

NIH Public Access

Author Manuscript

J Biopharm Stat. Author manuscript; available in PMC 2009 September 4.

Published in final edited form as: *J Biopharm Stat*. 2008 ; 18(6): 1063–1083. doi:10.1080/10543400802369004.

A Bayesian Meta-Analysis on Published Sample Mean and Variance Pharmacokinetic Data with Application to Drug-Drug Interaction Prediction

Menggang Yu1,* , **Seongho Kim**1, **Zhiping Wang**1, **Stephen Hall**2, and **Lang Li**1 ¹Division of Biostatistics, Department of Medicine, School of Medicine, Indiana University, Indianapolis, IN, 46032

²Division of Clinical Pharmacology, Department of Medicine, School of Medicine, Indiana University, Indianapolis, IN, 46032

Summary

In drug-drug interaction (DDI) research, a two drug interaction is usually predicted by individual drug pharmacokinetics (PK). Although subject-specific drug concentration data from clinical PK studies on inhibitor or inducer and substrate's PK are not usually published, sample mean plasma drug concentrations and their standard deviations have been routinely reported. Hence there is a great need for meta-analysis and DDI prediction using such summarized PK data. In this paper, an innovative DDI prediction method based on a three-level hierarchical Bayesian meta-analysis model is developed. The three levels model sample means and variances, between-study variances, and prior distributions. Through a ketoconazle-midazolam example and simulations, we demonstrate that our meta-analysis model can not only estimate PK parameters with small bias, but also recover their between-study and between-subject variances well. More importantly, the posterior distributions of PK parameters and their variance components allow us to predict DDI at both population-average and study-specific levels. We are also able to predict the DDI between-subject/study variance. These statistical predictions have never been investigated in DDI research. Our simulation studies show that our meta-analysis approach has small bias in PK parameter estimates and DDI predictions. Sensitivity analysis was conducted to investigate the influences of interaction PK parameters, such as the inhibition constant *Ki*, on the DDI prediction.

Keywords

Area under the concentration curve ratio (*AUCR*); Bayesian hierarchical model; Drug-drug interaction (DDI); Meta-analysis; Monte Carlo Markov chain (MCMC); Pharmacokinetics (PK); Prediction

1. Introduction

Pharmacokinetic approaches designed to characterize drug to absorption, distribution and elimination were well established (Rowland and Tozer 1995) and robust statistical methodologies were developed (Davidian and Giltinan 1995). Recently pharmacokinetic interactions among multiple drugs have received a great deal of attention because this phenomenon makes a significant contribution to adverse drug reaction profile of new drugs (Ito et al. 1998). The importance of drug-drug interactions (DDI) was exemplified by the

^{*}Corresponding author email: meyu@iupui.edu.

interaction of ketoconazole (KETO) and terfenadine, which caused potentially life-threatening ventricular arrhythmias (Monahan et al. 1990), and the interaction between sorivudine and fluorouracil that resulted in fatal toxicity (Watabe et al. 1996; Okuda et al. 1997).

In the DDI research, one of the central questions is whether two individual drugs' pharmacokinetics (PK) models and their *in-vitro* DDI parameters can predict their *in-vivo* DDI (Ito et al. 1998). All the prior PK models and their parameters are summarized from multiple data sources (i.e. published PK studies in either literatures or public databases). Data for drug concentrations are usually published as sample mean profiles, together with standard deviations. Meta-analysis utilizing such summarized data is needed to obtain information from a vast existing resource.

Li et al. (2007) did some pioneer work in fitting PK models to sample mean data and evaluate the DDI prediction based on estimated PK parameters. They showed that it was feasible to draw inference for important PK parameters with summarized data, which were usually only available resource during the early phase DDI research. They proposed a mean model (see Section 2.5) for estimating PK parameters. This paper extends their work to estimation of both PK parameters and variance components. It also aims at predicting the both mean and variance of DDI, which are both interesting in practice. Understanding inter-individual variability is of great importance in PK studies. Large inter-individual variation usually means large environmental or genetic effects, which usually lead to pharmacogenetics studies. Readers can refer to Li et al. (2007) for a more detailed discussion of related issues and challenges for DDI prediction.

It is well known that the majority of DDIs depend on drug metabolizing enzymes and/or cell membrane drug transporters ([www.Drug-Interactions.com\)](http://www.Drug-Interactions.com). The single most important locus of DDIs is the cytochrome P450 (CYP) family of drug monooxygenases located in the gutwall and liver (de Waziers et al. 1990). Among CYPs, CYP3A is the most abundant and accounts for approximately 30% and 90% of CYP protein in liver and intestine, respectively (Shimada et al. 1994). In addition, more than 60% of the drugs that are eliminated primarily by metabolism are metabolized by CYP3A. KETO is a potent and extensively characterized CYP3A inhibitor, and midazolam (MDZ) is a highly selective CYP3A substrate *in vivo* that does not depend on membrane transporters for intracellular access (Tsunoda et al. 1999). Therefore, the KETO-MDZ pair is employed as an inhibitor-substrate example to illustrate our model based approach to DDI prediction. This paper develops an innovative hierarchical Bayesian meta-analysis model for PK parameter and variance component estimation. In particular, we focus on performing DDI prediction based on meta-analysis. Actual computation is via a Monte Carlo Markov chain (MCMC) method.

2. Specification of Models for Meta-analysis

2.1 Pharmacokinetics Model Specification

KETO Pharmacokinetic Model—Table 1 and Figure 1 (a) summarize seven published KETO PK data sets that were used to establish the PK model of KETO. These data were obtained through digitalization from corresponding papers (Table 1). KETO was administered orally in all studies, although dosing forms and conditions could differ. Some studies (Figure 1a) with longer blood sample collection times demonstrated biphasic decline in plasma concentrations consistent with a two compartment PK model (Gascoigne et al. 1981;Huang et al. 1986;FDA 1999). Following the notation of Rowland and Tozer (1995), we have the following differential equation based models for drug quantities in two components, $(A1_I, A2_I)$, after an oral dose,

$$
\frac{dA_1}{dt} = F_1 \times \text{Dose} \times ka_1 \times e^{-ka_1 \times t} - CL_1 \times \frac{A_1}{V_1} + (-\frac{A_1}{V_1} + \frac{A_2}{V_2}) \times CL12_1, \n\frac{dA_2}{dt} = (\frac{A_1}{V_1} - \frac{A_2}{V_2}) \times CL12_1, (A_1, A_2) |_{t=0} = (0, 0).
$$
\n(1)

Here subscript *I* indicates the inhibitor, KETO; F_I is the bioavailability; $k a_I$ is the absorption rate constant; $(V1_I, V2_I)$ are the volumes of distribution in systemic and peripheral compartments; and $CL12_I$ is the inter-compartment rate constant. CL_I is the clearance and it is modeled as $CL_I = Q_h \times CLint_I/(Q_h + CLint_I)$, where hepatic blood-flow, $Q_h = 80$ l/h (Price et al. 2003) is known, and the intrinsic hepatic clearance $CLint_I = Vmax_I/(Km_I + A1_I/V1_I)$. As parameters F_I and Km_I are not readily identifiable from data, they were assumed to be known, *F*_{*I*}=0.7 (Cleary et al. 1992) and *Km*_{*I*}=0.5 (Li et al. 2007).

