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Intracranial and Abdominal Aortic Aneurysms: Similarities, Differences, and Need for a New Class of Computational Models

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Abstract

Intracranial and abdominal aortic aneurysms result from different underlying disease processes and exhibit different rupture potentials, yet they share many histopathological and biomechanical characteristics. Moreover, as in other vascular diseases, hemodynamics and wall mechanics play important roles in the natural history and possible treatment of these two types of lesions. The goals of this review are twofold: first, to contrast the biology and mechanics of intracranial and abdominal aortic aneurysms to emphasize that separate advances in our understanding of each disease can aid in our understanding of the other disease, and second, to suggest that research on the biomechanics of aneurysms must embrace a new paradigm for analysis. That is, past biomechanical studies have provided tremendous insight but have progressed along separate lines, focusing on either the hemodynamics or the wall mechanics. We submit that that there is a pressing need to couple in a new way the separate advances in vascular biology, medical imaging, and computational biofluid and biosolid mechanics to understand better the mechanobiology, pathophysiology, and treatment of these lesions, which continue to be responsible for significant morbidity and mortality. We shall refer to this needed new class of computational tools as Fluid-Solid-Growth (FSG) Models.

Keywords

hemodynamics; wall stress; saccular aneurysm; abdominal aortic aneurysm; growth and remodeling; cell and matrix turnover; patient-specific modeling

INTRODUCTION

The normal arterial wall consists of three layers: the intima, media, and adventitia. The innermost layer, or intima, consists of a monolayer of endothelial cells attached to a basement membrane composed of type IV collagen and laminin; the middle layer, or media, consists of smooth muscle cells embedded in an extracellular matrix composed of elastin, multiple types of collagen, and proteoglycans; the outermost layer, or adventitia, consists of fibroblasts embedded in an extensive plexus of type I collagen with admixed elastin. An aneurysm is defined as a focal dilatation of the arterial wall. Although the pathogenesis of aneurysms remains an enigma, the initial dilatation appears to be caused in part by degeneration of a portion of the arterial wall, often medial elastin and then smooth muscle. The two most common types of aneurysms are intracranial saccular aneurysms (ISAs) and abdominal aortic aneurysms

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(AAAs). ISAs occur in or near the circle of Willis, the major network of arteries that supplies blood to the brain; AAAs occur in the infrarenal aorta, the primary conduit that supplies blood to the legs. Genetics and risk factors play important roles in the natural history of aneurysms, but it is universally accepted that biomechanical factors (including increased pressures in hypertension) also play fundamental roles.

Advances in vascular biology, biomechanics, medical imaging, and computational methods collectively provide an unprecedented opportunity to develop and use mathematical models to understand better the progression of vascular diseases as well as to improve the design of medical devices and strategies for intervention. To realize this promise, however, there is a pressing need for a paradigm shift in our approach. Whereas the vast majority of prior modeling has focused on either the hemodynamics or the wall mechanics, and has focused on conditions at particular instants during disease progression or treatment, there is a need to develop coupled Fluid-Solid-Growth (FSG) Models of the evolving changes in vascular biology, geometry, wall properties, and hemodynamics. That is, most important vascular pathologies and most adverse responses to treatment manifest over extended periods, times during which the vascular wall grows and remodels in response to the altered mechanobiology. In this paper, we discuss the general need for FSG Models using as examples ISAs and AAAs. Toward that end, we briefly contrast the histopathology and mechanobiology of these aneurysms and review prior biomechanical analyses. We then outline a new approach for modeling these devastating lesions and conclude by suggesting particular needs for future research.

NATURAL HISTORY

Intracranial Saccular Aneurysm (ISA)

Two-to-five percent of people in the Industrialized World likely harbor an ISA, that is, a focal dilatation of a cerebral artery often characterized by a sac-like shape up to 30 mm in diameter (Figure 1). Histopathological and clinical studies reveal a higher prevalence in women (55– 65%) than men and that multiple lesions occur in 15–30% of aneurysm patients. Although these lesions may remain dormant for years, the small percentage that rupture tend to do so during the 5th to 7th decades of life (mean \sim 52 years old; 1). Ruptured ISAs are the leading cause of non-traumatic subarachnoid hemorrhage, and thus its devastating sequela (cerebral vasospasm), which causes significant morbidity and mortality.

The natural history of ISAs consists of three phases: pathogenesis, enlargement, and rupture. The genesis is a subject of considerable debate, but it is generally accepted that mechanical factors play fundamental roles. Cerebral arteries do not have an external elastic lamina, they have sparse medial elastin, they lack supporting perivascular tissue, and they have structural irregularities at the apex of their bifurcations (2,3). These factors appear to render these arteries susceptible to a local weakening under the persistent action of hemodynamic loads, particularly in hypertension (4). One theme, in particular, is the internal elastic lamina and muscular media must be markedly fragmented or degraded for an ISA to form (5,6). Risk factors include cigarette smoking, heavy alcohol consumption, and long-term use of analgesics or oral contraceptives, but these may play a lesser role (7,8). Increased familial incidence, up to 12% in some populations, suggests that genetics is also important. For example, cerebral arteries in patients harboring an ISA tend to have less type III collagen (9), and it has long been thought that a genetic defect renders these vessels more susceptible (10,11). Yet, gene studies have failed to link mutations in COL3A1 and aneurysms (12). Indeed, diverse studies have similarly not found a strong link to a mutation in any of a number of prime candidates, as, for example, genes for fibronectin, fibrillin-2, lysyl oxidase, α1-antitrypsin, matrix metalloproteinases (MMP-1,3,9,12), tissue inhibitors of MMPs (TIMP-1,2,3), endothelial derived nitric oxide synthase, or prostacyclin-stimulating factor (13–18).

ISAs enlarge from an initially small out-pouching of the wall, often to have diameters of 5 to 10 mm as well as complex shapes and composition. It was long thought that these lesions could enlarge rapidly due to structural instabilities, namely, a limit point instability or resonance (19–22). Whereas these studies assumed linear or rubber-like properties of the wall, more appropriate nonlinear analyses suggest that such instabilities are unlikely (23,24). Rather, ISAs appear to enlarge via mechano-regulated growth and remodeling (G&R) processes common to diverse vascular adaptations (25,26). That is, as in other arterial adaptations (e.g., responses to sustained alterations in blood flow, blood pressure, or axial stretch; 27–29), there is a significant increase in MMP activity, synthesis of extracellular matrix proteins, and apoptosis in ISAs (4,6,17,30–32). Indeed, Peters et al. (33) said it well: "aneurysmal dilation results in a highly dynamic cellular environment in which extensive wound healing and tissue/ extracellular matrix remodeling are taking place."

Allcock & Canham (34) reported large variations in "growth" rates: some lesions enlarged markedly over a few months while others remained nearly the same size for years. Based on a detailed statistical analysis, Mitchell & Jakubowski (35) suggested that ISAs go through a brief period of high growth and rupture-potential soon after formation, but those that survive may subsequently pose a much lower risk of rupture for an extended period. They estimated this high risk period to be less than 41 weeks, which is consistent with half-lives of arterial collagen (70 days or less in normalcy and much less in disease; 28). Note, therefore, that lesion strength is due primarily to fibrillar collagens, despite occasional small patches of smooth muscle of stellate form and dispersed fibroblasts, macrophages, and cellular debris (2,36). Kosierkiewicz et al. (37) identified an intimal-type thickening in some small ISAs and advanced plaques with lipid-laden macrophages in some large lesions, yet atherosclerosis appears to result from, not cause, ISA development. Rather enlargement appears to result primarily from the turnover of fibrillar collagens (including significant degradation by MMPs; 31–32), with orientation, cross-linking, and volume fractions being key factors. See work by Canham and colleagues on the layered orientation of aneurysmal collagen, which they suggest lies along great circle trajectories and renders lesions less distensible than the parent arteries $(38-40)$.

Rupture yields one of two outcomes: a catastrophic failure of a portion of the wall, with bleeding that is often lethal, or a transient leak, with less bleeding but clinical consequences nonetheless. Fortunately, the rate of rupture appears to be less than 0.1% per year. Failure mechanisms remain unknown, but rupture usually occurs at the fundus (pole) despite the neck often being thinner (2,41). Although this was long perplexing, finite element calculations suggest that the maximum biaxial stresses typically occur near the fundus, where intramural shear stress is small (23,42–43). For this reason, it appears that a rupture criterion should be based on tensile, not shearing, stresses or strains. Several studies associate other physical factors with rupture potential. Asari and Ohmoto (44) suggested that the combination of lesion location (e.g., middle cerebral artery), shape (i.e., multilobular), and excess loading (hypertension) indicates a high risk of rupture. Hademenos et al. (45) similarly suggested that multilobular lesions are more prone to rupture, but that less prevalent posterior lesions are more likely to rupture. Ruiz et al. (46) suggested further that aneurysms of the posterior communicating artery may be more prone to rupture because of increased nonsymmetrical contact with perianeurysmal tissue (they also suggested that symmetrical contact constraints may be protective, consistent with modeling predictions by 47). Despite differing opinions based on clinical observations, it is likely that the location of a lesion relative to the inlet flow stream is critical to all aspects of the natural history, including rupture potential, for this dictates intra-aneurysmal hemodynamics and associated mechanobiological responses (48). For example, heterogeneities in wall shear stress could locally alter collagen production (endothelin-1 is a promoter of collagen synthesis and nitric oxide is an inhibitor; 49) and give rise to a local weakening.

Notwithstanding these diverse factors, most studies still base rupture potential primarily on the maximum dimension, with estimates of the critical size ranging from 5 to 10 mm (e.g., 50, 51). From the perspective of mechanics, however, shape and wall thickness are more important determinants of stress in distended membranes than is overall size (23,43,52). Ujiie et al. (53) reported that 59% of ISAs are round, 24% oval, and 22% bar-like, but there has been little attempt to correlate rupture potential with specific shapes. That shape has not been considered is surprising since Crompton (54) showed long ago that saccular aneurysms in women tend to have a greater neck:height ratio and they are more likely to rupture. Lesion thickness can range from 30 to 500 μm in the unloaded configuration (55). It is believed that increasing thickness correlates primarily, but not exclusively, with continued enlargement (56).

Abdominal Aortic Aneurysm (AAA)

The normal adult human infrarenal aorta is approximately 12 cm long, 2 cm in diameter, and 2 mm thick. AAAs are typically defined by a 1.5 or more fold increase in diameter or simply a diameter greater than 3 cm; they tend to be diffuse, involving up to 9 cm of the infrarenal aorta (Figure 2). These lesions are six times more common in men, but three times more likely to rupture in women (57). Overall, AAAs affect 6 to 9% of people in the Industrialized World aged 65 and older. In addition to increasing age and male gender, risk factors include cigarette smoking, hypertension, chronic obstructive pulmonary disease, inflammation, and atherosclerosis. Some studies also suggest that up to 15% of the offspring of, and 13–32% of the siblings of, AAA patients also harbor a lesion (58). Of course, aging, smoking, hypertension, and atherosclerosis each correlate directly with increasing arterial wall stiffness (26), which in turn alters the hemodynamics. Like their intracranial counterparts, the natural history of AAAs includes pathogenesis, enlargement, and rupture.

Again, there is considerable debate over pathogenesis. Some suggest that AAAs initiate due to inflammation and a local weakening of the wall (e.g., early loss of elastin), which in turn leads to atherosclerotic involvement and intraluminal thrombus; others suggest that AAAs are a consequence of a ruptured atherosclerotic plaque and associated formation of a thrombus and neovascularization from the vasa vasorum. See Alexander (59), Silverstein et al. (57), Sakalihasan et al. (60), Choke et al. (61), Thompson (62), and references therein. Regardless, AAAs expand at rates up to 0.25 to 0.75 cm per year, initially slower, then faster. If untreated, many lesions continue to enlarge until rupture (e.g., 50% of untreated AAAs in patients with high surgical risk will rupture, often within 1 year; 63). Whereas patients with a ruptured anterolateral wall often succumb to sudden death, those with a ruptured posterolateral wall can survive to hospitalization. Overall, the rate of rupture of AAAs has been estimated at 3 to 9% per year, significantly higher than that for ISAs, with mortality between 65 and 85%.

