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FVB/N mice are highly resistant to primary infection with *Nippostrongylus brasiliensis*

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SUMMARY

Nippostrongylus brasiliensis larvae are particularly susceptible to immunological attack during the pre-lung stage of primary and secondary infections in mice. Whilst most of the common laboratory strains of mice are permissive hosts for the parasite, in this study we report for the first time, the strong resistance of naïve FVB/N mice to *N. brasiliensis*. Damage to larvae is evident within the first 24 h of infection and this may be critical to later larval development and reproductive success. Inflammatory responses in the skin, and larval escape from this tissue were comparable in susceptible CBA/Ca and resistant FVB/N mice, with most larvae exiting within 4 h of a primary infection. Lung larval burdens were also similar between strains, but larvae recovered from FVB/N mice were smaller and less motile. In FVB/N mice, larval colonization of the gut was impaired and worms produced very few eggs. However FVB/N mice did not show enhanced resistance to *Heligmosomoides bakeri* (also known as *Heligmosomoides polygyrus*), a nematode largely restricted to the gut. Damage done in the pre-lung or lung stages of infection with *N. brasiliensis* is likely to contribute to ongoing developmental and functional abnormalities, which are profoundly evident in the gut phase of infection.

Keywords

FVB/N mouse; *Nippostrongylus brasiliensis*; *Heligmosomoides bakeri*; *Heligmosomoides polygyrus*; primary resistance

INTRODUCTION

Parasitic helminth infections are a major problem worldwide and there are currently no vaccines available for use in humans. Tissue-invasive helminths employ immune evasion strategies that allow them to reside in the host for extended periods and, in some cases, for decades. Helminthic infections may result in a cellular response characterized by eosinophils, neutrophils and basophils (Kay *et al.* 1985). Eosinophils can be recruited to sites of infections in high numbers; however, the contribution they may make to an effective immune response

is controversial and not well understood (Behm and Ovington, 2000; Meeusen and Balic, 2000; Lee and Lee, 2005). Eosinophils appear to be important for protection in some parasitic infections (Sasaki *et al.* 1993; Ovington *et al.* 1998; Daly *et al.* 1999; Knott *et al.* 2007), while in others, their presence may be of little consequence (Matthaei *et al.* 1997; Takamoto *et al.* 1997; Dent *et al.* 1999) or may even be counter-productive (McCormick *et al.* 1996; Dent *et al.* 1997a, b).

To further understand and characterize the roles of eosinophils in parasitic helminth infections, transgenic (Tg) mouse models are often used. Mice with constitutive eosinophilia due to expression of an interleukin (IL)-5 transgene (Dent *et al.* 1990) are highly resistant to infections with *Nippostrongylus brasiliensis*, and entrapment of infective larvae in the skin is associated with the rapid recruitment of eosinophils (Dent *et al.* 1997b; Daly, 1999; Dent *et al.* 1999; Giacomini *et al.* 2008). Other studies have shown that IL-5 Tg mice are resistant to *Strongyloides stercoralis* infections and again, eosinophil recruitment into the skin is associated with the elimination of larvae (Herbert *et al.* 2000).

The intestinal phase of helminthic infections has been the focus of considerable research, since many species spend much of their life cycle in this organ. While the roles of IL-13, IL-4 and signal transduction and activator of transcription 6 (STAT6) in the intestinal phase of the *N. brasiliensis* life cycle have been well characterized (Urban *et al.* 1998; Fallon *et al.* 2002, 2006), the contributions that IL-5 and eosinophils make at this stage of infection are largely unknown. Recent data from our laboratory suggest that deletion of IL-5 may cause impairment of resistance in the intestinal phase of primary infections with *N. brasiliensis*, as well as the early pre-lung phase of secondary infections (Knott *et al.* 2007). Initially, a major objective of this study was to examine the consequences of intestine-specific expression of IL-5 (i-FABP-IL-5) or eotaxin (i-FABP-eotaxin) transgenes (Mishra *et al.* 2002) on helminth infections. Unexpectedly, the wildtype (WT) FVB/N animals used as controls were found to be highly resistant to primary infections with *N. brasiliensis*. This paper describes an investigation of the kinetics of nematode infections in FVB/N mice and the nature of the cellular inflammatory responses elicited.

MATERIALS AND METHODS

Animals

Heterozygous male and female CBA/Ca mice with approximately 49 copies (Tg5C2, abbreviated IL-5 Tg) of the IL-5 transgene (Dent *et al.* 1990) and their WT littermates were bred in the Medical School Animal House of the University of Adelaide. WT FVB/N male and female mice were bred by Laboratory Animal Services, University of Adelaide. Intestinal fatty acid binding protein (i-FABP)-IL-5 and i-FABP-eotaxin FVB/N Tg mice (Mishra *et al.* 2002) were bred at the John Curtin School of Medical Research, Australian National University. All mice were closely age-matched within experiments. Hooded Wistar rats used for passage of *N. brasiliensis* were obtained from Laboratory Animal Services at the University of Adelaide. All animals were handled according to the guidelines of the University of Adelaide Animal Ethics Committee.

Parasitological techniques

Parasite maintenance—Infective *N. brasiliensis* third-stage larvae (L3) were obtained by the faecal culture method after passage through Hooded Wistar rats as described previously (Daly, 1999), washed twice in MilliQ water and once in mouse osmolality phosphate-buffered saline (MPBS) (Sheridan and Finlay-Jones, 1977). Each mouse received 500 larvae for air pouch injections and 750 or 1000 larvae for subcutaneous (s.c.) injections. L3 were injected at the base of the neck or into air pouches in the middorsal skin using a 19-gauge needle. Mice

in secondary infection groups were injected s.c. with 750 L3 and 3 weeks later, after clearance of the primary infection, mice were challenged with 750 L3 and lung larval numbers enumerated.

Infective *Heligmosomoides bakeri* (*H. polygyrus*) (Cable *et al.* 2006) L3 were also obtained by the faecal culture method, but passage was performed in BALB/c mice. Larvae were stored at 4 °C until used, at which time the concentration was adjusted to 2000 L3/ml and each experimental animal received 200 L3 by oral gavage.

Parasite recovery from tissues—For *N. brasiliensis* experiments, lung larvae (L4) were recovered from animals on days 1 and/or 2 post-injection (p.i.) as previously described (Knott *et al.* 2007). The motility of larvae in minced lung preparations was assessed after 2 h of incubation at 37 °C by recording the number of major movements by individual larvae over 30 sec. Larvae were recovered from 3 mice/group and 30 larvae were examined/mouse. Motility data for larvae recovered from individual mice were combined to yield $n=90$ larvae/group. Faecal egg counts (day 6 p.i. for *N. brasiliensis* and various time-points from 13–116 days p.i. for *H. bakeri*) and intestinal worm burdens (day 7 p.i. for *N. brasiliensis* and days 41 and 117 p.i. for *H. bakeri*) were also performed as previously described (Giacomin *et al.* 2005; Knott *et al.* 2007).

Measurement of parasite length—Intestinal worms (day 7 p.i.) or lung larvae (day 2 p.i.) were collected from WT FVB/N and WT CBA/Ca mice and photographed using Konica Centuria ASA 400 film, a dissecting microscope (Olympus) and an Olympus c-35AD-4 camera. 6–19 larvae or worms were photographed from lungs or intestines at 56× magnification for each of 3–4 mice/group. The lengths of larvae and worms were determined from photographs using a cartographer's wheel (Kartenmesser, Germany).

Generation of skin air pouches

Air pouches were generated as previously described (Daly *et al.* 1999). Briefly, the shaved dorsal skin of anaesthetized mice was inflated subcutaneously with 2.5 ml of filtered air and 3 days later, pouches were re-inflated with 2 ml of sterile air and larvae were injected (500 L3/mouse) after a further 2 days. Air pouches were lavaged 4 or 5 times with 2 ml of MPBS, 0 and 4 h after injection of larvae. The viable cells and larvae recovered were enumerated and differential leukocyte counts were estimated from cytocentrifuged samples, with a minimum of 200 cells counted per sample using oil immersion light microscopy.