MDZ Pharmacokinetic Model—Table 1 and Figure 1 (b) summarize two published MDZ data sets obtained also through digitalization. They were used to establish the PK model of MDZ. It was assumed to follow a two-compartment model with an intravenous infusion.

$$
\frac{dA1_S}{dt} = -CL_s \times \frac{A1_S}{V1_S} + (-\frac{A1_S}{V1_S} + \frac{A2_S}{V2_S}) \times CL12_S
$$

\n
$$
\frac{dA2_S}{dt} = (\frac{A1_S}{V1_S} - \frac{A2_S}{V2_S}) \times CL12_S, (A1_S, A2_S)|_{t=0} = (Dose, 0)
$$
 (2)

Here subscript *S* indicates the inhibitor, MDZ; $(V1_S, V2_S)$ are the volumes of distribution in systemic and peripheral compartments; and $CL12_S$ is the inter-compartment rate constant. CL_S is the clearance and modeled as $CL_S = Q_h \times \text{Clint}_S / (Q_h + \text{Clint}_S)$, and $CLint_S = f\mu_S \times$ *Vmax_S* /(*Km_S* + *fu_S* × *A*1*s*/*V*1*s*). As the parameter *Km_S* was not readily identifiable from data, it was assumed to be known as 2 (Li et al. 2007). The unbound fraction parameter $f\mu_S$ is also fixed at 0.04 for MDZ.

KETO/MDZ Interaction Model—When KETO and MDZ are administrated separately, their corresponding PK data can be independently modeled by (1) and (2) respectively. However, when they are administrated simultaneously, models (1) and (2) need to be connected by (3) (Ito et al. 1998)

$$
\text{CLint}_{S} = \frac{fu_{S} \times V \text{max}_{S}}{Km_{S} \times (1 + \frac{fu_{I} \times A1_{I}}{MW_{I} \times K \text{ixV} I_{I})} + fu_{S} \times \frac{A1_{S}}{VI_{S}}}.
$$
\n(3)

As a result, $(A1_I, A2_I, A1_S, A2_S)$ need to be jointly solved. In (3), fu_I =0.03 (Martinez-Jorda 1990) is the unbound fraction of KETO in plasma, *MWI*=0.53 is the molecular weight of the inhibitor, and *Ki* is the inhibition constant. KETO is a potent inhibitor of CYP3A enzymes with a *Ki* value ranges from 0.0037 – 0.18 μM (Bourrie, et al. 1996;Gibbs, et al. 1999. von Moltke, et al. 1994,1996;Wrighten and Ring, 1994). In these in-vitro experiments, *Kⁱ* was obtained from incubations of KETO in human liver microsomes. Note that the equation (3) reduces to $CLint_S = f\mu_S \times V$ max_{*S*}/($Km_S + fu_S \times A1_S/V1_S$) when there is no inhibitor (that is, when $Ki = \infty$).

Denote $AUC_{S,W}$ as the substrate area under concentration curve (AUC) with inhibitor, and *AUCS,WO* is the substrate AUC without inhibitor. For any given set of PK parameters for both inhibitor and substrate and their dose-combination, the substrate's concentration profiles (both with and without inhibitors) can be predicted, and their $AUC_{S,W}$ and $AUC_{S,WO}$ can be calculated

via the trapezoidal rule (Rowland and Tozer 1995). A common criterion to evaluate the extent of interaction is the area under concentration ratio (*AUCR*) of substrate after and before inhibitor administration, $AUCR = AUC_S$, $W/AUC_{S,WO}$. Let $(A\tilde{U}C_{S, W}, A\tilde{U}C_{S, WO})$ be calculated *AUCs* based on PK parameter estimates from data, then *AUCR* can be calculated via $A\tilde{U}C_{S,W}/A\tilde{U}C_{S,WO}.$

Model Based DDI Prediction—In the early drug development stage, a decision of continuing clinical trials for a candidate drug partially depends on its interaction with the other drugs (inducers or inhibitors). PK model based DDI prediction becomes critical, and precise prediction can guide drug development. Practically, inhibitor or inducer's PK models and parameters are estimated from their PK studies, while substrate's PK model and parameters are estimated from substrate's PK studies. Their interaction parameter, *Ki*, in (3) is measured from *in-vitro* studies. Then, DDI are predicted by the joint models from (1), (2), and (3).

This model based DDI prediction has been successfully implemented for simulating and predicting the effect of non-simultaneous substrate and inhibitor administration (Yang et al. 2003). However all PK parameters were treated as known without uncertainty in performing DDI prediction. From the statistical point of view, this deterministic approach is incomplete, because PK parameters of inhibitor (or inducer) and substrate estimated from published studies must have standard errors and these uncertainties need to be translated into their DDI prediction. On the other hand, some interaction parameters measured from *in vitro* studies, such as *Ki*, usually do not have reported standard error. That doesn't mean that these interaction parameters can be accurately estimated. The influence due to inaccurate estimates from these parameters on DDI prediction needs to be evaluated. For example, the range of KETO's *Ki* is cited as to be between 0.18 and 0.0037 μM from an FDA guideline document on DDI (FDA 2006) as well as in published literature (Bouurie et al. 1996, Gibbs et al. 1999, von Moltke et al. 1994, von Moltke et al. 1996, Wrighton and Ring 1994). It is therefore worthwhile to check how sensitive our model (*in vivo*) prediction is for different values of *Ki*.

2.2 Hierarchical Bayesian Meta-Analysis Model Specification

Notations—Before describing various models, we use the following notations throughout our paper. The subscript *i* will be used for subject, *j* for time point, *k* for study and *h* for (dosing) phase. We assume that there are n_{kh} subjects in the dosing phase *h* of study *k*; study *k* has H_k phases and there are total of *K* studies for a given drug. For ease of presentation, we assume all subjects within a dosing phase of a specific study are measured at the same time points for plasma concentration data. Therefore we have T_{kh} time points in the dosing phase *h* of study *k*. A parameter without subscript will be invariant across studies while a parameter with subscript varies according to the subscript. We also use bolded symbols for vectors and matrices. As examples, *α* will be a vector of parameters whose values are invariant across different studies, and *βik* is a vector of parameter for subject *i* from study *k*. The need for consider some parameters as invariant across different studies is to reduce the number of parameters. For example, in KETO, there are five different dosing routes: tablet with meal (TM), tablet fasting (TF), solution with meal (SLM), solution fasting (SLF), suspension fasting (SPF). The absorption rates corresponding to them satisfies the constraints $ka_I_{TM} < ka_I_{SIM} <$ $ka_{I,SLF}$ and $ka_{I,TM} < ka_{I,TF} < ka_{I,SPF} < ka_{I,SLF}$. So to reduce the number of parameters, we only make *kaI,TM,k* to be study specific and introduce *α* =(*ΔkaI,TF, ΔkaI,SLM*, *ΔkaI,SPF, ΔkaI,SLF*), where the elements of α are restricted to be positive. In other words, we assume that the difference in absorption rates between different routes is study invariant.

