# Antigenic Similarity of Heat-Labile Enterotoxins from Diverse Strains of *Escherichia coli*

R. BRADLEY SACK\* AND JEAN L. FROEHLICH

Department of Medicine, Baltimore City Hospitals, Baltimore, Maryland 21224,\* and The Johns Hopkins University School of Medicine, Baltimore, Maryland 21205

Received for publication 9 February 1977

With use of the rabbit intestinal loop model, heat-labile enterotoxins from 21  $Escherichia\ coli$  strains isolated from a wide spectrum of patients with diarrheal diseases were all neutralized to high titer by two antisera prepared against enterotoxins of either  $E.\ coli$  or Vibrio cholerae. These findings suggest marked immunological similarity among heat-labile enterotoxins from a heterogenous group of  $E.\ coli$ .

Vibrio cholerae enterotoxin and the heat-labile enterotoxins (LTs) of Escherichia coli have been found to be similar physiologically (2, 9, 1)16), immunologically (12, 26), and biochemically (1). The enterotoxins produced by either of the two serotypes of V. cholerae (Inaba and Ogawa) are identical (7); similarly, the crude LTs produced by different E. coli serotypes are known to be at least immunologically related (12, 26). Whereas enterotoxin production is controlled by the chromosomal deoxyribonucleic acid of V. cholerae (27), enterotoxins of E. coli are genetically controlled by plasmids (13, 24, 25). Because of the assumed closely related antigenic structure of E. coli LTs, it has become the practice (3, 15, 22, 23) to use a single LT preparation to test for antitoxins that develop during infections caused by a variety of serotypes of enterotoxigenic E. coli (ETEC). Only a relatively few human strains of ETEC have been closely studied, however; these are mostly from the Indian subcontinent (4-6, 10-12, 26).

The present study was undertaken to test the hypothesis that LTs produced by  $E.\ coli$  of different serotypes, isolated from either children or adults with diarrheal disease in widely separated geographic areas, are immunologically similar.

## MATERIALS AND METHODS

Bacterial cultures. Twenty-one LT-producing E. coli isolated from patients who had diarrheal illness between 1968 and 1975 were studied. Some of the organisms produced heat-stable enterotoxin (ST) in addition to LT. Cultures were either maintained in nutrient agar stabs at room temperature or lyophilized. Serotyping was kindly done by F. Ørskov and I. Ørskov at the World Health Organization Collaborative Center for Reference and Research on Escherichia, Copenhagen (17).

Enterotoxin preparations. Crude LT preparations

from each of the strains were prepared in Syncase by methods previously described (18, 26). These dialyzed, lyophilized preparations were assayed in the 18-h rabbit intestinal loop model (14, 26) to determine the mean effective dose ( $ED_{50}$ ) as a function of dry weight. The slopes of the dose-response curves were similar to those previously described (18, 19, 26).

Antisera. Two antisera were used to neutralize the biological activity of these preparations. The antisera were: (i) purified equine cholera antitoxin prepared at the Swiss Serum Institute (SSVI) and supplied by the National Institutes of Health, and (ii) anti-E. coli serum prepared in rabbits against a single LT preparation of strain 408-3 (O78:H12), as previously described (18). The capacity of each antiserum to neutralize each of the LT preparations was determined in the rabbit intestinal loop model. Twofold dilutions of serum were mixed with  $3 ED_{50}$  of LT and shaken gently (60 shakes/min) for 1 h at 37°C. The antigen-antibody mixtures were then injected into rabbit intestinal loops (26), and fluid accumulation was read at 18 h. The end point of the titration was the highest dilution of serum that neutralized the fluid accumulation response by more than 50%. The titration curves were steep, and the end point usually represented complete neutralization. The geometric mean of at least two neutralization tests for each assay was calculated. (In most instances, three tests were done for each determination.)

