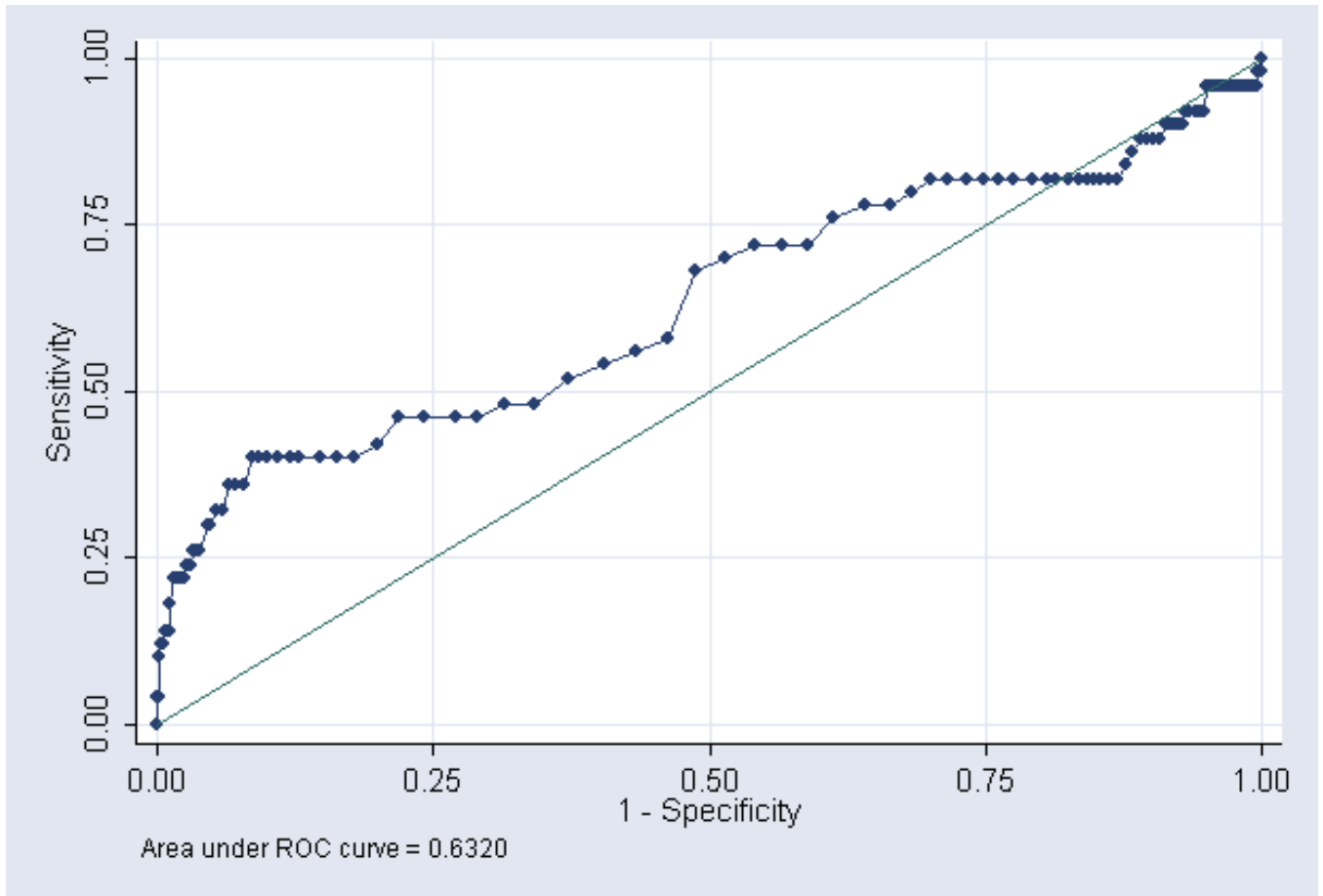


Figure 3. CSF glucose - ROC curve showing performance of CSF parameters as a diagnostic tool for predicting meningitis

Table 1

Mean values and ranges of CSF parameters previously reported normal CSF parameters for preterm neonates.

