

Physiological and Biochemical Tools Useful in Drought-Tolerance Detection in Genotypes of Winter Triticale: Accumulation of Ferulic Acid Correlates with Drought Tolerance

TOMASZ HURA^{1,*}, STANISŁAW GRZESIAK¹, KATARZYNA HURA², ELISABETH THIEMT³, KRZYSZTOF TOKARZ¹ and MARIA WĘDZONY^{1,4}

¹The Franciszek Górski Institute of Plant Physiology, Polish Academy of Sciences, Niezapominajek 21, 30-239 Kraków, Poland, ²Department of Plant Physiology, Faculty of Agriculture and Economics, Agricultural University, Podłużna 3, 30-239 Kraków, Poland, ³University of Hohenheim, State Plant Breeding Institute, D-70593 Stuttgart, Germany and ⁴Pedagogical University of Kraków, Podchorążych 2, 30-084 Kraków, Poland

Received: 20 March 2007 Returned for revision: 8 May 2007 Accepted: 8 June 2007 Published electronically: 7 August 2007

• *Background and Aims* The objectives of this study were to investigate whether a classification of triticale genotypes into drought-tolerant and drought-sensitive types based on field performance trials correlates with a classification based on measurements of some physiological and biochemical parameters in greenhouse conditions. In addition, an examination was carried out of whether ferulic acid, as the main origin of the blue fluorescence produced, contributes to drought tolerance.

• *Methods* Ten winter triticale genotypes were examined, five known to be drought tolerant and five drought sensitive. Measurements of the osmotic potential, leaf gas exchange, chlorophyll fluorescence, and blue and red fluorescence were performed. In addition, analysis of the total pool of phenolic compounds and ferulic acid as well as the measurements of PAL (L-phenylalanine ammonia-lyase) activity were carried out.

• *Key Results* In agreement with field trials, three out of five cultivars ('Lamberto', 'Timbo' and 'Piano') were classified as drought tolerant. However, in the case of cultivar 'Babor', included in the group of drought-sensitive cultivars, the values obtained for some measured parameters were close to (F_v/F_m') , phenolics content, osmotic potential) or even better than (non-photochemical quenching, red and blue fluorescence, ferulic acid content) those for drought-tolerant genotypes. Cultivars 'Imperial', 'Ticino', 'Trimaran' and 'Boreas' were included in the drought-sensitive group, whereas cultivars 'Focus' and 'Kitaro' were included in the moderately sensitive group. • *Conclusions* The experiments confirmed that the period of flowering, the critical phase for plants as far as water demand is concerned, is suitable for plant screening and differentiation due to their tolerance to drought. The most important criteria which enabled creation of the ranking list of plants, from those sensitive to drought to those tolerant to drought, were the ability to perform the process of osmoregulation, the efficiency of the utilization of excitation energy by the photosynthetic apparatus and the functioning of protective mechanisms involving the level of ferulic acid in leaf tissues.

Key words: Blue fluorescence, chlorophyll fluorescence, drought, ferulic acid, osmotic potential, phenolics, photosynthesis, red and far-red fluorescence, *Triticosecale*, winter triticale.

INTRODUCTION

In many regions water resources for agronomic uses are becoming scarce (Wardlaw, 2002; Llorens *et al.*, 2004; Flexas *et al.*, 2006); therefore, the development of droughttolerant lines is becoming increasingly more important. Field trials concerning natural drought stress are timeconsuming and expensive; furthermore, both space and the seed of genotypes, especially in early generations, are limited, so only a restricted number of genotypes can be tested in any one year. Another problem is that natural drought stress does not occur to the same extent every year, but for the selection process it is advantageous to be able to make selections within a few years under similar field conditions.

Nowadays, efforts are directed to finding new, cheap and physiologically trustworthy tools that can help in both the detection of drought stress and the selection of tolerant genotypes. Studying physiological and biochemical responses to water deficit may help in breeding plant cultivars of high yield and stability under drought conditions (Yordanov *et al.*, 2000).

During the acclimation to soil drought, plants initiate defence mechanisms against water deficit (Medrano *et al.*, 2002; Chaves and Oliveira, 2004). In the present work, fluorescence and spectrofluorescence methods were used to screen plants under drought-stress conditions. Measurement of chlorophyll fluorescence provides information about the function of the photosynthetic apparatus and is useful for stress detection (Bolhàr-Nordenkampf and Öquist, 1993; Schweiger *et al.*, 1996; Yordanov *et al.*, 1997).

