# *Microsporum equinum* in North America

JULIUS KANE,<sup>1</sup> ARVIND A. PADHYE,<sup>2\*</sup> AND LIBERO AJELLO<sup>2</sup>

Laboratory Services Branch, Ontario Ministry of Health, Toronto, Ontario M5W 1R5, Canada,<sup>1</sup> and Division of Mycotic Diseases, Center for Infectious Diseases, Centers for Disease Control, Atlanta, Georgia 30333<sup>2</sup>

Received 13 May 1982/Accepted 4 August 1982

*Microsporum equinum* was isolated in Ontario, Canada, from five human and two equine cases of ringworm infection. This dermatophyte was previously recovered from North American horses on several occasions, but was considered to be *M. canis*. We regard *M. equinum* as distinct from *M. canis*. It can be differentiated from *M. canis* by the smaller size of its macroconidia, its failure to perforate hair in vitro, its poor growth and sporulation on bromocresol purple casein dextrose agar, and its incompatibility with *Nannizzia otae*, the telemorph of *M. canis*.

*Microsporum equinum* (Delacroix and Bodin, 1898) Gueguen, 1904, is one of the causal agents of ringworm in horses and humans that was reported frequently in the early literature. It had been the cause of severe epizootics in large stables (7, 8) as well as an important agent of equine ringworm in army horses in Belgium (22) and France (8). Equine and human infections due to *M. equinum* have been reported from other countries, such as Algeria (2), Australia (12, 26), Czechoslovakia (24), Finland (1), Great Britain (23, 27), Germany (7), Norway (20), New Zealand (9), Rumania (5), Russia (21, 28, 30), and Zaire (13).

In spite of its wide geographic distribution, M. equinum has never been reported from North America. Recent studies on ringworm in race horses in Caracas, Venezuela (Gladys Tapia de Fossaert, personal communication), have confirmed its occurrence as one of the agents of equine ringworm. In the North American literature, Batte and Miller (6) cursorily mentioned M. equinum as one of the causal agents of equine ringworm. Even though they cited it as one of the causal agents, the authors failed to isolate it from any of the 54 horses they examined for ringworm infections. Another reason for its absence in the North American literature may be that most mycologists (3, 14, 15, 29) accepted Conant's conclusion (10, 11) that M. equinum was a later synonym of M. canis. For instance, during the animal ringworm survey conducted by the Mycology Division of the Centers for Disease Control in the 1950s, Georg et al. (15) studied 26 isolates from equine ringworm. Four of the 26 isolates, provided by P. K. C. Austwick, were isolated from equine ringworm in England and Scotland and were identified as M. equinum. One additional isolate that Georg

TABLE	1.	Isolation	data	on	М.	equinum and i	V.
			ota	e			

	olue
Organism and accession no.	Source
M. equinum	
	Skin scrapings, human; received as OMH" 1468
B-3343	Skin scrapings, human; received as OMH 257
	Skin lesion, horse; received as OMH 273
B-3585	Lesion on forehead, human; received as OMH 247
B-3586	Lesion on leg, human; received as OMH 258
B-3587	Lesion on leg, human; received as OMH 654
B-3588	Skin lesion, horse; received as OMH 1469
N. otae	
	Dessioned from A. Hosperson
B-2094 (+)	Received from A. Hasegawa, Tokyo, Japan, as VUT 74037
D 2005 ( )	Received from A. Hasegawa,
<b>D-209</b> 5 (-)	Tokyo, Japan, as VUT 74039
B-3580 (+)	F1 progeny monoascospore strain
	from B-2094 $\times$ B-2095 cross; received from I. Weitzman as
	Ha3
B-3581 (+)	Received from M. Hironaga,
2 5561 (*)	Otsu, Japan, as VUT 77054 (+)
<b>B-3582</b> (-)	Received from M. Hironaga,
	Otsu, Japan, as VUT 77055 (-)
B-3583 (+)	Received from M. Takashio,
	Antwerp, Belgium, as RV 42487 (+)
B-3584 (-)	Received from M. Takashio, Antwerp, Belgium, as RV 42488 (-)
	in Ministry of Health Taranta Cana

<sup>*a*</sup> OMH, Ontario Ministry of Health, Toronto, Canada.

