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Chronic Stress-Induced Hippocampal Vulnerability: The Glucocorticoid Vulnerability Hypothesis

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Synopsis

The hippocampus, a limbic structure important in learning and memory, is particularly sensitive to chronic stress and to glucocorticoids. While glucocorticoids are essential for an effective stress response, their oversecretion was originally hypothesized to contribute to age-related hippocampal degeneration. However, conflicting findings were reported on whether prolonged exposure to elevated glucocorticoids endangered the hippocampus and whether the primate hippocampus even responded to glucocorticoids as the rodent hippocampus did. This review discusses the seemingly inconsistent findings about the effects of elevated and prolonged glucocorticoids on hippocampal health and proposes that a chronic stress history, which includes repeated elevation of glucocorticoids, may make the hippocampus vulnerable to potential injury. Studies are described to show that chronic stress or prolonged exposure to glucocorticoids can compromise the hippocampus by producing dendritic retraction, a reversible form of plasticity that includes dendritic restructuring without irreversible cell death. Conditions that produce dendritic retraction are hypothesized to make the hippocampus vulnerable to neurotoxic or metabolic challenges. Of particular interest is the finding that the hippocampus can recover from dendritic retraction without any noticeable cell loss. When conditions surrounding dendritic retraction are present, the potential for harm is increased because dendritic retraction may persist for weeks, months or even years, thereby broadening the window of time during which the hippocampus is vulnerable to harm, called the Glucocorticoid Vulnerability Hypothesis. The relevance of these findings is discussed with regard to conditions exhibiting parallels in hippocampal plasticity, including Cushing's disease, Major Depressive Disorder (MDD), and Post-Traumatic Stress Disorder (PTSD).

Introduction

An individual's survival relies upon the successful activation and termination of a stress response. The stress response occurs following a real or perceived threat (stressor) and includes the activation of the sympathetic nervous system (SNS) and the hypothalamic-pituitary-adrenal (HPA) axis. The SNS is responsible for the rapid stress response, involving the release of catecholamines (epinephrine and norepinephrine) within seconds of the onset of the stressor. The HPA axis is responsible for the slower onset stress response, involving the release of glucocorticoids (Cortisol in humans and corticosterone in rodents) within several minutes of stressor onset. A critical feature of the stress response is that it terminates itself when the stressor has ended or is no longer perceived as a threat. The termination of the stress response is critical because the continued activation of the SNS and the HPA axis can wreak havoc on the body. A racing heart and high blood pressure triggered by epinephrine to maximize blood flow is critical for assisting in escaping the stressor, but a persistent racing heart and high blood pressure increases the risk of heart failure and arteriosclerosis. Elevated glucocorticoids are

essential for the redistribution of energy resources, but the long-term elevation of glucocorticoids starves some tissues of necessary resources and hinders immune function, increasing the susceptibility to disease. While the stress response is critical for the successful adaptation to a real or perceived threat, a persistent stress response can be detrimental.

In the 1930's, Hans Selye stumbled upon the discovery that chronic stress can shorten the lifespan. With the publication about the General Adaptation Syndrome (GAS) and the Diseases of Life, the connection that chronic stress can cause poor health received widespread acknowledgment /see 136/. In the years that followed the original publication, aspects of the GAS were inconsistent with new discoveries: among the notable findings was that glucocorticoids reduced inflammation instead of exacerbating it. Not until decades later did interest in chronic stress re-emerge to support a key concept of Selye's hypothesis that persistent stress can weaken the body.

Several findings beginning in the late 1960s revealed that prolonged exposure to stress or glucocorticoids harms the brain. Glucocorticoid function was first identified in peripheral tissue, hence, the name "glucocorticoid" reflects effects on energy metabolism. Subsequent studies revealed that the brain is an important target of glucocorticoids, with the forebrain and limbic structures containing a high proportion of specific receptors /87,88/. In line with this revelation was the finding that the brain responded poorly to extended and elevated glucocorticoid levels. Guinea pigs injected daily with glucocorticoids for four weeks exhibited brain damage in the hypothalamus and hippocampus /4/. In the late 1970's and early 1980's, studies began to link stress and glucocorticoids with poor hippocampal aging. Age-related hippocampal pathology, as measured by reactive astrocytes, positively correlated with plasma glucocorticoid levels in rats /74/. Importantly, reducing glucocorticoids by adrenalectomy at mid-age attenuated glial reactivity and neuronal loss commonly observed in aged rats /75/. These studies were followed by a series of reports showing that the dysfunctional hippocampus contributes to an impaired HPA axis response in aged rats /117–120,123/. These findings were consistent with reports showing that the hippocampus provides negative feedback to the HPA axis, with hippocampal stimulation and lesions causing decreases and increases in HPA axis activity, respectively /36,40,43,58,85/. Consequently, some investigators renamed the HPA axis, calling it the "LHPA" axis to acknowledge the influence of the limbic "L" region /as examples, see 78,106/. Collectively, these studies demonstrated that chronic stress and glucocorticoids impaired hippocampal function, which in turn contributed to the dysregulation of the HPA axis.

In 1986, the Glucocorticoid Cascade Hypothesis was proposed to describe the dynamic relationship between glucocorticoids and the hippocampus with aging /127/. This seminal hypothesis states that glucocorticoids secreted during periods of stress desensitize the hippocampus to further glucocorticoid exposure by downregulating glucocorticoid receptors, an effect that is self-correcting and therefore reversible. At some point, however, the downregulation of glucocorticoid receptors precipitates further hypersecretion of glucocorticoids until permanent hippocampal cell loss occurs. This irreversible hippocampal damage was proposed to make the hippocampus irreversibly insensitive to further glucocorticoid elevations, creating a feed-forward cycle of elevated glucocorticoids and continued hippocampal destruction as an individual ages. While elegant, the glucocorticoid cascade hypothesis had some inconsistencies /see 2/. The purpose of this review is to discuss the seemingly inconsistent findings concerning the effects of prolonged elevation of glucocorticoids on hippocampal health and to discuss the interpretation that a chronic stress history, which includes repeated elevation of glucocorticoids, may make the hippocampus vulnerable to potential injury.

Brief Historical Account of the Glucocorticoid Cascade Hypothesis and its Caveats

A series of studies starting in the 1980's showed that acute stress and the concurrent elevation of glucocorticoids exacerbated damage to the hippocampus caused by neurochemical or metabolic challenges. Glucocorticoids enhance hippocampal damage following neurotoxin insult /kanic acid, also known as 3-acetyl-pyridine, 125,126/, or metabolic challenge /hypoxia, 156/, /hypoglycemia, 156/, /ischemia, 124/. Glucocorticoids can aggravate hippocampal damage when steroid titers are elevated just prior to, during, or immediately after the neurotoxic challenge /122/. Reducing glucocorticoids via adrenalectomy or by metyrapone administration can attenuate glucocorticoid-induced exacerbation of hippocampal damage from a neurotoxin challenge /121,124,141,148/. Down-regulating the HPA axis with repeated glucocorticoid injections to produce low glucocorticoid titers during a hypoxia/ischemia challenge helps to protect against hippocampal damage /72/. While glucocorticoids can influence hippocampal susceptibility indirectly through their well-known effects on peripheral energy metabolism, in vitro tissue culture work demonstrates that glucocorticoids can also directly exacerbate hippocampal cell loss following hypoxia and hypoglycemia /128,156/, by compromising energy use /130,162/. Collectively, these studies show that acute glucocorticoid elevations exacerbate hippocampal damage when glucocorticoid elevations coincide with challenges that compromise energy use.

The Glucocorticoid Cascade Hypothesis illustrates the mechanism underlying the shift from reversible glucocorticoid receptor downregulation to permanent cell death in the hippocampus /127,130/. Under conditions of acute stress and high glucocorticoid levels, the hippocampus has reduced energy stores to respond effectively to a challenge, such as a metabolic event. Under most circumstances, high glucocorticoid levels occur without such a metabolic challenge and subsequently subside without any severe consequence to hippocampal health. Indeed, most young individuals rarely exhibit metabolic challenges, such as stroke, ischemia, hyperglycemia, hypoxia, etc., which could compromise hippocampal integrity. In contrast, aged individuals have a lifetime of experiences with elevated glucocorticoids and are more likely to experience a metabolic event that coincides with high glucocorticoid levels. Thus, the probability of any one of these elevated glucocorticoid events occurring around the same time as a metabolic event is greater in the aged than in the young. The transition from reversible glucocorticoid receptor downregulation to permanent cell death requires just one concurrent incident of glucocorticoid elevation and metabolic challenge. Once these two events coincide, hippocampal damage occurs, and HPA axis regulation by the hippocampus becomes progressively less effective, leading to ever-increasing glucocorticoid levels. Subsequent glucocorticoid elevations can become even more common, as each single stressful event is prolonged due to faulty the negative feedback, causing glucocorticoids to stay elevated longer in an individual with hippocampal damage. Consequently, because the duration of glucocorticoid elevation is prolonged, the probability of elevated glucocorticoids occurring around the same time as the next metabolic challenge is increased. Therefore, a damaged hippocampus makes itself even more susceptible to a metabolic challenge than a healthy hippocampus, creating a downward spiral of increased chances of potentiated hippocampal damage with each metabolic episode and this risk increases with age.

At the time that the Glucocorticoid Cascade Hypothesis was proposed, the supporting studies utilized rats. However, subsequent studies using rats and other models did not consistently find hippocampal damage with prolonged glucocorticoid or stress exposure. Some studies reported that chronic stress or glucocorticoids contributed to hippocampal cell death in adult rats /33, 123/ and non-human primates /129,157/, while others failed to find hippocampal cell loss in rats /8,21,142/, tree shrews /47,163/, non-human primates /76/, and humans /93/. The mounting

conflicting evidence clearly shows justification to question whether chronic stress and glucocorticoid elevations are sufficient to kill hippocampal neurons with age.

