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The detection of cytomegalovirus DNA in maternal plasma is associated with mortality in HIV-1 infected women and their

infants

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Abstract

Objective—Cytomegalovirus (CMV) is an important pathogen in healthy neonates and individuals with human immunodeficiency virus (HIV-1). The objective of this study was to determine whether the detection of CMV DNA (CMV DNAemia) in maternal plasma was associated with mortality in HIV-1 infected women or their infants.

Methods—A longitudinal study was designed to examine the relationship between maternal CMV DNAemia and maternal-infant mortality during two years *postpartum*. Sixty-four HIV-1 infected women and their infants were studied. CMV DNA loads were quantified in plasma from the mothers near the time of delivery. Baseline maternal CD4 counts, CD4%, HIV-1 RNA, and CMV DNAemia were evaluated as covariates of subsequent maternal or infant mortality in univariate and multivariate Cox regression.

Results—CMV DNA was detected in 11/64 (17%) of the HIV-1 infected women. HIV-1 and CMV viral load were strongly correlated in CMV DNAemic women (ρ =0.84, p=0.001). Detection of CMV DNAemia was associated with decreased maternal survival at 24 months *postpartum* (log-rank p=0.006). Additionally, HIV-1 infected infants born to CMV DNAemic women had a 4-fold increased risk of mortality during 24 months of follow-up. Maternal CMV DNAemia remained a significant risk factor for mortality in HIV-1 infected infants after adjusting for maternal CD4 cells/ mm³ (adjusted HR=4.3, CI=1.4–13), CD4% (HR=3.2, CI=1.0–10), HIV-1 viral load (HR=4.1, CI=1.4–12) or maternal death (HR=3.7, CI=1.0–13).

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Conflict of interest

The authors have no conflicts of interest to declare.

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Conclusions—Maternal plasma CMV DNAemia identified a subgroup of Kenyan women and infants at high risk for death in the two years following delivery.

Keywords

cytomegalovirus; vertical transmission; viral load; infant mortality

Introduction

Cytomegalovirus (CMV) is the most frequent viral cause of congenital disease globally, affecting 0.2–3% of live births in higher-income nations, with higher rates reported in sub-Saharan Africa [1–3]. Congenital CMV infection is associated with severe developmental disease such as microcephaly, sensorineural hearing loss and mental retardation [4,5]. Sequelae are present in ~10–20% of congenitally infected neonates who are born to women undergoing a primary infection, but maternal immunity confers significant protection from infection and disease in infants born to women with chronic CMV infection [6–8]. Postnatally acquired CMV is typically asymptomatic, but may be accompanied by transient mononucleosis and flu-like symptoms [9,10]. CMV is also major opportunistic pathogen in patients with human immunodeficiency virus (HIV-1) infection. In the absence of highly active antiretroviral therapy (HAART), patients with CD4 counts less than 50–100 cells/mm³ are at risk of CMV retinitis, gastrointestinal and neurological disease [11–14]. Vertical CMV transmission is more frequent in the setting of maternal HIV-1 infection and CMV infection has been associated with increased disease progression and mortality in HIV-1 infected infants [15–17].

Infants may acquire CMV close to the time they acquire HIV-1 infection, as a result of maternal primary CMV infection or recurrent infection [7,8,18]. Advancing maternal HIV-1 disease can also have important consequences for vertical HIV-1 transmission and infant survival. Maternal CD4 measurements, HIV-1 RNA viral load, and death have been shown to correlate with subsequent infant disease progression and mortality, though the precise mechanisms governing these relationships are not known [19–23]. Timing of vertical transmission, transfer of maternal antibody, exposure to co-infecting pathogens, and the ability to provide childcare may all be influenced by the mother's stage of HIV-1 disease. The role of maternal CMV replication and subsequent maternal-infant HIV-1 replication, immunosuppression and CMV replication on virus transmission and maternal-infant mortality.

Methods

Participants and study design

A longitudinal cohort study was designed to evaluate the relationship between maternal CMV DNA replication near the time of delivery and subsequent maternal and infant HIV-1 disease progression. The study protocol was approved by the Kenyatta National Hospital Ethics Review Committee and the Institutional Review Board of the University of Washington. A subset of 64 women and infants were selected from a previously described perinatal HIV-1 cohort [21,24,25]. HIV-1 seropositive pregnant women were recruited before their 28th week of gestation; the women received short-course zidovudine for prevention of HIV-1 transmission. Following delivery women and infants received no further antiretroviral therapy. Serial blood specimens were obtained in pregnancy, at delivery, and months 1, 3, 6, 9, 12, 15, 18, 21, and 24 *postpartum*. HIV-1 infected women and children were followed until death or exit from the study at two years *postpartum*. Infants who did not acquire HIV-1 during the study were followed for 1 year *postpartum*. The infants studied were part of a subset selected for intensive evaluation of immunological and viral factors as correlates of infant HIV-1 disease progression and survival. The selection of subjects from the original study cohort was therefore

based upon specimen availability and follow-up of the infants. Inclusion criteria for HIV-1 infected infants and their mothers were: 1) well defined timing of HIV-1 acquisition 2) availability of an infant plasma specimen by 1 month of age, and 3) infant survival to at least 3 months of age. Twenty HIV-1 uninfected infants and their HIV-1 infected mothers were also selected, based upon these criteria. Thirteen HIV-1 uninfected pregnant women were selected from a cohort with similar demographics as negative controls [26].

HIV-1 diagnosis and monitoring of immunologic parameters

HIV-1 RNA viral loads were measured in this cohort as previously described using the Gen-Probe transcription mediated assay [24,27,28]. Infant HIV-1 infection was diagnosed by nested PCR amplifying HIV-1 gag proviral DNA from dried blood spotted onto filter paper [29]. CD4 measurements were performed on whole blood using TriTest CD3FITC/CD4PE/CD45PerCP antibodies (BD Biosciences, San Jose, California, USA) and FACScan analysis with CELLQuest Software (BD Biosciences).