J. CLIN. MICROBIOL.

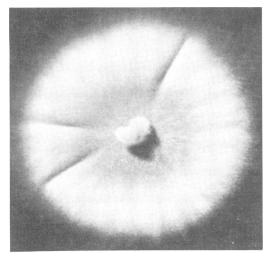


FIG. 1. Two-week-old colony of *M. equinum* on Sabouraud dextrose agar.

et al. studied was from Sabouraud's collection and also was identified originally as M. *equinum*. These five isolates were re-identified as M. *canis* by Georg et al. in accordance with Conant's conclusion. Many such isolates from ringworm in horses in the United States may have been incorrectly described in the literature as M. *canis*.

The discovery of Nannizzia otae, the teleomorph of *M. canis*, by Hasegawa and Usui (16) and the availability of their tester strains provided an opportunity to study compatibility relationships between M. canis and M. equinum. Mating studies between the tester strains of N. otae and M. equinum by Padhye et al. (25) showed that the isolates were sexually incompatible with each other. Among other tests that were done, the in vitro hair perforation test provided evidence of differences between the two species. All of the isolates of M. canis consistently perforated hair in vitro, whereas none of the M. equinum isolates did so. On the basis of the negative mating and perforation tests, they (25) concluded that *M. equinum* was a distinct and good species.

The question of the occurrence of *M. equinum* in the North American literature still remained unanswered. The present report documents the occurrence of *M. equinum* as the causal agent of ringworm in five human and two equine cases from Ontario, Canada.

#### MATERIALS AND METHODS

**Cultures.** Seven cultures of M. equinum were isolated from clinical specimens by one of us (J.K.) during a period of 8 years. The clinical material consisted of skin scrapings or epilated hair or both that had been

received from Ontario dermatologists and veterinarians by the Services Branch of the Ontario Ministry of Health, Toronto, Canada. In addition, seven tester strains of *N. otae* were included in the study for comparing the morphology and mating behavior of *M. equinum* with the anamorphs of *N. otae*. The isolation data on the *M. equinum* isolates and *N. otae* tester strains are summarized in Table 1.

**Morphology.** All cultures were grown on Sabouraud dextrose agar (1% neopeptone, 2% dextrose, 2% agar, and 0.05% chloramphenicol) and bromocresol purple (BCP) casein dextrose agar (19) for colony and micromorphology studies. The microscopic morphology of each isolate was also studied on slide cultures, using niger seed (*Guizotia abyssinica*) agar and medium 8 (17).

In vitro hair perforation (4) and urease broth tests (18) were performed on each isolate of M. equinum and M. canis.

Mating studies. Inocula from vigorously growing 10day-old colonies of M. equinum and the tester strains of N. otae were used in the mating studies. All crosses were done in duplicate, using 0.1% Sabouraud dex-

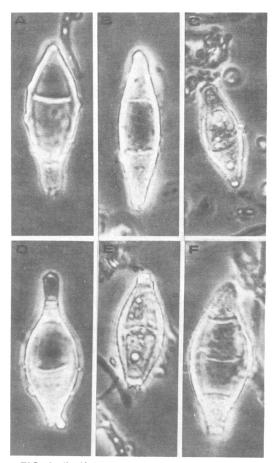


FIG. 2. Fusiform, vertucose macroconidia of M. equinum (×850).