Meta-Analysis Models for Individual Level Data—When individual level data are available for each study, summarizing PK parameters from multiple sources by meta-analysis has been addressed by applying Bayesian methods to construct a drug PK model from several

clinical study data sets. (see e.g. Wakefield and Rahman 2000). In general, the following set of models are used

• subject-specific PK model:

$$
[y_{ijkl}|\alpha, \beta_{ik}, z_{jkh}] \sim \text{MVN}[f(\alpha, \beta_{ik}, z_{jkh}), \sigma_0^2], \tag{4}
$$

• subject-specific PK parameter model in study *k*:

$$
[\beta_{ik}|\beta_k, \sum] \sim \text{MVN}(\beta_k, \sum), \tag{5}
$$

• study-specific PK parameter model:

$$
[\beta_k|\beta,\Omega] \sim \text{MVN}(\beta,\Omega) \tag{6}
$$

In (4), y_{iikh} represents log-transformed drug concentration at time t_{ikh} for subject *i* in phase *h* of study *k*. Here we use a generic notation *zjkh* to denote available data and constants for a study subject at time t_{jkh} . It includes dosage, dosing route, and fixed constants such as Q_h , F_l , Km_l , *fuI* , *MW^I* , *KmS, fuS*, and time point *tjkh* at which plasma concentration is measured, The mean value $f(\mathbf{a}, \mathbf{\beta}_{ik}, z_{ikh})$ is the predicted log-transformed drug concentration for subject *i* at *j*th time point from phase *h* of study *k*. Basically, *f*(*α***,***βik*,*zjkh*) corresponds to (part of) a numerical solution at given time points from (1) and (2) by input α , β_{ik} and z_{jkh} . So in the case that y_{ijkh} is the KETO concentration observed from the first compartment, then $f(\mathbf{a,}\mathbf{b}_{ik},z_{jkh}) = log(A1_I/\sqrt{2})$ *V1*_{*I*}) with *A1*_{*I*} solved from (1). The measurement error variation, σ_0^2 , is assumed equal across

studies. The variance components Σ and Ω are for subject-specific and study-specific PK parameters.

However, published data are often available in the form of sample average plasma

concentration. In other words, only $(\bar{y}_{\bullet jkh}, s_{jkh}^2)$, instead of y_{ijkh} are published. But model (4) obviously needs *yijkh*, and hence can not be directly implemented to perform meta-analysis

with summarized data. Reconstructing a drug PK model from $(\bar{y}_{\bullet jkh}, s_{jkh}^2)$ needs a new metaanalysis formulation and different Bayesian sampling algorithm.

Meta-Analysis Models with Sample Mean and Standard Deviation Data—As

individual data are not available, it is an unrealistic goal to estimate subject level PK parameters. The sample mean data $\bar{y}_{\cdot jkh}$ at most provide information about study specific PK parameters

 $\mathbf{\beta}_k$ and study to study variation parameters $\mathbf{\Omega}$. The sample variance data s_{jkh}^2 at most provide information about between-subject heterogeneity parameter Σ . As a result, models based on $(4) - (6)$ are only suitable when working on subject level data. When only sample mean and sample variance data are available, they need to be modified to make estimation feasible. In other words, the models should not involve *βik*. A natural approach is to base estimation methods on marginalized likelihood where *βik* is integrated out. However, as no analytic form is available for $f(\alpha, \beta_{ik}, z_{jkh})$, direct integration is not feasible. The alternative we take is to first derive an approximation for *f*(*α*, *βik,zjkh*) and then marginalize. Specifically, we adapt as follows.

By Taylor expansion at *β^k* , *f*(**α**,**β***ik,zjkh*) can be approximated as

$$
f(\alpha, \beta_{ik}, z_{jkh}) \approx f\left(\alpha, \beta_k, z_{jkh}\right) + \left(\frac{\partial f}{\partial \beta_{ik}}\Big|_{\beta_{ik} = \beta_k}\right)^T (\beta_{ik} - \beta_k).
$$
\n(7)

Denote $\partial \beta_{i_{k}}^T$, $\partial \beta_{i_{k-1}}^T$ Instead of individual level PK model (4), we assume the following *approximated* model $[y_{ijkh} | \alpha, \beta_{ik}, \beta_k, \sigma_0^2, z_{jkh}] \sim N(f(\alpha, \beta_k, z_{jkh}) + F_{ikh}^1(\beta_{ik} - \beta_k), \sigma_0^2)$. Now by integrating out β_{ik} using the conditional distribution of β_{ik} given β_k , we have

$$
[y_{\text{ijkl}}|\alpha, \beta_k, z_{\text{jkh}}, \sum, \sigma_0^2] \sim N\Big(f(\alpha, \beta_k, z_{\text{jkh}}), F_{\text{jkh}}^T \sum F_{\text{jkh}} + \sigma_0^2\Big). \tag{8}
$$

As a result,

$$
[\bar{y}_{\bullet jkh}|\alpha, \beta_k, z_{jkh}, \sum, \sigma_0^2] \sim N\left(f(\alpha, \beta_k, z_{jkh}), \frac{F_{jkh}^T \Sigma F_{jkh} + \sigma_0^2}{n_{kh}}\right)
$$
\n(9)

To obtain the approximate distribution for sample variance, $s_{jkh}^2 = \sum_{i=1}^{n_{kh}} (y_{ijkh} - \overline{y}_{\bullet jkh})^2 / (n_{kh} - 1)$, we use the known fact the normalized sample variance of *m* i.i.d. normal variables has a chisquare distribution with degree of freedom $m-1$. From the approximate model (8), we immediately obtain

$$
\frac{\sum_{i=1}^{n_{h_k}} (y_{ijkh} - \overline{y}_{\bullet jkh})^2}{F_{jkh}^T \Sigma F_{jkh} + \sigma_0^2} \sim \chi^2_{n_{kh}-1}, \text{ or equivalently, } \frac{(n_{kh}-1)}{F_{jkh}^T \Sigma F_{jkh} + \sigma_0^2} s_{jkh}^2 \sim \chi^2_{n_{kh}-1}
$$
\n(10)

Notice that (9) and (10) depend only on β_k , Σ , and σ_0^2 Hence estimation of these parameters is feasible with mean and standard deviation data. Our estimation will be based on the following hierarchical models.

• Sample mean PK model

$$
[\overline{y}_{\bullet jkh} | \alpha, \beta_k, z_{jkh}, \sum, \sigma_0^2] \sim N \left(f(\alpha, \beta_k, z_{jkh}), \frac{F_{jkh}^T \sum F_{jkh} + \sigma_0^2}{n_{kh}} \right)
$$
\n(11)

• Sample variance PK model

Yu et al. Page 7

$$
\frac{(n_{\rm kh} - 1)}{F_{\rm jkh}^T \Sigma F_{\rm jkh} + \sigma_0^2} s_{\rm jkh}^2 \sim \chi_{n_{\rm kh} - 1}^2
$$
\n(12)

• Study-specific PK parameter model:

$$
[\beta_k|\beta,\Omega] \sim \text{MVN}(\beta,\Omega) \tag{13}
$$

Many times, a main purpose of determining variability is to identify individuals at risk. However as our model is based on summarized data, no individual information is available. All we can do is to quantify variance for a given population. Of course if individual data are available, subject covariates can be used in explaining variability using individual level models.