In corroboration with this concern, some studies began to show inconsistencies in glucocorticoid responsivity in aged individuals. In a test of HPA axis activity, cortisol levels before and after a psychological test were compared between men in their twenties and sixties. Regardless of age, comparable endocrine response patterns were observed, which failed to support the hypothesis of the breakdown of the HPA axis with age /73/. In other studies, cortisol levels were measured in patients with Alzheimer's disease and healthy adults following an exogenous challenge with dexamethasone, a synthetic glucocorticoid that should inhibit the HPA axis via negative feedback, reducing the subsequent release of endogenous cortisol /45, 154/. Both studies found that control subjects and patients with Alzheimer's disease had comparable HPA axis response patterns. Moreover, Swanwick and colleagues (1998) repeated the dexamethasone challenge at 9-month intervals and were unable to predict the HPA axis response in the Alzheimer's disease patients based upon their previous history, suggesting that hypercortisolism and cell loss was not a cofactor in further degeneration. The inconsistencies between the rodent and human literature drew into question the relevance of the Glucocorticoid Cascade Hypothesis for humans /see 165/.

Some of the difficulty in understanding the mechanisms by which chronic glucocorticoids influence the hippocampus may arise from species-specific differences in stress steroid receptor expression. Glucocorticoids bind with a low affinity to the glucocorticoid receptor (GR) and with nearly ten fold higher affinity to the mineralocorticoid receptor /MR, 3/. While these names may appear counter-intuitive, the receptors for glucocorticoids were originally described after characterizing their peripheral actions; glucocorticoids produced robust effects on peripheral tissues expressing GR, without noticeable effects on tissues expression MR (i.e. kidney), mainly because the kidney contained 11 β -hydrosteroid dehydrogenase Type II to convert active glucocorticoids into inactive forms /48,135/ and to permit selective access of aldosterone to MR. Consequently, glucocorticoid receptors were named based upon ligand effectiveness in the peripheral tissues. In contrast to these tissues, 11 β -hydrosteroid dehydrogenase is nearly absent in the brain /134/, which expresses both GR and MR /113/. Moreover, the limbic regions highly express GR and MR, both of which confer functional consequences in response to glucocorticoids /37/. However, controversy arose because in the rodent brain, GR and MR were particularly concentrated within the hippocampus /113/, while differences were found in the nonhuman primate hippocampus /116/. Specifically, Rhesus monkeys express relatively few GR in the hippocampus, despite their relative abundance within the hypothalamus and pituitary /116/. This lack of robust hippocampal GR expression is problematic because glucocorticoids are proposed to bind to GR-sensitive tissue to mediate subsequent neuronal death within the hippocampus. Sánchez and colleagues /116/ hypothesized that MR may mediate the detrimental effects of chronic glucocorticoid exposure because MR are expressed in relatively high levels within the hippocampus. Alternatively, they suggested that the hypothalamus could be a putative target, given the high numbers of GR it contained. Unlike the Rhesus monkey, high levels of GR and/or MR are found within the hippocampus of the squirrel monkey /103/, macaque /17/, marmoset /63/ and human /131/. Patel and colleagues suggest that riboprobe sensitivities may have contributed to the differences in the Rhesus monkey, noting that human riboprobes were used in this study. For the human work, a potential confound is that the temporal lobes were excised to treat epileptic seizures, making it unclear whether GR expression represents the typical human hippocampus. The equivocal findings for primate GR levels within the hippocampus raise doubt as to whether chronic glucocorticoid exposure causes hippocampal damage via GR in humans. These conflicting outcomes contributed to the reduced interest in the Glucocorticoid Cascade Hypothesis to explain hippocampal decline with aging.

Chronic Stress Endangers the Hippocampus: The Glucocorticoid Vulnerability Hypothesis

If repeated elevation of glucocorticoids through a lifetime fails to kill neurons within the hippocampus, then do glucocorticoids contribute to hippocampal endangerment? Glucocorticoids may not be necessary for hippocampal damage, but they are an important contributing variable to hippocampal vulnerability to damage. A history of chronic stress is proposed to alter the hippocampus by elevating glucocorticoids and these alteration(s) extend the window of time, during which the hippocampus is susceptible to damage. A critical difference between the Glucocorticoid Cascade Hypothesis and the current Glucocorticoid Vulnerability Hypothesis is that glucocorticoids need not be elevated at the time of the metabolic challenge: a history of chronic stress is proposed to leave an “imprint” on the hippocampus, making the hippocampus vulnerable even when glucocorticoids are not elevated at the time of the metabolic challenge. As a consequence, chronic stress and glucocorticoids are not the sole determinant for hippocampal cell loss but play a critical role in priming the hippocampus to be more susceptible to subsequent insults.

A recent history of chronic stress is a significant variable contributing to the susceptibility of the hippocampus to degeneration. As described earlier, work in the 1980's showed that acute glucocorticoid elevations that coincided with a metabolic challenge produced more hippocampal damage than did metabolic challenge alone /130/, revealing the vulnerability of the hippocampus to simultaneous threats. Recently, work in our lab revealed that a chronic stress history could compromise hippocampal health as well. In one study, we exposed male rats to chronic immobilization stress for six hours/day for 21 days, then a few days later, we challenged the CA3 region of the hippocampus with ibotenic acid (IBO), a neurotoxin that is thought to act upon the glutamate system through NMDA receptors /67/. Rats with a history of chronic stress showed more CA3 damage from IBO compared to non-stressed controls /28/. To determine whether these effects were produced by circumstances surrounding the chronic stress history or from potential glucocorticoid elevation caused by the last day of restraint, we exposed rats to a single restraint episode (acute stress) at the time when chronically-stressed rats received their last day of restraint, then assessed IBO-induced hippocampal CA3 damage /28/. Only chronic stress, but not acute stress, exacerbated IBO-induced CA3 damage, which confirmed that waiting several days following the end of the last day of restraint permitted glucocorticoid elevations from the last restraint session to subside. These data support the interpretation that acute glucocorticoid elevations from restraint were unlikely to have exacerbated IBO-induced CA3 damage and emphasize that the history of chronic stress was critical for hippocampal susceptibility to the IBO challenge.

A caveat is that the surgical procedures to infuse IBO into the CA3 region of the hippocampus may have elevated stress hormones differently in chronically-stressed rats compared to controls. Such an interpretation would suggest that the surgical-induced elevations of stress hormones that were differentially primed from the chronic stress history mediated IBO-induced hippocampal damage. Specifically, the HPA axis response changes with chronic stress history /34,35/. Rats exposed repeatedly to the same chronic stress stimulus (homotypic stressor) begin to habituate to that stimulus and show attenuated HPA axis responses. In contrast, rats exposed repeatedly to the same chronic stress stimulus and then presented with a novel stressful stimulus (heterotypic stressor) exhibit potentiated HPA axis responses to the novel stimulus. In our paradigm, chronic restraint is homotypic and would produce a blunted HPA axis response to restraint, but infusing IBO under surgical anesthesia cause a heterotypic stress response. Consequently, the chronically-stressed rats may have had exacerbated CA3 damage in response to IBO because of a potentiated HPA axis response during the procedures to infuse IBO. To address this concern, we performed another study to infuse IBO at a time removed from the exposure to the potential heterotypic stressor of surgical anesthesia; cannulae were implanted

to target the CA3 region before treatment, and after recovery, rats were exposed to chronic glucocorticoids for 21 days, allowed to rest for several days, then infused with IBO through the pre-implanted cannula while alert /31/. We found that a history of chronic corticosterone exacerbated CA3 damage from the IBO challenge even without the potential heterotypic stressor of surgical anesthesia. The procedure of infusion through the cannula could still be construed to be a mild and novel heterotypic stressor for the rats. However, this seems unlikely as measurements of endogenous corticosterone responses to a mild and novel stressor in another circumstance (acute saline injection) produced similar corticosterone responses in the control and chronic corticosterone-treated rats. These findings are significant because they show that homotypic stress leaves an “imprint” on the hippocampus that extends the window of time during which the hippocampus is vulnerable to damage.

Conditions Producing Hippocampal Dendritic Retraction Confer Susceptibility to Damage

While the literature reveals inconsistencies with regards to chronic stress producing hippocampal neuronal death, many laboratories using a variety of species report reliable chronic stress effects on other hippocampal measures. These stress/glucocorticoid-induced hippocampal changes include decreasing neurogenesis /62,108,115/, altering dendritic spine density /97,153,155/, hindering synaptic plasticity /62,115/, impeding long-term potentiation /105/, changing the inhibitory and excitatory tone /62,102,112/, downregulating GR /69,170/, and reducing dendritic complexity /70,82,167/. Of these changes, hippocampal CA3 dendritic retraction, characterized by reduced dendritic complexity and total dendritic length, is an important indicator of a chronic stress history because it occurs following chronic stress or glucocorticoid exposure /30/, but not following acute stress, even when that acute stress is repeated for up to two weeks in some paradigms /80,91/. In contrast, the other chronic stress-induced changes within the hippocampus can be produced following acute stress/glucocorticoids /19,29,39,52,89,104/. Moreover, hippocampal CA3 dendritic retraction lasts for at least four days following the end of chronic stress /27,164/. Therefore, chronic stress alters the hippocampus, including CA3 dendritic retraction, and these changes are present relatively long (many days) after stress hormone elevations have subsided compared to stress-induced glucocorticoid elevations that return to baseline very quickly (within hours of the stressful event).

We investigated whether a history of chronic stress and the subsequent structural changes, as measured by dendritic retraction within the hippocampus, conveyed susceptibility to a neurotoxic challenge. We first compared the effects of an IBO challenge to the CA3 and CA1 hippocampal regions after chronic restraint stress. These two regions are critical because hippocampal CA3 neurons undergo dendritic retraction at three weeks following chronic stress or glucocorticoid treatment /91/, while CA1 neurons do not express dendritic retraction at this time /82,166,169/. We found that chronic stress exacerbated cell loss in the hippocampal CA3, but not CA1, region following an IBO challenge /28/. We also compared IBO-induced CA3 damage following chronic stress in both sexes because the extent of chronic stress-induced CA3 dendritic remodeling differs: chronic stress alters the larger apical region in males as opposed to the smaller basal region in females /49/ with evidence of ovarian hormones being neuroprotective /90/. We found that chronic stress exacerbated IBO-induced CA3 damage in males, but not females /28/. These findings suggest that chronic stress and its subsequent imprint on the hippocampus as measured by dendritic retraction may contribute to the vulnerability of the CA3 region to the IBO challenge.