The normal human abdominal aorta consists of 45.5% collagen (60% type I and 22% type III), 30.1% elastin, and 22% smooth muscle by dry weight (64,65). The layered media contains abundant smooth muscle cells delimited by 28 to 30 concentric elastic laminae, less than would be expected of an elastic artery of its wall thickness. It is thought that this structure, combined with augmented pressures due to pulse wave reflections, may render the abdominal aorta more susceptible to aneurysms. Regardless, AAAs tend to have up to 90% less elastin, much of which is fragmented, and concomitantly few smooth muscle cells (66). Note, therefore, that normal elastin attenuates smooth muscle migration, proliferation, and apoptosis (67–69), but potentially reparative elastogenesis appears to be ineffective (59), consistent with reports that structurally significant vascular elastin is produced primarily during development (28,70). Among others, Freestone et al. (71) suggest that as the media attenuates and the aorta dilates, the adventitia experiences a stress-induced thickening via a turnover of fibrillar collagen that reinforces the wall. Zarins et al. (72) suggested that such turnover may be a response to altered loads, not just a consequence of altered genetics. The combination of decreased elastin and

increased collagen and collagen cross-linking (66,73) increases wall stiffness and decreases distensibility to a degree that is evident clinically.

Whereas collagen turnover is likely a protective response to the loss of medial elastin and smooth muscle, localized imbalances between degradation and synthesis may be responsible in large part for eventual rupture (26,60). Key MMPs in AAAs are (59,74): MMP-1 (interstitial collagenase, acting primarily on fibrillar collagen), MMP-2 (gelatinase, acting primarily on elastin and cleaved collagen), MMP-9 (gelatinase, acting primarily on collagen IV), and MMP-3 (stromelysin, acting primarily on elastin). Although produced in latent forms and counteracted by TIMPS, MMPs are activated by plasmin, oxygen radicals, etc., and are significantly up-regulated in AAAs compared to normal aorta. Note, too, that MMP-9 is produced by inflammatory cells (e.g., macrophages and B-cells) that invade from the vasa vasorum or atherosclerotic intima.

Approximately 75% of AAAs have an intraluminal thrombus (61,75), yet the role of thrombus remains unclear. Stenbaek et al. (76) claim that rupture correlates best with the rate of growth of the thrombus and Kazi et al. (77) show dramatic differences in the portions of an AAA that are covered with a thrombus (e.g., thinner, less elastin, less smooth muscle, more T- and Bcells), yet Fillinger et al. (78) reported that the thickness of the thrombus does not correlate with rupture when based on matched overall diameters of the lesion. Nevertheless, thrombus appears to reduce overall wall stress slightly, it may be a barrier to the diffusion of oxygen and nutrients from the blood stream to the inner (avascular) portion of the wall, and it sequesters leukocytes and platelets that produce proteases and growth factors as well as control local levels of plasmin, which in turn activate MMPs (79,80). Hence, thrombus likely plays key chemomechanical roles in evolving AAAs and it must be understood and modeled better.

PRIOR BIOMECHANICAL ANALYSES

Intracranial Saccular Aneurysm (ISA)

Hemodynamics—There have been many in vitro experimental studies of fluid mechanics within and near an ISA, most using Newtonian fluids and rigid models of the arteries and aneurysms, with an emphasis on regions of elevated wall shear stress (e.g., 81,82). Alternatively, 3-D computational fluid dynamics (CFD) has been used increasingly to study cerebral blood flow, including flow in intracranial aneurysms (83–86). Indeed, there has been an important move to build computational models from in vivo medical imaging data in order to quantify patient-specific hemodynamics (87–95).

In contrast to the common hypothesis in experimental and CFD studies that locally elevated wall shear stress is important in pathogenesis, there has been no consensus as to which hemodynamic factors are important in enlargement or rupture. Cebral et al. (90) quantified flow velocity patterns in 62 patient-specific CFD models reconstructed from 3D rotational angiography and suggested that lesions most likely to rupture had complex or unstable flows and narrow jets and impingement sites. Increased flow complexity could be a sign of transition to turbulence and high-frequency pressure fluctuations, conditions thought to be responsible for post-stenotic dilatations (96). Alternatively, impingement due to a jet could cause steep gradients in wall shear stress and/or higher dynamic pressures on the wall. Yet, based on 26 patient-specific CFD simulations, Shojima et al. (97) reported that local rises in pressure due to flow impingement are less than 2 mmHg, which is small compared to nominal pressure levels in cerebral arteries, and concluded that dynamic pressures acting at bifurcations and on the walls of ISAs may be less significant to enlargement and rupture than previously assumed. In a longitudinal study of a basilar artery fusiform aneurysm, Acevedo-Bolton et al. (95) concluded that regions that continued to enlarge experienced low wall shear stress and

speculated that this might be due to increased residence time of "particles that degrade the aneurysm wall."

Finally, note that the wall shear stress in the cerebral vasculature (90,95) may normally be an order of magnitude higher than the 1 to 1.5 Pa value often cited as universal in large arteries (98). Of course, recent studies challenge the notion that wall shear stress is universally constant within large arteries within or across species (99,100). For example, Greve et al. (100) reported that wall shear stress τ_w in the abdominal aorta scales across mammalian species with body mass *M* according to the allometric relation $\tau_w \propto M^{-0.38}$. Thus, interpretation of what constitutes a "high" or "low" wall shear stress should be based on nominal values for that region, and care should be taken when interpreting data from animal models or cell culture studies.

Wall Mechanics—The biomechanics of ISAs has been studied using Laplace's equation: σ $= Pa/2h$, where σ is the uniform in-plane wall stress, *P* the transmural pressure, *a* the pressurized inner radius, and *h* the associated wall thickness. Examples range from Canham & Ferguson (101) who estimated a critical lesion diameter $d_c = (4\sigma_c V/\pi P)^{1/3} \sim 8.6$ mm, with σ_c the critical wall strength (1–10 MPa) and *V* the volume of wall to Meng et al. (102) who suggested that the rupture potential of a daughter aneurysm depends solely on two geometric parameters related to the size of its orifice. Laplace's relation has thus yielded useful insights, but it only holds for quasi-static inflations of spherical membranes and thereby cannot address elastodynamics, complex geometries, the propensity of rupture at the fundus, or how a lesion evolves. Conversely, early elasticity-based studies were limited by assumptions of linear or rubber-like behavior. In an attempt to address some of these shortcomings, Humphrey and colleagues began a series of nonlinear elasticity analyses in 1994 to analyze the then three primary biomechanical hypotheses: ISAs are materially unstable (21,22), they are dynamically unstable (19,20), and size alone is the critical predictor of rupture potential (7,50,54). Toward this end, one first needs information on the material behavior of human lesions.

Scott et al. (103) performed in vitro pressure-volume tests on 7 ISAs obtained at autopsy. Their data are fit well by a 2-D Fung-exponential strain energy function *w* (23):

$$
w = c(e^{Q} - 1), \text{ where } Q = c_1 E_{11}^2 + c_2 E_{22}^2 + 2c_3 E_{11} E_{22}, \tag{1}
$$

 c and c_i are material parameters, and E_{11} , E_{22} are principal values of the Green strain. Yet, Scott's data are global and essentially 1-D (i.e., pressure-volume), and thereby are not suitable to assess local properties or anisotropy. Whereas some investigators performed uniaxial stretching tests on strips of excised human aneurysms (5,56), Hsu et al. (104) developed a triplane video-based system to measure, in situ, local in-plane strains and curvatures at multiple pressures and Seshaiyer et al. (105) developed a sub-domain inverse finite element code (combining nonlinear finite elements with a Marquardt-Levenberg regression) to infer best-fit values of material parameters from these data. Based on data from 4 unruptured human lesions, they similarly found the Fung-exponential to be a reasonable descriptor, with typical values of the material parameters being $c = 12.6$ N/m, $c_1 = 10.74$, $c_2 = 13.08$, $c_3 = 11.02$ (which satisfy physical and mathematical constraints such as convexity). The meridional direction was often stiffer than circumferential, particularly away from the neck, thus suggesting a regional anisotropy.

Nonlinear finite element analyses (FEA) of idealized axisymmetric saccular aneurysms (23, 42) confirmed that Laplace's relation yields reasonable results only when lesions have a small neck and are nearly spherical. For this sub-class, however, one can prove analytically that lesions described by either Fung or Skalak-Tozeren-Zarda-Chien (STZC) type stress-strain relations cannot exhibit a limit point instability (23,106,107); this is in contrast to rubber-like

spherical membranes, which typically exhibit limit points (22). Because static instabilities tend to organize dynamic instabilities, it was not surprising that one can also show both analytically (given small perturbations about various equilibria) and numerically (using methods of nonlinear dynamics and a Runge-Kutta solution of a new nonlinear differential equation of the elastodynamics) that nearly spherical lesions tend to be dynamically stable (107,108). Hence, at least for this sub-class of ISAs, and probably more generally, neither quasi-static nor dynamic instabilities appear to be responsible for enlargement.

Recall from above that although a "critical diameter" remains controversial, lesion size continues to be one of the most common clinical metrics used to predict rupture potential. For axisymmetric membranes, however, the principal (i.e., meridional and circumferential) Cauchy stress resultants T_i can be found in closed form (109) and shown to depend directly on transmural pressure *P* and principal curvatures κ and κ_2 , not overall size:

$$
T_1 = \sigma_1 h = \frac{P}{2\kappa_2}, \qquad T_2 = \sigma_2 h = \frac{P}{\kappa_2} \left(1 - \frac{\kappa_1}{2\kappa_2} \right). \tag{2}
$$

Indeed, the continuing neglect of curvature is probably one reason why there is so much controversy over the elusive critical size (cf. 42,43). The significance of this general finding was illustrated, using FEA, for classes of axisymmetric lesions (23,42,47). For example, "small" lesions with a large neck:height ratio can have much higher stresses than "large" lesions with a small neck:height ratio; contact constraints due to interactions between a lesion and surrounding tissue can decrease the maximum stresses in an otherwise large lesion, however. Results also reveal that expected nonhomogeneous, non-equibiaxial distributions of stress in inflated, axisymmetric lesions can be restored to nearly homogeneous and equibiaxial (except for a boundary layer) by letting the material symmetry vary continuously from fundus to neck (25). These findings appear to be reasonable teleologically for they suggest that in response to local perturbations in properties (e.g., loss of elastin and smooth muscle), and thus geometry, ISAs can grow and remodel (via collagen turnover) so as to nearly recover the biaxial state of stress that was experienced by cells in the normal arterial wall. In summary, computations suggest that ISAs do not enlarge due to material or dynamic instabilities, and size is not the primary determinant of rupture potential. Rather, enlargement in response to an initial insult (first to elastin) appears to be yet another example of vascular G&R, one that seeks to alter structure locally so as to change the overall equilibrium geometry and thereby return stresses towards homeostatic values. Although idealized models are essential starting points, for they allow validation and they build intuition, there is a pressing need for geometrically and materially accurate finite element models of wall stress in saccular aneurysms. An important step in this direction was recently reported by Ma et al. (110).

Fluid-Solid Interactions—Following an earlier paper on the flow of a Newtonian fluid within 2-D, rigid saccular aneurysms, Low et al. (111) presented an important advance: a 3-D, fluid-solid interaction (FSI) model for two classes of lateral saccular aneurysms. Blood was modeled as a Casson fluid, the walls of the parent arteries and aneurysm were treated as linearly elastic, and flow was allowed to be unsteady. Among other conclusions, they found that "During the systolic acceleration phase where the aneurysm expands, inflow velocity near the downstream wall of the aneurysm is much higher in the distensible case, while at maximum flow rate, where the aneurysm already contracts, the inflow in the distensible case is significantly decreased." Given this initial finding on the importance of wall distensibility, it is surprising that most studies continue to focus on rigid wall models. This assumption precludes wave propagation phenomena and fundamentally changes the character of resultant solutions (112) as well as negating possible fluid-solid interactions.

The common prescription of a pressure field at the outlets of a computational model is likewise unrealistic when modeling wave propagation phenomena, especially for FSI studies or treatment planning where this pressure is unknown and part of the desired solution. Moreover, it is not feasible to prescribe pressure waveforms at each of the outlet boundaries in multibranched models since amplitude, shape, and phase depend on the solution in the modeled domain and cannot be assigned a priori. Consequences of the inattention to proper boundary conditions and wall deformability include uncertainty in global flow distributions, local velocity fields, and shear stresses. Indeed, although computed pressures are seldom reported, when they are reported the absolute values are inaccurate due to prescribed traction-free boundary conditions. For example, a number of papers on ISAs report elevated blood pressures within the lesion, yet closer inspection typically reveals values that differ only ~ 0.5 mmHg from those in nearby arteries. Such small differences would not be expected to be significant in the genesis and enlargement. Another unfortunate consequence of the inability to model well the pressure fields is, arguably, an over-emphasis on wall shear stress as the primary hemodynamic factor contributing to genesis, enlargement, and rupture. We conclude, therefore, that although the move to image-based, patient-specific models has yielded more realistic velocity and shear stress fields compared to idealized models, there has been little progress in measuring and then assigning physiologically realistic inlet flow waveforms, outlet boundary conditions, or wall properties.