Prior Specification and Posterior Distributions—Due to the nonlinear structure from PK models (1) - (2), no conjugate priors exists for all parameter except *Ω*. Vague priors are used for PK parameter population level PK parameters (a, β) and subject level heterogeneity parameter, $(\Sigma, \Omega, \sigma_0^2)$, where $\Sigma = \text{diag}\{\sigma_1^2\}$ and $\Omega = \text{diag}\{\omega_1^2\}$.

$$
p(\alpha, \beta) = U(0, 10^4); \ p(\sigma_i^2) = U(a, b) \iff p(\sigma_i^{-2}) = \frac{(\sigma_i^{-2})^{-2}}{(b-a)}, l = 0, 1, ..., q;
$$

\n
$$
p(\omega_i^2) = U(a, b) \iff p(\omega_i^{-2}) = \frac{(\omega_i^{-2})^{-2}}{(b-a)}, l = 1, ..., q.
$$
\n(14)

Variance components all follow a uniform prior because practically they rarely exceed the bound $(a, b) = (0.01^2, 2^2)$.

Let $\overline{Y} = {\{\bar{\mathbf{y}} \cdot jkh; j = 1, \cdots, T_{kh}; k = 1, \cdots, K; h = 1, \cdots, H_k\}}$ be the sample mean data and $S = \{s_{jkh}^2; j=1,\dots, T_{kh}; k=1,\dots, k; h=1,\dots, H_k\}$ the sample variance data. Then, the posterior probability distribution based on (11) - (13) and priors (14) is,

$$
p(\alpha, \beta_1, \ldots, \beta_K, \sum, \Omega, \sigma_0^2 | \overline{Y}_\bullet, S, z) \n\propto \left\{ \prod_{k=1}^K \prod_{h=1}^T \prod_{j=1}^T N \left(\overline{y}_{\bullet jkh} | f(\alpha, \beta_k, t_{jkh}), \frac{F_{jkh}^T \Sigma F_{jkh} + \sigma_0^2}{n_{kh}} \right) \right\} \n\times \left\{ \prod_{k=1}^K \prod_{h=1}^T \prod_{j=1}^T X_{n_{h_k}-1}^2 \left(\frac{(n_{kh}-1)s_{jkh}^2}{F_{jkh}^T \Sigma F_{jkh} + \sigma_0^2} \right) \right\} \n\times \left\{ \prod_{k=1}^K p(\beta_k | \beta, \Omega) \right\} \times p(\alpha, \beta) \times p(\Sigma) \times p(\Omega) \times p(\sigma_0^2)
$$
\n(15)

The estimation procedure is implemented with a Monte Carlo Markov chain method. The posterior probability functions, $p(\beta | \cdot)$, $p(\alpha | \cdot)$, $p(\beta_k | \cdot)$, $p(\sigma_0^{-2} | \cdot)$, $p(\sigma_1^{-2} | \cdot)$, and $p(\omega_1^{-2} | \cdot)$ are derived in the Appendix. We defer the implementation details of our computation to the data analysis section.

An Alternative Bayesian Hierarchical Model for the Sample Mean PK Profile—In Li et al. (2007), instead of modeling both $\bar{\mathbf{y}}_{\cdot jkh}$ and s_{jkh}^2 as in (10) and (11), only $\bar{\mathbf{y}}_{\cdot jkh}$ is modeled. The variance of $\bar{\mathbf{y}}_{\cdot jkh}$ is assumed to be s_{jkh}^2/n_k . So instead of (10), the following is used

$$
[\overline{y}_{\bullet jkh}|\alpha, \beta_k, Z_{jkh}, S_{jkh}^2] \sim N\left[f(\alpha, \beta_k, Z_{jkh}), S_{jkh}^2/n_{kh}\right]
$$
\n(16)

In the following context, (11)-(13) are referred as the *mean-variance model* and (13) and (16) are referred as the *mean model*. While both models are approximation models, there are two major differences between the mean-variance model (11)-(13) and the mean model, (13) and (16). Firstly, the mean model is not designed to recover the between-subject variance **Σ**, while

the mean-variance can. Secondly, by using observed sample variance s_{jkh}^2/n_k to represent the variance of $\bar{y}_{\cdot jkh}$, model (16) should be more robust to model specification. It does not require Taylor expansion for approximation of *f(α,βik,zjkh*). Of course the downside is observed sample variance can deviated from the true variance, depending on the number of subject in each study and the true variability of individual data. The MCMC algorithm for the mean model is described in the Appendix IV of Li et al. (2007).

DDI Prediction Based on Sample Mean and Standard Deviation Data—The key to estimate *AUCR* is to estimate $AUC_{S/W}$ and $AUC_{S/WO}$. When estimates of these two AUCs are carried out by using population level parameters α and β , the ratio of $AUC_{S,W}$ and $AUC_{S,WO}$ leads to population level DDI which we denote by AUCR(*α, β*, *Ki*). When instead of *β*, study level parameters *βk* are used, then we have study level DDI which we denote by AUCR (*α*, *βk*, *Ki*). By the same token, we can use *βik* and obtain individual level DDI, AUCR(*α*, *βik*, *Ki*).

Based on posterior distributions of PK parameters from the mean-variance model, we are able to obtain a posterior distribution of *AUCR* at different levels. For example, a posterior sample of the distribution of population level DDI can be obtained by $AUCR(\alpha^{(u)}, \beta^{(u)}, Ki)$, where ${\{\alpha^{(u)}, \beta^{(u)}\}}_{u=1,\ldots,U}$ are their posterior draws. Similarly, we can have posterior sample of study level DDI. However for subject-specific DDI, we can not proceed in the same way as no posterior draws of individual level β_{ik} are available (note that β_{ik} is not present in the meanvariance model). Instead we obtain samples of AUCR(α , β_{ik} , Ki) by first drawing β_{ik} from its prior distribution (4) with *Σ* replaced by its posterior draws. That is, given any specific draw of $(\alpha^{(u)}, \beta_k^{(u)}, \Sigma^{(u)})$, we draw $\beta_{ik}^{(v)}, v=1,...,V$, from $[\beta_{ik} | \beta_k^{(u)}, \Sigma^{(u)}]$. Corresponding to any draw of $\beta_{ik}^{(v)}$, a sample of AUCR($\alpha^{(u)}$, $\beta_{ik}^{(v)}$, Ki) can be obtained. For any specific *u*, a total of V samples $AUCR(\mathbf{\alpha}^{(u)}, \mathbf{\beta}_{ik}^{(v)}, K_i)$ are obtained. Due to the study level nature of our data, this is a reasonable way for estimating subject-specific DDI. By estimating DDI distribution, our approach provides more information about DDI prediction, compared with the deterministic approach (Yang et al. 2003).

3. KETO/MDZ Example

The PK parameters of both KETO and MDZ are estimated from published studies using the proposed mean-variance model and mean-model from Section 2.2. The predicted logtransformed drug concentrations for KETO are calculated from the differential equation model (1) and the predicted log-transformed drug concentrations for MDZ are calculated from the differential equation model (2). Based on (known) DDI parameters, (*Km^I* , *KmS*,*fu^I* , *Ki,MW^I*), the DDI outcome, *AUCR*, is then predicted by models (1) and (2) linked by (3).

From the appendix, only $p(\omega_1^{-2}|\cdot)$ has an explicit form and hence can be simulated directly.