To further investigate the contribution of chronic stress/glucocorticoid-induced dendritic retraction to hippocampal susceptibility to a neurotoxic challenge, we blocked CA3 dendritic retraction. Stress levels of corticosterone were administered to rats in the drinking water for

three weeks to produce CA3 dendritic retraction. Half of the rats were treated daily during the three weeks with the antiepileptic drug, phenytoin (Dilanton), to block chronic stress- and glucocorticoid-induced hippocampal CA3 dendritic retraction, as described by others /83, 166/. A few days after corticosterone exposure ended, rats received IBO infusions into the CA3 region via previously implanted cannulae. We confirmed that the glucocorticoid treatment produced CA3 dendritic retraction, which was prevented with phenytoin /31/. Importantly, we found that only conditions producing CA3 dendritic retraction (corticosterone + vehicle) showed exacerbated hippocampal CA3 cell loss by an IBO challenge /31/. Treatments that did not produce hippocampal CA3 dendritic retraction did not show IBO-induced exacerbation of hippocampal damage, including the chronic corticosterone group treated with phenytoin. Moreover, phenytoin did not exert its neuroprotective effect by altering the HPA axis because corticosterone profiles were similar among rats treated with corticosterone and phenytoin versus corticosterone and vehicle. Altogether, these findings show that chronic stress or glucocorticoid elevation that produces structural changes in hippocampal dendritic arbors also makes the hippocampus vulnerable to neurotoxic challenges, even when glucocorticoids are unlikely to be elevated during the neurotoxic challenge.

The mechanism by which chronic stress confers susceptibility to a neurotoxic challenge is unknown, but dendritic retraction may provide insight. Dendritic remodeling is essential for normal, healthy development and is an important part of plasticity in adult brains as well. In hibernating ground squirrels, for example, dendritic retraction occurs during hibernation, and dendritic complexity increases upon waking /109,110/. The decrease in dendritic complexity during a time of reduced forebrain function suggests that dendritic retraction may disrupt information processing. Consistent with this idea is the finding that dendritic complexity is correlated with synaptic input /111/. Hippocampal dendritic restructuring may be significant because a major source of input to the apical dendrites of CA3 neurons is glutamate, which is released from the commissural pathway (contralateral hippocampus), entorhinal cortex, and the mossy fibers of the dentate gyrus. However, unregulated glutamate can be neurotoxic /20/, and glucocorticoids increase extracellular glutamate levels /62,92,150/. Consequently, dendritic retraction may be an adaptive response to reduce the exposure of CA3 neurons to glucocorticoid-induced glutamate elevation. However, the price of dendritic retraction can be substantial. At the systems level, studies consistently show that CA3 dendritic retraction corresponds with impaired spatial ability /30,91/, a hippocampus-mediated function /98/. At the cellular level, slow functioning may hinder the ability of neurons to respond quickly or effectively to a neurotoxic or metabolic challenge. CA3 dendritic retraction is hypothesized to be a compensatory response to avoid over-exposure to elevated glucocorticoids and the subsequent increase in glutamate, but this dendritic restructuring has a cost: neurons that retract dendrites may be unable to respond adequately to an unexpected metabolic challenge, leading to exacerbated hippocampal damage.

If dendritic retraction has such a high cost of potentially irreversibly damaging the neurons, then why do neurons undergo this process? CA3 dendritic retraction is proposed to occur in response to an event that is certain: the bombardment of glutamate induced by elevated glucocorticoids, which makes adjusting to the potentially lethal exposure to glutamate paramount. In contrast, the potential for damage in response to a metabolic challenge is uncertain. Specifically, neuronal death from glutamate over-exposure is highly probable during increased glutamate release, whereas a metabolic event may not occur. Another factor to consider is that stress-induced CA3 dendritic retraction is reversible; neurons return to the prechronic stress condition within 10 days after the termination of chronic stress /27,164/. Consequently, CA3 dendritic retraction may be triggered by repeated, high glucocorticoid exposure, but then returns to baseline once glucocorticoid levels return to normal. Therefore, CA3 dendritic retraction is proposed to be a relatively low-risk, temporary neuroplastic response to combat the immediate looming threat of elevated glutamate.

Another interpretation is that chronic stress-induced changes in the MR/GR ratio within the hippocampus may confer susceptibility to a neurotoxin challenge. A dual role for MR and GR was proposed to describe HPA axis regulation /41,101/, then was extended to describe cognitive function as well /25,26,29,38,100/. Altering MR and/or GR expression can influence the HPA axis and perhaps susceptibility to a potential challenge to the hippocampus. Low MR expression within the CA2/3 region produced by chronic unpredictable stress corresponds with elevated basal levels of glucocorticoids /78/. Conversely, high MR expression within the hippocampus in a predisposed Lewis rat strain is associated with hyporesponsiveness to stress /101/. In either situation, the ability to detect changes in glucocorticoid levels is compromised because elevated basal glucocorticoids may reduce the sensitivity of MR to detect basal glucocorticoid fluctuations /38,41/ and over-expression of MR may hinder a robust stress response. Using our model of chronic restraint stress, we found reduced GR expression across the hippocampus and elevated glucocorticoids during cognitive testing /170/. This finding is consistent with reports showing impaired glucocorticoid feedback with chronic stress /5,119, 120/ or with glucocorticoid agonists /57/; however, not all studies report decreased MR or GR with chronic stress /59,143/. Instead, the ratio of MR/GR expression may be critical for regulating hippocampal function, such that a MR/GR ratio imbalance is hypothesized to contribute to cognitive dysfunction and susceptibility to disease /38,81/. Indeed, suicide victims exhibit lower MR/GR ratios within the hippocampus than controls /78/. Consequently, MR/GR ratio balance may be an important variable that contributes to hippocampal dysfunction.

The significance of these findings is that a history of chronic stress or glucocorticoid elevation, which produces alterations within the hippocampus, extends the window of time during which the hippocampus is susceptible to damage. Prior work demonstrated that a neurotoxic challenge that coincided with elevated glucocorticoids produced more damage than either event alone /121/. The current findings reveal that glucocorticoid elevations are necessary to produce hippocampal structural changes, but glucocorticoids need not be elevated at the time of the neurotoxic challenge to confer heightened susceptibility to damage. Importantly, the relatively long-lasting persistence of structural changes, including CA3 dendritic retraction in response to the transient stress-induced elevation of glucocorticoids is significant because it presents an extended opportunistic window during which the hippocampus is vulnerable. Therefore, chronically-stressed individuals are at increased risk for potential hippocampal damage because they are exposed to repeated elevation of glucocorticoids and the potential subsequent structural alterations within their hippocampus confer extended susceptibility.

An important factor to consider is the type of insult to the hippocampus. In our experimental studies, we used IBO neurotoxin. While the likelihood of an individual experiencing neurotoxicity is probably very low, this paradigm could be extrapolated to situations involving metabolic challenges. In the acute glucocorticoid exposure model, the presence of high glucocorticoid levels exacerbated hippocampal damage following exposure to a neurotoxin /kainic acid, 3-acetylpyridine, 121,122/, as well as to episodes of ischemia /124/, hypoxia /156/, or hypoglycemia /156/. These results illustrate that the hippocampus was susceptible to damage whether it was challenged with neurotoxin or a metabolic event, emphasizing that glucocorticoid-enhanced susceptibility can be generalized to a neurotoxic or metabolic insult. Since our current findings reveal that chronic stress conditions can also increase hippocampal susceptibility to a neurotoxic challenge, our results may be extrapolated to suggest that chronic stress conditions may also increase hippocampal susceptibility to metabolic events, although empirical testing will be important to address this issue.

Significance for Metabolic Challenges and Clinical Conditions

The finding that a history of chronic stress can exacerbate hippocampal damage in response to a neurotoxic/metabolic challenge has many implications for a variety of conditions that show

similar dynamics in hippocampal plasticity, including Cushing's disease, Major Depressive Disorder (MDD), and Post-Traumatic Stress Disorder (PTSD). Cushing's disease is a rare disorder characterized by the hypersecretion of glucocorticoids /95,96/. When undiagnosed, individuals show a high degree of susceptibility to mood disorders, such as MDD, and these symptoms improve with treatment /144/. MDD is among the most common psychiatric disorders /7,77/ and predominately involves changes in mood, but also includes alterations in cognition, HPA axis function, motivation, sleep, anhedonia, appetite and weight /42,56/. PTSD has received a great deal of attention recently due to current world events and is an anxiety disorder that can develop in response to real or perceived trauma. PTSD is characterized by hypo-HPA activity and episodes of reliving the trauma through intrusive memories (DSM-IV). Common features for all three conditions include small hippocampal volume and HPA axis disruption /13–15,46,54,61,94,137,145,149,160,161/. The relationship between hippocampal volume and glucocorticoid levels in these human conditions provides strong evidence that parallels our observations with chronic stress and dendritic retraction within the hippocampus of rodents.