Infant HIV-1 infection *in utero* was defined as the detection of either HIV-1 DNA or RNA within 48 hours of birth, followed by a positive specimen (viral RNA or DNA) at the subsequent clinic visit. The peak HIV-1 viral load was defined as the highest measurement obtained during the first 6 months of infection, and the set-point viral load was defined as the first viral load measured at least 6 weeks after the peak.

CMV diagnosis and quantification

Nucleic acids were extracted from 50–200 µl of plasma using the Qiagen UltraSens virus extraction kit (Qiagen, Valencia, California, USA). CMV DNA loads were measured using a real time PCR to detect the glycoprotein B gene (gB) as previously described [30]. The lower limit of detection was 1 copy/reaction. Negative (no DNA detection) and indeterminate (<1 copy/reaction) PCR assays were not included in calculations of median or peak viral load, and were categorized as negative. CMV DNAemia was defined as the detection of CMV DNA in plasma. Timing of CMV acquisition was estimated as the mid-point between the last negative and first positive measurement. The peak CMV viral load was defined as the highest measurement within the first 6 months of infection.

Statistical analyses

Stata SE v9 (Stata Corp. College Station, Texas, USA) was used for analysis. Viral loads were base 10 log-transformed (\log_{10}) before comparisons and inclusion in regression models. T tests were used to compare mean \log_{10} HIV-1 viral load and CD4 values between groups. Fisher exact tests were used to compare proportions between groups. Maternal factors were examined as covariates for maternal and infant survival in univariate and multivariate survival analyses. Cox regression was used to estimate time to death and time to CMV infection, and the log-rank test was used to compare time to death between groups of subjects. All reported p values are for two-tailed tests.

Results

Baseline parameters of HIV-1 infected women

CMV DNAemia was measured in 64 HIV-1 infected women and 13 HIV-1 negative controls. Women were screened near to the time of delivery (57 women screened at delivery, 5 at 32 weeks gestation and 2 at 1 month *postpartum*) and infants were screened longitudinally during the first two years of life. CMV DNA was detected in 11/64 (17%) of HIV-1 infected women and none of the HIV-1 negative women (0/13, p=0.2). The median CMV DNA load was low in CMV DNAemic women; median 1.8 log₁₀ DNA copies/ml (range=1.6–2.2). Comparisons

of the HIV-1 infected women at 32 weeks gestation revealed that neither baseline CD4 counts nor HIV-1 RNA levels differed significantly between CMV DNAemic and non-CMV DNAemic women (mean CD4 count 335 versus 420 cells/mm³ respectively, p=0.2; mean 5.2 vs 4.9 log₁₀ RNA copies/ml, respectively, p=0.2; Table 1). There was a trend for lower CD4% in the CMV DNAemic women (mean 16 vs 21, p=0.08).

Maternal disease progression and mortality

At 24 months, the group of CMV DNAemic women had a higher frequency of deaths than the non-CMV DNAemic women (3/11 vs 2/53, p=0.03), and shorter survival time (21 months vs 24 months, p=0.006). HIV-1 RNA and CMV DNA loads were highly correlated in the 11 HIV-1 infected women who were CMV DNAemic (ρ =0.84, p=0.001; Figure 1). Since CD4 measurements were not performed at delivery (the time at which most CMV viral loads were measured), we were unable to examine concurrent correlation between CD4 measurements and CMV loads.

Cox regression was used to examine the relationship between CD4 measurements, HIV-1 RNA viral load, CMV DNAemia, and survival in the HIV-1 infected women. To establish a constant sample size, only women with data for all covariates were included in the univariate and multivariate Cox regression (59 women with HIV-1 viral load, CD4 count, CD4% and CMV DNA measured). Univariate predictors of maternal death included CD4 cells/mm³ at 32 weeks gestation (HR=0.99, 95% CI=0.98–1.00, p=0.004), CD4% at 32 weeks gestation (HR=0.78, 95% CI=0.68–0.91, p=0.001), HIV-1 RNA viral load at 32 weeks gestation (HR=7.2, 95% CI=1.5–34, p=0.01) and maternal CMV DNAemia dichotomised as detected/not detected (HR=9.7, 95% CI=1.6–59, p=0.01). After adjusting for maternal CD4%, maternal CMV DNAemia was no longer a significant risk factor for death (adjusted HR=1.9, 95% CI=0.17–21, p=0.6). Results were similar if alternatively adjusting for maternal CD4 cells/mm³, or HIV-1 RNA viral load (data not shown).

Virus transmission

We next examined the association between maternal CMV DNAemia and mother-infant CMV transmission. The frequency of CMV detection at birth was similar between HIV-1 infected infants born to mothers with and without CMV DNAemia (Table 2, 20% vs 10%, respectively, p=0.5), however there was a trend for women who were CMV DNAemic to transmit CMV to their (HIV-1 infected) infants earlier than women who were not CMV DNAemic (mean 1.2 months for CMV DNAemic women, 3.1 months for non-DNAemic women, p=0.1). The cumulative prevalence of infant CMV detection at 12 months was similar between HIV-1 infected infants born to CMV DNAemic and non-DNAemic women (100% vs 92%, p=0.6). There was no difference between prevalence of infant CMV, *in utero* transmission of CMV, or timing of CMV transmission in the HIV-1 uninfected infants born to women with and without CMV DNAemia.

Infant HIV-1 disease progression

HIV-1 infected infants born to HIV-1 infected women with CMV DNAemia were similar to non-DNAemic women in their levels of HIV-1 replication and immunosuppression, as indicated by CD4 measurements and HIV-1 peak and set-point viral load (Table 2). CD4 counts were available for a 25 of the children at 6 months, and there was no significant difference between CD4% in children born to CMV DNAemic women and non-DNAemic women (mean 25% vs 25%, p=0.97). Peak CMV load was also similar in HIV-1 infected infants born to women with CMV DNAemia and without CMV DNAemia (median 3.0 vs 3.1 log₁₀ copies/ml, p=0.7).

Infant mortality

Though we did not detect differences in levels of HIV-1 replication, immunosuppression, or CMV replication between children grouped by maternal CMV DNAemia, the difference in infant mortality was striking. By 24 months *post partum* 6 of the 7 (86%) HIV-1 infected infants born to CMV DNAemic mothers had died, and 14 of the 37 (38%) HIV-1 infected infants born to the non-CMV DNAemic women had died (p=0.03). Infants born to CMV DNAemic women had a shorter mean survival time (10 months 95% CI=5.6–15) compared to infants born to non-CMV DNAemic women (19 months, 95% CI=16–21, p=0.003, Figure 2).

