

IDH1 mutations are present in the majority of common adult gliomas but rare in primary glioblastomas

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We screened exon 4 of the gene isocitrate dehydrogenase 1 (NADP+), soluble (*IDH1*) for mutations in 596 primary intracranial tumors of all major types. Codon 132 mutation was seen in 54% of astrocytomas and 65% of oligodendroglial tumors but in only 6% of glioblastomas (3% of primary and 50% of secondary glioblastomas). There were no mutations in any other type of tumor studied. While mutations in the tumor protein p53 gene (*TP53*) and total 1p/19q deletions were mutually exclusive, *IDH1* mutations were strongly correlated with these genetic abnormalities. All four types of mutant *IDH1* proteins showed decreased enzymatic activity. The data indicate that *IDH1* mutation combined with either *TP53* mutation or total 1p/19q loss is a frequent and early change in the majority of oligodendroglial tumors, diffuse astrocytomas, anaplastic astrocytomas, and secondary glioblastomas but not in primary glioblastomas. *Neuro-Oncology* 11, 341–347, 2009 (Posted to *Neuro-Oncology* [serial online], Doc. D08-00331, May 12, 2009. URL <http://neuro-oncology.dukejournals.org>; DOI: 10.1215/15228517-2009-025)

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A recent report indicated that the gene isocitrate dehydrogenase 1 (NADP+), soluble (*IDH1*) is somatically mutated predominantly among younger patients with glioblastomas and secondary glioblastomas (sGBs) compared with primary glioblastomas.¹ sGBs arise by progression of an astrocytic tumor of lower malignancy grade (a diffuse astrocytoma WHO grade II [A] or an anaplastic astrocytoma WHO grade III [AA]), while primary (or de novo) glioblastomas (pGB) have no known precursor lesions. Thus, the question arises of whether A and AA also show this mutation or whether it is involved in progression to sGB. Glioblastomas (GB) are the most malignant as well as the most common and, consequently, the best studied of the astrocytic tumors.² Many genetic abnormalities have been identified in GBs, including genes in the RB1 pathway (*CDKN2A/B*, *CDK4*, *CDK6*, *RB1*), the p53 pathway (*p14ARF*, *MDM2*, *MDM4*, *TP53*), the Akt pathway (*PTEN*, *PIK3CA*), and epithelial growth factor receptor gene *EGFR* (for review, see Collins³ and references therein). While these cellular pathways are almost always disrupted in both pGBs and sGBs, the frequency with which a particular gene is targeted differs among these GB types.⁴ Much less is known about other types of gliomas and other types of primary brain tumors. Rare mutations of the gene cyclin-dependent kinase inhibitor 2C (*CDKN2C* [*p18INK4C*]) or the tumor protein p53 gene (*TP53*) have been found in oligodendroglial tumors.⁵ Combined total codeletion of 1p and 19q, generally as a result of an unbalanced t(1;19) (q10;p10) translocation, is a common event among oligodendroglomas.⁶ However, the molecular consequences

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of this translocation are unknown. We hypothesized that *IDH1* mutations might be associated with A and/or AA, or even with a wider range of primary brain tumors, and carried out mutation screening of *IDH1* in a series of 596 intracranial primary tumors and 15 glioma cell lines. We found that *IDH1* is frequently mutated in A and AA as well as in all oligodendroglial tumor types. Among the astrocytic tumors, pilocytic astrocytomas (PAs) and pGBs are exceptions. We also demonstrate that mutated *IDH1* proteins showed decreased enzymatic activity.

Materials and Methods

Tumor Materials, DNA Extraction, and Array-Comparative Genomic Hybridization

The histopathological diagnosis was made according to WHO criteria.² The primary tumors consisted of 305 astrocytic tumors (including 38 PA, 22 A, 62 AA, 183 GB), 97 oligodendroglial tumors (including 34 oligodendrogliomas [O], 20 anaplastic oligodendrogliomas [AO], 20 oligoastrocytomas [OA], and 23 anaplastic oligoastrocytomas [AOA]), and 50 ependymal tumors (ependymoma WHO grade II, anaplastic ependymoma WHO grade III, subependymoma WHO grade I, and