Other posterior distributions, $p(a | \cdot)$, $p(\beta_k | \cdot)$, $p(\sigma_0^{-2} | \cdot)$, and $p(\sigma_1^{-2} | \cdot)$, are drawn through the Metropolis Hasting (MH) algorithm (Hastings et al. 1970). At each step, a random walk chain is used and the random pertubation is taken to mean normal with mean 0 and standard deviation of 10%. The mixing is well with such proposed density and the acceptance rate varies. For

population parameter α , the acceptance rate is about 40% and for most of the study specific parameters β_k , the acceptance rates are close to 25%. Five independent chains were run simultaneously to determine the convergence with dispersed starting values on population parameters. It needed less than 4000 iterations for the estimated potential scale reduction criterion of Gelman and Rubin (1992) to be less than 1.2. By visual inspection of trace plots, almost all chains start to mix well after 500 iterations. The final results are based on a single chain of 50000 iterations after a burn-in of 10000. Every $10th$ iteration after burn-in was extracted for summarizing results. The numerical strategies for solutions of the differential equations and the derivatives of differential equations were discussed in (Li et al. 2002; Li et al. 2004). Both MCMC algorithms and numerical solutions of differential equations are implemented in the statistical freeware R 2.2.1.

3.1 KETO Data Analysis

Table 1 and Figure 1 (a) summarize seven published KETO PK data sets that were used to establish the PK model of KETO. All studies employed oral (PO) dosing of the Nizoral (Janssen Pharmaceuticals) formulation. The participants in these seven studies were young and healthy. The model is defined as in $(11)-(13)$ together with PK model (1) . Here the study specific parameters are $\beta_k = (VI_{I,k}, V_{I,k}, CL12_{I,k}, V_{max,I,k}, k a_{I, TM,k})$ with corresponding population parameters $\beta = (VI_L, V_2, CL12_L, V_{maxL}, ka_{L,TM})$. We also take $\alpha = (\Delta ka_{L,TF}, \Delta ka_{L,SLM}, \Delta ka_{L,TM})$ *ΔkaI,SPF, ΔkaI,SLF*) where elements of *α* are restricted to be positive. Both the variance matrix *Σ* of study specific parameters and *Ω* of subject specific parameters are taken to be diagonal.

We use different diagonal elements $(\omega_{v_1}^2, \omega_{v_2}^2, \omega_{v_{\text{max}_I}}^2, \omega_{c}^2, \omega_{\text{kal}_I}^2)$ for Ω and

 for *Σ*. During the actual computation, we log-transform all PK parameters. The interpretation of the variance components is then the coefficient of variance (CV). The prior distributions follow the same formulations as those in (14).

Estimation results are reported in Table 2 and Figure 2. The 90% credit intervals of PK parameters are reported. Here are some highlights.

- Among the KETO PK parameter estimates, $V2_I$ and $CL12_I$ have wider 90% CI than the others, if the CIs are normalized by their means. This is because only three studies (Gascoigne et al. 1981; Huang et al. 1986; FDA 1999) had sampling time points late enough to detect the terminal phase of KETO elimination from plasma, while the other studies did not.
- **•** *V2^I* and *CL*12*^I* 's between-study CV estimates, 0.44 and 0.54 respectively, are larger than the other between-study CV estimates.
- *ka_I*, $V2_I$ and $CL12_I$'s between-subject CV estimates 0.92, 0.31 and 0.32 respectively, are larger than the other between-study CV estimates. One example of fitting to sample variances is illustrated by Figure 2(a) for a KETO study (Gascoigne et al. 1981). The within subject CV is estimated as 0.20.
- **•** Both mean-variance model (model 2 in table 2) and mean-model (model 1 in table 2) have comparable performance in estimating PK parameters and their between-study CVs. All their 90% CIs are overlapped.

3.2 MDZ Data Analysis

Table 1 and Figure 1 (b) summarize two published MDZ data sets. They were used to establish MDZ's PK model and estimate its PK parameters. MDZ (administered as Versed®, Roche Pharmaceuticals) was administered in the fasting state in all studies. The participants in these two studies were young and healthy. Its mean-variance model is (11)-(13) together with the PK model (2). Here the study specific parameters are $\beta_k = (V1_{S,k}, V2_{S,k}, CL12_{S,k}, Vmax_{S,k}).$

As there are only two studies used in the analysis, the between study variances were assumed to be the same crossing different PK parameters, $\omega_{s,1}^2$. The between subject variance,

. We also log-transform the PK parameters and hence the interpretation of all the variance components is coefficient of variance (CV). The prior distributions follow the same formulations as those in (14). Estimation results are reported in Table 2. Here are some highlights.

- **•** All of MDZ PK parameter estimates have comparable 90% CIs, if they are normalized by their means.
- The between-study CV is estimated as 0.32, within subject CV is estimated as 0.10. between-subject CV ranges from 0.11 to 0.20. The fitting of sample CV is displayed by an MDZ example (Lee et al. 2002) in Figure 2(b).
- **•** Both mean-variance model (model 2 in table 2) and mean-model (model 1 in table 2) have comparable performance in estimating PK parameters and their between-study CVs. All their 90% CIs are overlapped.

3.3 KETO/MDZ Interaction Prediction

Assuming a simultaneous oral dose KETO and an IV dose for MDZ, the interaction PK models follow equations (1) and (2) connected by (3). The inhibition constant *Ki* is assumed to take value of (0.0037, 0.01, 0.18) μM to assess their influence on DDI prediction. In the simulation, the blood sampling time points are $(0.25, 0.5, 1, 1.5, 2, 3, 4, 6, 12)$ hours after dose. MDZ is dosed at (2, 5, 10) mg levels, and KETO/MDZ is dosed at three combinations: 200/2mg, 400/5mg, 800/10mg. The predicted MDZ plasma concentrations are simulated both with and without KETO, and at three different *Ki* values. *AUC* and *AUCR* are calculated with the trapezoid rule (Rowland and Tozer 1995). Following our proposed DDI prediction procedures in section **2**, population-average DDI, subject-specific DDI, and between-subject/study variance of DDI are predicted and displayed in Table 3. Here are some highlights.

- **•** The smaller the *Ki*, the larger the predicted *AUCR*. The larger the dose combination, the larger the predicted *AUCR*. These results fit well to previously published simulation studies (Yang et al. 2003).
- The smaller the *Ki*, the smaller between-subject/study CV. The larger the dose combination, the larger the CVs of population-average and subject-specific *AUCR*s.
- **•** Subject-specific *AUCR* and population-average *AUCR* have very comparable mean estimates, but subject-specific *AUCR* has much higher CVs, which is due to the additional between-subject/study variations.
- The predicted mean between-subject/study CV is smaller than the CV of the subjectspecific *AUCR*, because it doesn't contain the variability due to the uncertainties of PK parameter estimates.

4. Simulation Studies for PK Parameter Estimation and DDI Prediction

Based on our proposed hierarchical Bayesian model, sample mean drug concentration data and their sample variances were fitted to estimate PK parameters and variance components for KETO and MDZ. It is not clear whether the estimates are biased, and consequently whether the predicted *AUCR* and its variance are biased. To answer these questions, statistical simulations were conducted. We carry out simulation study based on the mean-variance model in this paper. For simulation under the mean model, readers are referred to Li et al. (2007).