A consistent characteristic of these conditions is that symptom severity correlates with hippocampal size, which shows plasticity by improving with treatment. Patients with Cushing's disease show reduced hippocampal volume that inversely correlates with glucocorticoid levels /145/. Following intervention to reduce glucocorticoids, Cushing's disease patients show nearly 10% hippocampal volume increases after 1.5 to 2 years /11,60,146/ and improvement in function /147/. In patients with MDD, hippocampal volume is negatively correlated with symptom severity /137/. Antidepressant treatment can prevent or restore hippocampal volume loss because MDD individuals on long-term antidepressants show no significant differences in hippocampal volume compared to controls /138/. In Vietnam veterans with PTSD, smaller hippocampal volumes correlate with longer combat exposure /54,158/. Similarly, PTSD patients exposed to childhood sexual abuse or other trauma show smaller hippocampal volumes that are correlated with symptom severity /149,161/, demonstrating that these characteristics of PTSD extend to non-combat exposure. These findings are consistent with the animal literature showing that chronic antidepressant treatment increases neurogenesis /84/ and triggers synapse formation in hippocampal neurons /55/. It should be noted that some reports fail to find reduced hippocampal volumes in PTSD patients /9,107,133/, but a meta-analysis of 23 published studies on PTSD confirms that hippocampus volumetric differences covary with PTSD severity /65/. Importantly, antidepressant treatment can increase hippocampal volumes in PTSD patients /10,159/. In all conditions, the hippocampus is particularly vulnerable to volumetric changes, as many other brain regions are unaffected /15,54,61/. Taken together, the reversibility of hippocampal volume in all of these conditions suggests that neuron loss may be less important /32,79,152/, than other processes that may include changes in glial cells, connections and spine formation /93/, which are consistent with altered dendritic structure and are capable of plasticity. Consequently, hippocampal dendritic retraction produced by chronic stress in rodents and reduced hippocampal volumes observed in Cushing's disease, MDD, and PTSD are dynamic and have the capacity to recover.

Another important consideration is that individuals with Cushing's disease, MDD and PTSD show HPA axis disruption. For Cushing's disease and MDD, glucocorticoid levels correspond to symptom severity, such as mood /18,68/, changes in hippocampal volume /18,145/, and disturbance in hippocampal-dependent cognition /18,51,53,145/. Resolving the hypersecretion of glucocorticoids can improve symptoms /86,144,147/. Some have even suggested that antiglucocorticoid therapy may be a useful treatment for MDD /114/. Unlike Cushing's disease and MDD, individuals with PTSD are often characterized with a hyporesponsive HPA axis /172/, which may be an important symptom in the immediate aftermath of the trauma for developing PTSD /173/, and suggests that correcting the subthreshold glucocorticoid levels

may reduce the possibility of developing PTSD. A paradox is that glucocorticoids appear to be problematic when they are either too high (Cushing's disease or MDD) or too low (PTSD), but only high levels of glucocorticoids are traditionally thought to reduce hippocampal volumes. Also, studies on MDD typically find negative correlations with hippocampal volume and glucocorticoid levels, suggesting that low glucocorticoid levels are optimal for large hippocampal volume. However, a recent study on aged individuals with MDD reveals that hypo- and hyper-secretion of glucocorticoids can reduce hippocampal volumes /12/. Moreover, the animal literature shows that reduced glucocorticoids via adrenalectomy causes neural degeneration within the hippocampal dentate gyrus, /22–24,140/, which can be detected by volumetric measures /23/. Consequently, glucocorticoids can reduce hippocampal volume and plasticity when their levels are either too high or low. The overall message is that the effect of glucocorticoids on hippocampal size in these psychiatric conditions is described by an inverted U-shaped function, emphasizing the importance of regulating glucocorticoids at moderate levels so that extremes of glucocorticoid secretion are minimized.

Taken together, these findings show that patients with Cushing's disease, MDD, or PTSD exhibit a small hippocampus and dysregulated HPA axis, and these conditions are consistent with hippocampal dendritic retraction following chronic stress in rodents. If the conditions that apply to hippocampal susceptibility following chronic stress in rodents are extended to individuals with Cushing's disease, MDD, and PTSD, then the data suggest that the hippocampus of these untreated individuals is primed for susceptibility to permanent damage.

Metabolic events may be problematic when compounded with Cushing's disease, MDD, PTSD, or even a history of chronic stress. Metabolic events can include, for example, small blocked vessels or restricted blood flow, and/or low sugar or oxygen levels. While these may seem like ailments of older individuals, the demographics of the typical American suggest that young adults and children are also at risk. The average American is heavier than ever, with over half of adults considered to be overweight and nearly one-third obese /44/; children show obesity trends as well /99/. Health risks associated with obesity include arteriosclerosis and Type II diabetes, which contribute to restricted blood flow and blood sugar imbalance, respectively. Both types of risks reflect metabolic challenges that damage the brain and compromise function. Indeed, heart transplant patients show increased risk for hippocampal atrophy /168/, emphasizing that the hippocampus is susceptible to restricted blood flow. Some metabolic events may be innocuous in isolation, without necessarily causing noticeable brain damage. could be problematic in a susceptible individual. Specifically, subthreshold metabolic events that are benign to the hippocampus of a healthy individual may be harmful to people with Cushing's disease, MDD, PTSD or a history of chronic stress because these latter individuals exhibit a vulnerable hippocampus that may be less able to respond effectively to an anomalous metabolic challenge. Moreover, these individuals are potentially vulnerable for an extended period because hippocampal plasticity can be compromised for weeks, months, or even years. The longer a hippocampus is compromised, the higher the probability that a metabolic event will occur, and the end result may be permanent hippocampal cell loss and an inability to recover, despite antiglucocorticoid or antidepressant treatment. In summary, a metabolic challenge in isolation may be innocuous to the healthy hippocampus, but with a history chronic stress or predisposition to Cushing's Disease, MDD, or PTSD, the brain and hippocampus in particular may be less resilient.

Despite the potential for individuals with Cushing's disease, MDD, and PTSD to have increased susceptibility for irreparable damage to the hippocampus, there are many possibilities for intervention and prevention. As already described, antidepressants have positive effects on mood and hippocampal volume in MDD /138/ and PTSD /16/. While some studies suggest that small hippocampal volume may reflect predisposition to develop MDD /61/ and PTSD /71,174/, the finding that both respond to treatment shows that hippocampal plasticity is possible

regardless of whether or not hippocampal volume is a cause or a consequence. Diet may also be an important contributing variable, as obese individuals typically consume high fat foods, which has been linked to many of the potentially detrimental events that can cause hippocampal damage. In the animal literature, a history of a high fat diet exacerbates stress-induced hippocampal dendritic retraction /6/ and impairs hippocampal function /50,64/. Reducing fat intake and eating healthy foods may reduce the risk of hippocampal vulnerability. Finally, the theory of Cognitive Reserve is used to explain the finding that individuals with more mental activity develop better, more elaborate neuronal connections that can withstand a greater amount of pathological damage before succumbing to disease /66,151/. Individuals engaging in intellectually-stimulating activities throughout their lives maintain their cognitive abilities when others begin to show cognitive decline /1,132,139/. In the animal literature, housing rats in enriched environments to provide opportunities for social interactions, exercise, and novel visual stimulation prevents the detrimental effects of chronic stress on hippocampal function /171/, and studies are underway to investigate the neurobiological substrates underlying this effect.

Taken together, compelling findings suggest that a small hippocampus may confer susceptibility to potential damage, without necessarily reflecting neuronal loss. A small hippocampus may have altered neuronal morphology, which is dynamic and reversible, as emphasized by responding to treatments and interventions that include antidepressant therapy, diet, and cognitive challenges. The problem is that the probability for permanent neuronal damage within the hippocampus is increased because this vulnerable period can extend over weeks, months, or years. This interval during which the hippocampus is susceptible provides an opportunistic window for a seemingly minor, innocuous metabolic event to cause permanent harm in a small “compromised” hippocampus. On the other hand, some individuals may recover without incident because the vulnerable period may not have coincided with a metabolic challenge. Consequently, a lack of consistent findings in the literature regarding hippocampal cell death and aging may be more indicative of a small hippocampal volume conveying susceptibility to damage rather than neuronal loss.

Key Words

CA, Cornu Ammonis; DSM IV, Diagnostic and Statistical Manual of Mental Disorders Fourth Version; GAS, General Adaptation Syndrome; GR, Glucocorticoid Receptor; HPA, Hypothalamic-Pituitary-Adrenal; IBO, Ibotenic Acid; MDD, Major Depressive Disorder; MR, Mineralocorticoid Receptor; PTSD, Post Traumatic Stress Disorder; SNS, Sympathetic Nervous System.

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References

1. Alexander GE, Furey ML, Grady CL, Pietrini P, Brady DR, Mentis MJ, Schapiro MB. Association of premorbid intellectual function with cerebral metabolism in Alzheimer's disease: implications for the cognitive reserve hypothesis. *Am J Psychiat* 1997;154:165–172. [PubMed: 9016263]
2. Angelucci L. The glucocorticoid hormone: From pedestal to dust and back. *Eur J Pharmacol* 2000;405:139–147. [PubMed: 11033321]
3. Arriza JL, Simerly RB, Swanson LW, Evans RM. The neuronal mineralocorticoid receptor as a mediator of glucocorticoid response. *Neuron* 1988;1:887–900. [PubMed: 2856104]