myxopapillary ependymoma WHO grade I), as well as 144 other intracranial tumors: 36 medulloblastomas, 8 primitive neuroectodermal tumors (PNETs), 4 dysembryoplastic neuroepithelial tumors (DNETs), 48 vestibular schwannomas, and 48 meningiomas (see Table 1). Collection, handling, and DNA/RNA extraction of tumor tissues and the patients' blood samples were as described previously.⁷ The study was approved by the Ethical Committee of the Karolinska Hospital (no. 91:16), Sahlgrenska University Hospital (S339:01), and Cambridge Research Ethics Committee (Cambridge, UK; NRES Cambridgeshire 2 REC reference 03/115). In addition, 15 established glioma cell lines (U87MG, U118MG, U138MG, U178MG, U251MG, U373MG, TP265MG, TP365MG, TP483MG, TP276MG, TP336MG, T98G, H4, CCF-STTG1, A172) were included in the study.

Genetic Analysis

The genomewide copy number data were determined by comparative genomic hybridization on a 1-Mb array, as previously described.^{8,9} The details of the 1-Mb array data will be published elsewhere (K.I., unpublished data). Primers for *IDH1* exon 4 were according to Parsons et al.¹ Sequencing was performed as previously described.¹⁰ Mutation analysis of all other genes—*TP53*, retinoblas-

Table 1. Summary of genetic abnormalities

Tumor Type	WHO Grade	No. Cases	<i>IDH1</i> Mutation		<i>TP53</i> Mutation		Total 1p/19q Deletion	
Gliomas		452	119		142		54	
Astrocytomas		305						
Pilocytic astrocytoma	I	38	0		ND		0	
Diffuse astrocytoma	II	22	13	59%	11	50%	3	14%
Anaplastic astrocytoma	III	62	32	52%	42	68%	3	5%
Glioblastoma	IV	183	11	6%	65	36%	3	2%
Primary glioblastoma		173	6	3%	59	34%	3	2%
Secondary glioblastoma		10	5	50%	6	60%	0	
Oligodendrogliomas		97						
Oligodendroglioma	II	34	23	68%	2	6%	26	76%
Anaplastic oligodendroglioma	III	20	12	60%	3	15%	12	60%
Oligoastrocytoma	II	20	10	50%	8	40%	1	5%
Anaplastic oligoastrocytoma	III	23	18	78%	11	48%	6	26%
Ependymomas		50						
Ependymoma	II	23	0		ND		0	
Anaplastic ependymoma	III	7	0		ND		0	
Subependymoma	I	12	0		ND		0	
Myxopapillary ependymoma	I	8	0		ND		0	
Other brain tumors		144	0					
Medulloblastoma	IV	36	0		ND		0	
PNET	IV	8	0		ND		0	
DNET	I	4	0		ND		0	
Vestibular schwannoma	I	48	0		ND		ND	
Meningioma	I	48	0		ND		ND	
Glioma cell lines		15	0		10	67%	0	

Abbreviations: PNET, primitive neuroectodermal tumor; DNET, dysembryoplastic neuroepithelial tumor; ND, not determined.

toma 1 (*RB1*), phosphatase and tensin homolog (*PTEN*), and cyclin-dependent kinase inhibitor 2A (*CDKN2A*)—was performed as previously described.^{7,11}

IDH1 Enzymatic Assay

A wild-type *IDH1* expression vector construct with a 5' hemagglutinin (HA) tag was generated by amplifying cDNA synthesized from U251 cells (*IDH1* wild type; see below) using primers PC5779 (TAGAATTCCAC-CATGGCTTACCACATACGATGTTCCAGATTACGC-TATGTCCAAAATAATCATGTGGC) and PC5778 (GTATGAATTCAAAGTTGGCCTGAGCTAG) and subcloning the PCR product into pTracer-CMV2 (Invitrogen, Paisley, UK). All four *IDH1* mutants identified in this study (see below) were generated by site-directed mutagenesis using a QuikChange XL site-directed mutagenesis kit followed by confirmatory sequencing (Agilent Technologies, Santa Clara, CA, USA). COS-7 kidney cells were transfected with the constructs using Lipofectamine 2000 (Invitrogen), harvested 48 h post-transfection, and lysed with the CellLytic M reagent (Sigma-Aldrich, Gillingham, UK). The supernatant was used for both the enzymatic assay and Western blotting. The *IDH1* enzymatic assay was performed according to the manufacturer's recommendation (Sigma-Aldrich). Briefly, the substrate mix (42 mM glycylglycine buffer, 0.44 mM D,L-isocitric acid, 1 mM β -nicotinamide adenine dinucleotide phosphate [NADP], 0.6 mM MnSO₄) was preincubated at 37°C for 10 min. The cell lysate equivalent to 5 μ g total protein was then added to the premix, and the amount of NADPH produced was monitored by spectrophotometry at 340 nm at intervals for 25 min. The enzymatic activity was determined as $\Delta A_{340\text{nm}}/\text{min}$. Western blotting was performed as previously described¹⁰ using a rabbit anti-HA polyclonal antibody (Abcam, Cambridge, UK), a goat anti-*IDH1* monoclonal antibody (Santa Cruz Biotechnology, Santa Cruz, CA, USA), and a mouse anti- α -tubulin monoclonal antibody (Sigma-Aldrich).