	N	Birth weight (g)	WBC/mm ³	Protein (mg/dL)	Glucose (mg/dL)
Samson K, 1931	Unk		4 (<10)		
Otila E, 1948	66		10		
Wolf H, 1961	22	1880	2 (0-13)	105 (50-180)	
Gyllesward A, 1962	36	2012	7 (1-75)	132 (55-237)	
Sarff LD, 1976	30		9 (0-29)	115 (65-150)	50 (24-63)
Pappu LD, 1982	22	1437	7 (0-28)		
Rodriguez AF, 1990	43	1002	5 (0-44)	142 (45-370)	60 (29-217)

Table 2

Demographics of the cohort

	N (%)
Cohort	4632
Gestational Age (weeks)	
22-25	845 (18)
26-29	1965 (42)
30-33	1822 (39)
Birth weight (grams)	
366-500	64 (1)
501-750	772 (17)
751-1000	1025 (22)
1001-1250	844 (18)
1251-1500	663 (14)
1501-2000	851 (18)
>2000	413 (9)
Method of delivery	
Vaginal	1963 (43)
Caesarian section	2649 (57)
Race	
White	2394 (53)
Black	1102 (24)
Hispanic	817 (18)
Asian	76 (2)
Native American	36 (1)
Other	93 (2)
Sex	
Male	2577 (56)
Female	2055 (44)
Day of lumbar puncture	
0-3	875 (19)
4-7	434 (9)
8-14	933 (20)
15-21	764 (16)
22-28	542 (12)
29-60	824 (18)
60-257	260 (6)
Death	
Yes	192 (5)
No	3841 (95)

Table 3

CSF and blood culture results

	CSF result (%)	Blood result (%)
Total	4632	4351
Positive	95 (2)	428 (10)
Negative	4537 (98)	3557 (81)
Contaminants	0 (0)*	366 (8)
Gram positive	43	207
Group B <i>streptococcus</i>	9	30
<i>Enterococcus spp.</i>	2	39
<i>Listeria monocytogenes</i>	0	1
<i>Staphylococcus aureus</i>	19	100
Gram positive unspciated	13	37
Gram negative	33	156
<i>Citrobacter spp.</i>	0	2
<i>Escherichia coli</i>	16	58
<i>Enterobacter spp.</i>	3	23
<i>Haemophilus influenza</i>	0	3
<i>Klebsiella spp.</i>	5	36
<i>Morganella spp.</i>	0	2
<i>Pseudomonas aeruginosa</i>	5	7
<i>Salmonella spp.</i>	0	1
<i>Serratia spp.</i>	3	10
Fungi	19	65
<i>Candida spp.</i>	19	64
<i>Malassezia</i>	0	1

* CSF with contaminants (e.g. CoNS) were excluded from the analysis

Table 4
CSF parameter comparison – (WBC cells/mm³, glucose mg/dL, protein mg/dL)

	Sensitivity	Specificity	PPV	NPV	+ LR	- LR
WBC>0	95	12	2	99	1.1	0.4
WBC>10	80	67	5	99	2.4	0.3
WBC>20	73	79	7	99	3.5	0.3
*WBC>25	69	82	8	99	3.9	0.4
WBC>100	51	91	11	99	5.7	0.5
WBC>1000	24	97	17	98	8	0.8
glucose<10	18	99	33	98	23.0	0.8
* glucose<24	32	96	14	98	8.0	0.7
protein>90	96	18	2	99	1.2	0.2
* protein >170	61	75	5	99	2.4	0.5
All Harriet Lane values abnormal	26	97	13	98	8.7	0.8
Any Harriet Lane value abnormal	78	65	5	99	2.2	0.3
WBC>25, glucose<10, protein>250	18	99.5	41	98	164.0	0.8

* Harriet Lane value

Table 5

CSF parameter comparison for prediction of CoNS meningitis – (WBC cells/mm³, glucose mg/dL, protein mg/dL)

	Sensitivity	Specificity	PPV	NPV	+ LR	- LR
*WBC>25	37	78	2	99	1.7	0.8
* glucose<24	11	94	2	99	2.0	0.9
* protein >170	38	73	2	99	1.4	0.8

* Harriet Lane value