We next used Cox regression to examine the relationship between the detection of maternal CMV DNA and mortality in the HIV-1 infected infants. To establish a constant sample size in the univariate and multivariate analyses, we restricted the analysis to subjects who had measurements available for all covariates (42 HIV-1 infected mother-infant pairs). In univariate Cox regression, maternal factors significantly predicting time to infant death were maternal CD4% at 32 weeks gestation (HR=0.92, p=0.03) and maternal CMV DNAemia (HR=4.4, p=0.006, Table 3). Maternal CD4 cells/mm³ at 32 weeks gestation and maternal HIV-1 viral load at 32 weeks gestation were not significant predictors of mortality in this subset of patients. A trend for association was found between maternal death and infant survival (HR=3.5, p=0.06). Multivariate Cox regression models were subsequently constructed to examine maternal CMV DNAemia as a cofactor for infant mortality while controlling for maternal disease progression and HIV-1 replication (Table 3). Since CD4 measurements, HIV-1 viral load, and death are collinear [21], these were not included together in the same model. The detection of maternal CMV DNA remained a predictor of infant mortality after adjusting for either baseline maternal CD4 count (HR=4.3, p=0.009), maternal CD4% at 32 weeks (HR=3.2, p=0.05), maternal HIV-1 RNA viral load (HR=4.1, p=0.01) or maternal death (HR=3.7, p=0.04).

Discussion

Though CMV is typically asymptomatic when acquired postnatally in immunocompetent infants, CMV can cause disease and more rapid HIV-1 progression in HIV-1 infected infants [17]. At present, the role that maternal CMV replication plays in CMV transmission and outcome in the HIV-1 infected neonate is unknown. Our results demonstrate the important role that systemic CMV replication in the mother may play in mortality of the HIV-1 infected child. We found that women with CMV DNA detected in the plasma were more likely to die within 2 years *postpartum*, tended to transmit CMV to their infants earlier than non-DNAemic women, and their infants were less likely to survive the first 2 years of life.

We did not find a correlation between maternal CMV DNAemia and congenital CMV transmission. With so few *in utero* CMV transmission events, we are underpowered to show a difference between groups. However, other groups have reported that in healthy (HIV-1 negative) pregnant women with primary or chronic CMV infection, the presence of CMV DNA in the blood alone does not correlate well with *in utero* transmission [3,31]. The detection of CMV from cervical secretions and breastmilk have been shown to correlate better with early CMV transmission than CMV in the blood [3]. CMV DNA appears in the blood during acute infection of healthy immunocompetent adults and decreases steadily to undetectable levels during the 6 month period post-infection in most subjects [31–33] and the detection of CMV in the plasma of healthy individuals with chronic CMV infection is uncommon [3]. Therefore, the appearance of CMV DNA in the peripheral blood suggests either recent primary infection, or that the host's immune system is sufficiently compromised to enable systemic dissemination of CMV. From the information collected in this study we are unable to determine if the DNAemic women had primary infection, reactivation, or re-infection with a new strain of CMV.

The detection of CMV DNA in the blood of immunosuppressed subjects has previously been shown to predict CMV disease and mortality in transplant recipients [34], HIV-1 infected and uninfected infants [35,36], and HIV-1 infected adults [37–39]. The strong correlation between levels of HIV-1 RNA and CMV DNA suggest that both HIV-1 and CMV are able to rapidly take advantage of immunosuppression at this level to replicate. Additionally, CMV could potentially act as a direct co-factor to increase HIV-1 replication, as reviewed by Griffiths [40]. A correlation between HIV-1 and CMV levels have also been reported in adult studies of blood [41] and breastmilk [42]; and the presence of HIV-1 shedding also correlates with the detection of CMV in the cervix [43] and semen [44]. In our study, plasma CMV DNAemia was associated with maternal death during the 2-year post partum period. In the multivariate model, the effect of maternal CMV DNAemia lost significance when adjusting for CD4%. This result is consistent with CMV DNAemia being a marker for more advanced HIV-1 disease in the mothers, but not necessarily an independent contributing factor to mortality.

To the best of our knowledge, this is the first report demonstrating a link between maternal CMV replication and infant mortality. HIV-1 infected infants born to HIV-1 infected women with CMV DNAemia were at a 4-fold greater risk of mortality compared to those born to CMV DNA-negative women. In multivariate regression adjusting for maternal immunosuppression or HIV-1 viral load, maternal CMV DNAemia remained the strongest predictor of infant mortality, with a hazard ratio varying from to 3.2–4.0 depending on the model. The relationship between maternal CMV DNAemia and infant mortality did not seem to be related to infant HIV-1 viral load or HIV-1 set-point, which were both comparable between infants born to women with or without CMV DNAemia.

One explanation for the association we have observed between maternal CMV DNAemia and infant mortality is earlier transmission of CMV from the CMV DNAemic women. We have some evidence to support this hypothesis; we were underpowered to precisely examine the timing of infant CMV acquisition, but we observed a trend for earlier CMV transmission from women who were DNAemic. Several mechanisms may explain the proposed relationship. First, cellular activation initiated by acute CMV infection may influence HIV-1 replication and dissemination by increasing the pool of CCR5-expressing targets for HIV-1 infection [45], thereby accelerating CD4 depletion. As both CMV and HIV-1 are transmitted via breastmilk [46], CD4 depletion in the neonatal gut could conceivably be accelerated by the synchronous introduction of both viruses to this site of infection [47]. Additionally, if the neonatal gut is compromised by HIV-1 infection, CMV replication may occur more readily, and potentially lead to the development of CMV gastrointestinal disease.