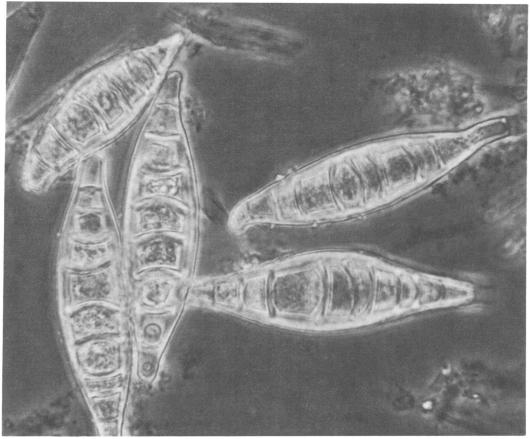


FIG. 3. Spindle-shaped, vertucose, thick-walled macroconidia of M. canis (×850).

trose agar with salts (31) and niger seed agar-medium 8 (17). They were incubated at  $25^{\circ}$ C in the dark for 4 to 6 weeks and were examined weekly for the presence of gymnothecia or pseudogymnothecia.

# RESULTS

Morphology on Sabouraud dextrose agar. Colonies of the seven isolates of M. equinum measured 60 to 70 mm in diameter after 2 weeks at 25°C. Their surfaces ranged from downy or velvety to finely powdery, especially in the central areas, and they were white or pale buff to pale salmon. Four of the 7 cultures developed radial grooves (Fig. 1). The reverse of the colonies varied from buff through amber to salmon.

Macroconidia were not readily produced by any of the isolates on Sabouraud dextrose agar. However, slide cultures with niger seed agarmedium 8 supported production of both microand macroconidia. The microconidia were pyriform to clavate, 3 to 8 by 2 to 3  $\mu$ m, one celled, smooth, and sessile or on short conidiophores, and they were borne laterally on simple hyphae. Macroconidia were generally numerous. They were elliptical to broadly fusiform, 18 to 60 by 5 to 13  $\mu$ m, with verrucose, thick walls (up to 3  $\mu$ m thick at the center) and predominantly two to four celled (Fig. 2A to F). They were borne terminally or on short lateral conidiophores, generally singly and rarely on more complexly branched conidiophores, to form clusters of macroconidia.

Colonies of *N. otae* tester strains measured 65 to 75 mm in diameter after 2 weeks at 25°C. At first, the colonies were mostly submerged, with a thin surface, strongly radiating, with a buff, powdery area developing in the center. The microconidia were clavate to pyriform, smooth, one celled, 3.5 to 8.0 by 1.5 to 3.5  $\mu$ m, and sessile or on short conidiophores. They were borne along the sides of simple hyphae. The macroconidia were fusiform, variable in size, 35 to 110 by 12 to 25  $\mu$ m, and up to 14 celled. Their verrucose walls were thick (up to 4  $\mu$ m at the center) (Fig. 3). They were borne terminally on short conidiophores at an acute angle to the parent hyphae.

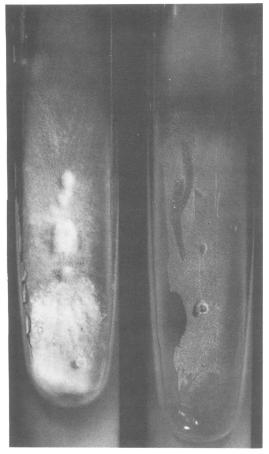


FIG. 4. Comparison of profuse growth of *M. canis* on BCP casein dextrose agar (left slant) with reduced growth of *M. equinum* (right slant) after 2 weeks.

Morphology in BCP casein dextrose agar. Colonies of *M. equinum* isolates grew poorly on BCP casein dextrose agar. The aerial mycelium was scanty and short. The growth induced a change in the pH of the medium, making it alkaline and resulting in a color change in the medium from grey to blue. Microscopically, *M. equinum* isolates did not sporulate on BCP casein dextrose agar.

Colonies of *N. otae* tester strains, on the other hand, showed good aerial growth on BCP casein dextrose agar. The mycelium was white to yellowish green and velvety to fine powdery. Microscopically, numerous macroconidia and a moderate number of microconidia were produced. The tester strains of *N. otae* did not change the pH of the medium or cause a change in color (Fig. 4).