4. Aus Der Mühlen K, Ockenfels H. Morphologische Veränderungen im Diencephalon und Telencephalon nach Störungen des Regelkreis Adenohypophyse-Nebennierenrinde. *Z Zellforsch* 1969;93:126–141. [PubMed: 5784535]
5. Avitsur R, Stark JL, Sheridan JF. Social stress induces glucocorticoid resistance in subordinate animals. *Horm Behav* 2001;39:247–257. [PubMed: 11374910]
6. Baran SE, Campbell AM, Kleen JK, Foltz CH, Wright RL, Diamond DM, Conrad CD. Synergy between high fat diet and chronic stress retracts apical dendrites in CA3. *NeuroReport* 2005;16:39–43. [PubMed: 15618887]
7. Blazer DG, Kessler RC, McGonagle KA, Swartz MS. The prevalence and distribution of major depression in a national community sample: the National Comorbidity Survey. *Am J Psychiat* 1994;151:979–986. [PubMed: 8010383]
8. Bodnoff SR, Humphreys AG, Lehman JC, Diamond DM, Rose GM, Meaney MJ. Enduring effects of chronic corticosterone treatment on spatial learning, synaptic plasticity, and hippocampal neuropathology in young and mid-aged rats. *J Neurosci* 1995;15:61–69. [PubMed: 7823152]
9. Bonne O, Brandes D, Gilboa A, Gomori JM, Shenton ME, Pitman RK, Shalev AY. Longitudinal MRI study of hippocampal volume in trauma survivors with PTSD. *Am J Psychiat* 2001;158:1248–1251. [PubMed: 11481158]
10. Bossini L, Tavanti M, Lombardelli A, Calossi S, Polizzotto NR, Galli R, Vatti G, Pieraccini F, Castrogiovanni P. Changes in hippocampal volume in patients with posttraumatic stress disorder after sertraline treatment. *J Clin Psychopharmacol* 2007;27:233–235. [PubMed: 17414261]
11. Bourdeau I, Bard C, Forget H, Boulanger Y, Cohen H, Lacroix A. Cognitive function and cerebral assessment in patients who have Cushing's syndrome. *Endocrinology and metabolism clinics of North America* 2005;34:357–369. [PubMed: 15850847]ix
12. Bremner MA, Deeg DJ, Beekman AT, Penninx BW, Lips P, Hoogendijk WJ. Major Depression in Late Life Is Associated with Both Hypo- and Hypercortisolemia. *Biol Psychiat* 2007;62:479–486. [PubMed: 17481591]
13. Bremner JD, Randall P, Scott TM, Bronen RA, Seibyl JP, Southwick SM, Delaney RC, McCarthy G, Charney DS, Innis RB. MRI-based measurement of hippocampal volume in patients with combat-related posttraumatic stress disorder. *Am J Psychiat* 1995;152:973–981. [PubMed: 7793467]
14. Bremner JD, Randall P, Vermetten E, Staib L, Bronen RA, Mazure C, Capelli S, McCarthy G, Innis RB, Charney DS. Magnetic resonance imaging-based measurement of hippocampal volume in posttraumatic stress disorder related to childhood physical and sexual abuse--a preliminary report. *Biol Psychiat* 1997;41:23–32. [PubMed: 8988792]
15. Bremner JD, Narayan M, Anderson ER, Staib LH, Miller HL, Charney DS. Hippocampal volume reduction in major depression. *Am J Psychiat* 2000;157:115–117. [PubMed: 10618023]
16. Bremner JD, Vermetten E. Neuroanatomical changes associated with pharmacotherapy in posttraumatic stress disorder. *Ann NY Acad Sci* 2004;1032:154–157. [PubMed: 15677402]
17. Brooke SM, de Haas-Johnson AM, Kaplan JR, Sapolsky RM. Characterization of mineralocorticoid and glucocorticoid receptors in primate brain. *Brain Res* 1994;637:303–307. [PubMed: 8180810]
18. Brown ES, Varghese FP, McEwen BS. Association of depression with medical illness: Does cortisol play a role? *Biol Psychiat* 2004;55:1–9. [PubMed: 14706419]
19. Cameron HA, Gould E. Adult neurogenesis is regulated by adrenal steroids in the dentate gyrus. *Neuroscience* 1994;61:203–209. [PubMed: 7969902]
20. Choi DW. Glutamate neurotoxicity and diseases of the nervous system. *Neuron* 1988;1:623–634. [PubMed: 2908446]
21. Coburn-Litvak PS, Tata DA, Gorby HE, McCloskey DP, Richardson G, Anderson BJ. Chronic corticosterone affects brain weight, and mitochondrial, but not glial volume fraction in hippocampal area CA3. *Neuroscience* 2004;124:429–438. [PubMed: 14980392]
22. Conrad CD, Roy EJ. Selective loss of hippocampal granule cells following adrenalectomy: Implications for spatial memory. *J Neurosci* 1993;13:2582–2590. [PubMed: 8501524]
23. Conrad CD, Roy EJ. Dentate gyrus destruction and spatial learning impairment after corticosteroid removal in young and middle-aged rats. *Hippocampus* 1995;5:1–15. [PubMed: 7787942]
24. Conrad CD, Leone D, Nemivant RR, Roy EJ. Long-term adrenalectomy can increase or decrease hippocampal dentate gyrus volumes. *J Neuroendocrinol* 1997;9:355–361. [PubMed: 9181489]

25. Conrad CD, Lupien SJ, Thanasoulis LC, McEwen BS. The effects of Type I and Type II corticosteroid receptor agonists on exploratory behavior and spatial memory in the Y-Maze. *Brain Res* 1997;759:76–83. [PubMed: 9219865]
26. Conrad CD, Lupien SJ, McEwen BS. Support for a bimodal role for Type II adrenal steroid receptors in spatial memory. *Neurobiol Learn Mem* 1999;72:39–46. [PubMed: 10371714]
27. Conrad CD, Magariños AM, LeDoux JE, McEwen BS. Repeated restraint stress facilitates fear conditioning independently of causing hippocampal CA3 dendritic atrophy. *Behav Neurosci* 1999;113:902–913. [PubMed: 10571474]
28. Conrad CD, Jackson JL, Wise L. Chronic stress enhances ibotenic acid-induced damage selectively within the hippocampal CA3 region of male, but not female rats. *Neuroscience* 2004;125:759–767. [PubMed: 15099689]
29. Conrad CD. The relationship between acute glucocorticoid levels and hippocampal function depends upon task aversiveness and memory processing stage. *Nonlinear Biol Toxicol Med* 2005;3:57–78.
30. Conrad CD. What is the functional significance of chronic stress-induced CA3 dendritic retraction within the hippocampus? *Behav Cognit Neurosci Rev* 2006;5:41–60. [PubMed: 16816092]
31. Conrad CD, McLaughlin KJ, Harman JS, Foltz C, Wiczorek L, Lightner E, Wright RL. Chronic glucocorticoids increase hippocampal vulnerability to neurotoxicity under conditions that produce CA3 dendritic retraction but fail to impair spatial recognition memory. *J Neurosci* 2007;27:8278–8285. [PubMed: 17670974]
32. Czéh B, Lucassen PJ. What causes the hippocampal volume decrease in depression? Are neurogenesis, glial changes and apoptosis implicated? *Eur Arch Psychiatry Clin Neurosci* 2007;257:250–260. [PubMed: 17401728]
33. Dachir S, Kadar T, Robinzon B, Levy A. Nimodipine's protection against corticosterone-induced morphological changes in the hippocampus of young rats. *Brain Res* 1997;748:175–183. [PubMed: 9067459]
34. Dallman, MF.; Bhatnagar, S.; Viau, V. Hypothalamo-Pituitary-Adrenal Axis. In: Fink, G., editor. *Encyclopedia of Stress*. New York NY: Academic Press; 2000. p. 468477
35. Dallman MF. Modulation of stress responses: How we cope with excess glucocorticoids. *Exp Neurol* 2007;206:179–182. [PubMed: 17628543]
36. Daniels WMU, Jaffer A, Engelbrecht AH, Russell VA, Taljaard JJF. The effect of intrahippocampal injection of kainic acid on corticosterone release in rats. *Neurochem Res* 1990;15:495–499. [PubMed: 2164646]
37. de Kloet ER. Brain corticosteroid receptor balance and homeostatic control. *Front Neuroendocrinol* 1991;12:95–164.
38. de Kloet ER, Oitzl MS, Joëls M. Stress and cognition: Are corticosteroids good or bad guys? *TINS* 1999;22:422–426. [PubMed: 10481183]
39. Diamond DM, Fleshner M, Ingersoll N, Rose GM. Psychological stress impairs spatial working memory: Relevance to electrophysiological studies of hippocampal function. *Behav Neurosci* 1996;110:661–672. [PubMed: 8864259]
40. Dunn JD, Orr SE. Differential plasma corticosterone responses to hippocampal stimulation. *Exp Brain Res* 1984;54:1–6. [PubMed: 6321219]
41. Evans RM, Arriza JL. A molecular framework for the actions of glucocorticoid hormones in the nervous system. *Neuron* 1989;2:1105–1112. [PubMed: 2696502]
42. Fava M, Kendler KS. Major depressive disorder. *Neuron* 2000;28:335–341. [PubMed: 11144343]
43. Feldman S, Conforti N. Participation of the dorsal hippocampus in the glucocorticoid feedback effect on adrenocortical activity. *Neuroendocrinology* 1980;30:52–55. [PubMed: 7354890]
44. Flegal KM, Carroll MD, Ogden CL, Johnson CL. Prevalence and trends in obesity among US adults, 1999–2000. *JAMA* 2002;288:1723–1727. [PubMed: 12365955]
45. Franceschi M, Airaghi L, Gramigna C, Truci G, Manfredi MG, Canal N, Catania A. ACTH and cortisol secretion in patients with Alzheimer's disease. *J Neurol Neurosurg Psychiatry* 1991;54:836–837. [PubMed: 1659618]
46. Frodl T, Meisenzahl EM, Zetsche T, Born C, Groll C, Jager M, Leinsinger G, Bottlender R, Hahn K, Moller HJ. Hippocampal changes in patients with a first episode of major depression. *Am J Psychiat* 2002;159:1112–1118. [PubMed: 12091188]

