

Effects of a Protein Preload on Gastric Emptying, Glycemia, and Gut Hormones After a Carbohydrate Meal in Diet-Controlled Type 2 Diabetes

JING MA, MBBS^{1,2}

JULIE E. STEVENS, BPHARM, BSC^{1,2}

KIMBERLY CUKIER, MBBS³

ANNE F. MADDOX, ASS DIP RAD TECH^{1,2}

JUDITH M. WISHART, BSC^{1,2}

KAREN L. JONES, PHD^{1,2}

PETER M. CLIFTON, MBBS, PHD^{3,4}

MICHAEL HOROWITZ, MBBS, PHD^{1,2,3}

CHRISTOPHER K. RAYNER, MBBS, PHD^{1,2}

OBJECTIVE — We evaluated whether a whey preload could slow gastric emptying, stimulate incretin hormones, and attenuate postprandial glycemia in type 2 diabetes.

RESEARCH DESIGN AND METHODS — Eight type 2 diabetic patients ingested 350 ml beef soup 30 min before a potato meal; 55 g whey was added to either the soup (whey preload) or potato (whey in meal) or no whey was given.

RESULTS — Gastric emptying was slowest after the whey preload ($P < 0.0005$). The incremental area under the blood glucose curve was less after the whey preload and whey in meal than after no whey ($P < 0.005$). Plasma glucose-dependent insulinotropic polypeptide, insulin, and cholecystokinin concentrations were higher on both whey days than after no whey, whereas glucagon-like peptide 1 was greatest after the whey preload ($P < 0.05$).

CONCLUSIONS — Whey protein consumed before a carbohydrate meal can stimulate insulin and incretin hormone secretion and slow gastric emptying, leading to marked reduction in postprandial glycemia in type 2 diabetes.

Diabetes Care 32:1600–1602, 2009

The rate of gastric emptying and the incretin, glucagon-like peptide 1 (GLP-1) and glucose-dependent insulinotropic polypeptide (GIP), response to a meal are known to be major determinants of postprandial blood glucose excursions (1,2). One strategy to minimize postprandial glycemia could be to administer a small load of protein or fat before a meal, so that the presence of nutrients in the small intestine induces the release of peptides such as GLP-1, GIP, and cholecystokinin (CCK) to slow gastric emptying and stimulate insulin secretion in advance of the main nutrient load (3,4).

We hypothesized that a protein preload would reduce the postprandial glycemic excursion in type 2 diabetic patients by these mechanisms.

RESEARCH DESIGN AND METHODS

The protocol included eight diet-controlled type 2 diabetic patients (seven male, mean \pm SE age 58 ± 3 years, BMI 28.6 ± 1.3 kg/m², duration of known diabetes 5.4 ± 1.1 years, and A1C $6.5 \pm 0.2\%$) who attended the laboratory after an overnight fast (14 h for solids and 12 h for liquids) on three separate occasions. Each patient con-

sumed beef-flavored soup (3.8 g noncaloric beef flavoring dissolved in 350 ml water) 30 min before a mashed potato meal containing 65 g powdered potato (Deb Instant Mashed Potato, Epping, Australia) with 20 g glucose (total: 59.1 g carbohydrate, 4.3 g fat, 5.2 g protein; 1,276.5 kJ), labeled with 20 MBq ^{99m}Tc-sulfur colloid (4). On one day, 55 g whey protein (876.7 kJ) was added to the soup. On another day, 55 g whey was mixed into the potato meal. On a third day, neither the preload nor the meal contained whey. Blood was sampled frequently for blood glucose and plasma hormone measurements.

Measurements

Gastric emptying was assessed by scintigraphy. Data were corrected for radioisotope decay, subject movement, and γ -ray attenuation, and the gastric half-emptying time (T50) was calculated (4).

Blood glucose concentrations were measured using a glucometer (Medisense Precision QID; Abbott Laboratories, Bedford, MA), which we have validated against the hexokinase technique (5). Plasma insulin was measured by enzyme-linked immunosorbent assay (ELISA; Diagnostic Systems Laboratories, Webster, TX). Total GLP-1 (GLPIT-36HK; Linco Research, St. Charles, MO), total GIP, and CCK-8 were measured by radioimmunoassay (6).

Cardiovascular autonomic function was assessed by the variation in R-R interval during deep breathing and the systolic blood pressure changes in response to standing (7).