Histology

Lungs were removed on day 2 p.i. and the post-caval lobe (right lung) was taken for histology. Tissues were fixed in 10% buffered formalin and embedded in paraffin. Five µm sections were cut and stained with haematoxylin & eosin (H&E) and photographed at 20× or 40× objective magnification.

Statistical analysis

Parasite burdens and size and inflammatory leukocyte numbers were compared using the non-parametric Kruskal-Wallis (KW) test, followed by Mann-Whitney U (MW) post-hoc testing or the parametric two-way ANOVA followed by Bonferroni post-hoc testing in Prism Version 5.01 (GraphPad Software Inc., La Jolla, CA, USA). Worm length was assessed in a linear mixed effects model followed by Bonferroni post-hoc testing with SAS Version 9.1 (SAS Institute Inc., Cary, NC, USA) software by Mr Thomas Sullivan (Statistician, Department of Public Health, University of Adelaide). In all tests $P<0.05$ was considered significant.

RESULTS

Intestinal phase of infection

In initial experiments with *N. brasiliensis* infections in i-FABP-IL-5 Tg and i-FABP-eotaxin Tg FVB/N mice, few worms were present in the small intestines of the control WT FVB/N mice compared with WT CBA/Ca mice. We further investigated the observation to explore the cause of this potent resistance to *N. brasiliensis*. Using WT CBA/Ca mice as a comparison strain, which have similar susceptibility to other commonly used mouse strains such as BALB/c (Knott *et al.* 2007) and C57BL/6 (Giacomin *et al.* 2005, 2008), we confirmed that fewer worms were present in the intestines of WT FVB/N mice (Fig. 1A) and fewer eggs were found (Fig. 1B). The numbers of both worms and eggs in WT FVB/N mice were comparable to those seen during primary infections in the highly resistant IL-5 Tg CBA/Ca mice (Fig. 1). We also observed that the few worms that were present in the intestines of WT FVB/N mice appeared to be smaller and much paler in colour (Fig. 2A) than worms found in the intestines of WT CBA/Ca mice (Fig. 2B). To test for differences in worm length for hosts from each of the WT CBA/Ca and FVB/N strains on day 5, 6 and 7 p.i., a linear mixed effects model was used. In the model, strain, day and the interaction between strain and day were fitted as fixed effects. A random mouse ID effect was also fitted in the model to account for dependence in repeated observations for individual worms from the same mouse. There were 228 observations in total and the linear mixed effects model used 24 degrees of freedom to estimate the fixed and random effects. The results of this test showed that the interaction effect between strain and day was not statistically significant. Thus there is no evidence to suggest that the difference in average worm length between the strains varied according to the day of measurement. In order to accurately interpret the main effects of strain and day, the linear mixed effects model was rerun but the interaction term was excluded. The results of the model showed a significant difference in worm lengths between the strains and over time. The model estimates of the mean worm length for each strain and for each day were then calculated. These estimates were adjusted for dependence within each mouse. Worm lengths were higher in the CBA/Ca strain and increased over the successive days. The increase in mean worm length from WT FVB/N hosts over 3 days was modest (12%), relative to that seen in worms recovered from WT CBA/Ca mice over the same period (23%).

The numbers of leukocytes in the small intestine were examined on days 6 and 7 p.i. Eosinophil numbers in the intestines of WT CBA/Ca and WT FVB/N mice both increased from day 6 to day 7, but there were no significant differences in the number of eosinophils between the 2 strains (data not shown). There were no other overt differences in the nature of the cellular inflammatory responses seen in the gut in the two strains.

Lung larval recovery, cellular response and migration to the small intestine

Under normal conditions, *N. brasiliensis* larvae migrate from the skin to the lungs and then to the small intestine. Since larval numbers and maturation were impaired at the intestinal phase of the infection in WT FVB/N mice, we next quantified the number and the size of larvae in the lungs, and assessed the accompanying cellular response. Lungs were removed from mice 1 and 2 days after injection of 750 L3 and lung-stage larvae were enumerated. We have previously found that in naïve IL-5 Tg CBA/Ca mice, the majority of larvae are trapped in the skin and pre-lung phase (Daly *et al.* 1999). WT CBA/Ca and WT FVB/N mice had higher lung larval burdens than the IL-5 Tg CBA/Ca mice, with values reaching statistical significance on day 2 p.i. (Fig. 3A). Interestingly, the numbers of lung larvae found in WT CBA/Ca and FVB/N mice were comparable on each of days 1 and 2 (Fig. 3A). However, on day 2 p.i., larvae recovered from WT FVB/N animals were less motile than larvae recovered from the lungs of WT CBA/Ca mice (mean \pm s.e.m.: 0.57 \pm 0.23 and 2.11 \pm 0.47 movements/30 sec respectively, $n=30$ larvae/group). The lengths of L4 were calculated to determine if impaired larval

development or damage in the pre-lung or lung phases of infection in WT FVB/N mice might restrict migration to the intestines. On days 1 and 2 p.i., the mean lengths of lung larvae recovered from WT FVB/N mice were significantly less than for larvae recovered from WT CBA/Ca mice on the same day (Fig. 3B) and on day 2, comparable to those few recovered from IL-5 Tg CBA/Ca mice. Larvae from WT FVB/N mice increased in size from day 1 to day 2 p.i. but growth was more substantial in parasites recovered from WT CBA/Ca mice (11% vs 29% increase in mean length respectively). This suggests that larvae migrating through WT FVB/N hosts may be damaged or suffer from impaired development prior to or upon arrival in the lungs. Histological analysis of parasite-infected lungs revealed no major inflammatory response in either strain of mice (Fig. 3C and D). Leukocytes were not present in high numbers in the tissue or lumen of the lungs and were not attached to and were rarely seen in close proximity to larvae. This is typical of the early lung inflammatory response in other strains of mice (Knott *et al.* 2007).

Since larvae or worms were not detected in the intestines of WT FVB/N mice in large numbers, the movement of parasites from the lungs to the intestine was assessed to determine if larvae were completing this phase of the migration. We enumerated larval or worm numbers in the lungs, trachea, oesophagus, stomach, small intestine and colon over days 2–6 p.i. (Table 1). As expected, larval numbers in the lungs were comparable between WT CBA/Ca and WT FVB/N mice on day 2 p.i. and numbers declined rapidly thereafter. Larvae were not found in high numbers when sampling the trachea, oesophagus or stomach in either strain (mean <10 larvae/site), but the detection of larvae from days 2–3 p.i. in most of these tissues suggests that larvae were migrating via this route from the lungs to the small intestine. On almost all days sampled, 2–3 times more larvae or worms were found in the small intestines of WT CBA/Ca mice than in WT FVB/N animals. The lower numbers of worms in WT FVB/N animals suggests a failure to colonize this compartment. No worms were seen in the colon in either strain on days 2–4, and only a small number (mean <10 worms/mouse) were found in the colon in WT CBA/Ca mice on days 5 and 6, suggesting that *N. brasiliensis* larvae and worms were rapidly lost from the entire length of the small intestine in WT FVB/N mice.