Statistical Analysis

Statistical analyses were performed using Statistical Package for Social Sciences (SPSS) version 16.0 (SPSS, Inc., Chicago, IL, USA). Significance of correlations between the parameters was assessed using either the chi-square test with Yates's correction or two-tailed Fisher's exact test. The log-rank test was used for univariate analysis, and Cox regression was used for multivariate analysis of potential association between *IDH1* mutations and overall patient survival.

Results

The entire exon 4 and adjacent intronic sequences of *IDH1* were sequenced in the 596 intracranial primary tumors and 15 glioma cell lines. Mutations were confirmed as somatic by sequencing matched normal DNA from peripheral white blood cells from individual

patients in selected cases. In total, 119 somatic mutations were identified. All mutations were missense mutations at codon 132 (arginine). Of these, 110 were c.395G>A (Arg132His), one c.394C>A (Arg132Ser), four c.394C>G (Arg132Gly), and four c.394C>T (Arg132Cys) (examples of the mutations are shown in Supplementary Fig. 1S). With the exception of six tumors (five with deletion of one allele and one with gain; data not shown), 113 of 119 mutations were associated with normal copy number at *IDH1* (2q33), and one retained the wild-type allele. No *IDH1* mutations were identified in any of the 17 matched-blood DNA from patients with *IDH1*-mutated tumors. Matched normal brain tissues surrounding tumors that harbored *IDH1* mutations in two cases (O24, OA9) showed no mutation, indicating that *IDH1* mutations were present only in glioma cells but not in the nonneoplastic cells (Supplementary Fig. 1S).

The prevalence of mutations in each tumor type is shown in Table 1. No PA had an *IDH1* mutation. A and AA showed high frequencies of mutations (59% and 52%), while GBs had a very low mutation rate overall (6%). However, when divided into primary and secondary GBs, *IDH1* mutations were found in only 6 of 173 (3%) of pGB, whereas they were present in 5 of 10 (50%) of sGBs ($p < 0.001$). Xenografts established from one pGB (GB181) retained the same mutation (c.395G>A). A majority of the four subtypes of oligodendroglial tumors (O, AO, OA, and AOA) also had *IDH1* mutations: 23 of 34 O (68%), 12 of 20 AO (60%), 10 of 20 OA (50%), and 18 of 23 AOA (78%). No mutations were found among the 50 ependymal tumors, 36 medulloblastomas, 8 supratentorial PNETs, 4 DNETs, 48 vestibular schwannomas, or 48 meningiomas. None of the 15 established glioma cell lines had *IDH1* mutations. The details of the *IDH1* mutations in individual tumors are presented in Supplementary Table 1S.

A majority of A, AA, and sGB tumors but only about 30% of pGBs are known to have *TP53* mutations, while a majority of O, AO, OA, and AOA are known to have total 1p/19q loss and very unusual *TP53* mutations.^{7,12} When all these tumors were included in one group (A, AA, GB, O, AO, OA, and AOA; total, 364 tumors), *TP53* mutations were present in 142 and combined total 1p/19q loss was present in 54 tumors (Table 1). The *TP53* mutations and total 1p/19q loss were, with the exception of three AOs, mutually exclusive (Supplementary Table 1S). When we looked for correlations between these two completely different genomic abnormalities and *IDH1* mutations, we found that the vast majority of *IDH1* mutations (109/119, 92%) were associated with *TP53* mutation, total 1p/19q loss, or both ($p < 0.001$; including two of the three AOs that had both *TP53* mutations and total 1p/19q loss). Only 10 tumors with *IDH1* mutations had neither *TP53* mutations nor total 1p/19q loss. However, 77 (54%) of the tumors with *TP53* mutations had no abnormality of *IDH1* (1 A, 14 AA, 1 sGB, 56 pGB, 1 O, 1 AO, 1 OA, and 2 AOA), and seven cases had total 1p/19q loss with no *IDH1* mutation (1 A, 1 AA, 1 GB, and 4 O). One AO had both *TP53* mutation and total 1p/19q loss but no *IDH1* mutation. Note that