In each simulated data set, eight studies for both KETO and MDZ were generated. KETO had 3 tablet dose levels (200mg, 400mg, 800mg) with meals and MDZ had IV bolus with 3 dosages

(2mg, 5mg, 10mg). There were 24 subjects for each study. Individual data were simulated first and then summarized as mean and standard deviation data for analysis. The blood sampling time points were (0.25, 0.5, 1, 1.5, 2, 3, 4, 6, 12, 24, 36) hours after dose for KETO and (0.25, 0.5, 1, 1.5, 2, 3, 4, 6, 12) for MDZ. At each KETO dose, its subject specific plasma concentrations were simulated with a two-compartment model (1) and subject specific models (4) – (6). The population PK parameters are *β* = log(*V*1*^I* ,*V*2*^I* , *CL*12*^I* ,*V* max*^I* ,*kaI,SLF*). We utilized the similar PK parameters from prescribed data analysis as their true values (Table 2). The between subject variance matrix is taken as $\sigma_1^2 I_{5\times 5}$ with σ_1^2 set to be 0.20². Study to study variance matrix is taken as $\omega_1^2 I_{5\times 5}$ with ω_1^2 set to be 0.30². The measurement error variance is set as 0.10^2 . Similar to the KETO, MDZ plasma concentrations were simulated with a two compartmental PK model (2) and subject specific models $(4) - (6)$. The PK parameters are $\beta = \log(V1_S, V2_S, V\text{max}_S, CL12_S)$, and the true PK parameters are listed in Table 3 and are chosen to be close to their estimates in the prescribed MDZ data analysis. The between subject variance matrix is taken as $\sigma_1^2 I_{4 \times 4}$ with σ_1^2 set to be 0.20². Study to study variance matrix is taken as $\omega_1^2 I_{4\times4}$ with ω_1^2 set to be 0.30². The measurement error variance σ_0^2 is set as 0.10². Totally, 100 simulated data sets were generated for both KETO and MDZ under each dose.

To fit the PK model in each simulation data set, an M-H algorithm was implemented as in the data example. To save computation time, initial values for PK parameters and variance components were chosen as their true values plus a random noise with a CV of approximately 10%. This increases the convergence rate of the algorithm, yet has minimum impact for the convergence itself. A total of 3000 iterations with 1000 burn-in were run to obtain PK parameter posterior distributions after the MCMC reached convergence. One in every five samples was picked to avoid autocorrelations. Both mean-variance model and mean-model were fitted to simulated data sets. Table 4 summarizes the PK parameters and variance components estimation biases for both approaches. Figure 3 summarizes the simulation results *for AUCR* prediction; and Figure 4 summarizes *the AUCR*'s variation prediction. Here are the highlights for simulation results.

- **•** In Table 4, except that *CL12*'s relative bias is around 7.5%, all the other PK parameters have relative bias less than 5%. *CL12* and *V2*'s estimates have larger bias than the other PK parameters. It is true in both mean-variance model and mean-model.
- In Table 4, both the between-subject and between-study variances are estimated with less than 5% relative bias.
- **•** In Figure 3, both subject-specific and population-average *AUCR* predictions have bias less than 5%. Their 90% CI coverage probabilities are close to their nominal levels.
- **•** In Figure 4, *AUCR*'s between-subject/study variation predictions have bias less than 5%, and their 90% CI coverage probabilities are close to their nominal levels.

5. Conclusions

In this paper, we have proposed a DDI prediction method based on an innovative three level hierarchical Bayesian model. It enables us to reconstruct a drug's PK model from multiple published PK studies. This approach can not only estimate PK parameters, but also can recover their between-study and between-subject variations. Most importantly, the posterior distributions of PK parameters and their variance components allow us to predict DDI at both population-average and study-specific levels. In addition, so we can also predict the DDI population and between-subject variance. These statistical predictions have never been investigated in a DDI research. Our simulation studies show that our meta-analysis approach

has small bias in PK parameter estimates and DDI prediction, when using the published sample mean and variance data.

Both data analysis and simulation studies demonstrate that the mean-variance model proposed in this paper and the mean-model in Li et al. (2007) have comparable performances in PK parameter estimations. In (Li et al. 2007), it was shown that PK parameter estimation is robust to prior distribution selection. In addition, it was shown in the paper that a second-order Taylor expansion approximation in (10) did not reduce the bias much. This is consistent with that of the marginal quasi-likelihood (MQL) approach in generalized linear mixed model (Molenberghs and Verbeke 2005) (Chap. 14, p270-273). Other approximation method, such as penalized quasi-likelihood approach (PQL) was reported to have better performance (Breslow and Clayton 1993; Wolfinger and O'Connell 1993), which is a first order Taylor expansion around subject-specific parameters. However, as the subject-specific level PK parameter information is not available in the sample mean data, PQL approach is not applicable in our meta-analysis.

The competitive inhibition interaction model (3) between MDZ/KETO has been well established in both in-vitro studies (von Moltke et al. 1996 and Gibbs et al.1999) and in-vivo studies (Yang et al. 2003 and Chien, et al. 2006). There are multiple MDZ/KETO DDI studies (McCrea, et al. 1999;Lam, et al. 2003;lkkola, et al. 1994;Lee et al. 2002; and Tsunoda et al. 1999). Our proposed drug interaction models (1) ∼ (3) predict reasonably well in MDZ/KETO interaction, given a single dose 200mg IV MDZ and a single dose $2mg$ KETO PO and $K_i =$ 0.0037 μM (Table 3). The predicted average AUCR = 3.02, and the reported average AUCR = 5.10 (Tsunoda et al. 1999). However, how to validate the model based DDI prediction is itself a highly challenging problem. Questions remain regarding how to determine the existence of DDI, how to test the equivalence between the predicted and the reported DDI, and how to expand the current two-compartmental interaction model to a physiological interaction model for possible improvement in DDI prediction. All these questions need significant joint efforts from statisticians and pharmacologists.

In this paper, we assume that the DDI PK model framework is known and correct. The unknown pieces are the pharmacokinetic parameters of the two drugs' two-compartment models. With a Bayesian approach, we are able to recover both the means and between-subject variances of the PK parameters from summarized data, which then leads to a valid DDI prediction. This paper establishes our Bayesian approach's feasibility and validity in recovering PK parameter means and variances from published summarized data. It serves as a fundamental building block for follow-up DDI model development and DDI prediction. The Bayesian models also provide a flexible framework to integrate prior knowledge into the estimation and prediction procedures.

Acknowledgments

Drs. Lang Li, Menggang Yu, and Stephen Hall's researches are supported by NIH grants, R01 GM74217 (LL), R01 GM67308 (SH), FD-T-001756(SH).

Appendix

Appendix Posterior Distributions

The following functions are conditional distributions to draw PK parameters and their variance components parameters in our three level hierarchical Bayesian model proposed in section 3.

Data analysis proposed in section 4 also utilized the following distributions. Assume the dimension of $\boldsymbol{\alpha}$ is p , of $\boldsymbol{\beta}_{ik}, \boldsymbol{\beta}_k$, and $\boldsymbol{\beta}$ is q .