47. Fuchs E, Flügge G, Ohl F, Lucassen P, Vollmann-Honsdorf GK, Michaelis T. Psychosocial stress, glucocorticoids, and structural alterations in the tree shrew hippocampus. *Physiol Behav* 2001;73:285–291. [PubMed: 11438353]
48. Funder, JW.; Pearce, PT.; Smith, R.; Smith, AI. *Science*. Vol. 242. New York, NY: 1988. Mineralocorticoid action: target tissue specificity is enzyme, not receptor, mediated; p. 583-585.
49. Galea LAM, McEwen BS, Tanapat P, Deak T, Spencer RL, Dhabhar FS. Sex differences in dendritic atrophy of CA3 pyramidal neurons in response to chronic restraint stress. *Neuroscience* 1997;81:689–697. [PubMed: 9316021]
50. Goldbart AD, Row BW, Kheirandish-Gozal L, Cheng Y, Brittan KR, Gozal D. High fat/refined carbohydrate diet enhances the susceptibility to spatial learning deficits in rats exposed to intermittent hypoxia. *Brain Res* 2006;1090:190–196. [PubMed: 16674930]
51. Gomez RG, Fleming SH, Keller J, Flores B, Kenna H, Debatista C, Solvason B, Schatzberg AF. The Neuropsychological Profile of Psychotic Major Depression and its Relation to Cortisol. *Biol Psychiat* 2006;60:472–478. [PubMed: 16483550]
52. Gould E, Tanapat P, McEwen BS, Flügge G, Fuchs E. Proliferation of granule cell precursors in the dentate gyrus of adult monkeys is diminished by stress. *PNAS* 1998;95:3168–3171. [PubMed: 9501234]
53. Grillon C, Smith K, Haynos A, Nieman LK. Deficits in hippocampus-mediated Pavlovian conditioning in endogenous hypercortisolism. *Biol Psychiatry* 2004;56:837–843. [PubMed: 15576060]
54. Gurvits TV, Shenton ME, Hokama H, Ohta H, Lasko NB, Orr SP, Kikinis R, Jolesz FA, McCarley RW, Pitman RK. Reduced hippocampal volume on magnetic resonance imaging in chronic post-traumatic stress disorder. *Biol Psychiat* 1996;40:1091–1099. [PubMed: 8931911]
55. Hajszan T, MacLusky NJ, Leranth C. Short-term treatment with the antidepressant fluoxetine triggers pyramidal dendritic spine synapse formation in rat hippocampus. 2005;21:1299–1303.
56. Henn FA, Vollmayr B. Stress models of depression: Forming genetically vulnerable strains. *Neurosci Biobehav Rev* 2005;29:799–804. [PubMed: 15925700]
57. Herman JP, Patel PD, Akil H, Watson SJ. Localization and regulation of glucocorticoid and mineralocorticoid receptor messenger RNAs in the hippocampal formation of the rat. *Mol Endocrinol* 1989;3:1886–1894. [PubMed: 2558306]
58. Herman JP, Schäfer MK-H, Young EA, Thompson R, Douglass J, Akil H, Watson SJ. Evidence for hippocampal regulation of neuroendocrine neurons of the hypothalamopituitary-adrenocortical axis. *J Neurosci* 1989;9:3072–3082. [PubMed: 2795152]
59. Herman JP, Spencer R. Regulation of hippocampal glucocorticoid receptor gene transcription and protein expression in vivo. *J Neurosci* 1998;18:7462–7473. [PubMed: 9736665]
60. Hook JN, Giordani B, Scheingart DE, Guire K, Giles J, Ryan K, Gebarski SS, Lagenecker SA, Starkman MN. Patterns of cognitive change over time and relationship to age following successful treatment of Cushing's disease. *J Intl' Neuropsychol Soc* 2007;13:21–29.
61. Janssen J, Hulshoff Pol HE, Lampe IK, Schnack HG, de Leeuw FE, Kahn RS, Heeren TJ. Hippocampal changes and white matter lesions in early-onset depression. *Biol Psychiat* 2004;56:825–831. [PubMed: 15576058]
62. Joëls M, Karst H, Alfarez D, Heine VM, Qin Y, van Riel E, Verkuyl M, Lucassen PJ, Krugers HJ. Effects of chronic stress on structure and cell function in rat hippocampus and hypothalamus. *Stress* 2004;7:221–231. [PubMed: 16019587]
63. Johnson EO, Brady L, Gold PW, Chrousos GP. Distribution of hippocampal mineralocorticoid and glucocorticoid receptor mRNA in a glucocorticoid resistant nonhuman primate. *Steroids* 1996;61:69–73. [PubMed: 8750435]
64. Kanoski SE, Meisel RL, Mullins AJ, Davidson TL. The effects of energy-rich diets on discrimination reversal learning and on BDNF in the hippocampus and prefrontal cortex of the rat. *Behav Brain Res* 2007;182:57–66. [PubMed: 17590450]
65. Karl A, Schaefer M, Malta LS, Dörfel D, Rohleder N, Werner A. A meta-analysis of structural brain abnormalities in PTSD. *Neurosci Biobehav Rev* 2006;30:1004–1031. [PubMed: 16730374]
66. Katzman R. Education and the prevalence of dementia and Alzheimer's disease. *Neurology* 1993;43:13–20. [PubMed: 8423876]

67. Keilhoff G, Wolf G, Stastny F, Schmidt W. Quinolate neurotoxicity and glutamatergic structures. *Neuroscience* 1990;34:235–242. [PubMed: 1970138]
68. Keller J, Flores B, Gomez RG, Solvason HB, Kenna H, Williams GH, Schatzberg AF. Cortisol circadian rhythm alterations in psychotic major depression. *Biol Psychiat* 2006;60:275–281. [PubMed: 16458262]
69. Kitraki E, Kremmyda O, Youlatos D, Alexis M, Kittas C. Spatial performance and corticosteroid receptor status in the 21-day restraint stress paradigm. 2004;1018:323–327.
70. Kleen JK, Sitomer MT, Killeen PR, Conrad CD. Chronic stress impairs spatial memory and motivation for reward without disrupting motor ability and motivation to explore. *Behav Neurosci* 2006;120:842–851. [PubMed: 16893290]
71. Koenen KC, Lyons MJ, Goldberg J, Simpson J, Williams WM, Toomey R, Eisen SA, True W, Tsuang MT. Co-twin control study of relationships among combat exposure, combat-related PTSD, and other mental disorders. *J Trauma Stress* 2003;16:433–438. [PubMed: 14584626]
72. Krugers HJ, Knollema S, Kemper RHA, Ter Horst GJ, Korf J. Down-regulation of the hypothalamo-pituitary-adrenal axis reduces brain damage and number of seizures following hypoxia/ischaemia in rats. *Brain Res* 1995;690:41–47. [PubMed: 7496805]
73. Kudielka BM, Schmidt-Reinwald AK, Hellhammer DH, Schürmeyer T, Kirschbaum C. Psychosocial stress and HPA functioning: No evidence for a reduced resilience in healthy elderly men. *Stress* 2000;3:229–240. [PubMed: 10938584]
74. Landfield PW, Waymire JC, Lynch G. Hippocampal aging and adrenocorticoids: Quantitative correlations. *Science* 1978;202:1098–1102. [PubMed: 715460]
75. Landfield PW, Baskin RK, Pitler TA. Brain aging correlates: Retardation by hormonal/pharmacological treatments. *Science* 1981;214:581–584. [PubMed: 6270791]
76. Leverenz JB, Wilkinson CW, Wamble M, Corbin S, Grabber JE, Raskind MA, Peskind ER. Effect of chronic high-dose exogenous cortisol on hippocampal neuronal number in aged nonhuman primates. *J Neurosci* 1999;19:2356–2361. [PubMed: 10066285]
77. Levinson DF. The genetics of depression: A review. *Biol Psychiat* 2006;60:84–92. [PubMed: 16300747]
78. López JF, Chalmers DT, Little KY, Watson SJ. Regulation of serotonin_{1A}, glucocorticoid, and mineralocorticoid receptor in rat and human hippocampus: Implications for the neurobiology of depression. *Biol Psychiat* 1998;43:547–573. [PubMed: 9564441]
79. Lucassen PJ, Muller MB, Holsboer F, Bauer J, Holtrop A, Wouda J, Hoogendijk WJ, De Kloet ER, Swaab DF. Hippocampal apoptosis in major depression is a minor event and absent from subareas at risk for glucocorticoid overexposure. *Am J Pathol* 2001;158:453–468. [PubMed: 11159183]
80. Luine V, Martinez C, Villegas M, Magariños AM, McEwen BS. Restraint stress reversibly enhances spatial memory performance. *Physiol Behav* 1996;59:27–32. [PubMed: 8848486]
81. Lupien SJ, Buss C, Schramek TE, Maheu F, Pruessner J. Hormetic influence of glucocorticoids on human memory. *Nonlin Biol Toxicol Med* 2005;3:23–56.
82. Magariños AM, McEwen BS. Stress-induced atrophy of apical dendrites of hippocampal CA3c neurons: Comparison of stressors. *Neuroscience* 1995;69:83–88. [PubMed: 8637635]
83. Magariños AM, McEwen BS. Stress-induced atrophy of apical dendrites of hippocampal CA3c neurons: Involvement of glucocorticoid secretion and excitatory amino acid receptors. *Neuroscience* 1995;69:89–98. [PubMed: 8637636]
84. Malberg JE, Eisch AJ, Nestler EJ, Duman RS. Chronic antidepressant treatment increases neurogenesis in adult rat hippocampus. *J Neurosci* 2000;20:9104–9110. [PubMed: 11124987]
85. Mandel AJ, Walter LF. Plasma corticosteroids: Changes in concentration after stimulation of hippocampus and amygdala. *Science* 1963;135:1212.
86. Mauri M, Sinforiani E, Bono G, Vignati F, Berselli ME, Attanasio R, Nappi G. Memory impairment in Cushing's disease. *Acta Neurol Scand* 1993;87:52–55. [PubMed: 8424312]
87. McEwen BS, Weiss JM, Schwartz LS. Selective retention of corticosterone by limbic structures in rat brain. *Nature* 1968;220:911–912. [PubMed: 4301849]
88. McEwen BS, Weiss JM, Schwartz LS. Uptake of corticosterone by rat brain and its concentration by certain limbic structures. *Brain Res* 1969;16:227–241. [PubMed: 5348850]