Larval retention and leukocyte recruitment in skin air pouches

In IL-5 Tg mice, low intestinal worm loads and poor fecundity may, in large part, be attributed to innate immune responses in the skin that are initiated within an hour of infection (Daly *et al.* 1999; Giacomini *et al.* 2008). To determine if this also applied in WT FVB/N mice, the inflammatory response to larvae injected into dorsal skin air pouches was assessed in WT FVB/N, WT CBA/Ca mice and IL-5 Tg CBA/Ca animals 0 and 4 h after infection. As expected, the total number of cells recovered from each group at 0 h was minimal and, as is typically the case, approximately 100% of larvae were recovered (data not shown). Larval retention in IL-5 Tg CBA/Ca mice at 4 h p.i. (Fig. 4A) was significantly higher than in the WT FVB/N group. There was no significant difference in the number of larvae recovered from air pouches in WT CBA/Ca and WT FVB/N mice. The strength and the composition of the cellular immune response were comparable in the two WT groups and significantly more cells were recovered from infected IL-5 Tg CBA/Ca mice (Fig. 4B). By 4 h p.i. neutrophils (Fig. 4C) were the dominant leukocyte subset, though eosinophils (Fig. 4D) were also present in very large numbers in IL-5 Tg mice. Lymphocytes and macrophages were not numerous in any strain (data not shown). These data suggest that impaired development of larvae and worms that can be detected in the lungs and gut in WT FVB/N mice is not due to obvious quantitative or qualitative enhancement of the cellular inflammatory response induced in the skin soon after infection.

CBA/Ca×FVB/N F1 offspring

It would appear that in WT FVB/N hosts, larvae were either being damaged prior to the intestinal phase of infection, resulting in impaired development, migration or colonization, or that factors required for these processes were not sufficient. F1 hybrids of WT CBA/Ca mice and WT FVB/N mice were infected to determine if components required for larval development that were lacking in FVB/N mice could be restored by introducing the susceptible CBA/Ca genotype. F1 mice showed the same pattern of infection as the parental WT FVB/N strain (Fig. 5). Lung larval numbers in the F1 crosses were similar to both parental strains (Fig. 5A). Lung larvae recovered from both the parental WT FVB/N strain and the F1 mice were significantly smaller in length, and were less motile than those recovered from parental WT CBA/Ca mice (data not shown). Relative to infections in parental WT CBA/Ca mice, significantly fewer eggs (Fig. 5B) and worms (Fig. 5C) were found in both the parental FVB/N and F1 mice.

Primary versus secondary infections

To determine if secondary immune responses to *N. brasiliensis* were enhanced in WT FVB/N mice compared with WT CBA/Ca mice, L3 were injected into animals 3 weeks after a primary infection and lung larval burdens were determined on day 2 of the challenge. The numbers of larvae recovered from the lungs after secondary infection of WT FVB/N mice were significantly lower than those recovered during primary infection (Fig. 6). The numbers of larvae recovered from the lungs of WT FVB/N and WT CBA/Ca mice during the secondary infection were not significantly different.

Resistance to *N. brasiliensis* in *i-FABP-IL-5* and *i-FABP-eotaxin Tg* mice

To investigate the role of eosinophils in the small intestines during infections with *N. brasiliensis*, FVB/N mice with intestine-specific expression of either IL-5 or eotaxin transgenes were used. Faecal egg (Fig. 7A) and intestinal worm (Fig. 7B) burdens were very low in primary infections of both of these Tg lines, but not significantly different to those recorded for WT FVB/N mice. Thus, expression of intestine-specific IL-5 or eotaxin transgenes, at least on this genetic background, did not provide additional resistance to *N. brasiliensis*.

H. bakeri infections in FVB/N mice

Since WT FVB/N mice were highly resistant to *N. brasiliensis*, these animals were assessed for resistance to primary infection with *H. bakeri*, another intestinal nematode. In WT CBA/Ca and WT FVB/N mice there were no significant differences in parasite egg production over days 13–116 p.i. (Fig. 8A) or worm numbers on days 41 and 117 p.i. (Fig. 8B). This suggests that the mechanisms that protect WT FVB/N mice against tissue-penetrating *N. brasiliensis* do not also provide effective resistance against a long-term parasitic nematode that is restricted to the gut.

DISCUSSION

When available in sufficient numbers, eosinophils are capable of providing potent resistance to primary infections with *N. brasiliensis*. IL-5 Tg mice with constitutive eosinophilia are highly resistant to this parasite and the majority of larvae are trapped in the skin, with eosinophils rapidly recruited in large numbers (Daly *et al.* 1999). Eosinophils are also necessary for protection in the early phase of secondary infections with *N. brasiliensis*. IL-5^{-/-} and ΔDdb1GATA (Yu *et al.* 2002) mice, both of which are deficient in eosinophils, have comparable lung larval burdens to those seen in primary infections (Knott *et al.* 2007). In the present study we have determined that WT FVB/N mice are also highly resistant to primary infections with *N. brasiliensis*, but not to *H. bakeri*. As with IL-5 Tg mice infected with *N. brasiliensis*, few worms were found in the small intestines of WT FVB/N mice and egg

production was minimal. While resistance to *N. brasiliensis* in WT FVB/N mice may be initiated in the pre-lung stage of infection as it is in IL-5 Tg mice, inflammation and larval retention in the skin was similar to that in WT CBA/Ca animals and eosinophils were not a particularly prominent subset within the leukocytes recruited. Inhibition of development at this early stage of infection may adversely affect the capacity of larvae to develop and colonize the gut.

IL-5 is important for eosinophil growth and development (Lopez *et al.* 1986; Enokihara *et al.* 1988; Yamaguchi *et al.* 1988a, b; Dent *et al.* 1990). Eotaxin is a chemotactic factor for eosinophils (Jose *et al.* 1994) and under normal conditions is expressed throughout the length of the gastrointestinal tract (Rothenberg *et al.* 1995). It would seem therefore that these factors are likely to be important in intestinal homeostasis and may play a significant role in producing and recruiting eosinophils into the gut during helminth infections. This is seen during infections with *N. brasiliensis*, where eosinophil numbers increase in the gut from days 6–11 in both WT and IL-5 Tg mice (Dent *et al.* 1999). The mechanisms responsible for this tissue-specific recruitment and the roles that eosinophils play in host protection during the gut phase of infection are not well understood, although eosinophilia may assist worm expulsion (Knott *et al.* 2007). Our initial studies were therefore designed to investigate the importance of both IL-5 and eotaxin for protection against *N. brasiliensis* at the level of the small intestine. In recent years, studies of immune responses to parasites have been aided by technology that permits selective over-expression of genes in mice. Pro-nuclear injections are often performed in FVB/N mice because fertilized eggs from this strain have prominent nuclei and this facilitates injection of DNA constructs (Taketo *et al.* 1991). While investigating infections in i-FABP-IL-5 and i-FABP-eotaxin Tg FVB/N mice (Mishra *et al.* 2002), we established that WT FVB/N mice had potent natural resistance to *N. brasiliensis* and the expression of either transgene did not further enhance this resistance to the parasite.

FVB/N mice have most frequently been cited in cancer, behavioural and neurological studies and are known to be resistant to atherosclerosis and collagen-induced arthritis. The immunobiology of FVB/N mice has not been extensively reported, but some minor abnormalities in thymus development have been noted (Nabarra *et al.* 2001). FVB/N mice share the *Hc⁰* allele with strains such as DBA/2, A/J, AKR and SWR and are deficient in the complement protein C5. Mice described as FVB or FVB/N have been compared to other laboratory mouse strains in several parasite studies. FVB mice eliminate *H. bakeri* faster than the C57BL/6 strain (Donskow-Schmelter *et al.* 2007), but FVB/N mice are more susceptible to *Clonorchis sinensis* than BALB/c, severe combined immunodeficiency (SCID) and athymic mice (Yoon *et al.* 2001). BALB/c, C57BL/6 and FVB mice are of comparable susceptibility to the fluke *Echinostoma hortense* and clear infections more rapidly than C3H/HeN and ICR mice (Lee *et al.* 2004).

