

Overexpression of polo-like kinase1 predicts a poor prognosis in hepatocellular carcinoma patients

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Abstract

AIM: To elucidate the role of overexpressed polo-like kinase1 (PLK1) in hepatocellular carcinoma (HCC).

METHODS: We prospectively collected clinicopathological, immunohistochemical and semi-quantitative reverse transcription-polymerase chain reaction (RT-PCR) data from 135 HCC patients undergoing successful hepatectomy. The correlations between PLK1 mRNA expression and clinicopathologic variables were analyzed by Mann-Whitney *U* test. Prognostic factors were identified by univariate and multivariate analyses.

RESULTS: Immunohistochemical results showed overexpression of PLK1 was mainly found in tumor tissues compared with tumor-free tissue. A similar mRNA result was obtained by semi-quantitative RT-PCR. A total of 111 samples were positive for PLK1 mRNA expression. The positive expression was correlated with venous invasion, tumor nodules and Edmondson grade. Furthermore, 1, 3, 5-year survival rates in the positive expression group were significantly lower than the negative control group. Multivariate analysis showed that positive PLK1 expression was an independent risk factor for HCC.

CONCLUSION: PLK1 could be a potential biomarker for diagnosis and therapy for HCC.

INTRODUCTION

Hepatocellular carcinoma (HCC) is one of the deadliest of all cancers, ranking third among all cancer-related mortalities^[1]. Surgical resection plays a major role in the treatment of HCC. However, less than 30% of HCC patients are surgical candidates^[2] due to limiting factors such as severe impairment of hepatic functional reserve, bilobar tumor distribution and extra-hepatic metastasis. No single-agent or combination chemotherapy regimen has been found to be particularly effective in HCC^[3]. Locoregional treatment is not the first choice for HCC^[4] and is reserved for non-surgical candidates.

The study of carcinogenesis mechanisms may provide new treatment regimens for cancer. Many key carcinogenetic pathways, such as increased angiogenesis, aberrant signal transduction and dysregulated cell cycle control, appear to be involved in tumor development^[5]. As a cell cycle control kinase, polo-like kinase1 (PLK1) and its overexpression are highly associated with many human cancers, including bladder^[6], breast^[7-10], colorectal^[11,12], endometrial^[13,14], esophageal^[15-17], gastric^[18-21], glioma^[22], hepatoblastoma^[23], hepatocellular carcinoma^[24], head and neck^[25], leukemia and lymphoma^[26,27], melanoma^[28,29], non-small-cell lung^[30], ovarian^[31], papillary^[32], pancreatic^[33,34], prostate^[35] and thyroid cancer^[36]. However, up to now, as far as we know, there are few studies available on the overexpression of PLK in HCC. The objective of our study was to understand the relevance of PLK1 expression in HCC.

MATERIALS AND METHODS

Patients

There were 135 successful hepatectomy procedures for HCC performed from January 2003 to September 2008 in our department. None of these patients were given preoperative transarterial chemoembolization as a neoadjuvant treatment, but they all regularly received postoperative chemoembolization by hepatic artery infusion. All specimens from 135 cases were collected, immediately frozen in liquid nitrogen and subsequently stored at -70°C for reverse transcription-polymerase chain reaction (RT-PCR). Five cases of liver regenerating nodule samples were obtained from patients with liver cirrhosis as controls. The clinicopathologic variables were recorded in detail including gender, age, liver cirrhosis, hepatitis B surface antigen, α -fetoprotein (AFP), venous invasion, Edmondson stage, tumor size (cm) and number of tumor nodules. Two expert pathologists who were blinded to the other results of the study, scored the HCC samples. The study protocol was approved by the Ethics Committee of Central South University, and written informed consent was provided by all participants prior to initiation of the study.

Follow-up

All patients were involved in our follow-up system, and were reviewed at 1-2 mo intervals. Routine post-operative medical examinations were carried out every 2 mo, including liver function, serum AFP level, B-ultrasound and CT. Follow-up ranged from 3-62 mo and ended on December 31, 2008. The median follow-up time was 18 mo.