there were also astrocytomas with no *TP53* or *IDH1* mutation and oligodendrogliomas with neither total loss of 1p/19q nor *IDH1* mutation, some of which had other detectable clonal genetic abnormalities.

When any abnormality of the p53 pathway was considered (*TP53* mutation, *MDM2/MDM4* amplification, or *p14ARF* homozygous deletion),⁷ it was significantly associated with *IDH1* mutation only in astrocytomas (A and AA, $p = 0.005$). Abnormalities of the RB1 pathway (*RB1* mutation, *CDKN2A* homozygous deletion or *Cyclins/CDK4/6* [cyclin-dependent kinase 4/6] amplification) were negatively associated with *IDH1* mutation in adult astrocytic tumors (all A, AA, and GB; $p < 0.001$) but not in oligodendroglial tumor groups. Among all the adult astrocytic tumors, *IDH1* mutations were also negatively associated with *PTEN* mutations ($p < 0.001$; no tumor had both *IDH1* and *PTEN* mutations) and with *EGFR* amplifications ($p < 0.001$; only one tumor had both an *IDH1* mutation and *EGFR* amplification).

The enzymatic activity of *IDH1* resulting in the production of NADPH was assayed using COS-7 cells transfected with the wild-type and all four mutant *IDH1* constructs. The Arg132His, Arg132Ser, and Arg132Cys mutants showed an 8- to 9-fold decrease of enzymatic activity. Arg132Gly showed a 2.4-fold decrease compared with wild type ($p < 0.001$, *t*-test; Fig. 1), thus showing a significantly higher level of activity than the other three mutants ($p < 0.001$). All transfectants showed higher levels of activity than the negative control (vector-alone transfectants; $p < 0.05$).

In addition to *IDH1* mutations occurring more frequently among secondary than among primary GBs ($p < 0.001$), such mutations were much more common in younger GB patients. The mean age of GB patients with *IDH1* mutations was 41 years, compared with 56 years for GB patients with no mutations of *IDH1* (*t*-test, $p = 0.002$), confirming the findings in a previous report.¹

Univariate analyses using the log-rank test were used to assess the impact of *IDH1* mutation on overall survival. *IDH1* mutations were associated with longer overall survival among all 364 tumors (including all A, AA, GB, O, AO, OA, and AOA; $p < 0.001$), all adult astrocytic tumors (including A, AA, and GB; $p < 0.001$), and even among all GBs ($p = 0.011$; Supplementary Fig. 2S). However, Cox regression multivariate analysis including age, histological diagnosis, WHO malignancy grade, *TP53*, 1p/19q status, and primary/secondary GB indicated that *IDH1* status was not an independent prognostic factor.

Discussion

In this study, we carried out an extensive screening for mutations of exon 4 of the *IDH1* gene in a large series of human brain tumors. We identified a high frequency of *IDH1* mutations in the majority of A, AA, O, AO, OA, AOA, and sGB tumors, contrasting with a mutation rate of only 3% in pGB tumors. These findings have been independently confirmed.¹³ No mutations were found in PAs, medulloblastomas, supratentorial PNETs, DNETs,