Abdominal Aortic Aneurysms (AAA)

Hemodynamics—Many investigators have similarly used CFD to study blood flow in AAAs (e.g., 113–117). In many cases, regions of high pressure or high wall shear stress are singled out as potentially important to pathogenesis, though with little direct reference to the underlying mechanobiology. Bluestein et al. (118) pointed out that recirculation zones may also promote thrombus formation, which in turn may affect rupture potential. Overall, focus has been largely on pulsatile flows through rigid, idealized axisymmetric models, with blood modeled as an incompressible Newtonian fluid and outlet boundary conditions prescribed to be traction-free or constant pressure. Moreover, these studies do not include major branches immediately proximal to AAAs, such as the celiac, superior mesenteric, and renal arteries. These branches significantly influence conditions in the infrarenal aorta in normal subjects (119–121) and likely influence conditions in AAAs. In summary, while prior applications of CFD have increased our understanding of hemodynamics within AAAs, more realistic analyses will have to await more anatomically and physiologically accurate models and inclusion of wall properties. Progress towards addressing this latter limitation is discussed below following a brief review of mechanical properties of the aneurysmal wall.

Wall Mechanics—Despite general recognition of the importance of wall mechanics in the natural history of AAAs (e.g., 59,63,122,123), there have been few studies of biomechanical properties. Early studies focused on gross measures of structural stiffness that can be useful in clinical correlations, but not biomechanical analyses. For example, Sumner et al. (124), MacSweeney et al. (125), and Lanne et al. (126) measured the pressure-strain modulus, $E_p =$ *ds* (*p^s* − *pd*)/(*d^s* − *dd*), where *p* and *d* denote luminal pressure and inner diameter, respectively, and subscripts *d* and *s* denote diastolic and systolic. Values for this modulus range from 140 to 337 kPa in age-matched "normals" versus 313 to 504 kPa in AAAs. In contrast, He & Roach (65) reported 1-D stress-strain data for 8 AAAs and 8 age-matched controls (all axially oriented samples); although the nonlinear responses were interpreted within the context of a linearized strain, they showed that AAAs are stiffer than the normal aorta. Thubrikar et al. (127) reported similar findings, but also that the circumferential direction was stiffer than the axial direction and stiffness varied around the lesion: despite being thicker (e.g., 2.7 vs. 2.1 mm), posterior portions were less stiff than anterior or lateral portions. Raghavan & Vorp (128) performed 1- D tests on 69 human AAA specimens oriented along either the circumferential or the axial

direction. They quantified data separately using a strain energy function of the form: $W = c_1$ $(I_B-3) + c_2 (I_B-3)^2$, where $I_B = tr \mathbf{B} = tr(\mathbf{F} \cdot \mathbf{F}^T)$ and **F** is the deformation gradient. Mean values of the material parameters were $c_1 = 186$ kPa, $c_2 = 2700$ kPa in the circumferential and $c_1 =$ 169 kPa, *c2* = 1570 kPa in the axial direction. Because 1-D data are not sufficient for determining multiaxial relations, Vande Geest et al. (129) performed biaxial tests on 26 AAAs and 8 age-matched controls (mean ages 72 and 71, respectively). The highly nonlinear responses were described well by a 2-D phenomenological Choi-Vito strain energy function, namely

$$
W = c \left(\exp[c_1 E_{\theta\theta}^2] + \exp[c_2 E_{zz}^2] + \exp[2c_3 E_{\theta\theta} E_{zz}] - 3 \right)
$$
\n(3)

where E_{AB} are principal values of Green strain $\mathbf{E} = 0.5(\mathbf{F}^T \cdot \mathbf{F} - \mathbf{I})$. Mean values of the material parameters were $c = 0.14$ kPa, $c_1 = 238.5$, $c_2 = 208.2$, and $c_3 = 204.1$ for AAAs and $c = 0.32$ kPa, c_1 = 70.6, c_2 = 71.7, and c_3 = 64 for age-matched non-diseased aortic samples. These values confirm that AAAs are indeed stiffer and more anisotropic (stiffer in the circumferential direction) than the normal aorta.

There has been less attention to properties of the thrombus that often lines AAAs. DiMartino et al. (130) performed 1-D extension tests on 21 samples of thrombus obtained from 6 patients; the behavior was modeled as linearly elastic, despite finite strains, with a reported Young's modulus of 130 kPa. Wang et al. (131) reported the most comprehensive study of the biomechanical properties of intraluminal thrombus from human AAAs. Briefly, each thrombus was removed from surgically resected human lesions and tested uniaxially within 24 hrs. Data from 50 specimens (mean lesion diameter of 5.6 cm) from 14 patients (69 years old) revealed that the behavior is mildly nonlinear over large strains, nearly isotropic (circumferential vs. axial), but highly nonhomogeneous (tissue from the luminal region was stiffer and stronger than that from the middle region). The data were fit well by a strain energy function of the form:

$$
W = b_1 (II_B - 3) + b_2 (II_B - 3)^2 \tag{4}
$$

where $2II_B = (tr\mathbf{B})^2 - tr\mathbf{B}^2$. Mean values of the material parameters were $b_I = 30.4$ kPa and b_2 = 30.1 kPa for the luminal region and b_1 = 21.4 kPa and b_2 = 21.7 kPa for the middle region. Measured strengths were on the order of 510 kPa for the luminal and 260 kPa for the middle regions, respectively.

The importance of wall stress in dictating the fate of individual lesions has also been recognized for decades. Most stress analyses have assumed idealized geometries, and are based on Laplace's equation (132,133), axisymmetric membrane theory (134), or linear elastic finite element analyses (130,135–138). Values of the Young's modulus range from 1 to 5 MPa. Linear analyses are inappropriate, however, due to the inherent nonlinear material behavior and large strains. In recent years, there has been increasing effort to develop more appropriate nonlinear FEA. The best models available (139,140,141) employ patient-specific geometries and treat the aneurysmal wall as nonlinearly elastic, albeit isotropic, homogeneous, uniformly thin, and unchanging. These authors acknowledge that many of their assumptions need to be relaxed, particularly that the in vivo diastolic geometry was assumed to be the stress-free reference. Nevertheless, Fillinger and colleagues showed, by comparing simulations for lesions that ruptured versus those that did not (100+ patients), that peak maximum normal stresses over ~440 kPa correlate strongly with rupture potential. This is an important finding; it reveals that computational models have real potential to improve our predictions of rupture potential

(see the excellent review by Vorp (142) for more on the wall mechanics). In closing, it is noted that many investigators (e.g., 137,138,141) use the von Mises stress to assess the "maximum stress" in AAAs. The von Mises stress has utility in classical engineering analyses of ductile materials, which yield due to excessive shear stresses. Available data on aortic and other arterial tissue suggest, however, that maximum normal stresses, not shears, govern failure (e.g., 143– 145). This issue of the most appropriate failure criteria must be addressed better.

Fluid-Solid Interactions—There has been increasing interest in FSI modeling of AAAs, yet most studies assume the arterial wall, AAA, and thrombus are linearly elastic, homogeneous, isotropic, nearly incompressible, and unchanging (146–148). Perhaps the most sophisticated FSI model to date is that by Wolters et al. (149). They used a shear thinning model for blood and a large strain, neo-Hookean model for the lesion ($W = c_1 (I_B - 3)$, with $c_1 = 1$ MPa). Nevertheless, the wall was assumed to be homogeneous, isotropic, uniformly thin (2 mm), and unchanging; outflow conditions were traction-free and wall deformation resulted from a uniform pressure boundary condition prescribed on the lateral surface rather than from the hemodynamic simulation. As such, this method cannot be regarded as fully coupled, and in particular, pressure fluctuations induced by turbulent or transitional flows in the lesion cannot be incorporated using this approach. Finally, as in most hemodynamic studies of AAAs, proximal branches of the abdominal aorta were neglected and a uniform velocity was prescribed in the infrarenal aorta.

NEED FOR A NEW CLASS OF COMPUTATIONAL TOOLS

Based on this brief review of the literature, it is evident that computational biomechanical models have become increasingly sophisticated over the years and they have contributed significantly to our understanding of the natural history of aneurysms. There is a pressing need, however, for a new class of models that can describe the *evolving* geometry, nonlinear wall properties, and hemodynamics, which in turn dictate the evolving cell mechanobiology that is responsible for matrix turnover and the possible rupture of aneurysms. In contrast to using computational fluid dynamics (CFD) to study the hemodynamics or finite element analyses (FEA) to study wall stress, we submit that there is a need to couple fluid-solid interaction (FSI) models over a cardiac cycle with long-term growth and remodeling (G&R) models of the evolving wall (Figure 3). We shall refer to this new class of coupled computational tools for studying evolving vascular changes as Fluid-Solid-Growth (FSG) Models.

FSI During a Cardiac Cycle

Simulations of G&R of the vasculature require, as input, the mechanical forces (or tractions) that the fluid exerts on the wall. We contend that these tractions should be computed using FSI simulations, which in turn should be based on realistic anatomic models, refined discretizations to resolve complex hemodynamics, physiologically realistic fluid boundary conditions, appropriate models of wall stiffness, and robust, stable solvers for generally large systems of coupled fluid-solid equations. In particular, the only way to infer mechanisms of enlargement is to model actual geometries, hence we must exploit the tremendous advances in clinical magnetic resonance imaging or computed tomography. Toward this end, it is essential to image branches proximal and distal to a lesion because hemodynamic simulations can be very sensitive to inlet boundary conditions, unless the region of interest is many diameters from the inlets and there is sufficient dissipation in the system (150), and it can be difficult to prescribe outlet conditions where the flow remains disturbed from the complex geometry of the lesion.

Prakash & Ethier (151) discuss requirements for mesh generation and note that "suitable mesh convergence studies are rarely reported in the computational hemodynamics literature." Unfortunately, this statement also characterizes the state-of-the art in aneurysm hemodynamics. As a result, many complex features observed in experimental studies of

aneurysm flow (152,153) have not been replicated in computational simulations. Muller et al. (154) and Sahni et al. (155) recently described anisotropic adaptive finite element methods that are well suited for resolving complex, 3-D pulsatile flow dynamics as occur in aneurysms. A posteriori error estimators, based on the Hessian of the velocity magnitude (speed) field, yield directional information on the solution error. Because gradients in the velocity field are typically much greater in the radial than the circumferential direction in arterial flows, mesh generation strategies that exploit this feature can generate anisotropic meshes having an order of magnitude fewer degrees of freedom but results comparable to standard isotropic meshes (155). In summary, adaptive mesh generation can enable mesh-independent solutions for even the largest simulations of aneurysm hemodynamics given contemporary computing resources.

Simulations of fluid-solid interactions necessitate special considerations of inlet and outlet boundary conditions for both the velocity and the pressure fields need to be computed accurately to prescribe tractions on the vessel wall. Yet, prior simulations of hemodynamics in intracranial and abdominal aortic aneurysms have prescribed either zero outlet pressure (i.e., traction-free) conditions or, in a few cases, pressure waveforms at exits of the computational domain. We submit that impedance outlet boundary conditions are necessary for 3-D hemodynamic simulations to yield physiologic pressure and flow waves in distensible multivessel models. Decades of research show that vascular impedance can accurately represent the effects of a distal vascular bed (156–159). Impedance can be represented by a lumped parameter model (157) or computed using linear wave theory on randomly-branched distributed models (156). For example, Olufsen (160) showed that vascular impedance can be calculated efficiently on binary fractal tree models and provide appropriate outlet conditions for 1-D nonlinear wave propagation models. Vignon & Taylor (161) described a coupled multi-domain method based on Dirichlet-to-Neumann and variational multiscale methods that simplifies downstream domains and integrates them within image-based models as outlet boundary conditions. Vignon-Clementel et al. (162) extended this approach to include impedance outlet conditions for 3-D rigid-wall models whereas Figueroa et al. (163) described effects of various outlet conditions in 3-D FSI simulations. Findings show the effect of varying outlet conditions on flow, pressure, and displacement waveforms for a single artery model wherein a velocity profile is prescribed at the inlet. Only the impedance outlet boundary condition resulted in physiologic waveforms, noting in particular the unrealistic consequences of a prescribed constant outlet pressure on the upstream pressure and displacement waveforms.