Although *N. brasiliensis* did successfully migrate to the lungs in WT FVB/N mice, there was evidence of developmental retardation, since the larvae recovered were smaller and less motile than those in the susceptible WT CBA/Ca mice. This was apparent even in lung larvae recovered 24 h post-primary infection and so the damage may have been inflicted in the pre-lung stage. There was also evidence of impaired development in the few larvae that belatedly migrated to the lungs of IL-5 Tg mice on day 2 p.i. However, unlike infections in mice expressing the IL-5 transgene on CBA/Ca (Daly *et al.* 1999 and this study), BALB/c (Knott *et al.* 2007) and C57BL/6 (Giacomin *et al.* 2008) genetic backgrounds, larvae were not trapped in the skin in significant numbers and eosinophils were only a small part of the inflammatory infiltrate recruited into the skin. Should larvae be trapped in the lungs of WT FVB/N mice, a strong cellular inflammatory response might be expected. However, as in other mouse strains (Daly *et al.* 1999; Reece *et al.* 2006; Knott *et al.* 2007), virtually no cellular infiltrate was seen in the lungs or around larvae during the period in which the majority of parasites were present

(1–2 days p.i.), suggesting that if larvae are being damaged at this site, cellular inflammatory mechanisms are not likely to be responsible. It is also important to note that relatively few larvae were actually trapped in the lungs of any of the mice used. While the majority of larvae have left the site of injection by 2–4 h p.i., few appear in the lungs before 16 h p.i. (Daly *et al.* 1999) and during the intervening period, the parasite is essentially invisible. It has long been assumed that during this time larvae are migrating via the lymphatics and/or blood vessels. However, given the likely significance of a pre-lung stage for innate resistance, it will be important to identify more accurately where the larvae are located during this period. In this way cellular and/or soluble resistance mechanisms are more likely to be identified.

Our investigations of the gut phase of the infection revealed that the few worms that were present in the small intestines of WT FVB/N mice were much paler in colour and significantly smaller in size than those recovered from WT CBA/Ca mice. It is feasible that damage acquired by *N. brasiliensis* larvae during migration from the skin, may be largely responsible for the ultimate failure of the parasite to feed and mature and to colonize the gut. Resistance mechanisms initiated in the pre-gut phase of infection may also be replicated and expanded upon in the gut. However, FVB/N mice are as susceptible as CBA/Ca mice to the nematode *H. bakeri*, which resides primarily in the gut. *H. bakeri* either suppresses or is resistant to the factors that protect FVB/N mice against *N. brasiliensis* or it is not exposed to them because it does not have a tissue migratory phase. Since *H. bakeri* is known to be immunosuppressive (Pritchard *et al.* 1984; Pritchard and Behnke, 1985; Crawford *et al.* 1989), it would be useful to explore the resistance of FVB/N mice to other intestinal parasites such as *Trichinella spiralis* and *Trichuris muris*, to determine if these parasites can be damaged in the gut in this host strain.

Eosinophils play a protective role in the gut in infections with *Strongyloides venezuelensis* (El-Malky *et al.* 2003) and *N. brasiliensis* (Knott *et al.* 2007), but as in the skin, these and other leukocytes were found in similar numbers in the intestines of infected WT FVB/N and WT CBA/Ca mice. We have yet to do an extensive analysis of leukocyte subsets in either the skin or gut in FVB/N mice during infection, but any differences to those in WT CBA/Ca mice were not overt. Although over-expression of IL-5 and eotaxin in the gut did not confer additional protection against *N. brasiliensis* on the naturally resistant FVB/N genetic background, it will be important to test the effect of these transgenes again on a more permissive strain. This will help to address the contribution that eosinophils can make to elimination of *N. brasiliensis* and other intestinal helminths from the gut.

The major histocompatibility complex (MHC) appears to be implicated in differential patterns of resistance to a number of parasitic helminths of sheep, cattle and mice (Else and Wakelin, 1989; Stear *et al.* 1990; Wassom and Kelly, 1990; Behnke and Wahid, 1991; Outteridge *et al.* 1996; Su and Dobson, 1997). However, as the resistance seen in WT FVB/N mice occurs within the first 2–3 days of primary infection, it is most likely due to innate rather than adaptive immune mechanisms and would therefore be less dependent on allelic differences in the MHC.

Many factors could contribute to resistance to tissue-invasive intestinal helminthic parasites. Although FVB/N mice are deficient in C5 and we have found complement and the C5a receptor to be important in WT and IL-5 Tg C57BL/6 mice in the first 2–3 h of infection with *N. brasiliensis*, complement deficiency does not ultimately cause major changes in lung or intestinal parasite burdens (Giacomin *et al.* 2008). Elements that influence expulsion of *N. brasiliensis* include STAT6, the IL-4 Receptor (R) α and IL-13 (Urban *et al.* 1998, 2001; Zhao *et al.* 2003). Similarly, the IL-4R α and STAT6 are required to expel adult *T. spiralis* worms (Urban *et al.* 2000).

Amphiregulin, a member of the epidermal growth factor family that is expressed by activated Th2 cells, is required for rapid expulsion of *T. muris* (Zaiss *et al.* 2006). Resistin-like molecules (RELM) are upregulated in inflammatory cells, including macrophages (Loke *et al.* 2002), and in the lungs and gut during *T. muris*, *N. brasiliensis* and *S. stercoralis* infections (Artis *et al.* 2004; Voehringer *et al.* 2004; Wang *et al.* 2005). RELM may contribute to both the pre-intestinal and intestinal phases of resistance to *N. brasiliensis* in WT FVB/N mice, since RELM- β binds to *T. muris* and may aid in expulsion of the parasite from the gut (Artis *et al.* 2004). Both RELM- α and - β may affect gastrointestinal function through actions on nerve and goblet cells (Artis *et al.* 2004; Wang *et al.* 2005; Nair *et al.* 2006). 129/SvJ mice are naturally resistant to *S. venezuelensis* and it is thought that higher concentrations of chondroitin sulphate in the intestine of infected mice may be responsible for this (Nakamura-Uchiyama *et al.* 2001). Relative to C57BL/6J mice, the CBA/Ca strain is highly resistant to primary infections with *Ascaris suum* and it is proposed that the parasites are most susceptible to damage during migration from the liver to the lungs (Lewis *et al.* 2006, 2007). The mechanisms responsible for this resistance in CBA/Ca mice have not yet been determined, but since the effect seems to be on migration within the first few days of infection, innate factors are likely to make major contributions.

These examples clearly demonstrate that the ways in which different parasites are eliminated from the host vary and are not necessarily directly associated with immunological mechanisms. Factors or pathways responsible for the resistance of WT FVB/N mice to *N. brasiliensis* may not be immunological in nature or at least not dependent on a cellular inflammatory response. Products of intestinal epithelial cells or other soluble factors found in the lungs and/or gut may play a crucial role. It is also possible that FVB/N mice lack essential nutrient/s required by *N. brasiliensis* larvae for development and colonization, but this seems less likely, given our observation that CBA/Ca \times FVB/N F1 mice are also resistant. Our data lead us to propose that resistance factors may be differentially expressed in the pre-lung phase of infection in WT FVB/N mice.

Although WT FVB/N mice are highly resistant to primary infections with *N. brasiliensis*, in our hands this strain proved to be quite susceptible to *H. bakeri* and this may be due to differences in the nature and duration of the life cycles of these parasites. In most mouse strains, *H. bakeri* resides in the gut for months, rather than just the few days seen in primary infections with *N. brasiliensis*. Variation in susceptibility to *H. bakeri* in different strains of mice has been well documented and candidate factors responsible for resistance have been identified. Mouse strains that effectively resist *H. bakeri* in a repetitive infection protocol include SWR, NIH and SJL. In contrast, BALB/c, CBA, A/J, C3H and C57BL/6 strains are susceptible to the parasite under the same regime and carry heavy worm burdens for months (Behnke *et al.* 2006b). Mice described as FVB were reported to eliminate *H. bakeri* by day 74 p.i. (Donskow-Schmelter *et al.* 2007), an outcome that is not consistent with our study, which was performed in WT C57BL/6, BALB/c mice (data not presented) as well as the CBA/Ca and FVB/N strains. We challenged our mice with approximately 70% more larvae than Donskow-Schmelter and colleagues and our FVB/N mice may prove to be a different substrain from the FVB animals used by this group (Cywinska *et al.* 2004; Donskow-Schmelter *et al.* 2007).