It has been shown that water deficit led to an increase in the intensity of the blue fluorescence originating from plant phenolics, mainly from ferulic acid (Lichtenthaler and Schweiger, 1998; Cerovic *et al.*, 2002; Meyer *et al.*, 2003; Morales *et al.*, 2005; Hura *et al.*, 2006). Phenolics can neutralize light falling on the leaves through its transformation into blue fluorescence, which is no longer damaging, and can even be used for photosynthetic quantum

* For correspondence. E-mail t.hura@ifr-pan.krakow.pl

© The Author 2007. Published by Oxford University Press on behalf of the Annals of Botany Company. All rights reserved. For Permissions, please email: journals.permissions@oxfordjournals.org conversion (Caldwell *et al.*, 1983; Schweiger *et al.*, 1996; Buschmann and Lichtenthaler, 1998; Bilger *et al.*, 2001). The phenolic compounds are very reactive structures due to the presence of the benzene ring. The benzene ring as a π -electron system can counteract UV and visible radiation (Bilger *et al.*, 1997, 2001). It is known that drought stress predisposes plants to injury of the photosynthetic apparatus through its interaction with UV or/and visible radiation (Garcia-Plazaola and Becerril, 2000). In the genotypes studied here, the contents of ferulic acid, as the main origin of the blue fluorescence, and total phenolics were measured, and the activity of PAL (L-phenylalanine ammonia-lyase), as the key enzyme in the synthesis of phenolics (Strack, 1997), was also analysed.

In the work presented here, attempts were made to estimate whether the division of triticale cultivars into droughttolerant and drought-sensitive genotypes based on field trials agrees with the division obtained from measurements of some physiological and biochemical parameters in greenhouse conditions, and whether ferulic acid as the main origin of the blue fluorescence may contribute to drought tolerance.

MATERIALS AND METHODS

Plant materials

The experiments were carried out on winter triticale $(\times Triticosecale, Wittmack)$ genotypes. Based on field trials, five genotypes were classified as drought sensitive ('Kitaro', 'Focus', 'Lamberto', 'Timbo' and 'Piano') and five as drought tolerant ('Babor', 'Boreas', 'Trimaran', 'Ticino' and 'Imperial'). Plants were grown during two successive cultivation seasons, 2002-2003. During 2002, field trials were performed at two locations with natural drought stress, in northern Germany and in the middle-east of Germany, whilst in 2003 they were performed at three locations, two in northern and one in southern Germany. Two water regimes were imposed (irrigated vs. non irrigated) with two replications each. Traits such as heading, height, spikes per m² and grain yield were measured (results not shown). The basis for the selection of the drought-susceptible and drought-tolerant genotypes was the results obtained in 2003, a year of severe drought; however, the results of both the two seasonal experiments were consistent with regard to the grouping of triticale genotypes into tolerant and susceptible. For the experiment in greenhouse conditions, five genotypes with the highest/ lowest grain yield in non-irrigated conditions were selected.

Plant growth conditions

Seedlings of each genotype were grown in vermiculite medium to approximately the first leaf stage in a greenhouse before being vernalized for 8 weeks in controlled conditions [maintained at 4 °C with 10 h illumination and with a photosynthetic photon flux density (PPFD) of 200 μ mol m⁻² s⁻¹]. After vernalization, seedlings of each genotype at the 3-leaf stage were planted in plastic pots (two seedlings per pot) of 15 cm diameter and 18 cm

height containing vermiculite as the medium, and were exposed in an air-conditioned greenhouse chamber to a 16 h light/8 h dark photoperiod, a temperature of 23/18 °C (± 2 °C) day/night, 60 ± 5 % relative air humidity and a PPFD of 350 µmol m⁻² s⁻¹.

For control plants 70% of FWC (field water capacity) and for water-stressed plants 30–35% of FWC were applied for 4 weeks. Drought treatment of triticale genotypes took place during heading and flowering. The plants were irrigated with full-strength Hoagland's nutrient solution once per week. The pots were weighed and the amount of the water lost was replaced to maintain the original weight of the pots in each treatment. Each treatment (control and drought) comprised 15 pots. In total, there were 30 samples within each treatment for each genotype. All the pots in each treatment, before measurements, were randomly divided into three subgroups, which were randomly assigned to measurements of physiological and biochemical parameters. The measurements for all genotypes were carried out on flag leaves during flowering.