Urease test. All of the M. canis strains hydrolyzed urea broth by 8 to 12 days. The seven isolates of M. equinum accomplished this within 8 to 14 days.

In vitro hair perforation test. All of the *M*. canis strains perforated hair in vitro, but none of the *M*. equinum isolates had perforated hair by the end of a 4-week incubation period.

Mating studies. None of the 49 crosses between seven M. equinum isolates and the seven tester strains of N. otae produced fertile gymnothecia or pseudogymnothecia or showed growth stimulation at the end of 6 weeks. These results confirmed our previous findings (25) that M. equinum was a species distinct from M. canis.

## DISCUSSION

The isolation of *M. equinum* from ringworm infections in humans and equines in Ontario, Canada, demonstrated that *M. equinum* occurs in North America. Georg et al. (15) recognized that *Trichophyton equinum* was the most common cause of equine ringworm in the United States, Canada, South America, and Europe. They recognized the following dermatophytes, in order of frequency, as being associated with equine ringworm: *M. canis*, *T. mentagrophytes* var. *mentagrophytes*, *T. verrucosum*, and *M.* gypseum. *M. equinum*, rather than *M. canis*, may prove to be the second most common incitant of equine ringworm in North America.

*M. equinum* can be easily differentiated from *M. canis* in a diagnostic laboratory by its morphological and physiological characteristics. The smaller size of the macroconidia produced by *M. equinum*, its failure to perforate hair in vitro, and its poor growth and lack of sporulation on BCP casein dextrose agar provide accurate tests to identify *M. equinum*.

#### ACKNOWLEDGMENTS

We thank Wilhelm Van der Kolk of the Ministry of Health for the photography and Margaret Kwok of the Media Department for the preparation of some of the media.

### LITERATURE CITED

- Aho, R. 1980. Studies of fungal flora in hair from domestic and laboratory animals suspected of dermatophytes. Acta Pathol. Microbiol. Scand. 88:79–83.
- Ainsworth, G. C., and P. K. C. Austwick. 1973. Fungal diseases of animals, p. 10–36. Commonwealth Agricultural Bureaux, Farnham Royal, Slough, England.
- 3. Ajello, L. 1968. A taxonomic review of the dermatophytes and related species. Sabouraudia 6:147-159.
- Ajello, L., and L. K. Georg. 1957. In vitro hair cultures for differentiating between atypical isolates of *Trichophyton* mentagrophytes and *Trichophyton rubrum*. Mycopathol. Mycol. Appl. 8:3-17.
- Alteras, I., and A. Avram. 1963. Consideratti asupra evolutiei dermatomicozelor pe 10 ani (1952–1961). Dermatol Venerol. (Bucurest) 8:41–48.
- Batte, E. G., and W. S. Miller. 1953. Ringworm of horses and its control. J. Am. Vet. Med. Assoc. 123:111-114.
- Bergner, K. 1942. Microsporum equinum und Achorion gypseum als Erreger von Flechtenerkrankungen bei Pferden. Zentralbl. Infektionskr. Haustiere 8:121-147.
- 8. Brocq-Rousseu, D., A. Urbain, and J. Barotte. 1927. Etude

des teignes du cheval et de l'immunite dans teignes experimentales. Ann. Inst. Pasteur Paris **41**:513-553.