Although models of the distal vasculature can be developed using distributed network or binary fractal tree approaches (156,160), it is preferable to use morphometric data to identify outlet boundary conditions. Such data are available for coronary and pulmonary circulations (164, 165), but there is a pressing need for morphometric data for the cerebral and abdominal aortic vasculatures. Until such data are available, distributed and lumped models should be tuned numerically by constricting or dilating the distal vascular beds until the overall level of pressure and distribution of flow in the image-based model matches available measurements. Note that one advantage of distributed models is that physiologic variations are modeled easily for example, effects of lower extremity exercise on abdominal aortic blood flow can be modeled by changing the inflow waveform, dilating the vessels in the lower extremities, and constricting the beds that supply the abdominal organs (166). Finally, note that methods to couple 3-D and 1-D computational models of blood flow with the aim of minimizing wave reflections from the outlet of the 3-D domain (167) are inadequate unless the 1-D model is itself terminated with an impedance outlet condition. In our experience, coupling the 3-D model directly to an impedance outlet boundary condition is simpler and yields excellent results for FSI simulations.

Efficient coupling of fluid and solid domains is also fundamental to successful FSI simulations. Standard FSI techniques, such as the arbitrary Lagrangian-Eulerian (ALE) method, remain problematic for realistic anatomic and physiologic models of large portions of the vasculature.

(6)

Hence consider a new method, the Coupled Momentum Method for Fluid Structure Interaction (CMM-FSI), which incorporates equations for the vascular wall at a variational level as a boundary condition for the fluid domain (163). Let three kinds of boundaries for an arbitrary geometry of a vessel be denoted as follows: Γ*g* represents an inflow surface where a velocity field is prescribed; Γ_h represents an outlet surface where either impedance or resistance is prescribed; and Γ*^s* represents the lateral surface of the fluid domain, which interacts with the vessel wall. Whereas a Dirichlet condition of zero displacement (i.e., no-slip) is typically prescribed on Γ*^s* in rigid wall models, this is no longer appropriate when dealing with

deformable vessels. Rather, letting Γ_s be subjected to a traction ${\bf t}_f^n$ originating from interactions with the vessel wall, the weak form of the governing equations for the fluid domain can be expressed as (163)

$$
\int_{\Omega} \left\{ \mathbf{w} \cdot (\rho \mathbf{v}_{n} + \rho \mathbf{v} \cdot \nabla \mathbf{v} - \mathbf{g}) + \nabla \mathbf{w} : (-p\mathbf{I} + \tau) - \nabla q \cdot \mathbf{v} \right\} d\mathbf{x} + \int_{\Gamma_{g}} q v_{n} ds + \int_{\Gamma_{h}} \left\{ -\mathbf{w} \cdot \mathbf{h} + q v_{n} \right\} d\mathbf{s} + \int_{\Gamma_{g}} \left\{ -\mathbf{w} \cdot \mathbf{t}_{f}^{n} + q v_{n} \right\} ds + \text{stabilization terms} = 0.
$$
\n(5)

Here, **v** and *p* are the fluid velocity and pressure (τ is the viscous Cauchy stress), **g** is a body force, **h** is a prescribed traction, and **w** and *q* are test functions. Details on the stabilization

terms can be found in Taylor et al. (119) and Figueroa et al. (163). The traction t_f^n is unknown a priori, but can be related to a fictitious body force governing the motion of the vessel wall using a thin-wall approximation similar to that used by Womersley (168), who coupled blood flow and wall motion in an analytical solution for pulsatile flow in an axisymmetric, deformable cylinder. Using this approach, one can write the term containing the unknown traction as a function of the wall stress and inertial terms whereby the governing equation for the fluid domain becomes

$$
\int_{\Omega} (\mathbf{w} \cdot (\rho \mathbf{v}_{n} + \rho \mathbf{v} \cdot \nabla \mathbf{v} - \mathbf{g}) + \nabla \mathbf{w} \cdot (-p\mathbf{I} + \tau) - \nabla q \cdot \mathbf{v}) d\mathbf{x} + \int_{\Gamma_g} q v_n ds - \int_{\Gamma_h} \mathbf{w} \cdot \mathbf{h} ds + \int_{\Gamma_h} q v_n ds + \zeta \int_{\Gamma_s} (\mathbf{w} \cdot \rho^s \mathbf{v}_{n} + \nabla \mathbf{w} \cdot \sigma(\mathbf{u})) ds - \zeta \int_{\partial \Gamma_s} \mathbf{w} \cdot \mathbf{h}^s dl + \int_{\Gamma_s} q v_n ds ++ \text{stabilization terms=0}.
$$

Boundary terms, defined on Γ*^s* , can then be expressed in terms of fluid variables velocity and pressure – using a 2-D model of the vessel wall (e.g., a membrane model enhanced with transverse shear) since fluid responses depend only on the structural stiffness of the wall. For example, if one assumes a linearly elastic response, wall stress **σ(u)**can be derived easily from linearized wall strains and the usual stiffness tensor ℂ.

It is essential, however, to account for finite wall strains in G&R (as well as those due to usual in vivo deformations from a reference configuration). Because of the aforementioned increased stiffness and decreased distensibility of aneurysms, one can incorporate information from nonlinear wall mechanics during G&R into FSI simulations over a cardiac cycle by exploiting a theory of "small deformations superimposed on large." Moreover, because timescales are so different between the cardiac cycle (seconds) and periods of G&R (days to years), one can solve the overall problem iteratively: determine tractions that are needed as inputs for the wall mechanics problem of finite deformation G&R by solving the hemodynamics problem assuming small deformations over a cardiac cycle, with each G&R solution providing an updated computational domain and structural stiffness of the wall for the next hemodynamic solution. Toward this end, Baek et al. (169) showed that complexities associated with an evolving aneurysm or artery (e.g., residual stress, nonlinear behavior, anisotropy, muscle tone, prior G&R, and finite deformations) can be so modeled. Let the position of a material point in

an artery or aneurysm be given by $\mathbf{x} = \mathbf{x}^0 + \mathbf{u}$, where \mathbf{x}^0 represents a finite motion from a suitable reference to any intermediate configuration between diastole and systole, and **u** represents a small additional displacement during the cardiac cycle. The total deformation gradient is $\mathbf{F} =$ $(I + H) \cdot F^{\circ}$, where $H = \partial u / \partial x^{\circ}$ is a small displacement gradient and F° is the initial finite deformation. It can be shown that the Cauchy stress due to the additional small deformation **σ** * is

$$
\sigma_{ij}^* = -p^* \delta_{ij} + \mathbb{C}_{ijkl} \varepsilon_{kl}^* + \mathbb{R}_{ijkl} \Omega_{kl}^*, \quad \mathbb{C}_{ijkl} = \delta_{ij} \widehat{\sigma}_{ij}^0 + \widehat{\sigma}_{il}^0 \delta_{jk} + 4F_{ik}^0 F_{jk}^0 F_{k}^0 F_{lQ}^0 \frac{\partial^2 W}{\partial C_{AB} \partial C_{PQ}}|_{C^0}
$$
\n(7)

where ϵ^* and Ω^* are the linearized strain and rotation tensors, \mathbb{C}_{ijkl} is the "stiffness" matrix, and the over-hat denotes an extra stress. Although this stress response function has a form similar to that in linearized elasticity, the stiffness depends explicitly on the initial finite deformations and stress, which can include residual stress and G&R, and a strain energy *W* that describes complex nonlinear, anisotropic behaviors at any point or any instant. Hence, one can use a fully nonlinear analysis for G&R, but pass updated values of C *ijkl* and ℝ *ijkl* to a usual FSI code to enable computationally efficient calculations of new hemodynamic loads for the adapting lesion and artery. We consider a method for finding appropriate strain energy functions for the wall in the next section. Finally, note that the variational FSI problem defined by equation (6) can be discretized using well-known, stabilized finite element techniques (e.g., 170,171) and the equations can be integrated in time using a modified version of the generalized-α method (172). This method provides an implicit, second-order accurate scheme with controllable numerical dissipation that shows excellent stability and has been applied to complex FSI problems with millions of degrees of freedom.

G&R Theory for Evolving Geometries and Material Properties

The last four decades have seen tremendous advances in vascular solid mechanics, from quantification of complex constitutive behaviors of tissues and cells to stress analyses of the same (26,173). Yet, most analyses have focused on geometries and properties at a particular instant, not how they evolve over time, and most constitutive relations have been phenomenological, describing overall behavior while assuming that the tissue or cell is materially uniform. An important advance, therefore, was the modeling of arterial development and adaptations to perturbed loads by Taber (174,175) and Rachev et al. (176,177). They employed the concept of kinematic growth introduced in 1981 by R. Skalak and showed that associated models can predict diverse manifestations of vessel growth. Two limitations, however, are the assumption that growth occurs in stress free configurations and that the tissue is materially uniform. Recall, for example, that the arterial wall consists of many different cell types and extracellular matrix proteins and proteoglycans, each of which can have individual material properties and rates of turnover in evolving, stressed configurations. A constrained mixture theory (178,179) can model evolving properties, turnover rates, and natural (i.e., stressfree) configurations of individual structural constituents that comprise a soft tissue, which is thereby modeled as materially nonuniform. Implementations of this G&R theory have predicted salient features of flow-, pressure-, and axial stretch-induced adaptations by carotid arteries (180–182) as well as enlargement of idealized intracranial aneurysms (183,184).

For illustrative purposes, consider an idealized thin-walled, axisymmetric aneurysm. Recall that intracranial aneurysms are largely devoid of elastin and smooth muscle, and thereby consist primarily of multiple families of collagen fibers. Because these fibers deform with the lesion, but are produced at different times and possibly with different orientations and natural configurations, they can be thought to constitute a constrained mixture (i.e., constituents co-

exist within neighborhoods over which continuum averaging holds and deform together despite having different natural configurations). Analogous to continuum theories of membranes (26,52), Cauchy membrane stresses, or tensions T_i (= $\sigma_i h$), for a thin-walled intracranial aneurysm can be computed at any time *t* via (in principal directions 1 and 2)

$$
T_1(t) \frac{1}{\lambda_2(t)} \frac{\partial w}{\partial \lambda_1} = \frac{1}{\lambda_2(t)} \frac{\partial}{\partial \lambda_1} \left(\sum_k w^k \right) = \frac{1}{\lambda_2(t)} \sum_k \frac{\partial w^k}{\partial \lambda^k(t)} \frac{\partial \lambda^k(t)}{\partial \lambda_1(t)},
$$

\n
$$
T_2(t) = \frac{1}{\lambda_1(t)} \sum_k \frac{\partial w^k}{\partial \lambda^k(t)} \frac{\partial \lambda^k(t)}{\partial \lambda_2(t)},
$$
\n(8)

where the strain energy function for the lesion (i.e., constrained mixture) $w = \sum w^k$ equals the sum of the energies stored in the *k* families of fibers, λ_1 and λ_2 are principal stretches experienced by the lesion, and

$$
\lambda^{k}(t) = \sqrt{\left(\lambda_1 \cos \alpha_o^{k}\right)^2 + \left(\lambda_2 \sin \alpha_o^{k}\right)^2}
$$
\n(9)

are stretches experienced by fibers in collagen family *k* relative to a common mixture reference configuration (with α_o^k the original angle between fiber family *k* and the 1-axis). To account for the constant deposition of new collagen fibers within evolving stressed configurations, it was assumed that there exists a target (homeostatic) deposition stretch $G_h^k(178)$, hence the stretch experienced by family *k*, relative to its unique natural configuration, is $\lambda_{n(\tau)}^k(t) = G_h^k \lambda^k(t) / \lambda^k(\tau)$, with *t* the current time and *τ* the past time at which family *k* was produced. Assuming locally parallel fibers within each family, a microstructurally-based constitutive relation for each family can be written in terms of the stored energy in a fiber

(185), $W^{k}(\lambda_{n(\tau)}^{k})$, which depends on stretches experienced by the fiber relative to its individual natural configuration rather than a reference configuration for the lesion as a whole. Constituents only contribute to the overall structural integrity of the lesion until they are degraded, however, thus constituent stored energies per area can be written as (184)

$$
w^{k}(t) = \frac{\rho^{k}(0)}{\rho} Q^{k}(t) W^{k}\left(\lambda_{n(0)}^{k}(t)\right) + \int_{0}^{t} \frac{m^{k}(\tau)}{\rho} q^{k}(t-\tau) W^{k}\left(\lambda_{n(\tau)}^{k}(t)\right) d\tau, \tag{10}
$$

where ρ^k (0) and Q^k (*t*) \in [0,1] represent the initial mass density, per unit area, and fraction of constituents produced prior to *t* = 0 that survive to current time *t*, and $m^k(\tau)$ and $q^k(\tau-\tau) \in$ [0,1] represent the mass density production rate and the fraction of constituents produced at time *τ*∈[0,*t*] that survives to time *t*. Of particular note, the mass density productions and survival functions can be related to mass balance relations for each constituent, which in turn can incorporate chemical reaction kinetics (e.g., matrix synthesis or proteolytic degradation) naturally (186). In this way, it may be possible to build multi-scale models wherein global hemodynamics can be related to local wall stress, which in turn can be coupled to molecularlevel biochemical reactions. Note, too, that the heredity integral contribution in equation (10) is similar to the concept of fading memory in nonlinear viscoelasticity; contributions to the overall structural integrity depend on the time since production. From equations (8)–(10), constituent principal Cauchy stress resultants are