#### RESULTS

Thirteen of the ETEC were from adults and eight were from children; one porcine strain is given for comparison. There were 11 different known "O" serogroups, one rough strain, and three untypable strains represented. Eight strains were from the Indian subcontinent and have been widely used in a number of laboratories (10407, 408-3, 339, 411-5). One was from the Middle East, one from Europe, eight from the United States, and three from Mexico. Nine were from patients hospitalized with nonspecific diarrhea (21), and four were from adults with traveler's diarrhea (15). Eleven strains produced both LT and ST; 10 produced LT only. The pig strain has been well characterized previously (16).

The results of the neutralizations are summarized in Table 1. Both anti-cholera and anti-*E. coli* neutralized each of the LT preparations to high, though somewhat variable, titers. The anti-cholera serum neutralized all but one of the LT preparations at higher dilutions than did the anti-*E. coli* antiserum, by an average factor of 5 (range, 0.97 to 12.6). Not shown on the table are neutralization titers of these sera assayed in identical fashion against crude *V. cholerae* enterotoxin, National Institutes of Health lot 001. The SSVI serum had a titer of 25,000, and the rabbit antiserum had a titer of 40, a strikingly different factor difference of 625 (26).

These neutralization titers should not be confused with standard antitoxin units previously assigned to these antisera. SSVI has been assigned the value of 1,000 U/ml against *E. coli* 408-3 LT (18), and the rabbit *E. coli* antiserum assayed at 280 U/ml against *E. coli* 408-3 LT (18); this difference in titers (a factor of 3.6) is also similar to that described from different enterotoxin preparations in this study.

The biological activity of the LT preparations also varied over a 20-fold range. There were no significant differences, however, between (i) strains from children versus adults, (ii) strains producing LT only versus strains producing both LT and ST, (iii) strains from different geographical areas, or (iv) strains isolated from adults with cholera-like illness or traveler's diarrhea.

### DISCUSSION

This study corroborates previous information that LTs from the majority of (if not all) ETEC are immunologically closely related. Since this antibody titration is based solely on neutralization of biologically active enterotoxin, the presence of naturally occurring toxoids or different amounts of enterotoxin subunits might cause considerable variation in the titer. This variation is not incompatible, however, with the possibility that the LTs may all be identical. These findings support the use of a single antigen to

Strain	Serotype	Adult (A) or child (C)	Geography	Clinical illness	Toxin(s)	LT ED <sub>50</sub>	Neutralizing anti- toxin titer <sup>a</sup>	
							Anti-E. coli rabbit serum	SSVI anti- cholera serum
10407	O78:H11	A	Dacca, Bangladesh	CLI	LT-ST	0.096	640	2500
408-3	O78:H12	A	Calcutta, India	CLI	LT-ST	0.125	783	2,000
339	O15:H11	A	Calcutta, India	CLI	LT-ST	0.082	160	625
J3-2	O85:H7	A	Calcutta, India	CLI	LT-ST	0.15	320	1,580
106-3	O78:H11	A	Calcutta, India	CLI	LT	0.200	640	625
411-5	O126:H12	A	Calcutta, India	CLI	LT	0.053	393	1,986
29487	O128:H21	A	Dacca, Bangladesh	CLI	LT-ST	0.75	1,416	5,000
1105-F	O6:H16	A	Calcutta, India	CLI	LT	0.54	500	3,784
I-439	O114:H-	A	Sana'a, Yemen	CLI	LT-ST	0.074	500	2,500
P-3	O8:H9	A	<b>Baltimore</b> <sup>c</sup>	TDd	LT	0.28	393	625
G-14850	O6:H16	С	Topolčany, Czechoslovakia	NSG	LT	1.05	62.5	
Deh-3	(O109):H21	C	Arizona	NSG	LT	0.20	1,000	ND
Joh-4	O64:H-	С	Arizona	NSG	LT	0.302	500	6,295
Lee-1	X	C	Arizona	NSG	LT-ST	0.31	314	1,574
Eth-6	O25:H-	C	Arizona	NSG	LT	0.165	198	1,574
Coo-10	O25:H-	C	Arizona	NSG	LT	0.06	314	993
Cos-6	Х	C	Arizona	NSG	LT-ST	0.14	706	3,140
Wcy-4	х	C	Arizona	NSG	LT-ST	0.35	250	1,574
206 C-2	Rough	A	Mexico City, Mexico	TD	LT	0.60	500	3,148
412 C-1	O6:H16	A	Mexico City, Mexico	TD	LT-ST	0.10	500	1,766
260 C-1	O6:H16	A	Mexico City, Mexico	TD	LT-ST	0.057	250	1,250
P-263"	O8:K87,K88a,b,:H19				LT	0.20	160	1,250