Semi-quantitative RT-PCR

Total RNA was extracted from tumor and tumor-free tissues using the TRIzol reagent (Gibco BRL) according to the manufacturer's instructions (Gibco BRL). cDNA synthesis performed using the reverse transcription system. The primers were as follows: PLK1, 5'-GATTCC ACGGCITTTTTCGAG-3', 5'-CCCACACAGGGTCTTC TTCC-3' (product size, 296 bp); β -actin: 5'-CGCGAGAA GATGACCCAGAT-3', 5'-GCACTGTGTTGGCGTAC AGG-3' (product size, 550 bp). The following PCR cycling parameters were employed: at 95°C for 5 min, followed by 35 cycles at 95°C for 45 s, 56°C for 1 min, at 72°C for 1 min and then 72°C for 7 min. The PCR products were resolved on a 1.5% agarose gel. All experiments were carried out in triplicate.

Immunohistochemical tissue slides

Immunohistochemical reaction against PLK1 (BD Transduction, a monoclonal mouse antibody) was performed in 5 μm paraffin sections. Negative controls were processed without primary antibody. For antigen retrieval, deparaffinized slides were placed in 0.01 mol/L sodium citrate buffer, pH 6.0 and boiled for 5 min in a pressure cooker. Then, slides were allowed to cool down for an additional 5 min in the same buffer. After

several rinses in TBS and pre-treatment with blocking reagent (Dako, Glostrup, Denmark) for 5 min, slides were incubated with primary antibody diluted 1:500 (PLK1) and 1:50 (Ki-67) in antibody diluent solution (Zymed, San Francisco, CA, USA) for 20 min at room temperature and then at 4°C overnight. After slides were washed in TBS, The slides were visualized using Aquatex (Merck, Gernsheim, Germany).

Statistical analysis

Quantitative values were presented as mean \pm SD or median (range). Independent Student's *t*-test was used to compare PLK1 mRNA expression in HCC and non-cancer samples. The Mann-Whitney *U* test was used for correlations between PLK1 mRNA expression and clinicopathologic variables. The Kaplan-Meier method was employed to calculate survival and the log-rank test to compare survival among two patient groups. Cox regression was adopted for multivariate analysis of prognostic predictors. The statistical software package SPSS16.0 (SPSS Inc, Chicago, IL) was applied for all analyses. A statistically significant *P* value was defined as < 0.05 .

RESULTS

Immunohistochemical analysis of PLK1 protein expression

The pattern of PLK1 protein expression was examined by means of immunohistochemical analysis. Overexpression was detected in tumor tissues, especially in cytoplasm, compared with the tumor-free tissue. A more varied morphology of the cells was reflected by high expression in the cytoplasm. (Figure 1A and B). Tumor tissues had sharp margins under the microscope (Figure 1B and D). No or sporadic expression was observed in adjacent normal tissue or regenerating nodules (Figure 1C and D).

Expression of PLK1 mRNA in HCC

To compare the expression levels of PLK1 mRNA between neoplasm, adjacent normal tissue in the experimental group and regenerating nodules in the control group, total RNA was extracted for RT-PCR. The results indicate that HCC tissues expressed a significantly higher level of PLK1 mRNA than adjacent normal tissue and regenerating nodule (0.53 ± 0.05 vs 0.23 ± 0.04 and 0.20 ± 0.02 , respectively, $P < 0.01$). When regenerating nodules were compared with adjacent normal tissue, the mRNA levels of regenerating nodules were slightly elevated, but the difference was not statistically significant (0.23 ± 0.04 vs 0.20 ± 0.02 , $P > 0.05$) (Figure 2).

There were 111 samples (111/135, 82.22%) in which the OD values of PLK1 mRNA in HCC tissues were higher than those of adjacent normal tissue ($P < 0.05$). These were named the PLK1 positive group. The remaining 24 samples (24/135, 17.78%) were named the PLK1 negative group. There were statistically significant differences in PLK1 mRNA levels between the positive and negative groups ($P = 0.003$).