Resistance, particularly in SWR mice, is associated with increases in serum tumour necrosis factor- α , intestinal goblet cells, mucosal mast cells and mucosal mast cell protease-1 levels (Behnke *et al.* 2003b). The killing of *H. bakeri* adult worms is dependent on the production of oxygen radicals such as hydrogen peroxide and acetaldehyde/xanthine oxidase (Ben-Smith *et al.* 2002). Furthermore, both Th1 and Th2 responses are strong in the resistant SWR and SJL strains during primary *H. bakeri* infections (Ben-Smith *et al.* 2003). Quantitative trait linkage mapping techniques have been used to extend these studies, with the identification of candidate resistance genes involved in immune regulation on chromosomes 1 and 17 (Behnke *et al.*

2003a; Iraqi *et al.* 2003; Menge *et al.* 2003; Behnke *et al.* 2006a). A similar approach may prove useful in identifying the resistance factors expressed by WT FVB/N mice infected with *N. brasiliensis*.

In summary, evidence of resistance to primary infections with *N. brasiliensis* in WT FVB/N mice is first manifested as a deleterious impact on the development and motility of lung larvae. WT FVB/N mice provide an excellent opportunity to explore potent and potentially novel mechanisms of early innate resistance to *N. brasiliensis* and possibly, to other tissue-invasive helminths. Further studies should include close scrutiny of the pre-lung phase of infection in this host.

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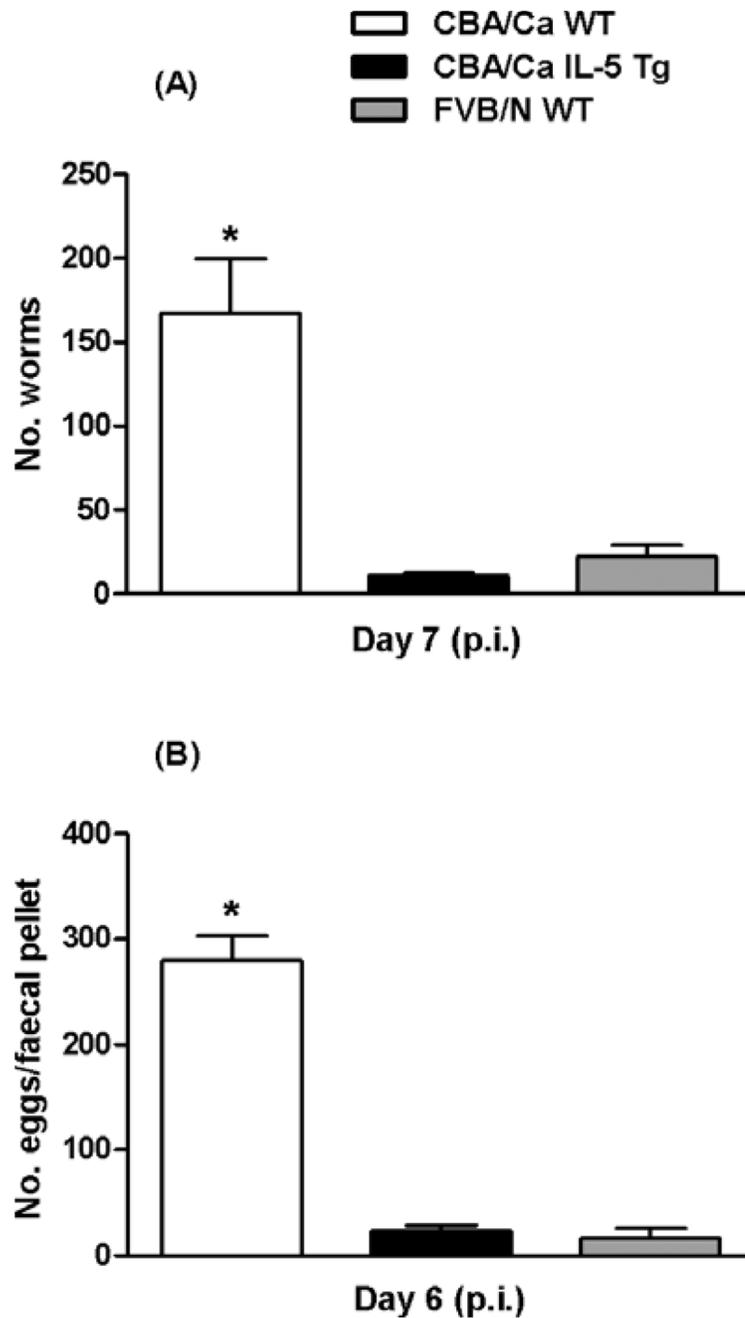


Fig. 1. *Nippostrongylus brasiliensis* intestinal worm burdens on day 7 (A) and faecal egg production on day 6 (B) in WT CBA/Ca, IL-5 Tg CBA/Ca and WT FVB/N mice after infection. 750 L3/mouse. Mean_{±SEM}, $n=4$ mice for all groups. Significant effect between strains (KW test: for (A) $H=8.192$, $P=0.0166$ and (B) $H=7.731$, $P=0.0210$). * Significantly greater than other groups (MW post-hoc tests, $P<0.05$). Data representative of 5 experiments.