Data were evaluated using repeated-measures ANOVA with treatment and time as factors (StatView 5.0; Abacus Concepts, Berkeley, CA). Data are shown as means \pm SE; $P < 0.05$ was considered significant.

RESULTS — Two of the eight subjects had definite autonomic dysfunction. The study was well tolerated.

From the ¹Discipline of Medicine, University of Adelaide, Royal Adelaide Hospital, Adelaide, Australia; the ²Centre of Clinical Research Excellence in Nutritional Physiology, Interventions and Outcomes, University of Adelaide, Adelaide, Australia; the ³Endocrine & Metabolic Unit, Royal Adelaide Hospital, Adelaide, Australia; and the ⁴Human Nutrition, Commonwealth Scientific and Industrial Research Organization, Adelaide, Australia.

Corresponding author: Christopher K. Rayner, chris.rayner@adelaide.edu.au.

Received 15 April 2009 and accepted 4 June 2009.

Published ahead of print at <http://care.diabetesjournals.org> on 18 June 2009. DOI: 10.2337/dc09-0723.

© 2009 by the American Diabetes Association. Readers may use this article as long as the work is properly cited, the use is educational and not for profit, and the work is not altered. See <http://creativecommons.org/licenses/by-nc-nd/3.0/> for details.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked "advertisement" in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