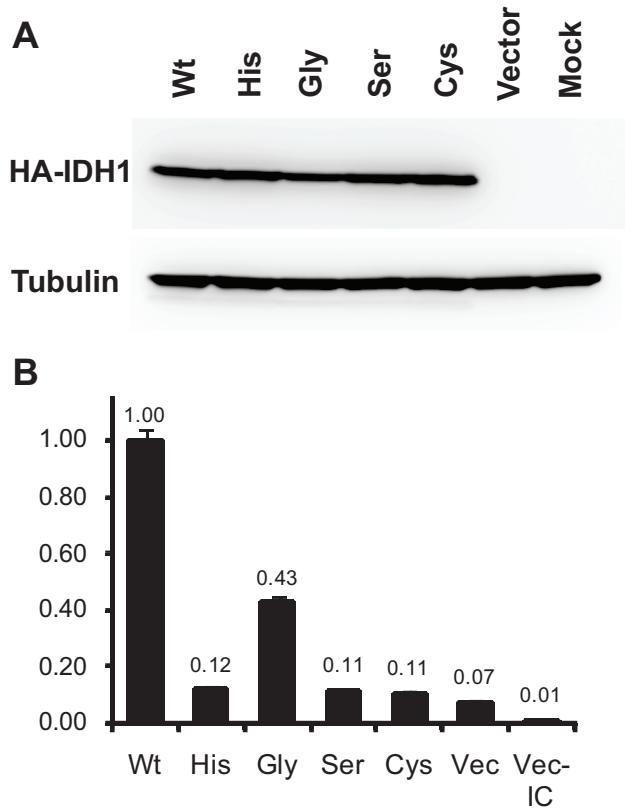


Fig. 1. Enzymatic assay of the wild-type and mutant *IDH1* (isocitrate dehydrogenase 1 [NADP⁺], soluble). (A) Western blotting of the COS-7 transfectants with an antihemagglutinin (anti-HA) antibody (top). The HA-tagged recombinant *IDH1* (46 kDa) is strongly expressed in the wild-type (Wt) and mutant transfectants (His, Arg132His; Gly, Arg132Gly; Ser, Arg132Ser; Cys, Arg132Cys). A comparable result was obtained using a goat anti-*IDH1* antibody (data not shown). The blot was reprobbed with anti- α -tubulin antibody to control for lane loading (bottom). (B) Enzymatic activity of wild-type and mutant *IDH1*. The relative activity is shown as a ratio to wild type (Wt, 1.00). The error bar shows standard error of the quadruplicated assays. The enzymatic activities in all four mutants were significantly decreased compared with wild-type ($p < 0.001$). The activity detected in the vector transfectants (Vec) compared with the vector transfectants without isocitrate (Vec-IC) represents the activity of endogenous *IDH1* in COS-7 cells. Reproducible results were obtained from independent transfection experiments.

ependymal tumors, schwannomas, or meningiomas. Thus, our results show that mutations of *IDH1* and/or *TP53* are the earliest common genetic abnormalities in the majority of diffuse astrocytomas (A). Single cases had one or the other, so a temporal order cannot be established. The situation is similar for the oligodendroglial tumors, where concurrent *IDH1*, total loss of 1p/19q, or both are the earliest common genetic abnormalities identified.

The finding that the majority of *IDH1* mutations are associated with either *TP53* mutation or total 1p/19q loss is striking. *TP53* mutations and total 1p/19q loss are gen-

erally mutually exclusive¹² and characterize WHO grade II diffuse astrocytomas and oligodendrogliomas, respectively, both of which are well known to show malignant progression. There are no known cellular consequences common to these two aberrations. However, based on these findings, the genetic models for development and progression of astrocytic and oligodendroglial tumors must be revised and include mutation of *IDH1* as an early event common to both types of glioma (Fig. 2, Supplementary Table 1S). In this model, the majority of oligodendrogliomas develop through acquisition of concurrent *IDH1* mutation and total 1p/19q loss (as a result of t[1;19] translocation), and grade II tumors may progress, some acquiring RB1 pathway abnormalities. The majority of diffuse astrocytomas arise through developing concomitant mutations of *IDH1* and *TP53*. These tumors generally acquire RB1 pathway alterations when they progress to sGBs. The alternative pathway for GB development, resulting in a pGB, is via synchronous RB1 and p53 pathway disruption,⁷ and this does not seem to be associated with *IDH1* mutation. Disruptions of the p53 pathway by mechanisms other than *TP53* mutation occur predominantly among pGB. Primary GBs may also have Akt pathway disruption (e.g., by *PTEN* mutation/deletion) and/or *EGFR* amplification (with or without rearrangement), unlike other astrocytomas or oligodendrogliomas. Oligoastrocytic tumors appear to develop through pathways similar to either the grade II/III astrocytomas or the oligodendrogliomas (i.e., *IDH1* mutation and either *TP53* mutation or total 1p/19q loss). Pilocytic astrocytomas do not progress, and the majority

have either a unique fusion gene containing the constitutively activated kinase domain of v-raf murine sarcoma viral oncogene homolog B1 (*BRAF*) or mutations of *BRAF* or neurofibromin 1 (*NF1*).¹⁰ They have no *IDH1* mutations or any other changes described above.