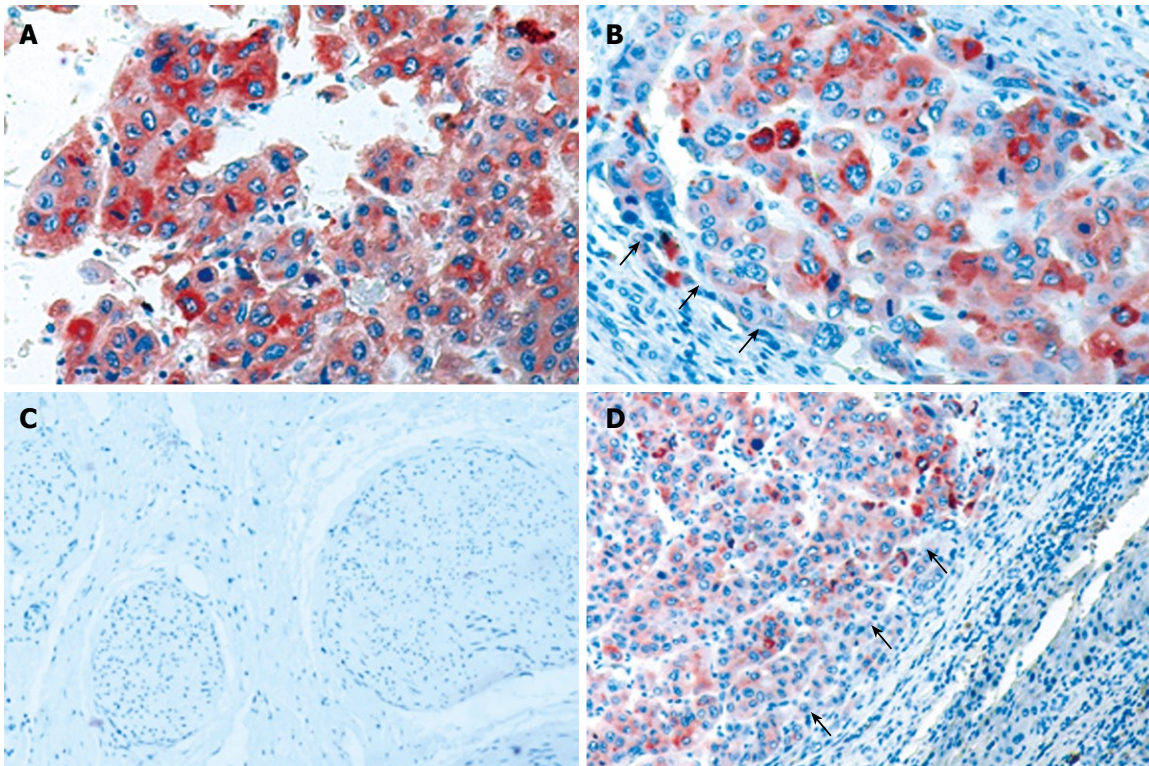


Figure 1 Expression of PLK1 in HCC tissue specimens. A: Expression of PLK1 in HCC tissue, especially in cytoplasm; B, D: Tumor tissue margins seen under the microscope (black arrows); C: Liver regenerating nodule sample; D: Adjacent normal tissue examined for PLK1 expression. Original magnifications, A and B $\times 200$; C and D $\times 100$.

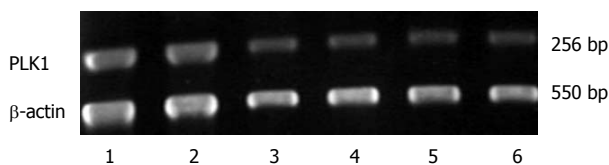


Figure 2 Expression of PLK1 mRNA in HCC, adjacent normal and regenerating nodule tissues. Lanes 1 and 2, HCC tissues; Lanes 3 and 4, adjacent normal tissue; Lanes 5 and 6, regenerating nodule tissues. RT-PCR for β -actin was used to monitor the quality of the RNA sample. RT-PCR was performed in triplicate.

Correlations between PLK1 positive expression group and clinicopathologic variables

The relationship between PLK1 positive expression groups and clinicopathologic data was analyzed by statistical software (Table 1). By the Mann-Whitney *U* test, the positive expression group with multiple tumor nodules had values significantly higher than that with solitary tumor nodules ($P = 0.002$). The high Edmondson-Steiner grade (III, IV) in the positive expression group was also significantly stronger than those of low grade tumors (I, II) ($P = 0.022$). Furthermore, venous invasion was significantly correlated with the positive expression group ($P = 0.042$). However, this study also showed that positive expression had no significant relationship with gender, age, liver cirrhosis, HBsAg, AFP or tumor size ($P > 0.05$) (Table 1).