89. McEwen BS. Stress and hippocampal plasticity. *Annu Rev Neurosci* 1999;22:105–122. [PubMed: 10202533]
90. McLaughlin KJ, Baran SE, Wright RL, Conrad CD. Chronic stress enhances spatial memory in ovariectomized female rats despite CA3 dendritic retraction: Possible involvement of CA1 neurons. *Neuroscience* 2005;135:1045–1054. [PubMed: 16165283]
91. McLaughlin KJ, Gomez JL, Baran SE, Conrad CD. The effects of chronic stress on hippocampal morphology and function: An evaluation of chronic restraint paradigms. *Brain Res* 2007;1161:56–64. [PubMed: 17603026]
92. Moghaddam B, Bolinao ML, Stein-Behrens B, Sapolsky R. Glucocorticoids mediate the stress-induced extracellular accumulation of glutamate. *Brain Res* 1994;655:251–254. [PubMed: 7812782]
93. Müller MB, Lucassen PJ, Yassouridis A, Hoogendijk WJG, Holsboer F, Swaab DF. Neither major depression nor glucocorticoid treatment affects the cellular integrity of the human hippocampus. *Eur J Neurosci* 2001;14:1603–1612. [PubMed: 11860455]
94. Neumeister A, Wood S, Bonne O, Nugent AC, Luckenbaugh DA, Young T, Bain EE, Charney DS, Drevets WC. Reduced hippocampal volume in unmedicated, remitted patients with major depression versus control subjects. *Biol Psychiat* 2005;57:935–937. [PubMed: 15820716]
95. Newell-Price J, Trainer P, Besser M, Grossman A. The diagnosis and differential diagnosis of Cushing's syndrome and pseudo-Cushing's states. *Endocr Rev* 1998;19:647–672. [PubMed: 9793762]
96. Newell-Price J, Bertagna X, Grossman AB, Nieman LK. Cushing's syndrome. *Lancet* 2006;367:1605–1617. [PubMed: 16698415]
97. Nichols NR, Zieba M, Bye N. Do glucocorticoids contribute to brain aging? *Brain Res Rev* 2001;37:273–286. [PubMed: 11744092]
98. O'Keefe, J.; Nadel, L. Oxford, England: Clarendon Press; 1978. *The Hippocampus as a Cognitive Map*.
99. Ogden CL, Flegal KM, Carroll MD, Johnson CL. Prevalence and trends in overweight among US children and adolescents, 1999–2000. *JAMA* 2002;288:1728–1732. [PubMed: 12365956]
100. Oitzl MS, de Kloet ER. Selective corticosteroid antagonists modulate specific aspects of spatial orientation learning. *Behav Neurosci* 1992;106:62–71. [PubMed: 1313244]
101. Oitzl MS, van Haarst AD, Sutanto W, de Kloet ER. Corticosterone, brain mineralocorticoid receptors (MRs) and the activity of the hypothalamic-pituitary-adrenal (HPA) axis: the Lewis rat as an example of increased central MR capacity and a hyporesponsive HPA axis. *Psychoneuroendocrinology* 1995;20:655–675. [PubMed: 8584606]
102. Orchinik M, Carroll SS, Li Y-H, McEwen BS, Weiland NG. Heterogeneity of hippocampal GABA_A receptors: Regulation by corticosterone. *J Neurosci* 2001;21:330–339. [PubMed: 11150350]
103. Patel PD, Lopez JF, Lyons DM, Burke S, Wallace M, Schatzberg AF. Glucocorticoid and mineralocorticoid receptor mRNA expression in squirrel monkey brain. *J Psychiat Res* 2000;34:383–392. [PubMed: 11165305]
104. Pavlides C, Watanabe Y, McEwen BS. Effects of glucocorticoids on hippocampal long-term potentiation. *Hippocampus* 1993;3:183–192. [PubMed: 8353605]
105. Pavlides C, Nivón LG, McEwen BS. Effects of chronic stress on hippocampal long-term potentiation. *Hippocampus* 2002;12:245–257. [PubMed: 12000121]
106. Pearson Murphy, BE. Glucocorticoids, Overview. In: Fink, G., editor. *Encyclopedia of Stress*. London, UK: Academic Press; 2000. p. 244-261.
107. Pederson CL, Maurer SH, Kaminski PL, Zander KA, Peters CM, Stokes-Crowe LA, Osborn RE. Hippocampal volume and memory performance in a community-based sample of women with posttraumatic stress disorder secondary to child abuse. *J Trauma Stress* 2004;17:37–40. [PubMed: 15027791]
108. Pham K, Nacher J, Hof PR, McEwen BS. Repeated restraint stress suppresses neurogenesis and induces biphasic PSA_{NCAM} expression in the adult rat dentate gyrus. *Eur J Neurosci* 2003;17:879–886. [PubMed: 12603278]

109. Popov VI, Bocharova LS. Hibernation-induced structural changes in synaptic contacts between mossy fibres and hippocampal pyramidal neurons. *Neuroscience* 1992;48:53–62. [PubMed: 1584425]
110. Popov VI, Bocharova LS, Bragin AG. Repeated changes of dendritic morphology in the hippocampus of ground squirrels in the course of hibernation. *Neuroscience* 1992;48:45–51. [PubMed: 1584424]
111. Purves D, Lichtman JW. Geometrical differences among homologous neurons in mammals. *Science* 1985;228:298–302. [PubMed: 3983631]
112. Reagan LP, Rosell DR, Wood GE, Spedding M, Munoz C, Rothstein J, McEwen BS. Chronic restraint stress up-regulates GLT-1 mRNA and protein expression in the rat hippocampus: reversal by tianeptine. *PNAS* 2004;101:2179–2184. [PubMed: 14766991]
113. Reul JM, de Kloet ER. Two receptor systems for corticosterone in rat brain: Microdistribution and differential occupation. *Endocrinology* 1985;117:2505–2511. [PubMed: 2998738]
114. Reus VI, Wolkowitz OM. Antiglucocorticoid drugs in the treatment of depression. *Expert Opin Investig Drugs* 2001;10:1789–1796.
115. Rosenbrock H, Koros E, Bloching A, Podhorna J, Borsini F. Effect of chronic intermittent restraint stress on hippocampal expression of marker proteins for synaptic plasticity and progenitor cell proliferation in rats. *Brain Res* 2005;1040:55–63. [PubMed: 15804426]
116. Sánchez MM, Young LJ, Plotsky PM, Insel TR. Distribution of corticosteroid receptors in the rhesus brain: Relative absence of glucocorticoid receptors in the hippocampal formation. *J Neurosci* 2000;20:4657–4668. [PubMed: 10844035]
117. Sapolsky RM, Krey LC, McEwen BS. The adrenocortical stress-response in the aged male rat: Impairment of recovery from stress. *Exp Gerontol* 1983;18:55–64. [PubMed: 6683660]
118. Sapolsky RM, Krey LC, McEwen BS. Corticosterone receptors decline in a site-specific manner in the aged-rat brain. *Brain Res* 1983;289:235–240. [PubMed: 6661643]
119. Sapolsky RM, Krey LC, McEwen BS. Stress down-regulates corticosterone receptors in a site-specific manner in the brain. *Endocrinology* 1984;114:287–292. [PubMed: 6690273]
120. Sapolsky RM, Krey LC, McEwen BS. Glucocorticoid-sensitive hippocampal neurons are involved in terminating the adrenocortical stress response. *PNAS* 1984;81:6174–6177. [PubMed: 6592609]
121. Sapolsky RM. A mechanism for glucocorticoid toxicity in the hippocampus: Increased neuronal vulnerability to metabolic insults. *J Neurosci* 1985;5:1228–1232. [PubMed: 3998819]
122. Sapolsky RM. Glucocorticoid toxicity in the hippocampus: Temporal aspects of neuronal vulnerability. *Brain Res* 1985;359:300–305. [PubMed: 4075151]
123. Sapolsky RM, Krey LC, McEwen BS. Prolonged glucocorticoid exposure reduces hippocampal neuron number: Implications for aging. *J Neurosci* 1985;5:1222–1227. [PubMed: 3998818]
124. Sapolsky RM, Pulsinelli WA. Glucocorticoids potentiate ischemic injury to neurons: Therapeutic implications. *Science* 1985;229:1397–1399. [PubMed: 4035356]
125. Sapolsky RM. Glucocorticoid toxicity in the hippocampus: Reversal by supplementation with brain fuels. *J Neurosci* 1986;6:2240–2244. [PubMed: 3746406]
126. Sapolsky RM. Glucocorticoid toxicity in the hippocampus: Temporal aspects of synergy with kainic acid. *Neuroendocrinology* 1986;43:440–444. [PubMed: 3736786]
127. Sapolsky RM, Krey LC, McEwen BS. The neuroendocrinology of stress and aging: The glucocorticoid cascade hypothesis. *Endocr Rev* 1986;7:284–301. [PubMed: 3527687]
128. Sapolsky RM, Packan DR, Vale WW. Glucocorticoid toxicity in the hippocampus: *In vitro* demonstration. *Brain Res* 1988;453:367–371. [PubMed: 3401775]
129. Sapolsky RM, Uno H, Rebert CS, Finch CE. Hippocampal damage associated with prolonged glucocorticoid exposure in primates. *J Neurosci* 1990;10:2897–2902. [PubMed: 2398367]
130. Sapolsky, RM. Cambridge, MA: MIT Press; 1992. *Stress, the Aging Brain, and the Mechanisms of Neuron Death*.
131. Sarrieau A, Dussailant M, Sapolsky RM, Aitken DH, Olivier A, Lal S, Rostene WH, Quirion R, Meaney MJ. Glucocorticoid binding sites in human temporal cortex. *Brain Res* 1988;442:157–160. [PubMed: 3359250]