Population PK parameters $(\alpha, \beta = (\beta_1, \ldots, \beta_q)^T)$ and study level PK parameters β_k :

$$
p(\alpha | \bullet) \propto \left\{ \prod_{h=1}^{H_k} \prod_{j=1}^{T_{kh}} N\left(f(\alpha, \beta_k, z_{jkh}), \frac{F_{jkh}^T \Sigma F_{jkh} + \sigma_0^2}{n_{kh}}\right) \right\}
$$
(A1)

$$
p(\beta_k|\bullet) \propto \left\{ \prod_{h=1}^H \prod_{j=1}^T N \left(f(\alpha, \beta_k, z_{jkh}), \frac{F_{jkh}^T \Sigma F_{jkh} + \sigma_{0k}^2}{n_k} \right) \right\} \times \left\{ \prod_{k=1}^K p(\beta_k|\beta, \Omega) \right\}
$$

$$
p(\beta_l|\bullet) \propto N \left(\left\{ \frac{1}{\tau_i^2} + \frac{K}{\omega_l^2} \right\}^{-1} \left\{ \frac{1}{\tau_i^2} \mu_l + \frac{K}{\omega_l^2} \overline{\beta}_l \right\}, \left\{ \frac{1}{\tau_i^2} + \frac{K}{\omega_l^2} \right\}^{-1} \right)
$$
 (A2)

for
$$
l=1, ..., q
$$
; where $\overline{\beta}_{l} = \sum_{k=1}^{K} \beta_{k1}$ (A3)

Variance components:

$$
Pr(\omega_1^2 | \bullet) \sim Inv - Gamma\left[\left(\frac{K}{2} - 1\right), \left(\frac{\sum_{k=1}^{K} (\beta_{k1} - \beta_l)^2}{2}\right) \right] \times 1_{\omega_1^2}(0.01^2, 2^2)
$$
\n(A4)

where $\frac{1}{\omega_i^2}$ (0.01², 2²) equals 1 when ω_i^2 is inside (0.01², 2²) and 0 otherwise.

$$
p(\sigma_i^2|\bullet)
$$

\n
$$
\propto \prod_{k=1}^{K} \prod_{h=1}^{H_k} \prod_{j=1}^{H_k} (F_{jkh}^T F_{jkh} \sigma_1^2 + \sigma_0^2)^{-\frac{n_{kh}}{2}+1}
$$

\n
$$
\times \exp \left\{-\frac{n_{kh} [\bar{y}_{\bullet jkh} - f(\alpha \beta k, z_{jkh})]^2 + \sum_{i=1}^{H_k h} (y_{ijkh} - \bar{y}_{\bullet jkh})^2}{2(F_{jkh}^T F_{jkh} \sigma_1^2 + \sigma_0^2)}\right\} \times 1_{\sigma_1^2} (0.01^2, 2^2)
$$
\n(A5)

$$
p(\sigma_0^2|\bullet)
$$

\n
$$
\propto \prod_{k=1}^{K} \prod_{h=1}^{T_{\rm kh}} [\Gamma_{jkh}^T F_{jkh} \sigma_1^2 + \sigma_0^2]^{-\frac{n_{\rm kh}}{2} + 1}
$$

\n
$$
\times \exp \left\{-\frac{n_{\rm kh} [\bar{y}_{\bullet jkh} - f(\alpha, \beta_k, z_{jkh})]^2 + \sum_{i=1}^{N_{\rm kh}} (y_{i,jkh} - \bar{y}_{\bullet jkh})^2}{2(F_{jkh}^T F_{jkh} \sigma_1^2 + \sigma_0^2)}\right\} \times 1_{\sigma_0^2} (0.01^2, 2^2)
$$
\n(A6)

Here $1_{\sigma_1^2}$ (0.01²,2²) and $1_{\sigma_1^2}$ (0.01²,2²) are similarly defined as $1_{\omega_1^2}$ (0.01²,2²).

References

- 1. Breslow NE, Clayton DG. Approximate inference in generalized linear mixed model. Journal of American Statistical Association 1993;88:9–25.
- 2. Bourrie M, Meunier V, Berger Y, Fabre G. Cytochrome P450 isoform inhibitors as a tool for the investigation of metabolic reactions catalyzed by human liver microsomes. Journal of Pharmacokinetics and Experimental Therapeutics 1996;277:321–332.
- 3. Chien JY, Lucksiri A, Ernest CS, Gorski JC, Wrighton SA, Hall SD. Stochastic prediction of CYP3Amediated inhibition of midazolam clearance by ketoconazole. Drug Metab Dispos 2006;34:1208– 1219. [PubMed: 16611859]
- 4. Cleary JD, Taylor JW, Chapman SW. Itraconazole in antifungal therapy. Ann Pharmacother 1992;26:502–509. [PubMed: 1315596]
- 5. Daneshmend TK, Warnock DW, Ene MD, Johnson EM, Parker G, Richardson MD, Roberts CJ. Multiple dose pharmacokinetics of ketoconazole and their effects on antipyrine kinetics in man. J Antimicrob Chemother 1983;12:185–188. [PubMed: 6311786]
- 6. Daneshmend TK, Warnock DW, Ene MD, Johnson EM, Potten MR, Richardson MD, Williamson PJ. Influence of food on the pharmacokinetics of ketoconazole. Antimicrob Agents Chemother 1984;25:1– 3. [PubMed: 6322675]
- 7. Daneshmend TK, Warnock DW, Turner A, Roberts CJ. Pharmacokinetics of ketoconazole in normal subjects. Journal of Antimicrobial Chemotherapy 1981;8:299–304. [PubMed: 6271723]
- 8. Davidian, M.; Giltinan, DM. Nonlinear models for repeated measurment data. Chapman and Hall; New York: 1995.
- 9. de Waziers I, Cugnenc PH, Yang CS, Leroux JP, Beaune PH. Cytochrome P 450 isoenzymes, epoxide hydrolase and glutathione transferases in rat and human hepatic and extrahepatic tissues. J Pharmacol Exp Ther 1990;253:387–394. [PubMed: 2329521]
- 10. FDA. Bioequivalence Reviews. 1999
- 11. FDA. Drug Drug Interaction Guideline. 2006. [http://wwwfdagov/cber/gdlns/interactionpdf](http://www.fdagov/cber/gdlns/interaction.pdf).
- 12. Gascoigne EW, Barton GJ, Michaels M, Meuldermans W, Heykans J. The kinetics of ketoconazole in animals and man. Clinical Research Reviews 1981;1:177–187.
- 13. Gelman A, Rubin DB. Inference From Iterative Simulation Using Multiple Sequences. Statistical Science 1992;7:457–472.
- 14. Gibbs MA, Thummel KE, Shen DD, Kunze KL. Inhibition of cytochrome P-450 3A (CYP3A) in human intestinal and liver microsomes: comparison of Ki values and impact of CYP3A5 expression. Drug Metab Dispos 1999;27(2):180–187. [PubMed: 9929500]
- 15. Huang YC, Colaizzi JL, Bierman RH, Woestenborghs R, Heykants J. Pharmacokinetics and dose proportionality of ketoconazole in normal volunteers. Antimicrob Agents Chemother 1986;30:206– 210. [PubMed: 3767339]
- 16. lkkola KT, Backman JT, Neuvonen PJ. Midazolam should be avoided in patients receiving the systemic antimycotics ketoconazole or itraconazole. Clin Pharmacol Ther 1994;55:481–5. [PubMed: 8181191]
- 17. Ito K, Iwatsubo T, Kanamitsu S, Ueda K, Suzuki H, Sugiyama Y. Prediction of pharmacokinetic alterations caused by drug-drug interactions: metabolic interaction in the liver. Pharmacol Rev 1998;50:387–412. [PubMed: 9755288]
- 18. Lam YW, Alfaro CL, Ereshefsky L, Miller M. Pharmacokinetic and pharmacodynamic interactions of oral midazolam with ketoconazole, fluoxetine, fluvoxamine, and nefazodone. J Clin Pharmacol 2003;43:1274–82. [PubMed: 14551182]
- 19. Lee JI, Chaves-Gnecco D, Amico JA, Kroboth PD, Wilson JW, Frye RF. Application of semisimultaneous midazolam administration for hepatic and intestinal cytochrome P450 3A phenotyping. Clin Pharmacol Ther 2002;72:718–728. [PubMed: 12496753]
- 20. Li L, Brown MB, Lee KH, Gupta S. Estimation and inference for a spline-enhanced population pharmacokinetic model. Biometrics 2002;58:601–611. [PubMed: 12229995]
- 21. Li L, Lin X, Brown M, Gupta S, Lee KH. A population pharmacokinetic model with time-dependent covariates measured with errors. Biometrics 2004;60:451–460. [PubMed: 15180671]