Survival analysis of prognostic factors

As mentioned above, 135 cases of HCCs were divided

into positive and negative expression groups. The Kaplan-Meier method was employed to analyze the correlation of PLK1 expression level and the prognosis of HCC patients. Our results indicated that the positive expression group correlated with a shorter survival time than the negative group. The median survival time was 22.53 mo *vs* 60.88 mo. In addition, the 1, 3 and 5 year survival rates for the patients with positive and negative expression in HCC were 91.3%, 67.1% and 50.3% and 67.8%, 42.3% and 20.9%, respectively. The overall survival rates in the two groups was significantly different ($P = 0.003$, log-rank test) (Figure 3). At the same time, all the clinicopathologic variables were also analyzed with the Kaplan-Meier method. The results showed that a high Edmondson grade, venous invasion and multiple tumor nodules were correlated with a poor prognosis in HCC while the other clinicopathologic variables did not provide any independent prognostic information (Table 2).

By multivariable Cox regression analysis, PLK1 mRNA positive expression (RR, 3.507; 95% CI: 1.386-8.874, $P = 0.008$), high Edmondson grade (RR, 1.929; 95% CI: 1.069-3.482, $P = 0.029$), multiple tumor nodules (RR, 2.377; 95% CI: 1.384-4.082, $P = 0.002$) and venous invasion (RR, 4.848; 95% CI: 2.649-8.871, $P < 0.001$) were found to be independent prognostic factors for survival (Table 3).

DISCUSSION

Polo-like kinase 1 (PLK1) belongs to a family of

Table 1 Correlations between PLK1 mRNA expression and clinicopathologic variables by Mann-Whitney *U* test

Factors	n	PLK1 mRNA		P
		Positive	Negative	
Gender				
Male	114	93	21	0.650
Female	21	18	3	
Age (yr)				
≤ 40	65	53	12	0.842
> 40	70	58	12	
Liver cirrhosis				
Yes	109	93	16	0.055
No	26	18	8	
HBsAg				
Positive	96	80	16	0.598
Negative	39	31	8	
AFP (ng/mL)				
≤ 400	51	45	6	0.156
> 400	84	66	18	
Venous invasion				
+ (cases)	53	48	5	0.042
- (cases)	82	63	19	
Edmondson stage				
I - II	56	41	15	0.022
III-IV	79	70	9	
Tumor size (cm)				
≤ 5.0	36	28	8	0.417
> 5.0	99	83	16	
Tumor nodule				
Single tumor nodule	77	70	7	0.002
Multiple tumor nodule	58	41	17	

HBsAg: Hepatitis B surface antigen; AFP: α -fetoprotein.

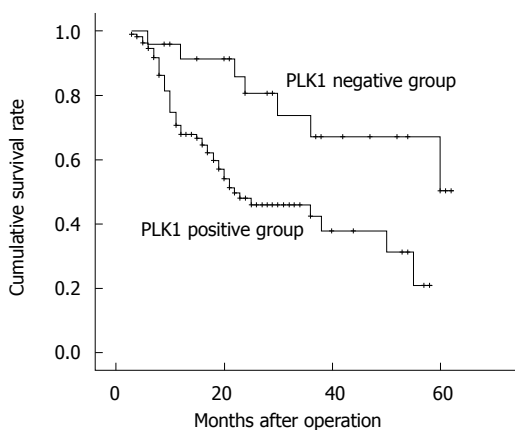


Figure 3 Log-rank test shows that HCC patients in the positive PLK1 mRNA expression group had a lower survival than those in the negative group.

conserved serine/threonine kinases involved in multiple mitotic processes^[37], including functional maturation of centrosomes, establishment of the bipolar spindle^[38], chromosome segregation^[39] and response to DNA damage^[40]. PLK1 is essential in the G₂/M-phase transition. The enzyme is able to activate CDC25c which in turn activates the CDC2/cyclin B1 complex leading to the import of cyclin B1 into the nucleus^[41], and it is also able to phosphorylate cyclin B1 directly^[42]. Moreover, PLK1 has been implicated in the regulation of anaphase-promoting complex/cyclosome^[43]. Interestingly, overexpression of PLK1 has been associated with tumor

Table 2 Correlation of several clinicopathological factors and of PLK1 mRNA expression group with patient survival (log-rank test)

Factors	P-value
Gender	0.155
Age	0.125
Liver cirrhosis	0.746
HBsAg	0.766
AFP	0.416
Venous invasion	< 0.001
Edmondson stage	0.001
Tumor size (cm)	0.183
Tumor nodule	0.018
PLK1 expression	0.003

Table 3 Multivariate survival analysis by Cox's proportional-hazard model for PLK1

	Relative risk	95.0% CI	P-value
Venous invasion	4.848	2.649-8.871	< 0.001
High Edmondson stage	1.929	1.069-3.482	0.029
Multiple tumor nodule	2.377	1.384-4.082	0.002
PLK1 positive expression	3.507	1.386-8.874	0.008

development and can serve as a prognostic marker for some cancers^[12,20,31]. However, few previous reports have examined the possible role of PLK1 in HCC.