Osmotic potential

Measurements were carried out with a dew point microvoltmeter (model HR-33T with C-52 sample chambers, Wescor Inc., Logan, UT, USA). Leaf discs (diameter = 5 mm) were taken for analysis from the middle part of leaves and were placed into an Eppendorf tube, frozen in liquid nitrogen and stored at -70 °C. For measurement, leaf samples were thawed at room temperature and the sap from leaf discs was extracted with a syringe and quickly transferred to a leaf chamber. The time needed for the saturation of leaf chambers was set to 40 min. The measurements for each genotype were done in the dew point mode at room temperature on five replicates.

Leaf gas exchange

Leaf gas exchange was measured on the fully expanded leaf using an infrared gas analyser (Li-6400, Portable Photosynthesis System, Lincoln, NE, USA), operated within an open system with a leaf chamber. Net photosynthesis, transpiration rated and stomatal conductance were determined under photosynthetically active radiation (PAR) of 1000 μ mol of photons m⁻² s⁻¹ and at a CO₂ concentration ranging from 380 to 400 ppm. The measurements for each genotype were done on seven replicates.

Chlorophyll fluorescence

Chlorophyll fluorescence was measured on the fully expanded leaf usng a pulse-modulated portable fluorometer FMS2 (Hansatech Instruments, King's Lynn, UK) at ambient temperature in the dark (dark-adapted leaf) and then, after 10 min of irradiation at a PPFD of 500 µmol of photons $m^{-2} s^{-1}$ (light-adapted leaf), QP (photochemical quenching coefficient) and NQP (non-photochemical quenching coefficient) were calculated according to Havaux *et al.* (1991): QP = $(F_m' - F_s)/(F_m' - F_o')$; NQP = $(F_m' - F_m)/(F_m - F_o')$.

The quantum efficiency of photosystem II (PSII) was calculated according to Genty *et al.* (1989): $\Phi_{PSII} = (F_{m}' - F_{s})/F_{m}'$. The measurements for each genotype were done on five replicates.

Spectrofluorescence

Fluorescence spectra were measured using a spectrofluorometer (Perkin-Elmer LS 50B, Norwalk, CT, USA). Fluorescence emission spectra of the red and far-red fluorescence were recorded between 650 and 800 nm. The leaves were excited at 450 nm. The spectral slit widths were set at 10 nm (excitation and emission). The fluorescence emission spectra of the blue fluorescence were recorded between 380 and 600 nm. The leaves were excited at 350 nm. The slit widths for excitation and emission monochromators were adjusted to 10 nm. The measurements of the blue, red and far-red fluorescence for each genotype were done on five replicates.

L-Phenylalanine ammonia-lyase activity

Measurement of PAL activity was carried according to modified methods described by Peltonen and Karjalainen (1995). All procedures were carried out at +4 °C. The reaction mixture contained 2.5 mL of 0.2 % solution of L-phenylalanine in 50 mM Tris–HCl (pH 8.5) and 0.5 mL of supernatant. The incubation of the reaction mixture was set at 24 h at 38 °C. Then the absorbance at 290 nm was measured. Enzyme activity was expressed as ng of cinnamic acid produced during 1 min per 1 mg of protein. In enzyme assays, protein contents were determined by the method of Bradford (1976). The measurements for each genotype were done on five replicates.

Phenolic analysis

For both total phenolics and ferulic acid measurements, the lyophilized material was homogenized in 80% ethanol. The total concentration of phenolics was determined using the Folin–Ciocalteau method of Singleton and Rossi (1965). The absorbance was measured at 760 nm. Chlorogenic acid was used as a standard.

Ferulic acid contents were analysed with a spectrofluorometer (Perkin-Elmer LS 50B, Norwalk, CT, USA). Before measurements, chlorophyll was removed by several extractions with hexane until no green colour was visible. The samples were excited at 243 nm and the detection was carried out at 434 nm. The slit widths for both excitation and emission monochromators were adjusted to 10 nm. The measurements of both total phenolics and ferulic acid concentrations for each genotype were done on five replicates.

Statistical analysis

Statistica 5.0 software for Windows was used. Data presented in the figures are means of five or seven replications \pm s.e. Correlations between measured parameters were tested at a probability of P < 0.05, and only statistically significant correlations are presented in the figures.