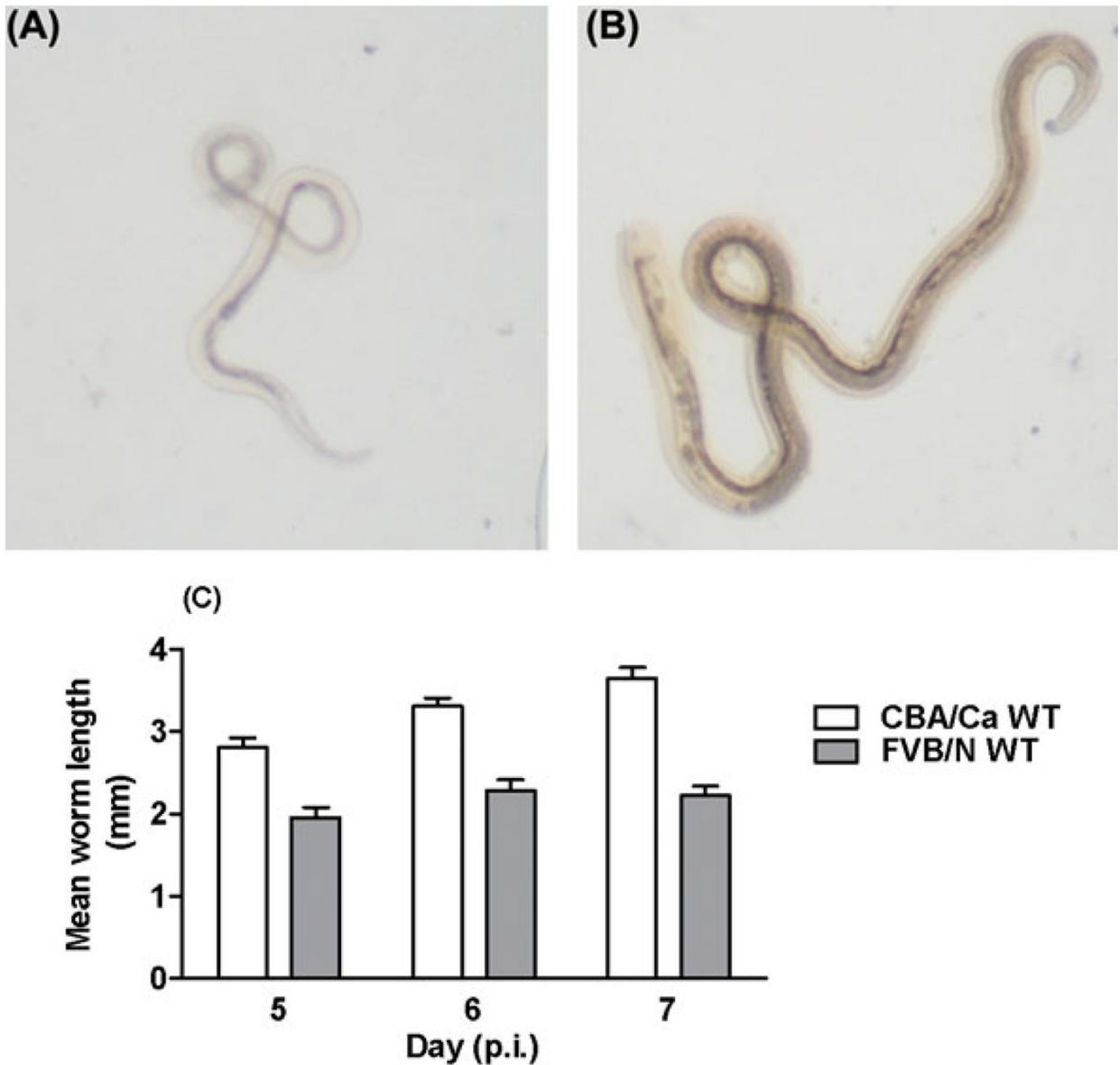


Fig. 2.

Representative photomicrographs (56 \times magnification) of *Nippostrongylus brasiliensis* worms recovered on day 6 p.i. from WT FVB/N mice (A) and WT CBA/Ca mice (B). Intestinal worms recovered 5–7 days p.i. from WT FVB/N mice were paler (A) and smaller (C) than those found in WT CBA/Ca hosts. Worm length (mm) for 4 mice/group/day, with 6–12 worms measured/mouse, $n=32$ –43 worms/group/day (C). A random mouse identity effect was fitted in the model to account for the dependence in repeated observations (worms) from the same mouse. The interaction effect between strain and day was not statistically significant (linear mixed effects model $F_{2, 204}=1.26$, $P=0.2871$). There was a significant difference in lengths of worms recovered from the two host strains and over time (linear mixed effects model; $F_{1, 204}=52.39$, $P<0.0001$ for strain and $F_{2, 204}=5.21$, $P=0.0062$ for day). On average, worm lengths were greater in the CBA/Ca strain (difference in means=1.0929, $P<0.0001$) and increased from day 5 to day 7 ($P=0.0062$). The mean worm length was greater at day 7 than at day 5 (Bonferroni post-hoc test, $t=3.13$, $P=0.0060$). There was no difference in mean worm length between days

5 and 6 and days 6 and 7 (Bonferroni post-hoc test, $t=2.29$, $P=0.0699$ and $t=0.85$, $P=1.1913$ respectively).

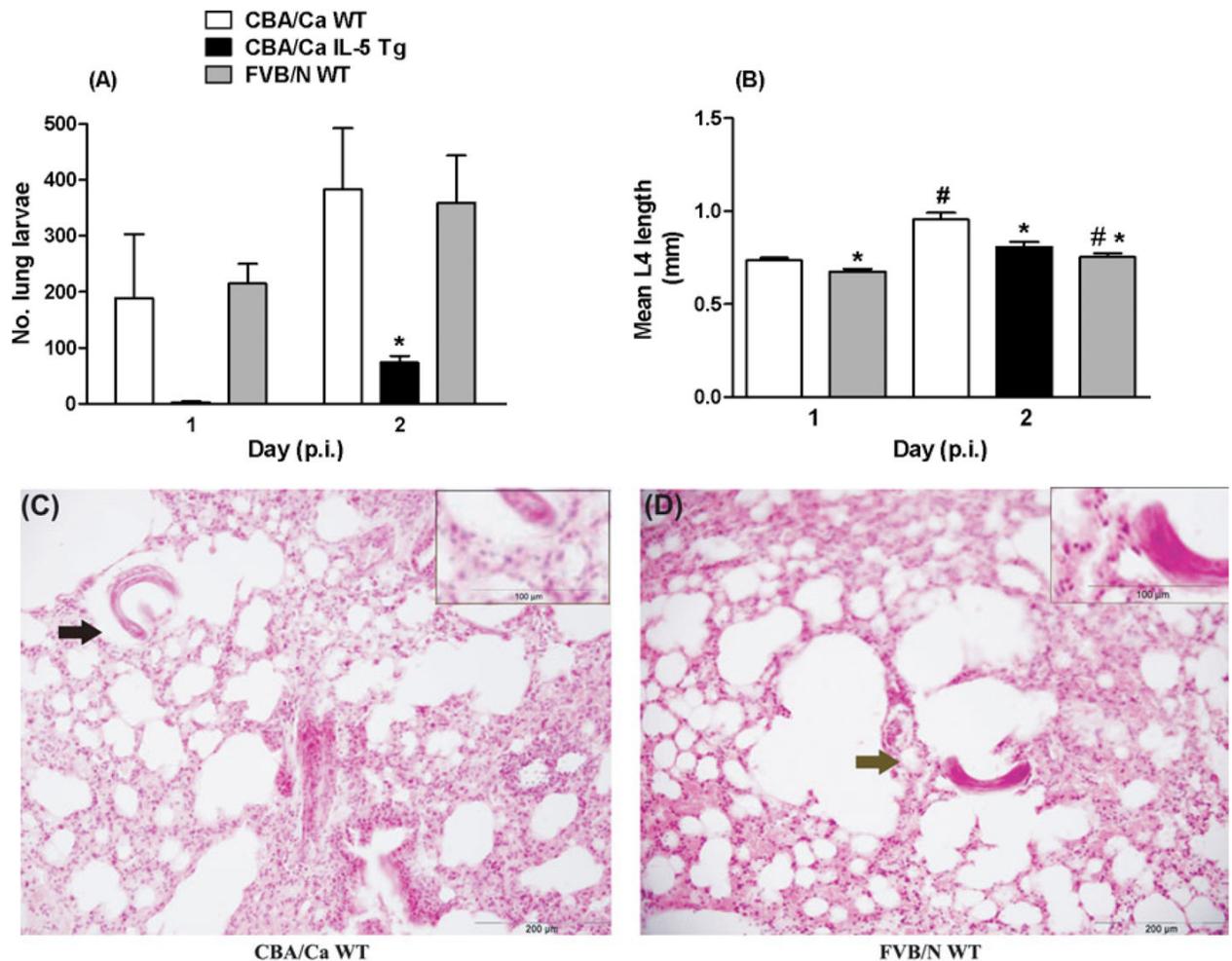


Fig. 3.

Number (A) and length (B) of *Nippostrongylus brasiliensis* larvae recovered from the lungs of WT CBA/Ca, IL-5 Tg CBA/Ca and WT FVB/N mice 1 and 2 days p.i. and lung histology for WT CBA/Ca (C) and WT FVB/N mice (D) 2 days p.i. 750 L3/mouse. (A) Mean number of lung larvae recovered_{+S.E.M.}, $n=3-4$ mice/group. There was a significant difference between strains on day 2 p.i. (KW test, $H=7.053$, $P=0.0294$). * Values for larvae from IL-5 Tg CBA/Ca mice were significantly less than those from WT CBA/Ca and FVB/N hosts on day 2 p.i. (MW post-hoc test, $P=0.0286$ and 0.0143 respectively). (B) Mean length (mm) of lung larvae_{+S.E.M.} from 3 mice/group for 10–19 larvae/mouse and $n=34-45$ larvae/group/day. Insufficient larvae recovered from IL-5 Tg mice to provide reliable data on day 1. Day 1: * significantly different from WT CBA/Ca values (MW test, $P=0.0067$). Day 2: significant difference between strains (KW test $H=23.97$, $P<0.0001$), * significantly different from WT CBA/Ca values (MW post-hoc test, $P=0.0013$ for WT CBA/Ca vs IL-5 Tg CBA/Ca; $P<0.0001$ for WT CBA/Ca vs WT FVB/N). (C and D) Lung sections stained with H&E, 150× magnification, inset 260× magnification and scale bars represent 200 μm and 100 μm respectively. *, lung larvae. ➡, area shown in inset.