IDH1 functions as a homodimer, is localized in peroxisomes and more diffusely in the cytoplasm, and catalyzes oxidative decarboxylation of isocitrate into α -ketoglutarate, generating NADPH from NADP⁺.¹⁴ NADPH is an important cofactor for regeneration of reduced glutathione, which is a critical component of the defense mechanism against oxidative damage.^{15,16} High levels of oxidative stress are reported in the brain compared with other organs. This is reflected in the common transition mutations (G:C → A:T) seen in gliomas, for example, of *TP53*.¹⁷ This was also the case in 81 of 142 (57%) of the *TP53* mutations and 114 of 119 (95%) of the *IDH1* mutations in our series, with only 5 of 119 (4%) mutations of *IDH1* being of any other type.

The 119 somatic mutations, almost all of which were heterozygous, resulted in the substitution of arginine with histidine, serine, or cysteine at codon 132. We showed that the ability to generate NADPH was significantly reduced in all four mutant *IDH1* proteins. These results indicate that Arg¹³² is critical for the enzymatic activity of *IDH1*. Jennings et al.¹⁸ reported that a substitution of arginine by glutamate at codon 132 in a rat ortholog of *IDH1* resulted in a significant reduction in catalytic activity. Although this mutation has not been seen in humans, their observations are in line with ours. Arg¹³² is one of the amino acids in *IDH1* that form

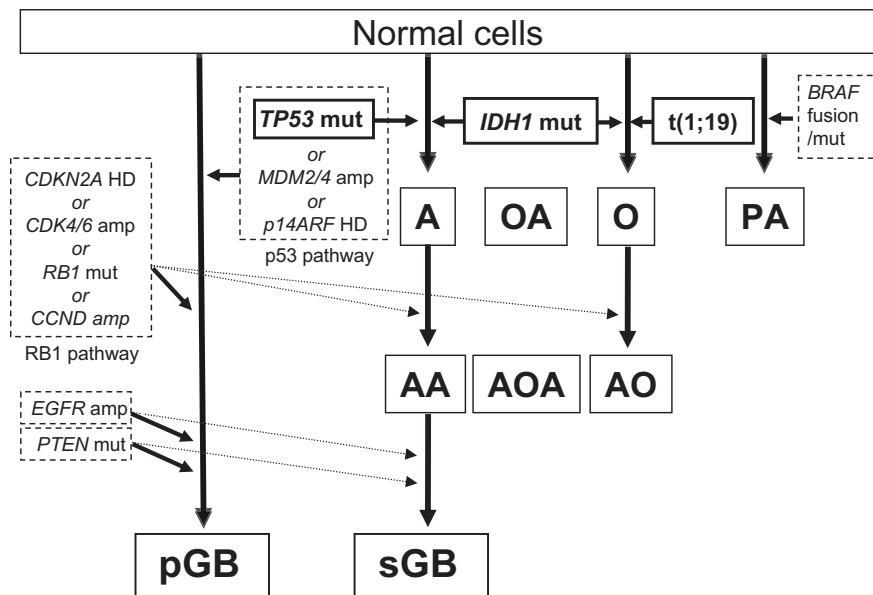


Fig. 2. A model for the development/progression of astrocytic and oligodendroglial tumors. Abbreviations: *CDKN2A*, cyclin-dependent kinase inhibitor 2A; HD, homozygous deletion; *CDK4/6*, cyclin-dependent kinase 4/6; amp, amplification; *RB1*, retinoblastoma 1; mut, mutation; *CCND*, cyclin D1/D2/D3; RB1 pathway, *CDKN2A*, *CDK4/6*, *RB1*, or *CCND*; *EGFR*, epithelial growth factor receptor; *PTEN*, phosphatase and tensin homolog; pGB, primary glioblastoma; *TP53*, tumor protein p53; *MDM2/4*, Mdm2/4 p53 binding protein homolog (mouse); *p14ARF*, the p14 alternative reading frame product of cyclin-dependent kinase inhibitor 2A; p53 pathway, *TP53*, *MDM2/4*, or *p14ARF*; A, diffuse astrocytoma; AA, anaplastic astrocytoma; sGB, secondary glioblastoma; *IDH1*, isocitrate dehydrogenase 1 (NADP⁺), soluble; OA, oligoastrocytoma; AOA, anaplastic oligoastrocytoma; O, oligodendrogloma grade II; AO, anaplastic oligodendrogloma; t(1;19), unbalanced t(1;19)(q10;q10) translocation; PA, pilocytic astrocytoma; *BRAF*, v-raf murine sarcoma viral oncogene homolog B1.