RESULTS AND DISCUSSION

In all tested genotypes a decrease in the osmotic potential (Ψ_o) during drought stress was observed (Fig. 1). The highest values of Ψ_o were detected in cultivars 'Timbo', 'Lamberto', 'Piano' and 'Babor', and were closer to those of the control plants. For the sensitive cultivar 'Babor', Ψ_o tended to be close to the values for the control, which suggests the existence of efficient osmoregulation.

Shangguan *et al.* (1999) found that the photosynthesis process during drought is possible due to the osmoregulation which affects the state of the leaf stomata and adaptation of the photosynthetic apparatus to drought conditions. Similar results had been obtained earlier in other studies (Matthews and Boyer, 1984; Sen Gupta and Berkowitz, 1988; Verslues and Bray, 2004). Ludlow (1987) and Weng (1993) report a positive correlation between photosynthesis and osmoregulation.

For the tested genotypes, a positive correlation between the photosynthesis rate and Ψ_{o} was found (Fig. 2A). In drought conditions, the highest photosynthesis rates were detected for cultivars 'Piano', 'Timbo' and 'Lamberto'. A significant correlation was also indicated between the transpiration rate and the osmotic potential (Fig. 2B). The transpiration rate tended to be highest in cultivars 'Piano', 'Timbo' and 'Lamberto'. The highest values of stomatal conductance, closest to the control value, were found for the drought-sensitive genotypes 'Babor' and 'Boreas' (Table 1). This points to severe disturbances in stomatal movement and lack of complete closure in drought conditions. For cultivars tolerant to water deficit, such as 'Timbo' and 'Piano', the stomatal conductance was high or close to that of the control, respectively. The experiments conducted in long-term drought conditions did not reveal any significant correlation between the stomatal conductance and the photosynthesis rate. This may suggest factors other than stomata-based factors inhibiting the photosynthesis process, most probably linked to the damage of the photosynthetic apparatus (Hura et al., 2006).

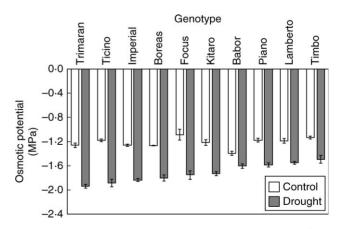


FIG. 1. Effect of drought treatment on the leaf osmotic potential (Ψ_0) of winter triticale genotypes. Data are means \pm s.e. of five replicates.

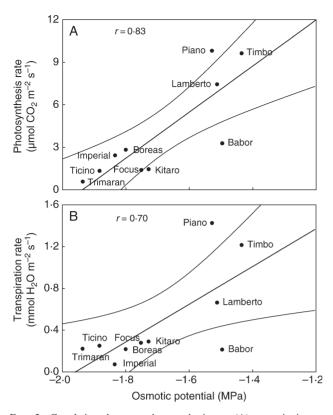


FIG. 2. Correlations between photosynthesis rate (A), transpiration rate (B) and osmotic potential (Ψ_{o}) for drought-treated genotypes of winter triticale.

Soil drought did not influence the photochemical efficiency of PSII (F_v/F_m) measured in dark-adapted leaves (Fig. 3). A distinct decrease in this parameter was observed only for the cultivar 'Imperial'. There are examples of studies showing resistance of PSII to water deficit in leaf tissues (Cornic and Briantais, 1991; Liang *et al.*, 1997; Noguès and Baker, 2000; Cornic and Fresneau, 2002). According to the authors, measurements of F_v/F_m do not provide enough information about PSII in drought conditions because a decrease in the activity of PSII may be a result of photoinhibition.

TABLE 1. Effect of drought stress on the stomatal conductance (mmol $CO_2 m^{-2} s^{-1}$) of winter triticale genotypes

Genotypes	Control	Drought
'Kitaro'	65.9 ± 1.0	54.3 ± 1.8
'Imperial'	194.2 ± 10.8	60.1 ± 3.7
'Focus'	74.7 ± 5.1	61.9 ± 5.2
'Ticino'	77.4 ± 4.1	64.2 ± 6.4
'Lamberto'	183.0 ± 4.7	67.9 ± 6.5
'Trimaran'	215.3 ± 5.9	69.3 ± 6.1
'Timbo'	190.4 ± 6.1	140.7 ± 1.6
'Piano'	197.3 ± 4.0	177.0 ± 5.6
'Boreas'	297.0 ± 7.1	273.3 ± 3.2
'Babor'	$302 \cdot 3 \pm 3 \cdot 8$	282.7 ± 1.9

Data are means \pm s.e. of seven replicates.