Our study has several strengths and some important limitations. Strengths include the prospective assessment of maternal factors and the collection of longitudinal data to assess the relative influence of maternal immunosuppression, HIV-1 viral load, mortality, and CMV DNAemia on infant mortality. An important limitation in the study was the use of plasma specimens for the measurement of CMV DNAemia. The use of whole blood or cell specimens would have increased our sensitivity to detect virus, but these specimens were not available. We are thus likely to have both underestimated levels of CMV replication in the blood, and to have underestimated the true frequency of individuals with active CMV replication. Secondly, we were not able to screen specimens from delivery for all of the infants; because of this we may have underestimated the true frequency of *in utero* CMV acquisition. Thirdly, because the original cohort was not enrolled to study CMV, we do not have data regarding infant CMVrelated morbidities. Finally, our exclusion of mother-infant pairs with less than 3 months of follow-up prohibits extrapolation of our results to infants who died in the first 3 months of life. Our selection criteria were designed to ensure adequate sampling and follow up for HIV-1 diagnosis, CMV diagnosis, and survival analysis, and these criteria may have led to selection bias. We found that maternal CMV strongly predicted mortality among infants who survived

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beyond 3 months. While it is likely that the same phenomenon occurs earlier in life, our selection criteria do not allow extrapolation to very early infant mortality.

Our study has important clinical implications. CMV DNAemia during pregnancy identified a subgroup of women and infants with a high risk of death in the two years following delivery. The value of CMV screening, prevention and therapy of pregnant HIV-1 infected women needs further study in regions where HIV-1/CMV co-infection is highly prevalent. Additionally, increased access to antiretrovirals will likely have indirect effects on the epidemiology of CMV in these regions, and the role of CMV in neonatal HIV-1 disease will need to be re-evaluated among women and infants receiving antiretroviral therapy.

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Figure 1. CMV and HIV-1 loads in co-infected women
Scatter-plot and linear fit line show data for HIV-1 infected, CMV DNAemic women (\rho=0.84, p=0.001).
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Kaplan-Meier curves showing time to death (log-rank p=0.003) for HIV-1 infected infants grouped by maternal CMV DNAemia. Solid line, infants born to mothers who were not CMV DNAemic; dashed line, infants born to mothers who were CMV DNAemic.

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	ae (months <i>post partum</i>) [95%CI] 21 [17–25] 24 [23–24] 0.006	nal mortality at 24 months <i>post partum</i>	27% (3/11)	3.8% (2/53)	0.03
Mean survival time (months post partum) [95%CI] 24 [23-24]		survival time (months post partum) [95%CI]	21 [17–25]	24 [23–24]	0.006

^aAt 32 weeks, CD4 measurements were available for 9 CMVDNA+ women and 50 CMVDNA- women, HIV-1 viral loads were available for 9 CMVDNA+ women and 52 CMVDNA- women.

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 Table 2

 Virus acquisition and disease progression of infants born to women with and without plasma CMV DNAemia.