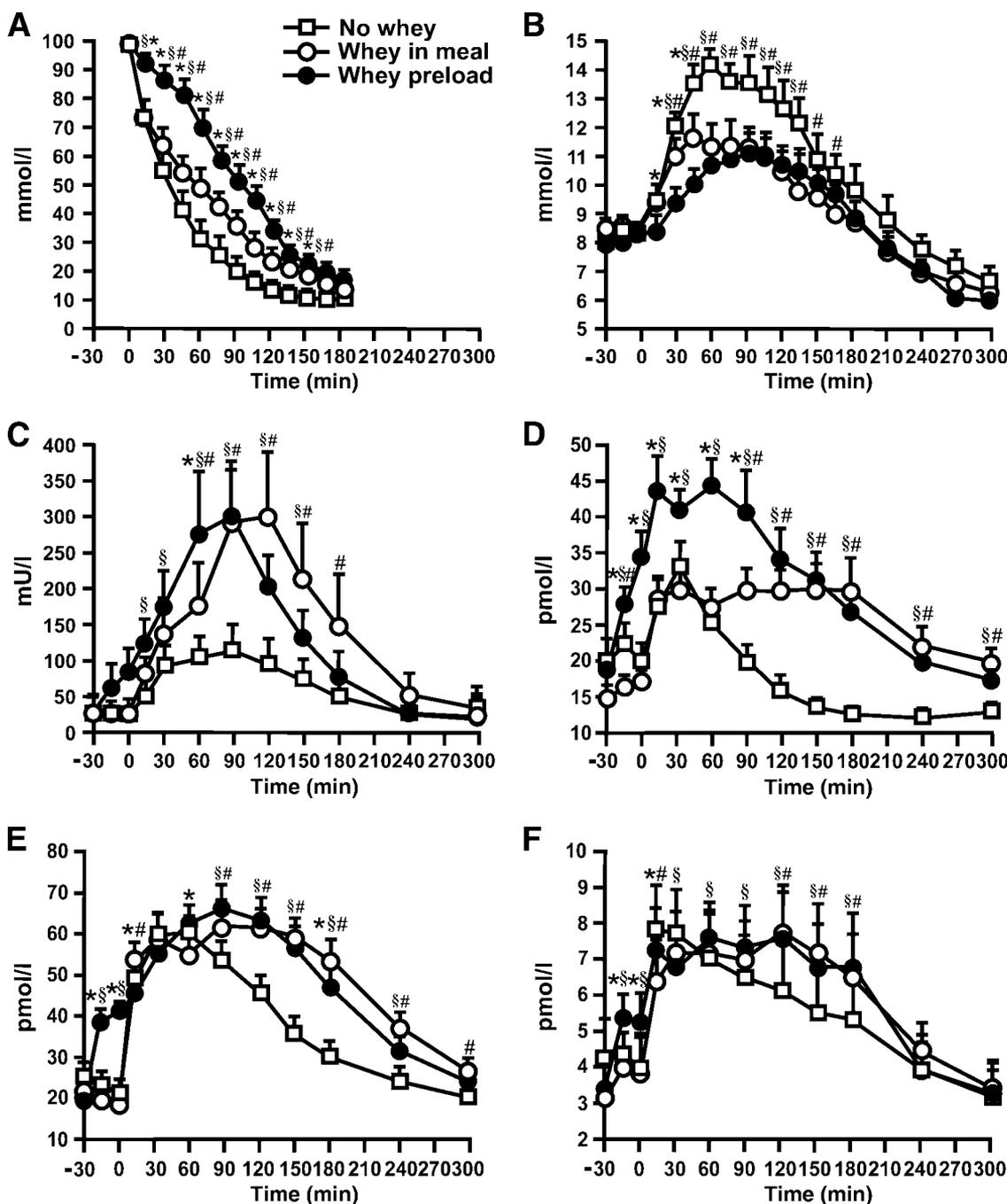


Figure 1—Gastric emptying (A), concentrations of blood glucose (B), plasma insulin (C), plasma GLP-1 (D), plasma GIP (E), and plasma CCK (F) in response to a mashed potato meal in eight type 2 diabetic patients. On each study day, subjects ingested 350 ml beef-flavored soup 30 min before a radiolabeled mashed potato meal; 55 g whey protein was added either to the soup (whey preload) or to the potato (whey in meal) or no whey was given (no whey). Data are means \pm SE. * $P < 0.05$, whey preload vs. whey in meal; # $P < 0.05$, whey in meal vs. no whey; \$ $P < 0.05$, whey preload vs. no whey.

On the no whey and whey in meal days, emptying was rapid initially and subsequently slower, whereas emptying after the whey preload approximated a linear pattern. Gastric emptying was slowest on the whey preload day (T50: 87.3 ± 5.4 min; $P = 0.0001$) and was slower with whey in the meal (53.0 ± 8.3

min; $P < 0.01$) than with no whey (39.0 ± 6.2 min).

There were no differences in baseline blood glucose, plasma insulin, GLP-1, GIP, or CCK concentrations (Fig. 1). The incremental area under the curve (iAUC) for blood glucose was less after the whey preload (363.7 ± 64.5 mmol \cdot min $^{-1} \cdot$

l $^{-1}$) and whey in meal (406.3 ± 85.9 mmol \cdot min $^{-1} \cdot$ l $^{-1}$) compared with no whey (734.9 ± 98.9 mmol \cdot min $^{-1} \cdot$ l $^{-1}$; $P < 0.005$ for both). The iAUCs for insulin, GLP-1, GIP, and CCK were greater when whey was given as a preload ($P < 0.05$ for all) or in the meal ($P < 0.005$ for all) compared with no whey. Despite an

earlier response, the iAUC for insulin did not differ between whey preload and whey in meal ($P = 0.50$). GLP-1 was greater between -15 min and 90 min with the whey preload compared with whey in meal ($P = 0.0001$), but the overall iAUC did not differ significantly.

CONCLUSIONS— We demonstrated that whey protein, when given before or with a high-carbohydrate meal, resulted in a substantial reduction in postprandial glycemia in diet-controlled type 2 diabetic patients. Given that the magnitude of the reduction was comparable with what would be hoped for using pharmacological therapy, such as sulfonylureas, these data have considerable implications for nutritional strategies in the management of diabetes.

The pivotal role of the gastrointestinal tract in determining postprandial glycemia has often been overlooked, but it is assuming increasing prominence, partly because of the development of gut peptide-based therapies for diabetes, such as the GLP-1 analog exenatide (8) and the amylin analog pramlintide (9), which may act predominantly by slowing gastric emptying. Similar to what we reported after an oil preload (4), whey slowed gastric emptying substantially, in particular when given before the meal, and is associated with the stimulation of GLP-1 and CCK. However, in contrast to the delayed insulin response observed after oil, whey augmented insulin secretion markedly, possibly by a combination of the incretin effect and the direct stimulation of the β -cells by absorbed amino acids (10). It is likely that the stimulation of insulin by whey was responsible for the much greater reduction in glycemia after whey than after oil, given that the effects on gastric emptying were comparable.