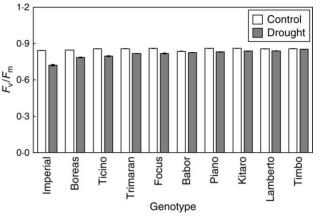


FIG. 3. Effect of drought treatment on the maximal efficiency of PSII photochemistry (F_v/F_m) of winter triticale genotypes. Data are means \pm s.e. of five replicates.

Statistically significant correlations were found between some parameters related to chlorophyll fluorescence $[\Phi_{PSII}, QP, ETR (electron transport rate), NQP and <math>F_v/F_m$] and the Ψ_o (Fig. 4). Leaf dehydration was accompanied by a decrease in actual fluorescence F_v/F_m for the sensitive genotypes 'Imperial', 'Ticino', 'Trimaran' and 'Boreas' (Fig. 4A). A decrease in F_v/F_m may result from low efficiency of the capture of excitation energy by PSII centres and low efficiency of light trapping by the PSII lightharvesting antenna due to photoinhibition. For tolerant cultivars, photoinhibition can be regarded as an adaptation to drought conditions, whereas in the case of sensitive cultivars such as 'Imperial', 'Ticino', 'Trimaran' and 'Boreas', it can be regarded as damage to PSII because photoinhibition itself can lead to the degradation of the D₁ protein (Barber and Andersson, 1991; Baker and Rosenqvist, 2004).

For the genotypes 'Piano', 'Timbo' and 'Lamberto', the highest values of QP under drought were recorded (Fig. 4B). The relatively high values of QP can be explained by the increased accumulation of non-reduced primary electron acceptors (Q_A) of PSII, which are ready to accept excitation energy in order to pass it further towards other photochemical processes (Maxwell and Johnson, 2000; Noguès and Baker, 2000).

The coefficient of NQP measures energy emitted as heat (Baker and Rosenqvist, 2004), and high emission points to the damaged photosynthetic apparatus (Loggini *et al.*, 1999). The drought-resistant cultivars 'Timbo' and 'Piano' and drought-sensitive 'Babor' showed the lowest values of this parameter (Fig. 4C).

The quantum efficiency of the PSII parameter (Φ_{PSII}) allows determination of the effectiveness of excitation energy utilization by chlorophyll *a* in the process of photosynthesis (Maxwell and Johnson, 2000; Noguès and Baker, 2000). On the basis of the measurements made, it is possible to distinguish genotypes with the lowest values of Φ_{PSII} , i.e. 'Imperial', 'Ticino', 'Trimaran', and the group with the highest values of this parameter, i.e. 'Lamberto', 'Timbo' and 'Piano' (Fig. 4D). The most important factor influencing the values of Φ_{PSII} could be different contributions of the open reaction centres of PSII to the

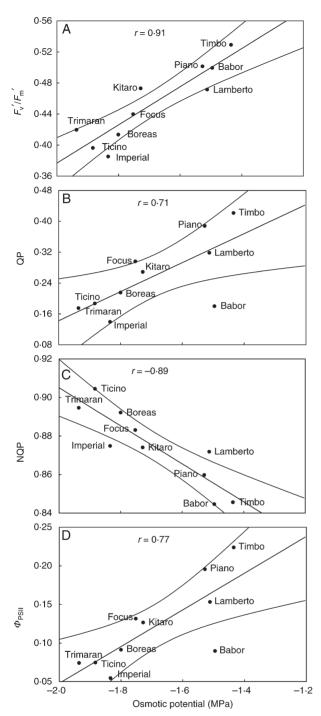


FIG. 4. Correlations between (A) actual fluorescence (F_v / IF_m') , (B) photochemical quenching coefficient (QP), (C) non-photochemical quenching coefficient (NQP), (D) quantum efficiency of PSII (Φ_{PSII}) and osmotic potential (Ψ_o) for drought-treated genotypes of winter triticale.

photochemical processes for different cultivars of winter triticale (Demmig-Adams and Adams, 1992; Loggini *et al.*, 1999).