In our study, overexpression of PLK1 by immunohistochemical analysis was detected in tumor tissues, while no or sporadic expression were observed in adjacent normal tissue or regenerating nodules. Further investigation showed that the degree of PLK1 mRNA expression was also higher in HCC tissues than in adjacent normal tissues and regenerating nodules; elevated mRNA levels of PLK1 were detected in 82.22% of tumor samples. These data indicate that overexpression of PLK1 was a frequent event in hepatocellular carcinoma.

In investigating the association between PLK1 expression and clinicopathological data, PLK1 positive expression was correlated with multiple tumor nodules, high Edmondson-Steiner grade (III, IV) and venous invasion; 1, 3 and 5 year survival rates of the positive expression group were 67.8%, 42.3% and 20.9%, respectively, lower than the negative group ($P < 0.01$); multivariate analysis showed that PLK1 was an independent prognostic factor. Thus, overexpression of PLK1 predicts a poor prognosis in HCC patients.

PLK1 is well known to be involved in cell proliferation. Was the PLK1 overexpression the cause of tumor formation or the consequence of high mitotic index during tumor cell proliferation? Others studies have shown that the overexpression of PLK1 in NIH3T3 leads to tumor formation^[44], and knockdown of PLK1 causes inhibition of growth and induction of apoptosis in human esophageal cancer cells^[17]. Furthermore, PLK1 was inhibited by DNA damage in the G₂ phase of mitosis. When the conserved threonine residue in the T-loop was changed to aspartic acid, expression of these mutants

was found to override the G2 arrest induced by DNA damage^[45]. In our studies, the level of PLK1 mRNA expression in HCC was obviously higher than in regenerating nodules, which also indicated that PLK1 might be a “proto-oncogene” for HCC.

In summary, this is the first attempt to clarify the relation of PLK1 expression and HCC. Our results indicate that high PLK1 expression predicts a poor prognosis in HCC patients. The data further suggest that PLK1 may serve as a potential biomarker for HCC. However, this is only a preliminary step. Further studies are warranted to understand the overexpression mechanism of PLK1 and to develop effective targeted interventions as a therapy for HCC.

COMMENTS

Background

Hepatocellular carcinoma (HCC) is one of the deadliest of all cancers with as yet incompletely elucidated causes. As a cell cycle control kinase, polo-like kinase 1 (PLK1) and its overexpression are highly associated with many human cancers. However, few studies are available on the overexpression of PLK1 in HCC.

Research frontiers

PLK1 is involved in cell proliferation. Its overexpression is associated with many human cancers. It is unclear whether the overexpression was the cause of tumor formation or the consequence of a high mitotic index during tumor cell proliferation. The trigger mechanisms of PLK1 overexpression in tumor formation are also unknown. Effective targeted interventions against PLK1 overexpression may be possible strategies as a therapy for HCC. All these questions are hotspots in the research field related to the article.

Innovations and breakthroughs

As a preliminary study of the relation between overexpression of PLK1 and HCC, it was found that overexpression of PLK1 was mainly found in tumor tissues compared with tumor-free tissue by immunohistochemical tissue slides and semi-quantitative reverse transcription-polymerase chain reaction analysis. In all HCC patients involved in the follow-up system, the survival rates in the PLK1 positive expression group were significantly lower than the negative control group. These results indicate that PLK1 could be a potential biomarker for diagnosis and therapy for HCC.

Applications

Overexpression of PLK1 is correlated with tumors. However the mechanism remains unclear. Their studies find overexpression of PLK1 in HCC, analyze the relationship between the prognosis of HCC and the overexpression, and provide the theoretical basis for targeted intervention against PLK1 as a means of treatment of HCC.

Terminology

PLK1: PLK1 is an important regulator of cell cycle progression during M-phase, involved in the assembly and dynamics of the mitotic spindle apparatus and in the activation and inactivation of CDK/cyclin complexes.