hydrophilic interactions with the α - and β -carboxylate groups of isocitrate.¹⁹ It has also been proposed that Arg¹³² may contribute to a semiopen conformation of *IDH1* during the transition from the inactive form to the active form of the enzyme.¹⁹ Substitution of Arg¹³² may therefore result in conformational changes with a consequent decrease in enzymatic activity.

Because the protein functions as a dimer, the consequences of mutation of one allele may be profound, since one mutated protein in a homodimer may result in a significant reduction in function—producing a dominant negative effect in a similar manner to p53 (functioning as a tetramer). Thus, mutation of one allele of *IDH1*, which results in the decreased ability to generate NADPH as shown in this study, is likely to lead to an increased susceptibility to oxidative damage and thus facilitate the accumulation of further, especially transition, mutations (G:C → A:T). Cells with reduced *IDH1* expression have increased levels of oxidative stress and DNA damage.¹⁶ The finding that *IDH1* mutations frequently accompany *TP53* mutations in the diffuse astrocytomas is particularly notable, because loss of p53 will circumvent normal oxidative-stress–induced apoptosis, as has been demonstrated in glioma cells.²⁰ This combination will therefore likely promote further mutations and tumor progression. It is more difficult to understand the association of *IDH1* mutation with the total concurrent loss of 1p/19q.

The fact that there were no mutations in the 15 established glioma cell lines is not surprising because almost all such lines are derived from GBs, and most of these were probably pGB. Most were derived many years before the differential classification of GBs into primary and secondary, and there are no data to indicate their status.

We confirmed that *IDH1* mutations occur more frequently among sGB and young patients with GB, as reported by Parsons et al.¹ Univariate analyses also showed that tumors with *IDH1* mutations were associated with longer overall survival of patients with any adult astrocytic or oligodendroglial tumor included in the series, with any adult astrocytic tumor alone, or with any GB. However, multivariate analysis failed to identify *IDH1* status as a prognostic factor independent of tumor type, grade, or age. These results are not surprising, because *IDH1* mutations are involved in both oligodendroglial and astrocytic tumors and represent only one of a diverse range of genetic abnormalities, and are therefore unlikely to be an independent prognostic factor.

IDH1 mutations appear to be rare in other common human tumors. One *IDH1* mutation (Arg¹³²Cys) in a colon cancer has been reported in a genomewide muta-

tion analysis of a series of colon and breast cancers,²¹ and no mutations were found in a series of pancreatic cancers.²² Bleeker et al.²³ found no mutations in *IDH1* exon 4 among 559 various nonglioma human cancers, including bladder, breast, colorectal, lung, melanoma, thyroid, ovary, and pancreas. High levels of oxidative stress in the brain compared with other organs may provide strong selective pressure for tumor cells that acquired *IDH1* mutations.

Our results indicate that *IDH1* mutation is common in A, AA, and sGB as well as all types of oligodendroglial tumors, but infrequent in pGB, thereby helping to distinguish pGBs from the others. Consequently, the models of astrocytic and oligodendroglial tumor development/progression require some revision. We also show that the missense mutations identified at Arg¹³² result in decreased enzymatic activity of *IDH1*. Further assessments of the cellular consequences of *IDH1* mutations are under way. We hope a more accurate understanding of the genetic changes and their consequences in these tumors will lead to improvements in the accuracy of diagnosis and prognosis, and the development of novel molecular targeted therapies.

It is important to note that after this paper was submitted for publication we identified somatic missense mutations at codon 172 (arginine) of *IDH2* in seven tumors that had either *TP53* mutation or total 1p/19q loss but no *IDH1* mutations. As a result, all oligodendroglial tumors with total 1p/19q loss harbored mutations of either *IDH1* or *IDH2*, while six oligodendroglial tumors with *IDH1* mutation had neither total 1p/19q loss nor *TP53* mutations. This strongly suggests that mutations of the NADP⁺-dependent *IDH* genes are the earliest changes in the development of oligodendroglial tumors.

Acknowledgments

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