Detected Not detected P Number of infants 11 53 7 HV-1 inflected 7 37 37 CVV acquisition (IIV-1 inflected inflants) 20% (J5) 10% (37) 05 CVV acquisition (IIV-1 inflected inflants) 20% (J7) 31 11.6.4.77 0.1 CVV acquisition (IIV-1 inflected inflants) 100% (77) 31 11.6.4.77 0.1 CVV acquisition (IIV-1 inflected inflants) 100% (77) 31 11.6.4.77 0.1 CVV acquisition (IIV-1 inflected inflants) 100% (77) 31 11.6.4.77 0.1 CVV acquisition (IIV-1 inflected inflants) 100% (77) 33% (11.2 0.1 0.6 CVV acquisition (IIV-1 inflected inflants) 100% (77) 20% (9.43) 0.1 0.6 CVV acquisition (IIV-1 inflected inflants) CVV acqq		Maternal	SMV DNA	
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Mean month of CMV acquisition [95%C1] 1.2 [0.57-1.7] 3.1 [1.6-4.7] 0.1 CMV acquisition (HIV-1 uninfected infants) $0\% (04)$ $8.3\% (1/12)$ 0.8 CMV acquisition (HIV-1 uninfected infants) $0\% (04)$ $8.3\% (1/12)$ 0.8 CMV detection at birth ^d $0\% (04)$ $8.3\% (1/12)$ 0.8 CMV detection by 12 months $100\% (444)$ $9.4\% (1/516)$ 0.8 Mean month of CMV acquisition [95%C1] $1.6 [1.0-2.3]$ $2.9 [1.3-4.4]$ 0.6 Nean month of CMV acquisition [95%C1] $1.6 [1.0-2.3]$ $2.9 [1.3-4.4]$ 0.6 Mean month of CMV acquisition [95%C1] $1.6 [1.0-2.3]$ $2.9 [1.3-4.4]$ 0.7 Mean Peak log ₁₀ HIV-1 viral load (\pm SE) 6.7 ± 0.30 6.7 ± 0.30 6.9 ± 0.11 0.7 Mean Set-point log ₁₀ HIV-1 viral load (\pm SE) 6.7 ± 0.30 6.7 ± 0.30 6.7 ± 0.30 0.7 Mean Set point log ₁₀ (HV V) $Mean CP4\%$ at 6 months <i>post partum</i> (\pm SE) 5.5 ± 3.9 2.5 ± 2.0 0.7 Mean Ret log ₁₀ (MV V) $Mean CP4\%$ at 6 months <i>post partum</i> (\pm SE) 2.5 ± 3.3 2.5 ± 2.0 <td< td=""><td>CMV detection by 12 months</td><td>100% (7/7)</td><td>95% (34/37)</td><td>0.6</td></td<>	CMV detection by 12 months	100% (7/7)	95% (34/37)	0.6
CMV acquisition (HIV-1 uninfected infants) $0\% (0.4)$ $8.3\% (1/2)$ 0.8 CMV attraction at bitrul ⁴ $0\% (0.4)$ $8.3\% (1/2)$ 0.8 CMV detection by 12 months $100\% (4/4)$ $9.4\% (15/16)$ 0.8 Mean month of CMV acquisition (95%CI) $100\% (4/4)$ $9.4\% (15/16)$ 0.8 Mean month of CMV acquisition (95%CI) $1.6(1.0-2.3)$ $2.911.3.4.4$ 0.6 Nean month of CMV acquisition (95%CI) $1.6(1.0-2.3)$ $2.911.3.4.4$ 0.6 Nean month of CMV acquisition (95%CI) 6.7 ± 0.30 6.9 ± 0.11 0.7 Mean Sterpoint log ₁₀ HIV-1 viral load (\pm SE) 6.7 ± 0.30 6.3 ± 0.34 6.1 ± 0.15 0.7 Mean Sterpoint log ₁₀ HIV-1 viral load (\pm SE) 6.3 ± 0.34 6.1 ± 0.15 0.7 Mean Sterpoint log ₁₀ HIV-1 viral load (\pm SE) 6.3 ± 0.34 6.1 ± 0.15 0.7 Mean CD4% at 6 months <i>post partum</i> (\pm SE) 2.5 ± 3.9 2.5 ± 2.0 0.7 Mean CD4% at 6 months <i>post partum</i> (\pm SE) $7.4.3.5$ 2.5 ± 3.9 2.5 ± 2.0 0.7 Mean CD4% at 6 months <i>post partum</i> (\pm SE) 2.5 ± 3.9 2.5 ± 3.0 0.7 <	Mean month of CMV acquisition [95%CI]	1.2 [0.57–1.7]	3.1 [1.6-4.7]	0.1
CWV detection at birth ^d 0% $(0/4)$ 8.3% $(1/12)$ 0.8 CWV detection by 12 months 100% $(4/4)$ 9.4% $(15/16)$ 0.8 CWV detection by 12 months 100% $(4/4)$ 9.4% $(15/16)$ 0.8 Mean month of CMV acquisition [95%CI] 1.6 [1.0–2.3] 2.9 [1.3–4.4] 0.6 Nean month of CMV acquisition [95%CI] 1.6 [1.0–2.3] 2.9 [1.3–4.4] 0.6 Nean month of CMV acquisition [95%CI] 6.7 ± 0.30 6.7 ± 0.30 6.9 ± 0.11 0.7 Nean Peak log in HV-1 viral load (\pm SE) 6.7 ± 0.30 6.3 \pm 0.14 0.7 0.7 Mean Set-point log in HV-1 viral load (\pm SE) 6.3 \pm 0.34 6.1 \pm 0.15 0.7 Mean CD4% at 6 months <i>post partum</i> (\pm SE) ^b 2.5 ± 3.9 2.5 ± 3.0 0.7 Mean CD4% at 6 months <i>post partum</i> (\pm SE) ^b 2.5 ± 3.0 0.7 0.7 Mean CD4% at 6 months <i>post partum</i> (\pm SE) ^b 2.5 ± 3.0 0.7 0.7 Mean CD4% at 6 months <i>post partum</i> (\pm SE) 2.5 ± 3.0 0.7 0.7 Mean CD4% at 6 months <i>post partum</i> (\pm SE) 2.5 ± 3.0 0.7 0.7	CMV acquisition (HIV-1 uninfected infants)			
CMV detection by 12 months100% (4/4)94% (15/16)0.8Mean month of CMV acquisition [95%CI]1.6 [1.0–2.3]2.9 [1.3–4.4]0.6Summary immunologic parameters (HV-infected infants) $1.6 [1.0–2.3]$ $2.9 [1.3–4.4]$ 0.6Nean Peak log ₁₀ HIV-1 viral load (\pm SE) 6.7 ± 0.30 6.7 ± 0.30 6.9 ± 0.11 0.7 Mean Peak log ₁₀ HIV-1 viral load (\pm SE) 6.3 ± 0.34 6.1 ± 0.15 0.7 Mean Set-point log ₁₀ HIV-1 viral load (\pm SE) 5.3 ± 0.34 5.3 ± 0.34 6.1 ± 0.15 0.7 Mean Set-point log ₁₀ HIV-1 viral load (\pm SE) 5.3 ± 0.34 5.5 ± 2.0 0.7 Mean CD4% at 6 months <i>post param</i> (\pm SE) 5.3 ± 0.34 5.5 ± 3.9 2.5 ± 2.0 0.7 Median peak log ₁₀ CMV DNA copies/mlHIV-1 infected infants (QR) $3.0 (2.6-3.5)$ $3.1 (2.6-3.5)$ 0.7 Mortality in HIV-1 infected infants (QR) $3.2 (2.6-3.5)$ $2.5 (2.0-3.0)$ 0.7 Mortality in HIV-1 infected infants 0.67 $3.2 (2.6-3.5)$ 0.7 Mortality in HIV-1 infected infants 0.67 0.67 0.67	CMV detection at birth ^{a}	0% (0/4)	8.3% (1/12)	0.8