 $T_1^k(t)$

 $T_2^k(t)$

$$
= \frac{1}{\lambda_2(t)} \left(\frac{\rho^k(0)Q^k(t)G_h^k}{\rho \lambda^k(0)} \frac{\partial W^k}{\partial \lambda_{n(0)}^k(t)} \frac{\partial \lambda^k(t)}{\partial \lambda_1(t)} + \int_0^t \frac{m^k(\tau)q^k(t-\tau)G_h^k}{\rho \lambda^k(\tau)} \frac{\partial W^k}{\partial \lambda_{n(\tau)}^k(t)} \frac{\partial \lambda^k(t)}{\partial \lambda_1(t)} d\tau \right),
$$

\n
$$
= \frac{1}{\lambda_1(t)} \left(\frac{\rho^k(0)Q^k(t)G_h^k}{\rho \lambda^k(0)} \frac{\partial W^k}{\partial \lambda_{n(0)}^k(t)} \frac{\partial \lambda^k(t)}{\partial \lambda_2(t)} + \int_0^t \frac{m^k(\tau)q^k(t-\tau)G_h^k}{\rho \lambda^k(\tau)} \frac{\partial W^k}{\partial \lambda_{n(\tau)}^k(t)} \frac{\partial \lambda^k(t)}{\partial \lambda_2(t)} d\tau \right).
$$
\n(11)

In the case of axisymmetry, the sum over all *k* constituents yields the stress resultants for the lesion, which by equilibrium are given by (cf. equation 2)

$$
T_1 = \frac{P}{2\kappa_2} = \sum_k T_1^k, \qquad T_2 = \frac{P}{\kappa_2} \left(1 - \frac{\kappa_1}{2\kappa_2} \right) = \sum_k T_2^k \tag{12}
$$

where the *κⁱ* are principal curvatures. In cases of non-axisymmetry, equations for the full stress resultant tensor **T** can be computed similarly, though typically only via finite elements. Moreover, the Cauchy stress can be computed from the Cauchy stress resultant tensor via the deformed thickness. Illustrative examples of lesion G&R, independent of fluid-solid coupling, can be found in Baek et al. (183,184); see, too, the recent study by Kroon and Holzapfel (187).

Although Feng et al. (188) and Chatziprodromou et al. (189) support a similar FSG approach in principle, they assume the wall is linearly elastic and model changes in wall properties by simply prescribing decreases in a Young's modulus that allow the wall to evolve. In contrast, we submit that there is a pressing need to embed rational growth and remodeling equations within a robust fluid-solid interaction model. This new paradigm was first presented by Figueroa et al. (190) and Baek et al. (191), but its full development must be our next focus. Indeed, although the overall theory is reasonably general, able to account for the evolution of properties, orientations, natural configurations, and mass fractions of multiple constituents, the primary challenge is to prescribe individual constitutive relations, particularly for the mass density productions, survival functions, and individual strain energy functions. Hence, there is further motivation for additional revealing experiments on both ISAs and AAAs.

CLOSURE

It is estimated that up to 12 million people harbor an intracranial aneurysm in the U.S.A. alone, over 30,000 of whom present yearly with a subarachnoid hemorrhage following rupture. It is expected that (a) advances in genetic research and a better understanding of risk factors will allow improved identification of patients at risk of developing saccular aneurysms and (b) advances in medical imaging will allow routine screening of high-risk patients, which together will result in a larger number of diagnosed unruptured lesions. Immediate prophylactic intervention for every diagnosed aneurysm will be unthinkable, however, because well less than 1% of these lesions rupture annually $(1,51)$, and this low yearly risk appears to remain nearly constant as the patient ages (35,192). Rather, there will be an increasingly greater need to predict better which lesions will likely rupture, and thus require immediate intervention, and which will allow periodic monitoring. Such a decision can be aided by rational decision analysis algorithms (e.g., 193), but we must first understand better the biomechanics of enlargement that precedes rupture. Advances in neurointerventional technologies will similarly demand a better understanding of lesion stability. For example, the decision whether to perform a clip ligation, coil embolization, stenting, or any of a number of future endovascular techniques could be based in part on an assessment of an individual lesion and an understanding of how

treatment may alter that lesion as well as the surrounding (adapting) vasculature. Computational FSG models have much to offer in this regard.

It is estimated that 175,000 people are diagnosed each year with an abdominal aortic aneurysm in the U.S.A. alone. Two current methods used to treat AAAs are (a) open surgical repair by replacing the diseased segment with a synthetic arterial graft or (b) endovascular repair by deploying a stent that is designed to shield the aneurysmal wall from the hemodynamic loads. Particularly provocative is the observation that some endovascularly treated AAAs actually regress, with decreases in size up to 8% (194,195). Given that an AAA actively grows and remodels during most of its natural history, this suggests that a stent can alter the applied loads in such a way that G&R may reverse the natural course of enlargement, which usually ends in rupture. Computational FSG models could help predict the rupture potential, help in the design of stents that exploit this advantageous regression, or predict if specific pharmacologic interventions could augment positive effects of a stent (59,60).

SUMMARY POINTS

- **1.** Precise mechanisms by which intracranial and abdominal aortic aneurysms initiate, enlarge, or rupture remain unknown, but biomechanical factors play fundamental roles. Indeed, assessment of rupture-potential should include the geometry (shape, thickness), properties (anisotropy, strength), and applied loads (hemodynamic, perivascular), not be based primarily on the maximum dimension.
- **2.** Following an early loss of elastin and subsequent loss of smooth muscle, a dynamic turnover of collagen can reinforce an aneurysm as it thins and enlarges; this turnover is controlled, in part, by cellular responses to changes in hemodynamic-induced wall shear and intramural stresses.
- **3.** Atherosclerosis and intraluminal thrombus appear to play key roles in the development of AAAs whereas they appear to be a consequence of the development of some ISAs. That AAAs tend to be less stable biologically than ISAs suggests that the chronic inflammation associated with atherosclerotic lesions and intraluminal thrombus can be critical to the natural history.

FUTURE ISSUES

- **1.** Continuing advances in vascular biology, medical imaging, biomechanics, and computational methods present a special opportunity to build integrated Fluid-Solid-Growth (FSG) Models that can account for 3-D patient-specific geometries, the full biofluid mechanics (shear and pressure), and the nonlinear mechanics of the wall (properties, biaxial stress), including cell-mediated changes in wall structure, properties, and geometry that will require multiscale modeling.
- **2.** There is a pressing need for appropriate data and constitutive functions to enable us to exploit theoretical and computational advances. For example, there is a pressing need for morphological data on distal resistance beds. Moreover, amongst other substances, it is known that endothelin-1 and transforming growth factor-beta increase collagen synthesis by smooth muscle cells whereas nitric oxide and tissue necrosis factor-alpha decrease collagen synthesis, yet there are no quantitative descriptors of these relationships.

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References

- 1. Juvela S. Natural history of unruptured intracranial aneurysms: risks for aneurysm formation, growth, and rupture. Acta Neurochir Suppl 2002;82:27–30. [PubMed: 12378985]
- 2. Stehbens WE. Pathology and pathogenesis of intracranial berry aneurysms. Neurol Res 1990;12:29– 34. [PubMed: 1970622]
- 3. Finlay HM, Whittaker P, Canham PB. Collagen organization in the branching region of human brain arteries. Stroke 1998;29:1595–1601. [PubMed: 9707199]
- 4. Inci S, Spetzler RF. Intracranial aneurysms and arterial hypertension: A review and hypothesis. Surg Neurol 2000;53:530–542. [PubMed: 10940419]
- 5. Toth M, Nadasy GL, Nyary I, Kerenyi T, Orosz M, Molnarka G, Monos E. Sterically inhomogeneous viscoelastic behavior of human saccular cerebral aneurysms. J Vasc Res 1998;35:345–355. [PubMed: 9789115]
- 6. Zhang B, Fugleholm K, Day LB, Ye S, Weller RO, Day INM. Molecular pathogenesis of subarachnoid hemorrhage. Int J Biochem Cell Biol 2003;35:1341–1360. [PubMed: 12798348]
- 7. Wiebers DO, et al. Unruptured intracranial aneurysms risk of rupture and risks of surgical intervention. International study of unruptured intracranial aneurysms investigators. N Engl J Med 1998;339:1725– 1733. [PubMed: 9867550]
- 8. Mitchell P, Gholkar A, Vindlacheruvu RR, Mendelow AD. Unruptured intracranial aneurysms: benign curiosity or ticking bomb? Lancet Neurol 2004;3:85–92. [PubMed: 14747000]
- 9. Kilic T, Sohrabifar M, Kurtkaya O, Yildirim O, Elmaci I, Gunel M, Pamir MN. Expression of structural proteins and angiogenic factors in normal arterial and unruptured and ruptured aneurysm walls. Neurosurg 2005;57:997–1007.
- 10. Neil-Dwyer G, Bartlett JR, Nicholls AC, Narcisi P, Pope FM. Collagen deficiency and ruptured cerebral aneurysms. J Neurosurg 1983;59:16–20. [PubMed: 6864273]
- 11. Hegedus K. Some observations on reticular fibers in the media of the major cerebral arteries. Surg Neurol 1984;22:301–307. [PubMed: 6205460]
- 12. Ruigrok YM, Rinkel GJE, Wijmenga C. Genetics of intracranial aneurysms. Lancet Neurol 2005;4:179–189. [PubMed: 15721828]
- 13. van den Berg JS, Limburg M, Pals G, Arwert F, Westerveld A. Type III collagen deficiency in a family with intracranial saccular aneurysms. Cerebrovas Dis 2001;11:92–94.
- 14. Krex D, Rohl H, Konig IR, Ziegler A, Schackert HK, Schackert G. Tissue inhibitor of metalloproteinases-1, -2, and -3 polymorphisms in a white population with intracranial saccular aneurysms. Stroke 2003;34:2817–2821. [PubMed: 14605322]
- 15. Yoneyama T, Kasuya H, Onda H, Akagawa H, Jinnai N, Nakajima T, Hori T, Inoue I. Association of positional and functional candidate genes FGF1, FBN2, and LOX on 5q31 with intracranial aneurysms. J Hum Genet 2003;48:309–314. [PubMed: 12750963]
- 16. Hofer A, Ozkan S, Hermans MJ, Kubassek N, Sitzer M, Burtscher J, Knopp U, Schoch B, Wanke I, Huebner F, Raabe A, Steinmetz H, Auburger G. Mutations in the lysyl oxidase gene not associated with intracranial aneurysm in central European families. Cerebrovas Dis 2004;18:189–193.
- 17. Kassam AB, Horowitz M, Chang Y-F, Peters D. Altered arterial homeostasis and cerebral aneurysms: A molecular epidemiology study. Neurosurg 2004;54:1450–1462.
- 18. Akagawa H, Kasuya H, Onda H, Yoneyama T, Sasahara A, Kim CJ, Lee JC, Yang TK, Hori T, Inoue I. Influence of endothelial nitric oxide synthase T-786C single nucleotide polymorphism on aneurysm size. J Neurosurg 2005;102:68–71. [PubMed: 15658098]
- 19. Simkins TE, Stehbens WE. Vibrational behavior of arterial aneurysms. Lett Appl Engr Sci 1973;1:85– 100.
- 20. Hung EJN, Botwin MR. Mechanics of rupture of cerebral saccular aneurysms. J Biomech 1975;8:385– 392. [PubMed: 1206041]
- 21. Austin GM, Schievink W, Williams R. Controlled pressure-volume factors in the enlargement of intracranial saccular aneurysms. Neurosurg 1989;24:722–730.
- 22. Akkas, N. Aneurysms as a biomechanical instability problem. In: Mosora, F., editor. Biomechanical Transport Processes. Plenum Press; 1990. p. 303-311.