Peer review

This is a good descriptive study in which authors correlated polo-like kinase 1 expression in human HCC with clinicopathological findings. The data are very informative and deserve publication in the journal. The results are interesting and suggest that PLK1 could be a potential biomarker for diagnosis and therapy for HCC.

REFERENCES

- 1 **Block TM**, Mehta AS, Fimmel CJ, Jordan R. Molecular viral oncology of hepatocellular carcinoma. *Oncogene* 2003; **22**: 5093-5107
- 2 **Belghiti J**, Kianmanesh R. Surgical treatment of hepatocellular carcinoma. *HPB (Oxford)* 2005; **7**: 42-49
- 3 **Zhu AX**. Systemic therapy of advanced hepatocellular carcinoma: how hopeful should we be? *Oncologist* 2006; **11**: 790-800
- 4 **Yang Y**, Nagano H, Ota H, Morimoto O, Nakamura M, Wada H, Noda T, Damdinsuren B, Marubashi S, Miyamoto A, Takeda Y, Dono K, Umeshita K, Nakamori S, Wakasa K, Sakon M, Monden M. Patterns and clinicopathologic features of extrahepatic recurrence of hepatocellular carcinoma after curative resection. *Surgery* 2007; **141**: 196-202
- 5 **Ueno Y**, Moriyama M, Uchida T, Arakawa Y. Irregular regeneration of hepatocytes is an important factor in the hepatocarcinogenesis of liver disease. *Hepatology* 2001; **33**: 357-362
- 6 **Yamamoto Y**, Matsuyama H, Kawauchi S, Matsumoto H, Nagao K, Ohmi C, Sakano S, Furuya T, Oga A, Naito K, Sasaki K. Overexpression of polo-like kinase 1 (PLK1) and chromosomal instability in bladder cancer. *Oncology* 2006; **70**: 231-237
- 7 **Wolf G**, Hildenbrand R, Schwar C, Grobholz R, Kaufmann M, Stutte HJ, Strebhardt K, Bleyl U. Polo-like kinase: a novel marker of proliferation: correlation with estrogen-receptor expression in human breast cancer. *Pathol Res Pract* 2000; **196**: 753-759
- 8 **Spankuch B**, Heim S, Kurunci-Csacsco E, Lindenau C, Yuan J, Kaufmann M, Strebhardt K. Down-regulation of Polo-like kinase 1 elevates drug sensitivity of breast cancer cells in vitro and in vivo. *Cancer Res* 2006; **66**: 5836-5846
- 9 **Spankuch B**, Steinhauser I, Wartlick H, Kurunci-Csacsco E, Strebhardt KI, Langer K. Downregulation of Plk1 expression by receptor-mediated uptake of antisense oligonucleotide-loaded nanoparticles. *Neoplasia* 2008; **10**: 223-234
- 10 **Spankuch B**, Kurunci-Csacsco E, Kaufmann M, Strebhardt K. Rational combinations of siRNAs targeting Plk1 with breast cancer drugs. *Oncogene* 2007; **26**: 5793-5807
- 11 **Takahashi T**, Sano B, Nagata T, Kato H, Sugiyama Y, Kunieda K, Kimura M, Okano Y, Saji S. Polo-like kinase 1 (PLK1) is overexpressed in primary colorectal cancers. *Cancer Sci* 2003; **94**: 148-152
- 12 **Weichert W**, Kristiansen G, Schmidt M, Gekeler V, Noske A, Niesporek S, Dietel M, Denkert C. Polo-like kinase 1 expression is a prognostic factor in human colon cancer. *World J Gastroenterol* 2005; **11**: 5644-5650
- 13 **Takai N**, Miyazaki T, Fujisawa K, Nasu K, Hamanaka R, Miyakawa I. Polo-like kinase (PLK) expression in endometrial carcinoma. *Cancer Lett* 2001; **169**: 41-49
- 14 **Tang L**, Wang TT, Wu YT, Zhou CY, Huang HF. High expression levels of cyclin B1 and Polo-like kinase 1 in ectopic endometrial cells associated with abnormal cell cycle regulation of endometriosis. *Fertil Steril* 2009; **91**: 979-987
- 15 **Tokumitsu Y**, Mori M, Tanaka S, Akazawa K, Nakano S, Niho Y. Prognostic significance of polo-like kinase expression in esophageal carcinoma. *Int J Oncol* 1999; **15**: 687-692
- 16 **Feng YB**, Lin DC, Shi ZZ, Wang XC, Shen XM, Zhang Y, Du XL, Luo ML, Xu X, Han YL, Cai Y, Zhang ZQ, Zhan QM, Wang MR. Overexpression of PLK1 is associated with poor survival by inhibiting apoptosis via enhancement of survivin level in esophageal squamous cell carcinoma. *Int J Cancer* 2009; **124**: 578-588
- 17 **Bu Y**, Yang Z, Li Q, Song F. Silencing of polo-like kinase (Plk) 1 via siRNA causes inhibition of growth and induction of apoptosis in human esophageal cancer cells. *Oncology* 2008; **74**: 198-206
- 18 **Chen XH**, Lan B, Qu Y, Zhang XQ, Cai Q, Liu BY, Zhu ZG. Inhibitory effect of Polo-like kinase 1 depletion on mitosis and apoptosis of gastric cancer cells. *World J Gastroenterol* 2006; **12**: 29-35
- 19 **Weichert W**, Ullrich A, Schmidt M, Gekeler V, Noske A, Niesporek S, Buckendahl AC, Dietel M, Denkert C. Expression patterns of polo-like kinase 1 in human gastric cancer. *Cancer Sci* 2006; **97**: 271-276
- 20 **Kanaji S**, Saito H, Tsujitani S, Matsumoto S, Tatebe S, Kondo A, Ozaki M, Ito H, Ikeguchi M. Expression of polo-like kinase 1 (PLK1) protein predicts the survival of patients with gastric carcinoma. *Oncology* 2006; **70**: 126-133
- 21 **Jang YJ**, Kim YS, Kim WH. Oncogenic effect of Polo-like