132. Schaie KW. The Seattle longitudinal studies of adult intelligence. *Curr Dir Psychol Sci* 1993;26:129–137.
133. Schuff N, Neylan TC, Lenoci MA, Du AT, Weiss DS, Marmar CR, Weiner MW. Decreased hippocampal N-acetylaspartate in the absence of atrophy in posttraumatic stress disorder. *Biol Psychiat* 2001;50:952–959. [PubMed: 11750891]
134. Seckl JR, Walker BR. 11beta-hydroxysteroid dehydrogenase type 1 as a modulator of glucocorticoid action: from metabolism to memory. *TEM* 2004;15:418–424. [PubMed: 15519888]
135. Seckl JR.; Yau, JLW.; Homes, MC. The role of 11-hydroxysteroid dehydrogenases in the regulation of corticosteroid activity in the brain. In: Steckler, T.; Kaline, NH.; Reul, JMHM., editors. *Handbook of Stress and the Brain*. New York, NY: Elsevier; 2005. p. 313–328.
136. Selye, H. N.Y.: McGraw-Hill Book Co; 1976. *The Stress of Life*.
137. Sheline YI, Wang PW, Gado MH, Csernansky JC, Vannier MW. Hippocampal atrophy in recurrent major depression. *PNAS* 1996;93:3908–3913. [PubMed: 8632988]
138. Sheline YI, Gado MH, Kraemer HC. Untreated depression and hippocampal volume loss. *Am J Psychiat* 2003;160:1516–1518. [PubMed: 12900317]
139. Shimamura AP, Berry JM, Mangels JA, Rusting CL, Jurica PJ. Memory and cognitive abilities in university professors: Evidence for successful aging. *Psychol Sci* 1995;6:271–277.
140. Sloviter RS, Valiquette G, Abrams GM, Ronk EC, Sollas AL, Paul LA, Neubort S. Selective loss of hippocampal granule cells in the mature rat brain after adrenalectomy. *Science* 1989;243:535–538. [PubMed: 2911756]
141. Smith-Swintosky VL, Pettigrew LC, Sapolsky RM, Phares C, Craddock SD, Brooke SM, Mattson MP. Metyrapone, an inhibitor of glucocorticoid production, reduces brain injury induced by focal and global ischemia and seizures. *J Cereb Blood Flow Metab* 1996;16:585–598. [PubMed: 8964797]
142. Sousa N, Madeira MD, Paula-Barbosa MM. Effects of corticosterone treatment and rehabilitation on the hippocampal formation of neonatal and adult rats. An unbiased stereological study. *Brain Res* 1998;794:199–210. [PubMed: 9622630]
143. Spencer RL, McEwen BS. Adaptation of the hypothalamic-pituitary-adrenal axis to chronic ethanol stress. *Neuroendocrinology* 1990;52:481–489. [PubMed: 2126355]
144. Starkman MN, Scheingart DE, Schork MA. Cushing's syndrome after treatment: Changes in cortisol and ACTH levels, and amelioration of the Depressive syndrome. *Psychiat Res* 1986;19:177–188.
145. Starkman MN, Gebarski SS, Berent S, Scheingart DE. Hippocampal formation volume, memory dysfunction, and cortisol levels in patients with Cushing's syndrome. *Biol Psych* 1992;32:756–765.
146. Starkman MN, Giordani B, Gebarski SS, Berent S, Schork MA, Scheingart DE. Decrease in cortisol reverses human hippocampal atrophy following treatment of Cushing's Disease. *Biol Psychiat* 1999;46:1595–1602. [PubMed: 10624540]
147. Starkman MN, Giordani B, Gebarski SS, Scheingart DE. Improvement in learning associated with increase in hippocampal formation volume. *Biol Psychiat* 2003;53:233–238. [PubMed: 12559656]
148. Stein BA, Sapolsky RM. Chemical adrenalectomy reduces hippocampal damage induced by kainic acid. *Brain Res* 1988;473:175–180. [PubMed: 3208122]
149. Stein MB, Koverola C, Hanna C, Torchia MG, McClarty B. Hippocampal volume in women victimized by childhood sexual abuse. *Psychol Med* 1997;27:951–959. [PubMed: 9234472]
150. Stein-Behrens BA, Lin WJ, Sapolsky RM. Physiological elevations of glucocorticoids potentiate glutamate accumulation in the hippocampus. *J Neurochem* 1994;63:596–602. [PubMed: 7913489]
151. Stern Y. What is cognitive reserve? Theory and research application of the reserve concept. *J Int Neuropsychol Soc* 2002;8:448–460. [PubMed: 11939702]
152. Stockmeier CA, Mahajan GJ, Konick LC, Overholser JC, Jurjus GJ, Meltzer HY, Uylings HB, Friedman L, Rajkowska G. Cellular changes in the postmortem hippocampus in major depression. *Biol Psychiatry* 2004;56:640–650. [PubMed: 1552247]
153. Sunanda, Rao MS, Raju TR. Effect of chronic restraint stress on dendritic spines and excrescences of hippocampal CA3 pyramidal neurons-A quantitative study. *Brain Res* 1995;694:312–317. [PubMed: 8974660]

154. Swanwick GR, Kirby M, Bruce I, Buggy F, Coen RF, Coakley D, Lawlor BA. Hypothalamic-pituitary-adrenal axis dysfunction in Alzheimer's disease: lack of association between longitudinal and cross-sectional findings. *The American journal of psychiatry* 1998;155:286–289. [PubMed: 9464214]
155. Tata DA, Marciano VA, Anderson BJ. Synapse loss from chronically elevated glucocorticoids: relationship to neuropil volume and cell number in hippocampal area CA3. *J Comp Neurol* 2006;498:363–374. [PubMed: 16871536]
156. Tombaugh GC, Yang SH, Swanson RA, Sapolsky RM. Glucocorticoids exacerbate hypoxic and hypoglycemic hippocampal injury in vitro: Biochemical correlates and a role for astrocytes. *J Neurochem* 1992;59:137–146. [PubMed: 1613495]
157. Uno H, Tarara R, Else JG, Suleman MA, Sapolsky RM. Hippocampal damage associated with prolonged and fatal stress in primates. *J Neurosci* 1989;9:1705–1711. [PubMed: 2723746]
158. Vasterling JJ, Duke LM, Brailey K, Constans JI, Allain AN Jr, Sutker PB. Attention, learning, and memory performances and intellectual resources in Vietnam veterans: PTSD and no disorder comparisons. *Neuropsychology* 2002;16:5–14. [PubMed: 11853357]
159. Vermetten E, Vythilingam M, Southwick SM, Charney DS, Bremner JD. Long-term treatment with paroxetine increases verbal declarative memory and hippocampal volume in posttraumatic stress disorder. *Biol Psychiat* 2003;54:693–702. [PubMed: 14512209]
160. Videbech P, Ravnkilde B. Hippocampal volume and depression: A meta-analysis of MRI studies. *J Neurosci* 2004;161:1957–1966.
161. Villarreal G, Hamilton DA, Petropoulos H, Driscoll I, Rowland LM, Griego JA, Kodituwakku PW, Hart BL, Escalona R, Brooks WM. Reduced hippocampal volume and total white matter volume in posttraumatic stress disorder. *Biol Psychiat* 2002;52:119–125. [PubMed: 12114003]
162. Virgin CEJ, Ha TP-T, Packan DR, Tombaugh GC, Yang SH, Horner HC, Sapolsky RM. Glucocorticoids inhibit glucose transport and glutamate uptake in hippocampal astrocytes: Implications for glucocorticoid neurotoxicity. *J Neurochem* 1991;57:1422–1428. [PubMed: 1680166]
163. Vollmann-Honsdorf GK, Flugge G, Fuchs E. Chronic psychosocial stress does not affect the number of pyramidal neurons in tree shrew hippocampus. *Neurosci Lett* 1997;233:121–124. [PubMed: 9350847]
164. Vyas A, Pillai AG, Chattarji S. Recovery after chronic stress fails to reverse amygdaloid neuronal hypertrophy and enhanced anxiety-like behavior. *Neuroscience* 2004;128:667–673. [PubMed: 15464275]
165. Wang PS, Lo M-J, Kau M-M. Glucocorticoids and aging. *J Formos Med Assoc* 1997;96:792–801. [PubMed: 9343978]
166. Watanabe Y, Gould E, Cameron HA, Daniels DC, McEwen BS. Phenytoin prevents stress-and corticosterone-induced atrophy of CA3 pyramidal neurons. *Hippocampus* 1992;2:431–436. [PubMed: 1308199]
167. Watanabe Y, Gould E, McEwen BS. Stress induces atrophy of apical dendrites of hippocampal CA3 pyramidal neurons. *Brain Res* 1992;588:341–345. [PubMed: 1393587]
168. Wilner AP, de Varennes B, Gregoire PA, Lupien S, Pruessner JC. Glucocorticoids and hippocampal atrophy after heart transplantation. *The Annals of thoracic surgery* 2002;73:1965–1967. [PubMed: 12078806]
169. Woolley CS, Gould E, McEwen BS. Exposure to excess glucocorticoids alters dendritic morphology of adult hippocampal pyramidal neurons. *Brain Res* 1990;531:225–231. [PubMed: 1705153]
170. Wright RL, Lightner EN, Harman JS, Meijer OC, Conrad CD. Attenuating corticosterone levels on the day of memory assessment prevents chronic stress-induced impairments in spatial memory. *Eur J Neurosci* 2006;24:595–605. [PubMed: 16903861]
171. Wright RL, Conrad CD. Enriched environment prevents chronic stress-induced spatial learning and memory deficits. *Behav Brain Res* 2008;187:41–47. [PubMed: 17904657]
172. Yehuda R. Psychoneuroendocrinology of post-traumatic stress disorder. *Psychiat Clin North Am* 1998;21:359–379.

173. Yehuda R, McFarlane AC, Shalev AY. Predicting the development of posttraumatic stress disorder from the acute response to a traumatic event. *Biological psychiatry* 1998;44:1305–1313. [PubMed: 9861473]
174. Yehuda R, Schmeidler J, Wainberg M, Binder-Brynes K, Duvdevani T. Vulnerability to posttraumatic stress disorder in adult offspring of Holocaust survivors. *The American journal of psychiatry* 1998;155:1163–1171. [PubMed: 9734537]