- 23. Kyriacou SK, Humphrey JD. Influence of size, shape and properties on the mechanics of axisymmetric saccular aneurysms. J Biomech 1996;29:1015–1022. [PubMed: 8817368]Erratum, 30:761, 1997
- 24. David G, Humphrey JD. Further evidence for the dynamic stability of intracranial saccular aneurysms. J Biomech 2003;36:1143–1150. [PubMed: 12831740]
- 25. Ryan JM, Humphrey JD. Finite element based predictions of preferred material symmetries in saccular aneurysms. Ann Biomed Engr 1999;27:641–647.
- 26. Humphrey, JD. Cardiovascular Solid Mechanics: Cells, Tissues, and Organs. Springer; Verlag, NY: 2002.
- 27. Gibbons GH, Dzau VJ. The emerging concept of vascular remodeling. Mech of Disease 1994;330 (20):1431–1438.
- 28. Langille BL. Arterial remodeling: relation to hemodynamics. Can J Physiol Pharmacol 1996;74:834– 841. [PubMed: 8946070]
- 29. Jackson ZS, Gotlieb AI, Langille BL. Wall tissue remodeling regulates longitudinal tension in arteries. Circ Res 2002;90:918–925. [PubMed: 11988494]
- 30. Mimata C, Kitaoka M, Nagahiro S, Iyama K, Hori H, Yoshioka H, Ushio Y. Differential distribution and expressions of collagens in the cerebral aneurysmal wall. Acta Neuropathol 1997;94:197–206. [PubMed: 9292688]
- 31. Bruno G, Todor R, Lewis I, Chyatte D. Vascular extracellular matrix remodeling in cerebral aneurysms. J Neurosurg 1998;89:431–440. [PubMed: 9724118]
- 32. Gaetani P, Tartara F, Grazioli V, Tancioni F, Infuso L, Baena RR. Collagen cross-linkage, elastolytic and collagenolytic activities in cerebral aneurysms: A preliminary investigation. Life Sci 1998;63:285–292. [PubMed: 9698037]
- 33. Peters DG, Kassam AB, Feingold E, Heidrich-O'Hare E, Yonas H, Ferrell RE, Brufsky A. Molecular anatomy of an intracranial aneurysm. Coordinated expression of genes involved in wound healing and tissue remodeling. Stroke 2001;32:1036–1042. [PubMed: 11283408]
- 34. Allcock JM, Canham PB. Angiographic study of the growth of intracranial aneurysms. J Neurosurg 1976;45:617–612. [PubMed: 978239]
- 35. Mitchell P, Jakubowski J. Estimate of the maximum time interval between formation of cerebral aneurysm and rupture. J Neurol Neurosurg Psychiatry 2000;69:760–767. [PubMed: 11080228]
- 36. Lang ER, Kidd M. Electron microscopy of human cerebral aneurysms. J Neurosurg 1965;22:554– 562. [PubMed: 5319276]
- 37. Kosierkiewicz TM, Factor SM, Dickson DW. Immunocytochemical studies of atherosclerotic lesions of cerebral berry aneurysms. J Neuropath Exp Neurol 1994;53:399–406. [PubMed: 8021714]
- 38. Canham PB, Finlay HM, Tong SY. Stereological analysis of the layered structure of human intracranial aneurysms. J Microsc 1996;183:170–180. [PubMed: 8805828]
- 39. Canham PB, Finlay HM, Kiernan JA, Ferguson GG. Layered structure of saccular aneurysms assessed by collagen birefringence. Neurol Res 1999;21:618–626. [PubMed: 10555180]
- 40. MacDonald DJ, Finlay HM, Canham PB. Directional wall strength in saccular brain aneurysms from polarized light microscopy. Ann Biomed Eng 2000;28:533–542. [PubMed: 10925951]
- 41. Sekhar LN, Heros RC. Origin, growth and rupture of saccular aneurysms: A review. Neurosurg 1981;8:248–260.
- 42. Shah AD, Harris JL, Kyriacou SK, Humphrey JD. Further roles of geometry and properties in saccular aneurysm mechanics. Comp Meth Biomech Biomed Engr 1997;1:109–121.
- 43. Ma B, Harbaugh RE, Raghavan ML. Three-dimensional geometrical characterization of cerebral aneurysms. J Biomech 2004;32:264–273.
- 44. Asari S, Ohmoto T. Growth and rupture of unruptured cerebral aneurysms based on the intraoperative appearance. Acta Med Okayama 1994;48:257–262. [PubMed: 7863797]
- 45. Hademenos GJ, Massoud TF, Turjman F, Sayre JW. Anatomical and morphological factors correlating with rupture of intracranial aneurysms in patients referred for endovascular treatment. Neuroradiol 1998;40:755–760.
- 46. Ruiz DSM, Yilmaz H, Dehdashti AR, Alimenti A, de Tribolet N, Rufenacht DA. The perianeurysmal environment: Influence on saccular aneurysm shape and rupture. Am J Neuroradiol 2006;27:504– 512. [PubMed: 16551985]

- 47. Seshaiyer P, Humphrey JD. On the potentially protective role of contact constraints on saccular aneurysms. J Biomech 2001;34:607–612. [PubMed: 11311701]
- 48. Hassan T, Timofeev EV, Satto T, Shimizu H, Ezura M, Matsumoto Y, Takayama K, Tominaga T, Takahashi A. A proposed parent vessel geometry-based categorization of saccular intracranial aneurysms: computational flow dynamics analysis of the risk factors for lesion rupture. J Neurosurg 2005;103:662–680. [PubMed: 16266049]
- 49. Rodriguez-Feo JA, Sluijter JPG, de Kleijn DPV, Pasterkamp G. Modulation of collagen turnover in cardiovascular disease. Curr Pharmacol Design 2005;11:2501–2514.
- 50. Ujiie H, Sato K, Onda H, Oikkawa A, Kagawa M, Atakakura K, Kobayashi N. Clinical analysis of incidentally discovered unruptured aneurysms. Stroke 1993;24:1850–1856. [PubMed: 8248967]
- 51. Wiebers DO, et al. Unruptured intracranial aneurysms: natural history, clinical outcome, and risks of surgical and endovascular treatment. Lancet 2003;362:103–110. [PubMed: 12867109]
- 52. Humphrey JD. Computer methods in membrane biomechanics. Comp Meth Biomech Biomed Engr 1998;1:171–210.
- 53. Ujiie H, Tamano Y, Sasaki K, Hori T. Is the aspect ratio a reliable index for predicting the rupture of a saccular aneurysm? Neurosurg 2001;48:495–502.
- 54. Crompton MR. The pathology of ruptured middle-cerebral aneurysms with special reference to differences between the sexes. Lancet 1962;2:421–425. [PubMed: 13882322]
- 55. Suzuki J, Ohara H. Clinicopathological study of cerebral aneurysms. J Neurosurg 1978;48:505–514. [PubMed: 632875]
- 56. Steiger HJ, Aaslid R, Keller S, Reulen HJ. Strength, elasticity and viscoelastic properties of cerebral aneurysms. Heart Vessels 1986;5:41–46. [PubMed: 2584177]
- 57. Silverstein MD, Pitts SR, Chaikof EL, Ballard DJ. Abdominal aortic aneurysm (AAA) : Costeffectiveness of screening, surveillance of intermediate-sized AAA, and management of symptomatic AAA. BUMC Proceedings 2005;18:345–367. [PubMed: 16252027]
- 58. Davies MJ. Aortic aneurysm formation Lessons from human studies and experimental models. Circ 1998;98:193–195.
- 59. Alexander JJ. The pathobiology of aortic aneurysms. J Surg Res 2004;117:163–175. [PubMed: 15013727]
- 60. Sakalihasan N, Limet R, Defawe OD. Abdominal aortic aneurysm. Lancet 2005;365:1577–1589. [PubMed: 15866312]
- 61. Choke E, Cockerill G, Wilson WRW, Sayed S, Dawson J, Loftus I, Thompson MM. A review of biological factors implicated in abdominal aortic aneurysm rupture. Eur J Vasc Endovasc Surg 2005;30:227–244. [PubMed: 15893484]
- 62. Thompson RW. Aneurysm treatments expand. Nat Med 2005;11:1279–1281. [PubMed: 16333264]
- 63. Fillinger MF, Racusin J, Baker RK, Cronenwett JL, Teutelink A, Schermerhorn ML, Zwolak RM, Powell RJ, Walsh DB, Rzucidlo EM. Anatomic characteristics of ruptured abdominal aortic aneurysm on conventional CT scans: Implications for rupture risk. J Vasc Surg 2004;39:1243–1252. [PubMed: 15192565]
- 64. Fisher GM, Llaurado JG. Collagen and elastin content in canine arteries selected from functionally different vascular beds. Circ Res 1966;19:394–399. [PubMed: 5914851]
- 65. He CM, Roach MR. The composition and mechanical properties of abdominal aortic aneurysms. J Vasc Surg 1994;20:6–13. [PubMed: 8028090]
- 66. Carmo M, Colombo L, Bruno A, Corsi FRM, Roncoroni L, Cuttin MS, Radice F, Mussini E, Settembrini PG. Alteration of elastin, collagen and their cross-links in abdominal aortic aneurysms. Eur J Vasc Endovasc Surg 2002;23:543–549. [PubMed: 12093072]
- 67. Krettek A, Sukhova GK, Libby P. Elastogenesis in human arterial disease: A role for macrophages in disordered elastin synthesis. Arterioscler Thromb Vasc Biol 2003;23:582–587. [PubMed: 12615674]
- 68. Brooke BS, Bayes-Genis A, Li DY. New insights into elastin. Trends Cardiovasc Med 2003;13:176– 183. [PubMed: 12837579]
- 69. Mithieux SM, Weiss AS. Elastin. Adv Protein Chem 2005;70:437–461. [PubMed: 15837523]