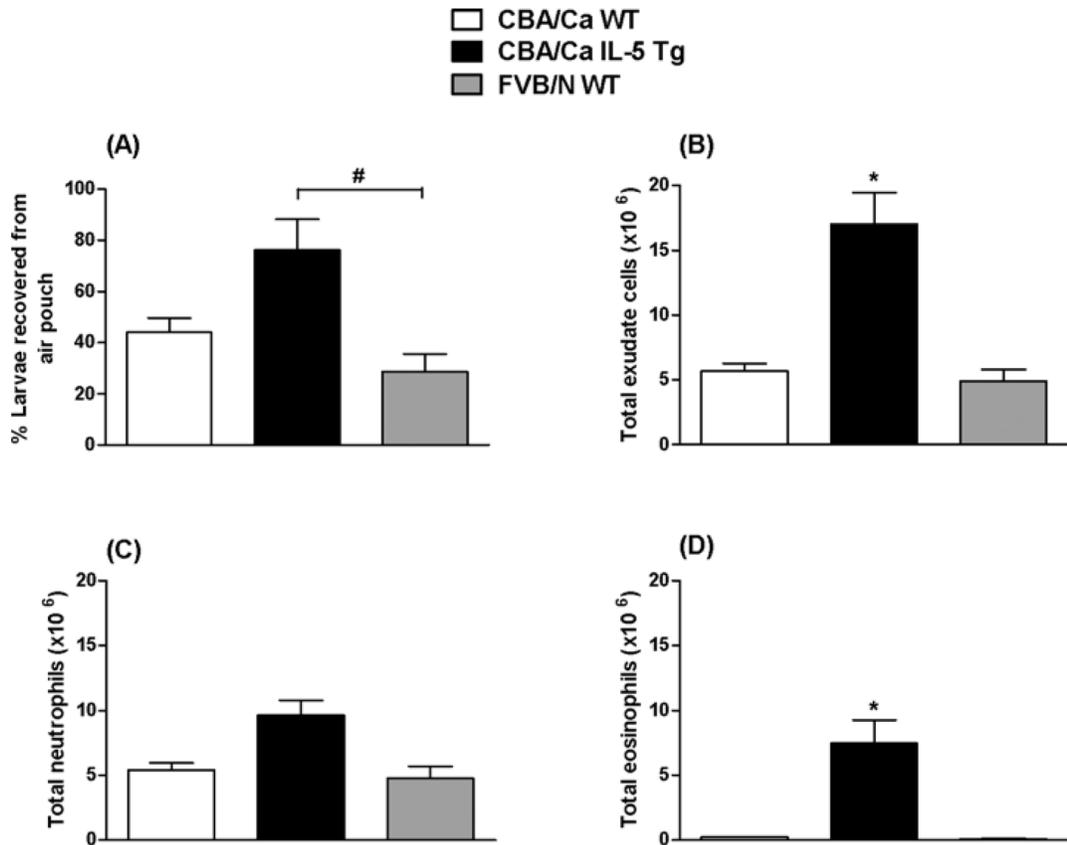


Fig. 4. Larvae (A), total leukocytes (B), neutrophils (C) and eosinophils (D) recovered by lavage of air pouches in WT CBA/Ca, IL-5 Tg CBA/Ca and WT FVB/N mice 4 h after injection of 500 *Nippostrongylus brasiliensis* L3/mouse. Data in A represent larvae recovered as percentage of inoculum_{+S.E.M.} Data in B, C and D are mean number of leukocytes (10⁶)_{+S.E.M.}, n=4 mice for all groups. Significant effect between strains for A, B and D (KW test (A) H=6.615, P=0.0366, (B) H=7.538, P=0.0231, (D) H=9.846, P=0.0073). No significant effect between strains in C (KW test H=5.808, P=0.0548). * Significantly greater than WT CBA/Ca and FVB/N values (MW post-hoc test, P<0.05). # Significantly greater than WT FVB/N value (MW post-hoc test, P<0.05). Representative of 3 experiments.

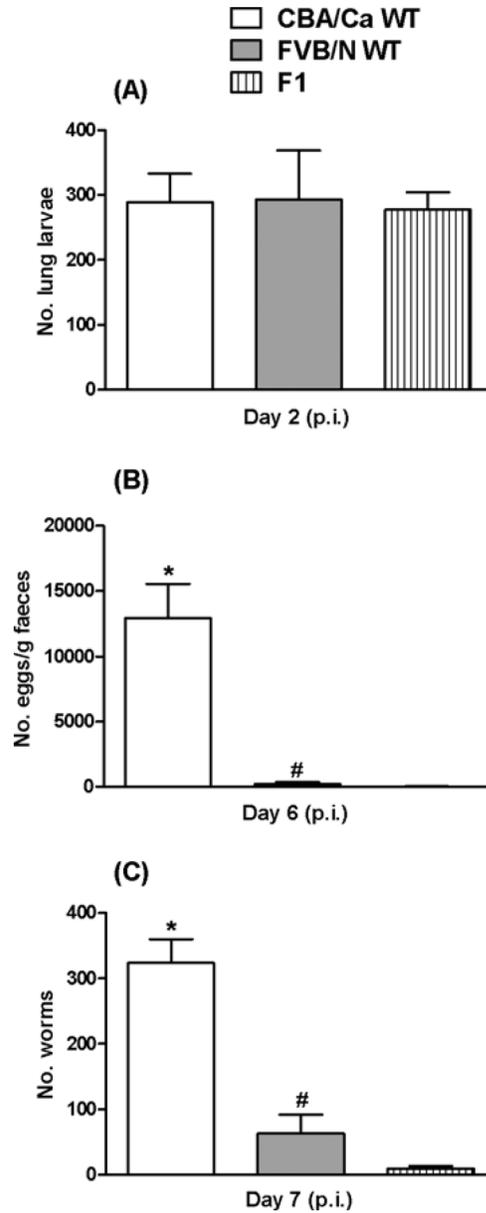


Fig. 5. Recovery of *Nippostrongylus brasiliensis* larvae from the lungs (A) of WT CBA/Ca, WT FVB/N and WT FVB/N \times CBA/Ca F1 hybrid mice 2 days p.i. Faecal eggs (B) and intestinal worms (C) for day 6 and 7 p.i. respectively. 750 L3/mouse. Data represent mean number of larvae, eggs/g faeces or worms recovered \pm S.E.M., $n=4-6$ mice/group. Significant effect between strains in B and C (KW test (B) $H=12.60$, $P=0.0018$ (C) $H=12.38$, $P=0.0021$). * Significantly greater than WT FVB/N and F1 mice (MW post-hoc test, $P<0.05$). # Significantly greater than F1 mice (MW post-hoc test, $P<0.05$). Representative of 2 experiments.

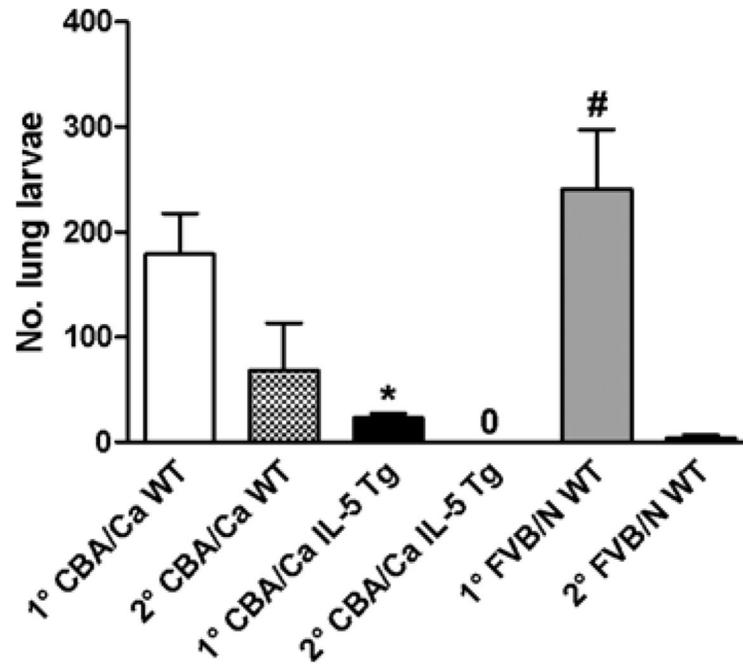


Fig. 6.