- kinase 1 expression in human gastric carcinomas. *Int J Oncol* 2006; **29**: 589-594
- 22 **Dietzmann K**, Kirches E, von Bossanyi, Jachau K, Mawrin C. Increased human polo-like kinase-1 expression in gliomas. *J Neurooncol* 2001; **53**: 1-11
- 23 **Yamada S**, Ohira M, Horie H, Ando K, Takayasu H, Suzuki Y, Sugano S, Hirata T, Goto T, Matsunaga T, Hiyama E, Hayashi Y, Ando H, Suita S, Kaneko M, Sasaki F, Hashizume K, Ohnuma N, Nakagawara A. Expression profiling and differential screening between hepatoblastomas and the corresponding normal livers: identification of high expression of the PLK1 oncogene as a poor-prognostic indicator of hepatoblastomas. *Oncogene* 2004; **23**: 5901-5911
- 24 **Wang XQ**, Zhu YQ, Lui KS, Cai Q, Lu P, Poon RT. Aberrant Polo-like kinase 1-Cdc25A pathway in metastatic hepatocellular carcinoma. *Clin Cancer Res* 2008; **14**: 6813-6820
- 25 **Knecht R**, Elez R, Oechler M, Solbach C, von Ilberg C, Strebhardt K. Prognostic significance of polo-like kinase (PLK) expression in squamous cell carcinomas of the head and neck. *Cancer Res* 1999; **59**: 2794-2797
- 26 **Mito K**, Kashima K, Kikuchi H, Daa T, Nakayama I, Yokoyama S. Expression of Polo-Like Kinase (PLK1) in non-Hodgkin's lymphomas. *Leuk Lymphoma* 2005; **46**: 225-231
- 27 **Sun Q**, Zhang Y, Liu F, Zhao X, Yang X. Identification of candidate biomarkers for hepatocellular carcinoma through pre-cancerous expression analysis in an HBx transgenic mouse. *Cancer Biol Ther* 2007; **6**: 1532-1538
- 28 **Strebhardt K**, Kneisel L, Linhart C, Bernd A, Kaufmann R. Prognostic value of pololike kinase expression in melanomas. *JAMA* 2000; **283**: 479-480
- 29 **Kneisel L**, Strebhardt K, Bernd A, Wolter M, Binder A, Kaufmann R. Expression of polo-like kinase (PLK1) in thin melanomas: a novel marker of metastatic disease. *J Cutan Pathol* 2002; **29**: 354-358
- 30 **Wolf G**, Elez R, Doermer A, Holtrich U, Ackermann H, Stutte HJ, Altmannsberger HM, Rubsamen-Waigmann H, Strebhardt K. Prognostic significance of polo-like kinase (PLK) expression in non-small cell lung cancer. *Oncogene* 1997; **14**: 543-549
- 31 **Weichert W**, Denkert C, Schmidt M, Gekeler V, Wolf G, Kobel M, Dietel M, Hauptmann S. Polo-like kinase isoform expression is a prognostic factor in ovarian carcinoma. *Br J Cancer* 2004; **90**: 815-821
- 32 **Ito Y**, Miyoshi E, Sasaki N, Kakudo K, Yoshida H, Tomoda C, Uruno T, Takamura Y, Miya A, Kobayashi K, Matsuzuka F, Matsuura N, Kuma K, Miyauchi A. Polo-like kinase 1 overexpression is an early event in the progression of papillary carcinoma. *Br J Cancer* 2004; **90**: 414-418