- 70. Stenmark KR, Mecham RP. Cellular and molecular mechanisms of pulmonary vascular remodeling. Ann Rev Physiol 1997;59:89–144. [PubMed: 9074758]
- 71. Freestone T, Turner RJ, Coady A, Higman DJ, Greenhalgh RM, Powell JT. Inflammation and matrix metalloproteinases in the enlarging abdominal aortic aneurysm. Arterioscler Thromb Vasc Biol 1995;15:1145–1151. [PubMed: 7627708]
- 72. Zarins CK, Weisenberg E, Kolettis G, Stankunavicius R, Glagov S. Differential enlargement of artery segments in response to enlarging atherosclerotic plaques. J Vasc Surg 1988;7:386–394. [PubMed: 3346952]
- 73. Sobolewski K, Wolanska M, Bankowski E, Gacko M, Glowinski S. Collagen, elastin and glycosaminoglycans in aortic aneurysms. Acta Biochimica Polonica 1995;42:301–308. [PubMed: 8588480]
- 74. Lijnen HR. Plasmin and matrix metalloproteinases in vascular remodeling. Thromb Haemost 2001;86:324–333. [PubMed: 11487021]
- 75. Swedenborg J, Kazi M, Eriksson P, Hedin U. Influence of the intraluminal thrombus in abdominal aortic aneurysms. Acta Chir Belg 2004;104:606–608. [PubMed: 15663260]
- 76. Stenbaek J, Kalin B, Swedenborg J. Growth of thrombus may be a better predictor of rupture than diameter in patients with abdominal aortic aneurysms. Eur J Vasc Endovasc Surg 2000;20:466–469. [PubMed: 11112467]
- 77. Kazi M, Thyberg J, Religa P, Roy J, Eriksson P, Hedin U, Swedenborg J. Influence of intraluminal thrombus on structural and cellular composition of abdominal aortic aneurysm wall. J Vasc Surg 2003;38:1283–1292. [PubMed: 14681629]
- 78. Fillinger MF, Racusin J, Baker RK, Cronenwett JL, Teutelink A, Schermerhorn ML, Zwolak RM, Powell RJ, Walsh DB, Rzucidlo EM. Anatomic characteristics of ruptured abdominal aortic aneurysm on conventional CT scans: Implications for rupture risk. J Vasc Surg 2004;39:1243–1252. [PubMed: 15192565]
- 79. Vorp DA, Lee PC, Wang DHJ, Makaroun MS, Nemoto EM, Ogawa S, Webster MW. Association of intraluminal thrombus in abdominal aortic aneurysm with local hypoxia and wall weakening. J Vasc Surg 2001;34:291–299. [PubMed: 11496282]
- 80. DiMartino ES, Vorp DA. Effect of variation in intraluminal thrombus constitutive properties on abdominal aortic aneurysm wall stress. Ann Biomed Engr 2003;31:804–809.
- 81. Lieber BB, Stancampiano AP, Wakhloo AK. Alteration of hemodynamics in aneurysm models by stenting: Influence of stent porosity. Ann Biomed Engr 1997;25:460–469.
- 82. Tateshima S, Murayama Y, Villablanca JP, Morino T, Nomura K, Tanishita K, Vinuela F. In vitro measurement of fluid induced wall shear stress in unruptured cerebral aneurysms harboring blebs. Stroke 2003;34:187–192. [PubMed: 12511772]
- 83. Nichita CC, Lieber BB, Pitman EB. Numerical simulation of flow in a stented and nonstented cerebral arterial segment with a side wall aneurysm using the immersed boundary technique. Annl Biomed Engr 2000;28:61.
- 84. Jou LD, Quick CM, Young WL, Lawton MT, Higashida R, Martin A, Saloner D. Computational approach to quantifying hemodynamic forces in giant cerebral aneurysms. Am J Neuroradiol 2003;24:1804–10. [PubMed: 14561606]
- 85. Hoi Y, Meng H, Woodward SW, Bendok BR, Hanel RA, Guterman LR, Hopkins LN. Effects of arterial geometry on aneurysm growth: three-dimensional computational fluid dynamics study. J Neurosurg 2004;101:676–681. [PubMed: 15481725]
- 86. Stuhne GR, Steinman DA. Finite-element modeling of the hemodynamics of stented aneurysms. J Biomech Engr 2004;126:382–387.
- 87. Oshima M, Torii R, Kobayashi T, Taniguchi N, Takagi K. Finite element simulation of blood flow in the cerebral artery. Comp Meth Appl Mech Engr 2001;191:661–671.
- 88. Cebral JR, Castro MA, Soto O, Lohner R, Alperin N. Blood-flow models of the circle of Willis from magnetic resonance data. J Engr Math 2003;47:369–386.
- 89. Cebral JR, Hernandez M, Frangi A, Putman C, Pergolesi R, Burgess J. Subject-specific modeling of intracranial aneurysms. Proceedings of SPIE Intern Soc Optical Engr 2004;5369:319–327.
- 90. Cebral JR, Castro MA, Burgess JE, Pergolizzi RS, Sheridan MJ, Putman CM. Characterization of cerebral aneurysms for assessing risk of rupture by using patient-specific computational hemodynamics models. Am J Neuroradiol 2005;26:2550–2559. [PubMed: 16286400]
- 91. Steinman CA, Milner JS, Norley CJ, Lownie SP, Holdsworth DW. Image-based computational simulation of flow dynamics in a giant intracranial aneurysm. Am J Neuroradiol 2003;24:559–566. [PubMed: 12695182]
- 92. Shojima M, Oshima M, Takagi K, Torii R, Hayakawa M, Katada K, Morita A, Kirino T. Magnitude and role of wall shear stress on cerebral aneurysm. Stroke 2004;35:2500–2505. [PubMed: 15514200]
- 93. Jou L-D, Dispensa B, Lawton M, Higashida R, Young W, Saloner D. Correlation Between Lumenal Geometry Changes and Hemodynamics in Fusiform Intracranial Aneurysms. AJNR Am J Neuroradiol 2005;26:2357–63. [PubMed: 16219845]
- 94. Moore S, David T, Chase JG, Arnold J, Fink J. 3D models of blood flow in the cerebral vasculature. J Biomech 2006;39:1454–1463. [PubMed: 15953607]
- 95. Acevedo-Bolton G, Jou L-D, Dispensa BP, Lawton MT, Higashida RT, Martin AJ, Young WL, Saloner D. Estimating the hemodynamic impact of interventional treatments of aneurysms: Numerical simulation with experimental validation: Technical case report. Neurosurg 2006;59:429– 430.
- 96. Boughner D, Roach M. Effect of low frequency vibration on the arterial wall. Circ Res 1971;29:136– 144. [PubMed: 5105817]
- 97. Shojima M, Oshima M, Takagi K, Torii R, Nagata K, Shirouzu I, Morita A, Kirino T. Role of the bloodstream impacting force and the local pressure elevation in the rupture of cerebral aneurysms. Stroke 2005;36:1933–8. [PubMed: 16081858]
- 98. Kamiya A, Togawa T. Adaptive regulation of wall shear stress to flow change in the canine carotid artery. Am J Physiol 1980;239:H14–H21. [PubMed: 7396013]
- 99. Langille BL. Remodeling of developing and mature arteries: endothelium, smooth muscle, and matrix. J Cardiovasc Pharmacol 1993;21:S11–S17. [PubMed: 7681126]
- 100. Greve JM, Les AS, Tang BT, Draney Blomme MT, Wilson NM, Dalman RL, Pelc NJ, Taylor CA. Allometric scaling of wall shear stress from mice to humans: quantification using cine phasecontrast MRI and computational fluid dynamics. Am J Physiol 2006;291:H1700–1708.
- 101. Canham PB, Ferguson GG. A mathematical model for the mechanics of saccular aneurysms. Neurosurg 1985;17:291–295.
- 102. Meng H, Feng Y, Woodward SH, Bendok BR, Hanel RA, Guterman LR, Hopkins LN. Mathematical model of the rupture mechanism of intracranial saccular aneurysms through daughter aneurysm formation and growth. Neurol Res 2005;27:459–465. [PubMed: 15978170]
- 103. Scott S, Ferguson GG, Roach MR. Comparison of the elastic properties of human intracranial arteries and aneurysms. Can J Physiol and Pharmacol 1972;50:328–332. [PubMed: 5038350]
- 104. Hsu FPK, Liu AMC, Downs J, Rigamonti D, Humphrey JD. A triplane video-based experimental system for studying axisymmetrically inflated biomembranes. IEEE Trans Biomed Engr 1995;42:442–450.
- 105. Seshaiyer P, Shah AD, Kyriacou SK, Humphrey JD. Multiaxial mechanical behavior of human saccular aneurysms. Comp Meth Biomech Biomed Engr 2001;4:281–289.
- 106. Humphrey JD, Kyriacou SK. The use of Laplace's equation in aneurysm mechanics. Neurol Res 1996;18:204–208. [PubMed: 8837052]
- 107. Haslach HW, Humphrey JD. Dynamics of biological soft tissue or rubber: Internally pressurized spherical membranes surrounded by a fluid. Int J Nonlin Mech 2003;39:399–420.
- 108. Shah AD, Humphrey JD. Finite strain elastodynamics of saccular aneurysms. J Biomech 1999;32:593–599. [PubMed: 10332623]
- 109. Hsu FPK, Schwab C, Rigamonti D, Humphrey JD. Identification of response functions for nonlinear membranes via axisymmetric inflation tests: Implications for biomechanics. Int J Sol Struct 1994;31:3375–3386.
- 110. Ma B, Lu J, Harbaugh RE, Raghavan ML. Nonlinear anisotropic stress analysis of anatomically realistic cerebral aneurysms. J Biomech Engr 2007;129:88–96.
- 111. Low M, Perktold K, Raunig R. Hemodynamics in rigid and distensible saccular aneurysms: A numerical study of pulsatile flow conditions. Biorheol 1993;30:287–298.

- 112. Nichols, W.; O'Rourke, MF. Theoretical, experimental and clinical principles. Vol. 4. Lea & Febiger; Philadelphia: 1998. McDonald's Blood Flow in Arteries.
- 113. Perktold K. On the paths of fluid particles in an axisymmetrical aneurysm. J Biomech 1987;20:311– 317. [PubMed: 3584155]
- 114. Taylor TW, Yamaguchi T. Three-dimensional simulation of blood flow in an abdominal aortic aneurysm: Steady and unsteady flow cases. J Biomech Engr 1994;116:89–97.
- 115. Yu SC, Chan WK, Ng BT, Chua LP. A numerical investigation on the steady and pulsatile flow characteristics in axisymmetric abdominal aortic aneurysm models with some experimental conditions. J Med Eng Technol 1999;23:228–239. [PubMed: 10738687]
- 116. Finol EA, Amon CH. Blood flow in abdominal aortic aneurysms: Pulsatile flow hemodynamics. ASME J Biomech Engr 2001;123:474–484.
- 117. Finol EA, Keyhani K, Amon CH. The effect of asymmetry in abdominal aortic aneurysms under physiologically realistic pulsatile flow conditions. ASME J Biomech Engr 2003;125:207–217.
- 118. Bluestein D, Niu L, Schoephroerster RT, Dewanjee MK. Steady flow in an aneurysm model: Correlation between fluid dynamics and blood platelet deposition. J Biomech Engr 1996;118:280– 294.
- 119. Taylor CA, Hughes TJR, Zarins CK. Finite Element Modeling of 3-dimensional Pulsatile Flow in the Abdominal Aorta: Relevance to Atherosclerosis. Annl Biomed Engr 1998;26:1–13.
- 120. Taylor CA, Hughes TJR, Zarins CK. Effect of exercise on hemodynamic conditions in the abdominal aorta. J Vasc Surg 1999;29:1077–89. [PubMed: 10359942]
- 121. Tang BT, Cheng CP, Draney MT, Wilson NM, Tsao PS, Herfkens RJ, Taylor CA. Abdominal aortic hemodynamics in young healthy adults at rest and during lower limb exercise: Quantification using image-based computer modeling. Am J Physiol 2006;291:H668–H676.
- 122. Miller FJ. Aortic aneurysms It's all about the stress. Arterioscler Thromb Vasc Biol 2002;22:1948– 1949. [PubMed: 12482817]
- 123. Vorp DA, Vande Geest JP. Biomechanical determinants of abdominal aortic aneurysm rupture. Arterioscler Thromb Vasc Biol 2005;25:1558–1566. [PubMed: 16055757]
- 124. Sumner DS, Hokanson DE, Strandness DE. Stress-strain characteristics and collagen-elastin content of abdominal aortic aneurysms. Surg Gyn Ob 1970:459–466.
- 125. MacSweeney STR, Young G, Greenhalgh RM, Powell JT. Mechanical properties of the aneurysmal aorta. Br J Surg 1992;79:1281–1284. [PubMed: 1486417]
- 126. Lanne T, Sonesson B, Bergqvist D, Bengtsson H, Gustafsson D. Diameter and compliance in the male human abdominal aorta: Influence of age and aortic aneurysm. Eur J Vasc Surg 1992;6:178– 184. [PubMed: 1572458]
- 127. Thubrikar MJ, Labrosse M, Robicsek F, Al-Soudi J, Fowler B. J Med Eng Technol 2001;25:133– 142. [PubMed: 11601439]
- 128. Raghavan ML, Vorp DA. Toward a biomechanical tool to evaluate rupture potential of abdominal aortic aneurysm: identification of a finite strain constitutive model and evaluation of its applicability. J Biomech 2000;33:475–482. [PubMed: 10768396]
- 129. Vande Geest JP, Sacks MS, Vorp DA. The effects of aneurysm on the biaxial mechanical behavior of human abdominal aorta. J Biomech 2006;39:1324–1334. [PubMed: 15885699]
- 130. Di Martino E, Mantero S, Inzoli F, Melissano G, Astore D, Chiesa R, Fumero R. Biomechanics of abdominal aortic aneurysm in the presence of endoluminal thrombus: Experimental characterisation and structural static computational analysis. Eur J Vasc Endovasc Surg 1998;15:290–299. [PubMed: 9610340]
- 131. Wang DHJ, Makaroun M, Webster MW, Vorp DA. Mechanical properties and microstructure of intraluminal thrombus from abdominal aortic aneurysm. J Biomech Engr 2001;123:536–539.
- 132. McGiffin DC, McGiffin PB, Galbraith AJ, Cross RB. Aortic wall stress profile after repair of coarctation of the aorta. J Thorac Cardiovasc Surg 1992;104:924–931. [PubMed: 1405691]
- 133. Marston WA, Criado E, Baird CA, Keagy BA. Reduction of aneurysm pressure and wall stress after endovascular repair of abdominal aortic aneurysm in a canine model. Ann Vasc Surg 1996;10:166– 173. [PubMed: 8733869]