Although our study involved a small number of subjects who had well-controlled, predominantly uncomplicated type 2 diabetes, the improvement in postprandial glycemia was marked and highly consistent. Further evaluation is

now required in poorly controlled patients and those taking oral hypoglycemic agents in order to determine whether the acute effects are sustained in the longer term. It would also be important to confirm whether the effects are evident with a smaller load of protein in order to minimize additional energy intake. Although concerns have been raised about hyperinsulinemia as a risk factor for vascular disease (11), it is more likely that it represents a marker for other risk factors (12), and in the UK Prospective Diabetes Study (UKPDS), stimulation of insulin by sulfonylureas was not associated with increased cardiovascular events (13).

The concept of using dietary manipulations to treat type 2 diabetes, based on our knowledge of the contribution of gastric emptying and gut peptides to postprandial glycemic responses, appears to hold much promise.

Acknowledgments— This work was supported by the National Health and Medical Research Council (NHMRC) of Australia. The salary of K.L.J. is also funded by the NHMRC.

No potential conflicts of interest relevant to this article were reported.

Parts of this study were presented in abstract form at Digestive Diseases Week, San Diego, California, 17–22 May 2008. Complete data have been submitted for presentation at the annual meeting of the European Association for the Study of Diabetes, Vienna, Austria, 29 September–2 October 2009.

We thank Murray Goulburn for supply of the whey protein isolate and Jane Bowen for advice about formulating the preloads.

References

1. Chaikomin R, Rayner CK, Jones KL, Horowitz M. Upper gastrointestinal function and glycemic control in diabetes mellitus. *World J Gastroenterol* 2006;12:5611–5621
2. Rayner CK, Samsom M, Jones KL, Horowitz M. Relationships of upper gastrointestinal motor and sensory function with glycemic control. *Diabetes Care* 2001;24:371–381
3. Bowen J, Noakes M, Trenerry C, Clifton PM. Energy intake, ghrelin, and cholecys-

tokinin after different carbohydrate and protein preloads in overweight men. *J Clin Endocrinol Metab* 2006;91:1477–1483

4. Gentilcore D, Chaikomin R, Jones KL, Russo A, Feinle-Bisset C, Wishart JM, Rayner CK, Horowitz M. Effects of fat on gastric emptying of and the glycemic, insulin, and incretin responses to a carbohydrate meal in type 2 diabetes. *J Clin Endocrinol Metab* 2006;91:2062–2067
5. Horowitz M, Edelbroek MA, Wishart JM, Straathof JW. Relationship between oral glucose tolerance and gastric emptying in normal healthy subjects. *Diabetologia* 1993;36:857–862
6. Santangelo A, Peracchi M, Conte D, Fraquelli M, Porrini M. Physical state of meal affects gastric emptying, cholecystokinin release and satiety. *Br J Nutr* 1998;80:521–527
7. Ewing DJ, Clarke BF. Diagnosis and management of diabetic autonomic neuropathy. *Br Med J (Clin Res Ed)* 1982;285:916–918
8. Linnebjerg H, Park S, Kothare PA, Trautmann ME, Mace K, Fineman M, Wilding I, Nauck M, Horowitz M. Effect of exenatide on gastric emptying and relationship to postprandial glycemia in type 2 diabetes. *Regul Pept* 2008;151:123–129
9. Kong MF, King P, Macdonald IA, Stubbs TA, Perkins AC, Blackshaw PE, Moyses C, Tattersall RB. Infusion of pramlintide, a human amylin analogue, delays gastric emptying in men with IDDM. *Diabetologia* 1997;40:82–88
10. Fieseler P, Bridenbaugh S, Nustede R, Martell J, Orskov C, Holst JJ, Nauck MA. Physiological augmentation of amino acid-induced insulin secretion by GIP and GLP-1 but not by CCK-8. *Am J Physiol* 1995;268:E949–E955
11. Reaven GM. Insulin resistance and compensatory hyperinsulinemia: role in hypertension, dyslipidemia, and coronary heart disease. *Am Heart J* 1991;121:1283–1288
12. Wingard DL, Barrett-Connor EL, Ferrara A. Is insulin really a heart disease risk factor? *Diabetes Care* 1995;18:1299–1304
13. UK Prospective Diabetes Study (UKPDS) Group. Intensive blood-glucose control with sulphonylureas or insulin compared with conventional treatment and risk of complications in patients with type 2 diabetes (UKPDS 33). *Lancet* 1998;352:837–853