- 33 **Gray PJ Jr**, Bearss DJ, Han H, Nagle R, Tsao MS, Dean N, Von Hoff DD. Identification of human polo-like kinase 1 as a potential therapeutic target in pancreatic cancer. *Mol Cancer Ther* 2004; **3**: 641-646
- 34 **Weichert W**, Schmidt M, Jacob J, Gekeler V, Langrehr J, Neuhaus P, Bahra M, Denkert C, Dietel M, Kristiansen G. Overexpression of Polo-like kinase 1 is a common and early event in pancreatic cancer. *Pancreatol* 2005; **5**: 259-265
- 35 **Weichert W**, Schmidt M, Gekeler V, Denkert C, Stephan C, Jung K, Loening S, Dietel M, Kristiansen G. Polo-like kinase 1 is overexpressed in prostate cancer and linked to higher tumor grades. *Prostate* 2004; **60**: 240-245
- 36 **Salvatore G**, Nappi TC, Salerno P, Jiang Y, Garbi C, Ugolini C, Miccoli P, Basolo F, Castellone MD, Cirafici AM, Melillo RM, Fusco A, Bittner ML, Santoro M. A cell proliferation and chromosomal instability signature in anaplastic thyroid carcinoma. *Cancer Res* 2007; **67**: 10148-10158
- 37 **Zhou T**, Aumais JP, Liu X, Yu-Lee LY, Erikson RL. A role for Plk1 phosphorylation of NudC in cytokinesis. *Dev Cell* 2003; **5**: 127-138
- 38 **Seki A**, Coppinger JA, Jang CY, Yates JR, Fang G. Bora and the kinase Aurora cooperatively activate the kinase Plk1 and control mitotic entry. *Science* 2008; **320**: 1655-1658
- 39 **Kang YH**, Park JE, Yu LR, Soung NK, Yun SM, Bang JK, Seong YS, Yu H, Garfield S, Veenstra TD, Lee KS. Self-regulated Plk1 recruitment to kinetochores by the Plk1-PBIP1 interaction is critical for proper chromosome segregation. *Mol Cell* 2006; **24**: 409-422
- 40 **Yuan JH**, Feng Y, Fisher RH, Maloid S, Longo DL, Ferris DK. Polo-like kinase 1 inactivation following mitotic DNA damaging treatments is independent of ataxia telangiectasia mutated kinase. *Mol Cancer Res* 2004; **2**: 417-426
- 41 **Toyoshima-Morimoto F**, Taniguchi E, Nishida E. Plk1 promotes nuclear translocation of human Cdc25C during prophase. *EMBO Rep* 2002; **3**: 341-348
- 42 **Jackman M**, Lindon C, Nigg EA, Pines J. Active cyclin B1-Cdk1 first appears on centrosomes in prophase. *Nat Cell Biol* 2003; **5**: 143-148
- 43 **Li M**, Zhang P. The function of APC/CCdh1 in cell cycle and beyond. *Cell Div* 2009; **4**: 2
- 44 **Smith MR**, Wilson ML, Hamanaka R, Chase D, Kung H, Longo DL, Ferris DK. Malignant transformation of mammalian cells initiated by constitutive expression of the polo-like kinase. *Biochem Biophys Res Commun* 1997; **234**: 397-405
- 45 **Smits VA**, Klompmaker R, Arnaud L, Rijksen G, Nigg EA, Medema RH. Polo-like kinase-1 is a target of the DNA damage checkpoint. *Nat Cell Biol* 2000; **2**: 672-676

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