- 134. Elger DF, Blackketter DM, Budwig RS, Johansen KH. The influence of shape on the stresses in model abdominal aortic aneurysms. J Biomed Engr 1996;118:326–332.
- 135. Stringfellow MM, Lawrence PF, Stringfellow RG. The influence of aorta-aneurysm geometry upon stress in the aneurysm wall. J Surg Res 1987;42:425–433. [PubMed: 3573768]
- 136. Mower WR, Baraff LJ, Sneyd J. Stress distributions in vascular aneurysms: Factors affecting risk of aneurysm rupture. J Surg Res 1993;55:155–161. [PubMed: 8412094]
- 137. Inzoli F, Boschetti F, Zappa M, Longo T, Fumero R. Biomechanical factors in abdominal aortic aneurysm rupture. Eur J Vasc Surg 1993;7:667–674. [PubMed: 8270069]
- 138. Vorp DA, Wang DHJ, Webster MW, Federspiel WJ. Effect of intraluminal thrombus thickness and bulge diameter on the oxygen diffusion in abdominal aortic aneurysm. J Biomech Engr 1998;120:579–583.
- 139. Fillinger, Raghavan ML, Marra SP, Croonenwett JL, Kennedy FE. In vivo analysis of mechanical wall stress and abdominal aortic aneurysm rupture risk. J Vasc Surg 2002;36:589–597. [PubMed: 12218986]
- 140. Fillinger MF, Marra SP, Raghavan ML, Kennedy FE. Prediction of rupture risk in abdominal aortic aneurysm during observation: Wall stress versus diameter. J Vasc Surg 2003;37:724–732. [PubMed: 12663969]
- 141. Raghavan ML, Fillinger MF, Marra SP, Naegelein BP, Kennedy FE. Automated methodology for determination of stress distribution in human abdominal aortic aneurysm. J Biomech Engr 2005;127:868–871.
- 142. Vorp D. Biomechanics of abdominal aortic aneurysm. J Biomech 2007;40:1887–1902. [PubMed: 17254589]
- 143. Mohan M, Melvin JW. Failure properties of passive human aortic tissue. II. Biaxial tension tests. J Biomech 1983;16:31–44. [PubMed: 6833308]
- 144. Lee MC, Haut RC. Strain rate effects on tensile failure properties of the common carotid artery and jugular veins of ferrets. J Biomech 1992;25:925–927. [PubMed: 1639836]
- 145. Purslow PP. Positional variations in fracture toughness,, stiffness, and strength of descending thoracic pig aorta. J Biomech 1983;16:947–953. [PubMed: 6654923]
- 146. Di Martino ES, Guadagni G, Fumero A, Ballerini G, Spirito R, Biglioli P, Redaelli A. Fluid-structure interaction within realistic three-dimensional models of the aneurysmatic aorta as a guidance to assess the risk of rupture of the aneurysm. Med Engr Phys 2001;23:647–655.
- 147. Li Z, Kleinstreuer C. A new wall stress equation for aneurysm-rupture prediction. Annl Biomed Engr 2005;33:209–213.
- 148. Scotti CM, Shkolnik AD, Muluk SC, Finol EA. Fluid-structure interaction in abdominal aortic aneurysms: Effects of asymmetry and wall thickness. BioMed Engr Online 2005;4:1–22.
- 149. Wolters BJBM, Rutten MCM, Schurink GWH, Kose U, de Hart J, van de Vosse FN. A patientspecific computational model of fluid-structure interaction in abdominal aortic aneurysms. Med Engr Phys 2005;27:871–883.
- 150. Moyle K, Mallinson G, Cowan B. Volumetric methods for evaluating irreversible energy losses and entropy production with application to bioengineering flows. Int J Num Meth in Flds 2006;50:1357– 1368.
- 151. Prakash S, Ethier CR. Requirements for mesh resolution in 3D computational hemodynamics. J Biomech Engr 2001;123:134–144.
- 152. Peattie RA, Riehle TJ, Bluth EI. Pulsatile flow in fusiform models of abdoiminal aortic aneurysms: flow fields, velocity patterns and flow-induced wall stresses. J Biomech Engr 2004;126:438–46.
- 153. Salsac AV, Sparks SR, Lasheras JC. Hemodynamic changes occurring during the progressive enlargement of abdominal aortic aneurysms. Ann Vasc Surg 2004;18:14–21. [PubMed: 14712379]
- 154. Muller J, Sahni O, Li X, Jansen KE, Shephard MS, Taylor CA. Anisotropic adaptive finite element method for modelling blood flow. Comp Meth Biomech Biomed Engr 2005;8:295–305.
- 155. Sahni O, Muller J, Jansen KE, Shephard MS, Taylor CA. Efficient anisotropic adaptive discretization of the cardiovascular system. Comp Meth Appl Mech Engr 2006;195:5634–5655.
- 156. Taylor MG. Wave transmission through an assembly of randomly branching elastic tubes. Biophys J 1966;6:697–716. [PubMed: 5972372]

- 157. Westerhof N, Bosman F, De Vries CJ, Noordergraaf A. Analog studies of the human systemic arterial tree. J Biomech 1969;2:121–43. [PubMed: 16335097]
- 158. Avolio AP. Multi-branched model of the human arterial system. Med Biol Engr Comput 1980;18:709–718.
- 159. Stergiopulos N, Young DF, Rogge TR. Computer simulation of arterial flow with applications to arterial and aortic stenoses. J Biomech 1992;25:1477–1488. [PubMed: 1491023]
- 160. Olufsen MS. Structured tree outflow condition for blood flow in larger systemic arteries. Am J Physiol 1999;276:H257–268. [PubMed: 9887040]
- 161. Vignon IE, Taylor CA. Outflow Boundary Conditions for One-Dimensional Finite Element Modeling of Blood Flow and Pressure Waves in Arteries. Wave Motion 2004;39:361–374.
- 162. Vignon-Clementel IE, Figueroa CA, Jansen KJ, Taylor CA. Outflow boundary conditions for finite element modeling of blood flow and pressure in arteries. Comp Meth Appl Mech Engr 2006;195:3776–3796.
- 163. Figueroa CA, Vignon-Clementel IE, Jansen KE, Hughes TJR, Taylor CA. A coupled momentum method for modeling blood flow in three-dimensional deformable arteries. Comp Meth Appl Mech Engr 2006;195:5685–5706.
- 164. Kassab GS, Rider CA, Tang NJ, Fung YC. Morphometry of pig coronary arterial trees. Am J Physiol 1993;265:H350–365. [PubMed: 8342652]
- 165. Huang W, Yen RT, McLaurine M, Bledsoe G. Morphometry of the human pulmonary vasculature. J Appl Physiol 1996;81:2123–2133. [PubMed: 8941537]
- 166. Steele BN, Olufsen M, Taylor CA. Fractal network model for simulating abdominal and lower extremity blood flow during resting and exercise conditions. Comp Meth Biomech Biomed Engr 2007;10:39–51.
- 167. Formaggia L, Gerbeau JF, Nobile F, Quarteroni A. On the coupling of 3D and 1D Navier-Stokes equations for flow problems in compliant vessels. Comp Meth Appl Mech Engr 2001;191:561– 582.
- 168. Womersley JR. Oscillatory motion of a viscous liquid in a thin walled elastic tube I: the linear approximation for long waves. Phil Mag 1955;7:199–221.
- 169. Baek S, Gleason RL, Rajagopal KR, Humphrey JD. Theory of small on large for computing fluidsolid interactions in arteries. Comp Meth Applied Mech Engr 2006;196:3070–3078.
- 170. Brooks AN, Hughes TJR. Streamline upwind/Petrov-Galerkin formulations for convection dominated flows with particular emphasis on the incompressible Navier-Stokes equations. Comp Meth Appl Mech Engr 1982;32:199–259.
- 171. Hughes TJR, Franca LP, Hulbert GM. A new finite element formulation for fluid dynamics: VIII. The Galerkin/Least-Squares method for advective-diffusive equations. Comp Meth Appl Mech Engr 1989;73:173–189.
- 172. Jansen KE, Whiting CH, Hulbert GM. A generalized alpha-method for integrating the filtered Navier-Stokes equations with a stabilized finite element method. Comp Meth Appl Mech Engr 2000;190:305–319.
- 173. Fung, YC. Biomechanics: Mechanical Properties of Living Tissues. Springer; Verlag, NY: 1993.
- 174. Taber LA. Biomechanics of growth, remodeling, and morphogenesis. Appl Mech Rev 1995;48:487– 545.
- 175. Taber LA. A model of aortic growth based on fluid shear and fiber stresses. ASME J Biomech Engr 1998;120:348–354.
- 176. Rachev A, Stergiopulos N, Meister J-J. A model for geometric and mechanical adaptation of arteries to sustained hypertension. ASME J Biomech Engr 1998;120:9–17.
- 177. Rachev A. A model of arterial adaptation to alterations in blood flow. J Elast 2000;61:83–111.
- 178. Humphrey JD, Rajagopal KR. A constrained mixture model for growth and remodeling of soft tissues. Math Model Meth Appl Sci 2002;12:407–430.
- 179. Humphrey JD, Rajagopal KR. A constrained mixture model for arterial adaptations to a sustained step-change in blood flow. Biomech Model Mechanobiol 2003;2:109–126. [PubMed: 14586812]
- 180. Gleason RL, Taber LA, Humphrey JD. A 2-D model of flow-induced alterations in the geometry, structure and properties of carotid arteries. ASME J Biomech Engr 2004;126:371–381.

- 181. Gleason RL, Humphrey JD. A mixture model of arterial growth and remodeling in hypertension: Altered muscle tone and tissue turnover. J Vas Res 2004;41:352–363.
- 182. Gleason RL, Humphrey JD. Effects of a sustained extension on arterial growth and remodeling: A theoretical study. J Biomech 2005;38:1255–1261. [PubMed: 15863110]
- 183. Baek S, Rajagopal KR, Humphrey JD. Competition between radial expansion and thickening in the enlargement of an intracranial saccular aneurysm. J Elast 2005;80:13–31.
- 184. Baek S, Rajagopal KR, Humphrey JD. A theoretical model of enlarging intracranial fusiform aneurysms. ASME J Biomech Engr 2006;128:142–149.
- 185. Lanir Y. Constitutive equations for fibrous connective tissues. J Biomech 1983;16:1–12. [PubMed: 6833305]
- 186. Slattery, JC. Momentum, Energy, and Mass Transfer in Continua. Krieger Publishing; NY: 1981.
- 187. Kroon M, Holzafpel GA. A model for saccular cerebral aneurysm growth by collagen fibre remodeling. J Theor Biol 2007;247:775–787. [PubMed: 17482213]
- 188. Feng Y, Wada S, Tsubota K-I, Yamaguchi T. The application of computer simulation in the genesis and development of intracranial aneurysms. Tech & Health Care 2005;13:281–291.
- 189. Chatziprodromou I, Tricoli A, Poulikakos D, Ventikos Y. Haemodynamics and wall remodeling of a growing cerebral aneurysm: A computational model. J Biomech 2007;40:412–426. [PubMed: 16527284]
- 190. Figueroa, CA.; Baek, S.; Vignon-Clementel, IE.; Humphrey, JD.; Taylor, CA. Towards patientspecific modeling I. Hemodynamics in a growing aneurysm. SPIE Meeting; San Diego, CA. 2006. Conference Proceeding CD
- 191. Baek, S.; Figueroa, CA.; Taylor, CA.; Humphrey, JD. Towards patient-specific modeling II. Biomechanics of a growing aneurysm. SPIE Meeting; San Diego, CA. 2006. Conference Proceeding CD
- 192. Juvela S, Porras M, Heiskanen O. Natural history of unruptured intracranial aneurysms: a long term follow-up study. J Neurosurg 1993;79:174–182. [PubMed: 8331397]
- 193. van Crevel H, Habbema JDF, Braakman R. Decision analysis of the management of incidental intracranial saccular aneurysms. Neurol 1986;36:1335–1339.
- 194. Matsumura JS, Pearce WH, McCarthy WJ, Yao JS. Reduction in aortic aneurysm size: Early results after endovascular graft placement. EVT investigators. J Vasc Surg 1997;25:113–123. [PubMed: 9013914]
- 195. Bertges DJ, Chow K, Wyers MC, Landsittel D, Frydrych AV, Stavropoulos W, Tan WA, Rhee RY, Fillinger MF, Fairman RM, Makaroun MS. Abdominal aortic aneurysm size regression after endovascular repair is endograft dependent. J Vasc Surg 2003;37:716–723. [PubMed: 12663968]

Figure 1.

Computational model of a large portion of the human circle of Willis including an intracranial saccular aneurysm. Shown too are estimated velocity distributions based on a FSI simulation.

Figure 2.

Computational model of a large portion of the human infrarenal aorta including an abdominal aortic aneurysm. Shown too are estimated velocity distributions based on a FSI simulation of both rest and exercise.

HEMODYNAMICS

BIOMECHANICS

Figure 3.

Schema of the proposed Fluid-Solid-Growth (FSG) modeling approach: fluid-solid interaction (FSI) computations of the hemodynamics (time scale of seconds) provide updated information on wall tractions for the biosolid mechanics computations of G&R (time scale of days to weeks or years), which in turn provide updated information on changing geometry and material properties for the FSI code. The solution iterates between the hemodynamics and wall mechanics until either the vasculature achieves a steady state or a vessel or lesion ruptures. Modified from Figueroa et al. (190).