Mean month of CMV acquisition [95%C1] $1.6 [1.0-2.3]$ $2.9 [1.3-4.4]$ 0.6 Summary immunologic parameters (HIV-infected infants) 6.7 ± 0.30 6.9 ± 0.11 0.7 Mean Peak log ₁₀ HIV-1 viral load (\pm SE) 6.7 ± 0.30 6.9 ± 0.11 0.7 Mean Set-point log ₁₀ HIV-1 viral load (\pm SE) 6.3 ± 0.34 6.1 ± 0.15 0.7 Mean Set-point log ₁₀ HIV-1 viral load (\pm SE) 5.3 ± 0.34 6.1 ± 0.15 0.7 Mean CD4% at 6 months <i>post partum</i> (\pm SE) ^b 2.5 ± 3.9 2.5 ± 2.0 0.97 Mean CD4% at 6 months <i>post partum</i> (\pm SE) ^b $3.0 (2.6-3.5)$ 2.5 ± 2.0 0.97 Mein peak log ₁₀ CWV DNA copies/ml $3.0 (2.6-3.5)$ $3.1 (2.6-3.5)$ 0.7 HIV-1 infected infants (IQR) $3.0 (2.6-3.5)$ $2.5 (2.0-3.0)$ 0.7 Morality in HIV-1 infected infants 0.67 $3.0 (2.6-3.5)$ 0.7 Morality in HIV-1 infected infants $0.86\% (677)$ $3.0 (2.6-3.5)$ 0.3 Morality in HIV-1 infected infants 0.67 0.67 0.03	CMV detection by 12 months	100% (4/4)	94% (15/16)	0.8
Summary immunologic parameters (HIV-infected infants) 6.7 ± 0.30 6.9 ± 0.11 0.7 Mean Peak log ₁₀ HIV-1 viral load (\pm SE) 6.7 ± 0.30 6.9 ± 0.11 0.7 Mean Set-point log ₁₀ HIV-1 viral load (\pm SE) 6.3 ± 0.34 6.1 ± 0.15 0.7 Mean Set-point log ₁₀ HIV-1 viral load (\pm SE) 5.3 ± 0.34 6.1 ± 0.15 0.7 Mean CD4% at 6 months <i>post partum</i> (\pm SE) 2.5 ± 3.9 2.5 ± 3.9 2.5 ± 2.0 0.97 Median peak log ₁₀ CMV DNA copies/ml 1.7 2.5 ± 3.9 2.5 ± 2.0 0.97 HIV-1 infected infants (IQR) $3.0 (2.6-3.5)$ $3.1 (2.6-3.5)$ 0.7 HIV-1 unificeted infants (IQR) $3.2 (2.6-3.5)$ $2.5 (2.0-3.0)$ 0.3 Mortality in HIV-1 infected infants 1.07 ± 0.13 0.03 Mortality at 24 months $86\% (6/7)$ 0.03 0.03 Mortality at 24 months 0.02 0.03 0.03	Mean month of CMV acquisition [95%CI]	1.6 [1.0–2.3]	2.9 [1.3-4.4]	0.6
Mean Peak log $_{10}$ HIV-1 viral load (± SE)6.7 ± 0.306.9 ± 0.110.7Mean Set-point log $_{10}$ HIV-1 viral load (± SE)6.3 ± 0.346.1 ± 0.150.7Mean Set-point log $_{10}$ HIV-1 viral load (± SE)6.3 ± 0.346.1 ± 0.150.7Mean CD4% at 6 months <i>post partum</i> (± SE)25 ± 3.925 ± 2.00.97Median peak log $_{10}$ CMV DNA copies/ml25 ± 3.925 ± 2.00.97Median peak log $_{10}$ CMV DNA copies/ml3.0 (2.6-3.5)3.1 (2.6-3.5)0.7HIV-1 infected infants (IQR)3.2 (2.6-3.5)3.2 (2.6-3.5)0.3Mortality in HIV-1 infected infantsMortality at 24 months86% (6/7)3.86% (6/7)0.03Mortality at 24 months86% (6/7)0.6670.16 ± 0.110.03	Summary immunologic parameters (HIV-infected infants)			
Mean Set-point \log_{10} HIV-1 viral load (± SE) 6.3 ± 0.34 6.1 ± 0.15 0.7 Mean CD4% at 6 months <i>post partum</i> (± SE) 25 ± 3.9 25 ± 2.0 0.97 Median peak \log_{10} CMV DNA copies/ml $3.0 (2.6-3.5)$ $3.1 (2.6-3.5)$ 0.7 HIV-1 infected infants (IQR) $3.0 (2.6-3.5)$ $3.1 (2.6-3.5)$ 0.7 HIV-1 uninfected infants (IQR) $3.2 (2.6-3.5)$ $2.5 (2.0-3.0)$ 0.3 Mortality in HIV-1 infected infants $86\% (67)$ $3.80\% (14.37)$ 0.03 Mortality at 24 months 0.01 ± 0.012 0.02 0.02	Mean Peak log ₁₀ HIV-1 viral load (\pm SE)	6.7 ± 0.30	6.9 ± 0.11	0.7
Mean CD4% at 6 months <i>post partum</i> (\pm SE) ^b 25 \pm 3.9 25 \pm 2.0 0.97 Median peak log ₁₀ CMV DNA copies/ml Median peak log ₁₀ CMV DNA copies/ml 1 </td <td>Mean Set-point $\log_{10} \rm HIV$ -1 viral load $(\pm \rm SE)$</td> <td>6.3 ± 0.34</td> <td>6.1 ± 0.15</td> <td>0.7</td>	Mean Set-point $\log_{10} \rm HIV$ -1 viral load $(\pm \rm SE)$	6.3 ± 0.34	6.1 ± 0.15	0.7
Median peak log ₁₀ CMV DNA copies/ml 3.0 (2.6–3.5) 0.7 HIV-1 infected infants (IQR) 3.0 (2.6–3.5) 0.7 HIV-1 uninfected infants (IQR) 3.2 (2.6–3.5) 0.3 Mortality in HIV-1 infected infants 108 86% (6/7) 0.3 Mortality at 24 months 86% (6/7) 38% (14/37) 0.03	Mean CD4% at 6 months <i>post partum</i> $(\pm SE)^b$	25 ± 3.9	25 ± 2.0	0.97
HIV-1 infected infants (IQR) 3.0 (2.6–3.5) 3.1 (2.6–3.5) 0.7 HIV-1 uninfected infants (IQR) 3.2 (2.6–3.5) 2.5 (2.0–3.0) 0.3 Mortality in HIV-1 infected infants 3.2 (2.6–3.5) 2.5 (2.0–3.0) 0.3 Infant mortality at 24 months 86% (6/7) 38% (14/37) 0.03	Median peak log ₁₀ CMV DNA copies/ml			
HIV-1 uninfected infants (IQR) 3.2 (2.6-3.5) 2.5 (2.0-3.0) 0.3 Mortality in HIV-1 infected infants 86% (6/7) 38% (14/37) 0.03 Infant mortality at 24 months 86% (6/7) 38% (14/37) 0.03	HIV-1 infected infants (IQR)	3.0 (2.6–3.5)	3.1 (2.6–3.5)	0.7
Mortality in HIV-1 infected infants Infant mortality at 24 months 86% (6/7) 38% (14/37) 0.03	HIV-1 uninfected infants (IQR)	3.2 (2.6–3.5)	2.5 (2.0–3.0)	0.3
Infant mortality at 24 months 86% (6/7) 38% (14/37) 0.03 Management of the information of the inform	Mortality in HIV-1 infected infants			
	Infant mortality at 24 months	86% (6/7)	38% (14/37)	0.03
	Mean survival time in months [95%CI]	10 [5.6–15]	19 [16–21]	0.003