TABLE 1. Neutralization of E. coli LTs by antiserum prepared against enterotoxins of E. coli or V. cholerae

<sup>a</sup> Reciprocal of geometric mean dilution of serum that neutralized 3  $ED_{50}$  of LT.

<sup>•</sup> Cholera-like illness.

<sup>c</sup> Recently returned from Pakistan.

<sup>d</sup> Traveler's diarrhea.

Nonspecific gastroenteritis.

' ND, Not done.

Porcine strain.

## 572 SACK AND FROEHLICH

The observation that the biological activity of the different LT preparations is similar, regardless of the origin of the strain, suggests that the differences in severity of diarrheal illness seen in patients may be largely a function of the human host, or possibly other non-LT bacterial factors, rather than that of the LT itself.

It should be pointed out that this study was begun at a time before the development of the tissue culture assays. Although the rabbit loop model is cumbersome and time-consuming, we believed it worthwhile to complete the study with a single assay. There is every reason to suspect that results from other assay systems would be similar (3, 22).

### ACKNOWLEDGMENTS

Acknowledgment is made to the Gerontology Research Center, National Institute on Aging, for facilities extended under its Guest Scientist Program. We thank Eva Aldova for supplying strain C-14850 and S. M. Sadique for supplying strain I-439.

This work was supported by contract DADA 17-73-C-3055 from the U. S. Army Medical Research and Development Command, and grants 7 R22 AI11358 and 2 R22 AI07625 from the National Institute of Allergy and Infectious Disease.

#### LITERATURE CITED

- 1. Dafni, Z., and J. B. Robbins. 1976. Purification of heatlabile enterotoxin from *Escherichia coli* 078:H11 by affinity chromatography with antiserum to Vibrio cholerae toxin. J. Infect. Dis. 133:S138-S141.
- Donta, S. T., H. W. Moon, and S. C. Whipp. 1974. Detection of heat-labile *Escherichia coli* enterotoxin with the use of adrenal cells in tissue culture. Science 183:334-336.
- Donta, S. T., D. A. Sack, R. B. Wallace, H. L. DuPont, and R. B. Sack. 1974. Tissue-culture assay of antibodies of heat-labile *Escherichia coli* enterotoxins. N. Engl. J. Med. 291:117-121.
- Evans, D. G., D. J. Evans, Jr., and N. F. Pierce. 1973. Differences in the response of rabbit small intestine to heat-labile and heat-stable enterotoxins of *Escherichia coli*. Infect. Immun. 7:873-880.
- Evans, D. G., D. J. Evans, Jr., and S. L. Gorbach. 1973. Identification of enterotoxigenic *Escherichia coli* and serum antitoxin activity by the vascular permeability factor assay. Infect. Immun. 8:731-735.
- Etkin, S., and S. L. Gorbach. 1971. Studies on enterotoxin from *Escherichia coli* associated with acute diarrhea in man. J. Lab. Clin. Med. 78:81-87.
- Finkelstein, R. A. 1970. Antitoxic immunity in experimental cholera: observations with purified antigens and the ligated ileal loop model. Infect. Immun. 1:464-467.
- Gorbach, S. L., J. G. Banwell, B. D. Chatterjee, B. Jacobs and R. B. Sack. 1971. Acute undifferentiated human diarrhea in the tropics. I. Alterations in intestinal microflora. J. Clin. Invest. 50:881-889.
- Guerrant, R. L., U. Ganguly, A. G. Casper, E. J. Moore, N. F. Pierce, and C. C. Carpenter. 1973. Effect of Escherichia coli on fluid transport across canine

small bowel. Mechanisms and time-course with enterotoxin and whole bacterial cells. J. Clin. Invest. 22:1707-1714.