Yu et al. Page 15

- 22. Li L, Yu M, Chin R, Lucksiri A, Flockhart D, Hall S. Drug-Drug Interaction Prediction: A Bayesian Meta-Analysis Approach. Statistis in Medicine 2007;26:3700–3721.
- 23. Martinez-Jorda R, Rodriguez-Sasianin JM, Calvo R. Serum Binding of Ketoconazole in Health and Disease. International Journal fo Clinical Pharmacology Research 1990;5:271–276.
- 24. McCrea J, Prueksaritanont T, Gertz BJ, Carides A, Gillen L, Antonello S, Brucker MJ, Miller-Stein C, Osborne B, Waldman S. Concurrent administration of the erythromycin breath test (EBT) and oral midazolam as in vivo probes for CYP3A activity. J Clin Pharmacol 1999;39:1212–20. [PubMed: 10586386]
- 25. Molenberghs G, Verbeke G. Models for discrete Longintudinal data. 2005
- 26. Monahan BP, Ferguson CL, Killeavy ES, Lloyd BK, Troy J, Cantilena LR Jr. Torsades de pointes occurring in association with terfenadine use. Jama 1990;264:2788–2790. [PubMed: 1977935]
- 27. Okuda H, Nishiyama T, Ogura K, Nagayama S, Ikeda K, Yamaguchi S, Nakamura Y, Kawaguchi Y, Watabe T. Lethal drug interactions of sorivudine, a new antiviral drug, with oral 5-fluorouracil prodrugs. Drug Metab Dispos 1997;25:270–273.
- 28. Price PS, Conolly RB, Chaisson CF, Gross EA, Young JS, Mathis ET, Tedder DR. Modeling interindividual variation in physiological factors used in PBPK models of humans. Crit Rev Toxicol 2003;33:469–503. [PubMed: 14594104]
- 29. Rowland, M.; Tozer, TN. Clinical Pharmacokinetics Concept and Applications. Lippincott Williams & Wilkins; London: 1995.
- 30. Shimada T, Yamazaki H, Mimura M, Inui Y, Guengerich FP. Interindividual variations in human liver cytochrome P-450 enzymes involved in the oxidation of drugs, carcinogens and toxic chemicals: studies with liver microsomes of 30 Japanese and 30 Caucasians. J Pharmacol Exp Ther 1994;270:414–423. [PubMed: 8035341]
- 31. Tsunoda SM, Velez RL, von Moltke LL, Greenblatt DJ. Differentiation of intestinal and hepatic cytochrome P450 3A activity with use of midazolam as an in vivo probe: effect of ketoconazole. Clin Pharmacol Ther 1999;66:461–471. [PubMed: 10579473]
- 32. von Moltke LL, Greenblatt DJ, Duan SX, Harmatz JS, Shader RI. In vitro prediction of the terfenadineketoconazole pharmacokinetics interaction. Journal of Clinical Pharmacology 1994;34:1222–1227. [PubMed: 7738219]
- 33. von Moltke LL, Greenblatt DJ, Schmider DJ, Duan SX, Wright CE, Harmatz JS, Shader RI. Midazolam hydroxylation by human liver microsomes in vitro: inhibition by fluoxitine, norfluxetine, and by azole antifungal agents. Journal of Clinical Pharmacology 1996;36:783–791. [PubMed: 8889898]
- 34. Wakefield JC, Rahman N. The combination of population pharmacokinetic studies. Biometrics 2000;56:263–270. [PubMed: 10783805]
- 35. Watabe T. Strategic proposals for predicting drug-drug interactions during new drug development: based on sixteen deaths caused by interactions of the new antiviral sorivudine with 5-fluorouracil prodrugs. J Toxicol Sci 1996;21:299–300. [PubMed: 9035040]
- 36. Wolfinger R, O'Connell M. Generalized linear mixed models: a pseudo-likelihood approach. Journal of statistical Computation and Simulation 1993;48:233–243.
- 37. Wrighten SA, Ring BJ. Inhibition of human CYP3A catalyzed 1′-hydroxy midazolam formation by ketoconazole, nifedipine, erythromycin, cimetidine, and nizatidine. Pharm Res 1994;11:921–924. [PubMed: 7937537]
- 38. Yang J, Kjellsson M, Rostami-Hodjegan A, Tucker GT. The effects of dose staggering on metabolic drug-drug interactions. Eur J Pharm Sci 2003;20:223–232. [PubMed: 14550889]

Yu et al. Page 16

Figure 1.

(a) published ketoconazole studies; (b) published midazolam studies. Every curve represents a sample mean drug concentration profile; multiple curves from one study represent multiple phases of a pharmacokinetics study.

Figure 2.

(a) The fitting of sample coefficient of variance from a ketoconazole study (Gascoigne et al. 1981). The solid line is the sample coefficient variance, and dots are the posterior draws of the predicted coefficient of variance. (b) The fitting of sample coefficient of variance from a midazolam study (Lee et al. 2002). The solid line is the sample coefficient variance, and dots are the posterior draws of the predicted coefficient of variance.

Yu et al. Page 19

Figure 3.

(a), (b), and (c) are population-average *AUCR* prediction estimates, relative bias, and 90% credit interval coverage probability respectively. (d), (e), and (f) are subject-specific *AUCR* prediction estimates, relative bias, and 90% credit interval coverage probability respectively. These results are based on 100 simulated data sets.

Yu et al. Page 20

Figure 4.

(a), (b), and (c) are between-subject/study *AUCR* coefficient of variance estimates, relative bias, and 90% credit interval coverage probability respectively. These results are based on 100 simulated data sets.

Table 1

Published KETO and MDZ Data Sets

† In the original paper, MDZ was through IV fusion in a very short period to 2mg, we used the portion of data after fusion and assume IV bolus administration.

Pharmacokinetics Parameter Estimates for KETO and MDZ

J Biopharm Stat. Author manuscript; available in PMC 2009 September 4.

V max*S* 5125.4 (3035.2, 7936.4) 4488 (3051.8, 7001.3)

Model 1 *** : a Bayesian three-level hierarchical model for the sample mean. Model 1** : a Bayesian three-level hierarchical model for the sample mean. 90% Cl***: a 90% credit interval. It is represented by its relative scale to the mean. 90% CI*** : a 90% credit interval. It is represented by its relative scale to the mean.

NIH-PA Author Manuscript NIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH-PA Author Manuscript

Table 3 Population-Average and Subject-Specific AUCR Prediction

Note: the results are presented as mean \times / \div (1+CV)*100%.

Bias of PK Parameter Estimates

Bias of PK Parameter Estimates

Yu et al. Page 25

Model 1** : a Bayesian three-level hierarchical model for the sample mean.

Model 1 *** : a Bayesian three-level hierarchical model for the sample mean.

Yu et al. Page 26