- Carman, M. G., F. M. Rush-Munro, and M. Carter. 1979. Dermatophytes isolated from domestic and feral animals. N. Z. Vet. J. 27:136, 143–144.
- 10. Conant, N. F. 1936. Studies in the genus *Microsporum*. Arch. Dermatol. Syphilol. 33:665–683.
- 11. Conant, N. F. 1941. A statistical analysis of spore size in the genus *Microsporum*. J. Invest. Dermatol. 4:265–278.
- Connole, M. D. 1977. Current status of the ecology and epidemiology of animal mycoses with special reference to Queensland, Australia, p. 123–128. *In* K. Iwata (ed.), Recent advances in medical and veterinary mycology. University Park Press, Baltimore, Md.
- Dekeyser, J., L. Delcambe, and D. Thienpont. 1960. Activité therapeutique de l'iturine et du chinsol sur le teigne du cheval a *Microsporum equinum*. Bull. Epizoot. Dis. Afr. 8:279–288.
- Georg, L. K., W. Kaplan, and L. B. Camp. 1957. Trichophyton equinum a reevaluation of its taxonomic status. J. Invest. Dermatol. 29:27–37.
- Georg, L. K., W. Kaplan, and L. B. Camp. 1957. Equine ringworm with special reference to *Trichophyton* equinum. Am. J. Vet. Res. 18:798-810.
- Hasegawa, A., and K. Usui. 1975. Nannizzia otae sp. nov., the perfect state of Microsporum canis Bodin. Jpn. J. Med. Mycol. 16:148–153.
- Hironaga, M., K. Nozaki, and S. Watanabe. 1980. Ascocarp production by *Nannizzia otae* on keratinous and nonkeratinous agar media and mating behaviour of *N. otae* and 123 Japanese isolates of *M. canis*. Mycopathologia 72:135–141.
- Kane, J., and J. B. Fischer. 1971. The differentiation of *Trichophyton rubrum* and *T. mentagrophytes* by use of Christensen's urea broth. Can. J. Microbiol. 17:911–913.
- Kane, J., and C. Smitka. 1978. Early detection and identification of *Trichophyton vertucosum*. J. Clin. Microbiol. 8:740–747.

- Lindqvist, K. 1960. Ringworms hos husdyr i Norge. Nord. Veterinaermed. 12:21-28.
- Medvedeva, E. A. 1964. O microsporii volosistol chasti golovy obuslovlennoi *Microsporum equinum*. Vestn. Dermatol. Venerol. 38:25-26.
- 22. Neefs and Gillain. 1931. Contribution a l'etude de la teigne. Ann. Med. Vet. 76:193-209.
- O'Grady, K. J., M. P. English, and R. P. Warin. 1972. Microsporum equinum infection of the scalp in an adult. Br. J. Dermatol. 86:175–176.
- Otčenášek, M., K. Krivanee, J. Dvořak, J. Komárek, and A. Černa. 1975. *Microsporum equinum* als Erreger einer dermatophytose des Pferdes. Zentralbl. Veterinaermed. 22:833-841.
- Padhye, A. A., I. Weitzman, and L. Ajello. 1979. Mating behaviour of *Microsporum equinum* with *Nannizzia otae*. Mycopathologia 69:87-90.
- Pascoe, R. R. 1976. Studies on the prevalence of ringworm among horses in racing and breeding stables. Aust. Vet. J. 52:419–421.
- Pepin, G. A., and P. K. C. Austwick. 1968. Skin diseases of domestic animals. II. Skin diseases, mycological origin. Vet. Rec. 82:208-214.
- Petrovich, S. V. 1975. Vozbuditeli dermatomikozov loshadei. Veterinariya (Moscow) 10:49-51.
- 29. Silva-Hutner, M., I. Weitzman, and S. A. Rosenthal. 1981. Cutaneous mycoses (Dermatomycoses), p. 863–929. *In* A. Balows and W. J. Hausler (ed.), Diagnostic procedures for bacterial, mycotic and parasitic infections, 6th ed. American Public Health Association, Washington, D.C.
- Spesivtzeva, N. A. 1957. Forty years of veterinary mycology. J. Agric. Sci. 34:37–43.
- 31. Takashio, M. 1981. Production. maintenance and utilization of tester strains by sexual crossings of dermatophytes. A propos of *Nannizzia otae*, p. 69–80. *In* R. Vanbreuseghem and C. De Vroey (ed.), Sexuality and pathogenicity of fungi. Masson Publishing U.S.A., Inc., New York.