- Gyles, C. L. 1974. Immunological study of the heatlabile enterotoxins of *Escherichia coli* and *Vibrio cholerae*. Infect. Immun. 9:564-570.
- Gyles, C. L. 1974. Relationships among heat-labile enterotoxins of *Escherichia coli* and *Vibrio cholerae*. J. Infect. Dis. 129:227-283.
- Gyles, C. L., and D. A. Barnum. 1969. A heat-labile enterotoxin from strains of *Escherichia coli* enteropathogenic for pigs. J. Infect. Dis. 120:419-426.
- Gyles, C. L., M. So, and S. Falkow. 1974. The enterotoxin plasmids of *Escherichia coli*. J. Infect. Dis. 130:40-49.
- Kasai, G. J., and W. Burrows. 1966. The titration of cholera toxin and antitoxin in the rabbit ileal loop. J. Infect. Dis. 116:606-614.
- Merson, M. H., G. K. Morris, D. A. Sack, J. G. Wells, J. C. Feeley, R. B. Sack, W. B. Creech, A. Z. Kapikian, and E. J. Gangarosa. 1976. Travelers' diarrhea in Mexico. A prospective study of physicians and family members attending a congress. N. Engl. J. Med. 294:1299-1305.
- Moon, H. W., S. C. Whipp, and A. L. Baetz. 1971. Comparative effects of enterotoxins from *Escherichia coli* and *Vibrio cholerae* on rabbits and swine small intestine. Lab. Invest. 25:133-140.
- Ørskov, F., and I. Ørskov. 1975. Escherichia coli O:H serotypes isolated from human blood. Prevalence of the K1 antigen with technical details of O and H antigenic determination. Acta Pathol. Microbiol. Scand. 83:595-600.
- Sack, R. B. 1973. Immunization with Escherichia coli enterotoxin protects against homologous enterotoxin challenge. Infect. Immun. 8:641-644.
- Sack, R. B. 1975. Human diarrheal disease caused by enterotoxigenic *Escherichia coli*, p. 333-353. *In* M. P. Starr (ed.), Annual review of microbiology, vol. 29. Annual Reviews Inc., Palo Alto, Calif.
- Sack, R. B., S. L. Gorbach, J. G. Banwell, B. Jacobs, B. D. Chatterjee, and R. C. Mitra. 1971. Enterotoxigenic *Escherichia coli* isolated from patients with severe cholera-like disease. J. Infect. Dis. 123:378-385.
- Sack, R. B., N. Hirschhorn, I. Brownlee, R. A. Cash, W. E. Woodward, and D. A. Sack. 1975. Enterotoxigenic *Escherichia coli*-associated diarrheal disease in Apache children. N. Engl. J. Med. 292:1041-1045.
- Sack, R. B., N. Hirschhorn, W. E. Woodward, D. A. Sack, and R. A. Cash. 1975. Antibodies to heat-labile *Escherichia coli* enterotoxin in Apaches in Whiteriver, Arizona. Infect. Immun. 12:1475-1477.
  Sack, R. B., B. Jacobs, and R. Mitra. 1974. Antitoxin
- Sack, R. B., B. Jacobs, and R. Mitra. 1974. Antitoxin responses to infections with enterotoxigenic *Esche*richia coli J. Infect. Dis. 129:330-335.
- Skerman, F. J., S. B. Formal, and S. Falkow. 1972. Plasmid-associated enterotoxin production in a strain of *Escherichia coli* isolated from humans. Infect. Immun. 35:622-624.
- Smith, H. W., and S. Halls. 1968. The transmissible nature of the genetic factor in *Escherichia coli* that controls enterotoxin production. J. Gen. Microbiol. 52:319-334.
- Smith, N. W., and R. B. Sack. 1973. Immunologic crossreactions of enterotoxins from *Escherichia coli* and *Vibrio cholerae*. J. Infect. Dis. 127:164-170.
- Vasil, M. L., R. K. Holmes, and R. A. Finkelstein. 1975. Conjugal transfer of a chromosomal gene determining production of enterotoxin in Vibrio cholerae. Science 187:849-850.