aRestricted to infants tested for CMV DNA at birth.

b at 6 months, CD4 measurements were available for 5 infants born to CMV DNA+ women and 20 infants born to CMV DNA- women.

Material CP4 cellsimu ³ 0.999 0.997-1.01 0.5 Material CD4% 0.22 (0.95-1.03) 0.03 Material CD4% 0.22 (0.73-3.31) 0.03 Material CD4% 1.6 (0.73-3.31) 0.03 Material HV-1 RNA viral load 1.6 (0.73-3.13) 0.06 Material CD4 Second 1.6 (0.73-3.13) 0.06 Material CNV DNAemia 4.4 [1.5-13] 0.06 Material CNV DNAemia 1.4 [1.5-13] 0.06 Material CNV DNAemic 1.0 [1.5-13] 0.06 Material CNV DNAemic 1.0 [1.5-13] 0.06 Material CNV DNAemic 2.3 [1.4-13] 0.00 Material CP4 1.6 [1.4-13] 0.00 Material CP4% 0.35 [1.4-13] 0.00 Material CP4% 0.35 [1.4-13] 0.00 Material CP4% 0.35 [1.4-13] 0.00 Material CP4% 1.4 0.05 0.01 Material CP4% 0.0	Univariate ^d	HR	[95%CI]	d
Material CA etastimation 0.03 0.037-1.01 0.0 Material CD46, 16 0.03-3.31 0.03 Material CD46, 1 0.03 0.03 Material HP1 RNs vial load 3.5 0.056-131 0.06 Material detail 1.4 1.5-131 0.066 Material detail Material detail 0.15-3.51 0.06 Material CVV DNAenic HR 0.55%Cl 0.06 Material CVV DNAenic HR 0.95%Cl 0.06 Material CVV DNAenic 1.0 1.0 1.0 0.06 Material CD4 cells/m ³ 1.0 1.0-1.01 0.09 0.09 Model 1: Material CD4, cells/m ³ 1.0 0.09 0.09 Material CD4, cells/m ³ 1.0 1.0 0.09 0.09 Material CD4, cells/m ³ 1.0 0.05 0.06 0.00 Material CD4, cells/m ³ 0.05 0.05 0.09 0.09 Material CD4, cells/m ³ 0.05 0.06 0.06 0.06	Mattern 1 (17) 1-1-1-3			2 0
Material CD4% 02 03 03 Material CD4% 1.6 (0.86-0.9) 0.3 Material HIV-1 RNA viral load 1.6 (0.73-3.3) 0.05 Material CNV DNAenia 3.3 (0.96-13) 0.06 Material CVV DNAenia 4.4 (1.5-13) 0.06 Material CVV DNAenia HR 95%CI 0.06 Material CVV DNAenia HR 95%CI 0.06 Model 1: 1.0 1.0 1.0 0.06 Material CP4 cells/mu ³ 1.0 1.0 1.0 0.0 Model 1: Material CP4 cells/mu ³ 0.05 0.05 0.0 Model 2: Model 2: 1.0 1.0 0.0 0.0 Model 2: Model 2: 0.05 0.05 0.05 0.05 Model 2: Model 2: Model 2: 0.05 0.05 0.05 Model 2: Model 2: Model 2: 0.05 0.05 0.05 Model 2: Model 2: Model 2: 0.05		666.0	[0.1–1.66.0]	C.U
Maternal HrV-1 RNA viral load 1.6 (0.73-3.5) 0.3 Maternal death 3.5 (0.96-13) 0.06 Maternal death 4.4 (1.5-13) 0.06 Maternal death 4.9 (1.5-13) 0.06 Maternal CVV DNAtoria HR (9.96-13) 0.06 Maternal CVV DNAtoria HR (9.95%CI 0.06 Maternal CVA collocation 1.0 (1.0-10) 0.06 Model 1: Model 2: (1.0-10) 0.09 0.09 Model 2: Model 2: (1.0-10) 0.01 0.00 Model 2: Model 2: (1.0-10) 0.01 0.01 Maternal CD45 0.05 (0.64-30) 0.05 0.01 Model 2: Model 3: (1.0-10) 0.05 0.01 Model 3: Model 3: (1.0-10) 0.05 0.05 Model 4: Model 4: (1.0-10) 0.01 0.01 Model 4: Model 4: (1.0-12) 0.01 0.01 Model 4:	Maternal CD4%	0.92	[0.86-0.99]	0.03
Maerral death 3.5 (0.96-13) 0.06 Maerral CWV DNAmia 4.4 [1.5-13] 0.06 Material CWV DNAmia HR [1.5-13] 0.06 Material CWV DNAmia HR [1.5-13] 0.06 Material CW DNAmia HR [1.5-13] 0.06 Material CM eals/m ³ [1.2 [1.2-10] [0.06 Material CM eals/m ³ [1.0 [1.4-13] 0.09 Material CM eals/m ³ [1.4-13] [0.06 [0.06 Material CM eals/m ³ [1.4-13] [0.09 [0.06 Material CM eals/m ³ [1.4-13] [0.09 [0.06 Material CM eals [1.4-13] [0.06 [0.06 Material CM eals [1.4-13] [0.06 [0.06 Material CM eals [1.4-13] [0.06 [0.06 Material LM IV IRNA viral load [1.4-12] [0.06 [0.06 Material CM eal [1.4-12] [0.14-12] [0.06 Material CM eal [1.4-12] [0.06 [0.06	Maternal HIV-1 RNA viral load	1.6	[0.73 - 3.3]	0.3