Recovery of *Nippostrongylus brasiliensis* lung larvae during primary (1°) and secondary (2°) infections of WT CBA/Ca, IL-5 Tg CBA/Ca and WT FVB/N mice on day 2 p.i. 750 L3/mouse for 1° and 2° infections. 2° challenges were delivered 3 weeks after the 1° infection. Mean number of larvae recovered/mouse^{±s.e.m.}, $n=3-4$ mice/group. 0=no lung larvae detected. The interaction between strain and infection (1° or 2°) was significant (2-way ANOVA, $F_{2,16}=3.98$, $P=0.0397$), indicating that the difference in L4 numbers between the strains depended on infection (1° or 2°). * 1° IL-5 Tg CBA/Ca values significantly less than in 1° infections of WT CBA/Ca and FVB/N groups (Bonferroni post-hoc test, WT CBA/Ca vs IL-5 Tg CBA/Ca $t=2.852$, $P<0.05$; WT FVB/N vs IL-5 Tg CBA/Ca $t=3.99$, $P<0.01$). # 1° WT FVB/N values significantly greater than 2° infection of the same genotype (Bonferroni post-hoc test, $t=4.688$, $P<0.001$).

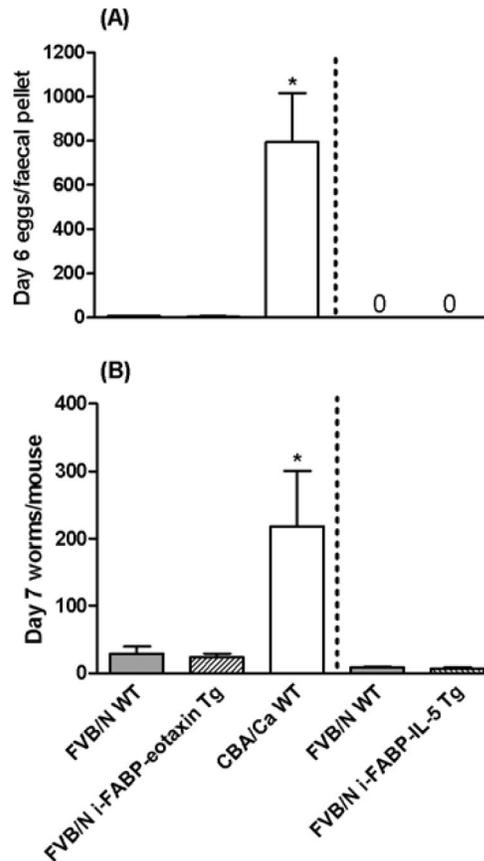


Fig. 7.

Egg (A) and intestinal worm (B) numbers from WT FVB/N, i-FABP-Eotaxin Tg FVB/N, i-FABP-IL-5 Tg FVB/N and WT CBA/Ca mice. 1000 *Nippostrongylus brasiliensis* L3/mouse. Mean \pm S.E.M. for eggs enumerated on day 6 and worms recovered on day 7 p.i., $n=4-8$ mice/group. Separate experiments delineated by dashed line. 0=no eggs detected. Representative of 2-3 experiments for each of the transgenic lines. Significant effect between groups to the left of the dashed line (KW test (A) $H=9.770$, $P=0.0076$ (B) $H=8.779$, $P=0.0124$). * Significantly greater than other groups in the same experiment (MW post-hoc test, $P<0.05$).

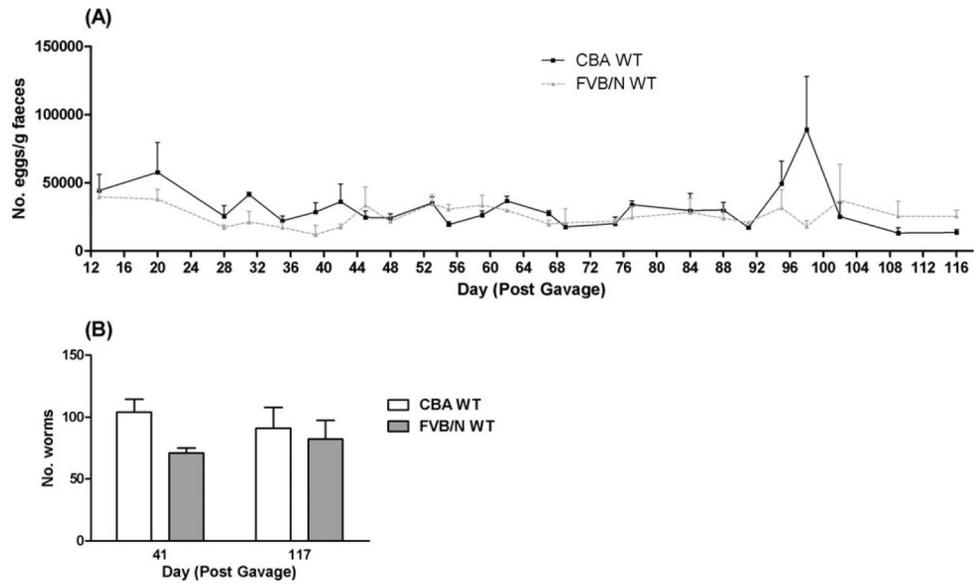


Fig. 8. Egg (A) and intestinal worm (B) numbers for WT CBA/Ca and WT FVB/N mice infected by oral gavage with 200 *Heligmosomoides bakeri* L3. Mean \pm S.E.M., n=3–7 mice/group.

Table 1

Nippostrongylus brasiliensis recovery from infected tissues of CBA/Ca WT and FVB/N WT mice (Larvae and worms were enumerated after s.c. infection with 750 *N. brasiliensis* L3. Mean \pm S.E.M. for 3 mice/group, ND=not done.)

Tissue	Day 2 (p.i.)		Day 3 (p.i.)		Day 4 (p.i.)		Day 5 (p.i.)		Day 6 (p.i.)	
	CBA/Ca WT	FVB/N WT	CBA/Ca WT	FVB/N WT	CBA/Ca WT	FVB/N WT	CBA/Ca WT	FVB/N WT	CBA/Ca WT	FVB/N WT
Lungs	135.3 \pm 14.6	185.0 \pm 58.2	12.3 \pm 5.8	46.3 \pm 13.7	2.3 \pm 0.3	16.0 \pm 4.0	0.7 \pm 0.3	5.3 \pm 4.3	ND	ND
Trachea	9.3 \pm 3.3	6.0 \pm 2.6	0.3 \pm 0.3	1.3 \pm 0.81	0	0	ND	ND	ND	ND
Oesophagus	0.3 \pm 0.3	0.3 \pm 0.3	0	0	0	0	ND	ND	ND	ND
Stomach	0	5.7 \pm 1.7	0.7 \pm 0.7	0.3 \pm 0.3	0	0	ND	ND	ND	ND
Small intestine	23.0 \pm 11.4	41.0 \pm 14.2	191.3 \pm 87.2	86.0 \pm 21.6	221.3 \pm 106.5	63.0 \pm 1.5	119.3 \pm 10.1	43.7 \pm 41.2	102.0 \pm 8.0	57.0 \pm 20.0
Colon	0	0	0	0	0	0	4.67 \pm 4.67	0	6.0 \pm 5.0	0