Material CWD NAemia 4.4 [1.5-13] 0.006 Multivariae ^b HR [95%CI] p Multivariae HR [95%CI] p Model 1: 10 [1.0-1.0] p Model 2: 1.0 [1.0-1.0] [0.95%CI] 0.95 Mother CNV DNAemic 1.0 [1.4-1.3] 0.009 Mother CNV DNAemic 0.95 [1.4-1.3] 0.009 Mother CNV DNAemic 0.95 [1.4-1.3] 0.01 Mother CNV DNAemic 3.2 [1.4-1.3] 0.05 Mother CNV DNAemic 1.4 [1.4-1.2] 0.01 Model 3: Mother CNV DNAemic 0.1 0.01 Mother CNV DNAemic 1.4 0.01 0.01	Maternal death	3.5	[0.96–13]	0.06
$\begin{tabular}{ c c c c c c } \hline HR & [95\%CI] & p & & & & & & & & & & & & & & & & & $	Maternal CMV DNAemia	4.4	[1.5–13]	0.006
Model I: 1.0 [1.0-1.0] 0.9 Maternal CD4 cells/mm ³ 1.0 [1.4-13] 0.009 Mother CMV DNAemic 4.3 [1.4-13] 0.009 Mother CMV DNAemic 0.95 [0.88-1.0] 0.1 Maternal CD4% 3.2 [1.0-10] 0.05 Mother CMV DNAemic 3.2 [1.0-10] 0.05 Mother CMV DNAemic 1.4 [0.64-3.0] 0.4 Mother CMV DNAemic 1.4 [0.64-3.0] 0.4 Mother CMV DNAemic 1.5 [0.33-7.1] 0.4 Mother CMV DNAemic 1.5 [0.33-7.1] 0.4	Multivariate ^b	HR	[95%CI]	d
Maternal CD4 cells/mm³1.0 $(1.0-1.0]$ 0.9 Modrer CWV DNAemic4.3 $(1.4-13]$ 0.009 Model 2: $(1.4-13]$ 0.009 Model 2: $(0.88-1.0]$ 0.00 Maternal CD4% 0.95 $(0.88-1.0]$ 0.1 Maternal CD4% 3.2 $(1.0-10]$ 0.1 Model 3: $(1.0-10]$ 0.1 0.05 Model 3: $Model 3:$ $(1.0-10]$ 0.05 Model 3: $Model 3:$ $(1.4-12)$ 0.01 Model 4: $Modrer CMV DNAemic1.4(1.4-12)0.01Model 4:Modrer CMV DNAemic1.5(0.33-7.1]0.4Modrer CMV DNAemic1.5(1.0-13]0.4$	Model 1:			
Moduer CWV DNAemic 4.3 [14–13] 0.009 Model 2: 0.001 Model 2: 0.001 Maternal CD4% 0.95 0.01 Maternal CD4% 0.95 0.1 <td< td=""><td>Maternal CD4 cells/mm³</td><td>1.0</td><td>[1.0-1.0]</td><td>0.9</td></td<>	Maternal CD4 cells/mm ³	1.0	[1.0-1.0]	0.9
Model 2: 0.95 [0.88–1.0] 0.1 Maternal CD4% 0.95 [0.88–1.0] 0.1 Mother CMV DNAemic 3.2 [1.0–10] 0.05 Model 3: 1.4 0.64–3.0] 0.4 Maternal HIV-1 RNA viral load 1.4 [0.64–3.0] 0.4 Mother CMV DNAemic 4.1 [1.4–12] 0.01 Model 4: 1.5 [0.33–7.1] 0.4 Maternal death 1.5 [0.33–7.1] 0.4 Mother CMV DNAemic 3.7 [1.0–13] 0.4	Mother CMV DNAemic	4.3	[1.4–13]	0.009
Maternal CD4% 0.95 [0.88-1.0] 0.1 Mother CMV DNAemic 3.2 [1.0-10] 0.05 Model 3: 1.4 0.1 0.05 Model 3: 1.4 0.64-3.0] 0.4 Maternal HIV-1 RNA viral load 1.4 [1.4-12] 0.4 Mother CMV DNAemic 4.1 [1.4-12] 0.01 Model 4: 1.5 [0.33-7.1] 0.4 Maternal death 1.5 [0.33-7.1] 0.4 Mother CMV DNAemic 3.7 [1.0-13] 0.4	Model 2:			
Mother CMV DNAemic 3.2 [1.0-10] 0.05 Model 3: <	Maternal CD4%	0.95	[0.88-1.0]	0.1
Model 3: Model 3: 0.04 Maternal HIV-1 RNA viral load 1.4 [0.64–3.0] 0.4 Maternal HIV-1 RNA viral load 4.1 [1.4–12] 0.01 Mother CMV DNAemic 4.1 [1.4–12] 0.01 Model 4: 1.5 [0.33–7.1] 0.4 Maternal death 1.5 [0.33–7.1] 0.4 Mother CMV DNAemic 3.7 [1.0–13] 0.04	Mother CMV DNAemic	3.2	[1.0-10]	0.05
Maternal HIV-1 RNA viral load 1.4 [0.64–3.0] 0.4 Mother CMV DNAemic 4.1 [1.4–12] 0.01 Model 4: 1.5 (0.33–7.1] 0.4 Maternal death 1.5 [0.33–7.1] 0.4 Mother CMV DNAemic 3.7 [1.0–13] 0.04	Model 3:			
Mother CMV DNAemic 4.1 [1.4-12] 0.01 Model 4: 0.01 0.01 Maternal death 1.5 [0.33-7.1] 0.4 Mother CMV DNAemic 3.7 [1.0-13] 0.04	Maternal HIV-1 RNA viral load	1.4	[0.64 - 3.0]	0.4
Model 4: 1.5 [0.33-7.1] 0.4 Maternal death 3.7 [1.0–13] 0.04	Mother CMV DNAemic	4.1	[1.4–12]	0.01
Maternal death 1.5 [0.33-7.1] 0.4 Mother CMV DNAemic 3.7 [1.0-13] 0.04	Model 4:			
Mother CMV DNAemic 3.7 [1.0–13] 0.04	Maternal death	1.5	[0.33-7.1]	0.4
	Mother CMV DNAemic	3.7	[1.0–13]	0.04

HR, hazard ratio.

 a Maternal CD4 cells/mm³, CD4% and HIV-1 viral load at 32 weeks gestation, death by 24 months.

^b Adjusted HR, [95%CI] and p values are shown for four Cox regression models including maternal CMV DNAemia and either maternal CD4 cells/mm³, CD4%, HIV-1 RNA viral load or death.

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Table 3