Figure 5 shows changes in the ETR, which reflects the activity of CO_2 assimilation. The increase in the value of this parameter, which was noticed for the cultivars

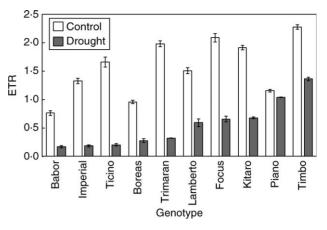


FIG. 5. Effect of drought treatment on the electron transport rate (ETR) of winter triticale genotypes. Data are means \pm s.e. of five replicates.

'Timbo' and 'Piano', reflects the best utilization, among the studied genotypes, of the products of the light phase of photosynthesis in the light-independent phase of photosynthesis (Medrano *et al.*, 2002).

Statistically significant correlations between the rate of photosynthesis and the chlorophyll fluorescence parameters $(F_v/F_m, QP, NQP, \Phi_{PSII} \text{ and ETR})$ were found (Fig. 6A–E). This result illustrates the fact that in drought conditions the photosynthetic apparatus of the genotypes 'Piano', 'Timbo' and 'Lamberto' were the most efficient in excitation-energy capture by PSII centres followed by the delivery of this energy onwards to the next photochemical reactions in the photosynthesis process, with the lowest loss of the energy as heat. Moreover, for tolerant cultivars, the products of the light phase were utilized more efficiently during the light-independent phase of photosynthesis.

Figure 7 shows significant correlations between Ψ_o and the intensity of the red fluorescence (Fig. 7A) and far-red fluorescence (Fig. 7B). The photosynthetic apparatus is capable of dissipation of the excess energy as heat, but is also able to neutralize high-energy states through the emission of red fluorescence (Schweiger *et al.*, 1996; Hura *at al.*, 2006). This phenomenon may be part of a defence mechanism, but it may also result from damage to the photosynthetic apparatus in the cultivars 'Ticino', 'Trimaran' and 'Imperial'.

Significant correlations have been found between NQP and the intensity of the red fluorescence (Fig. 8A) and far-red fluorescence (Fig. 8B). The results point to the fact that in drought conditions the cultivars 'Piano', 'Timbo' and 'Lamberto' utilized the excitation energy in the optimal way in comparison with sensitive cultivars. The cultivar 'Babor', which was drought sensitive, emitted fluorescence of the lowest intensity in both ranges. The increase in the red fluorescence emission in drought conditions, observed especially for sensitive cultivars, occurs at the cost of the photosynthetic conversion of absorbed light and is related to damage of PSII and lightharvesting complexes and changes in the activity of electron transport molecules (Lichtenthaler, 1996; Šesták and Šiffel, 1997; Buschmann and Lichtenthaler, 1998).

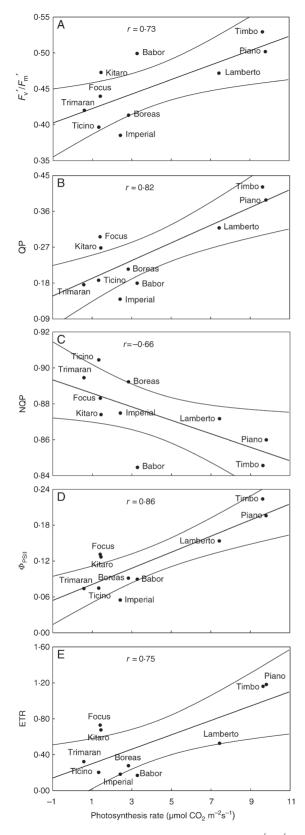


FIG. 6. Correlations between (A) actual fluorescence (F_v'/F_m') , (B) photochemical quenching coefficient (QP), (C) non-photochemical quenching coefficient (NQP), (D) quantum efficiency of PSII (Φ_{PSII}), (E) electron transport rate (ETR) and photosynthesis rate for drought-treated genotypes of winter triticale.

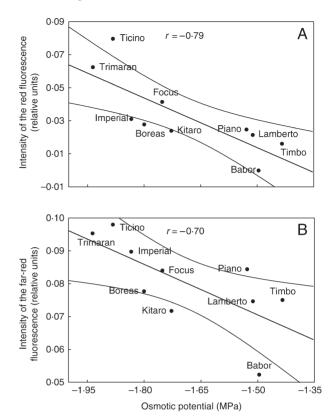


FIG. 7. Correlations between (A) emission of red fluorescence, (B) emission of far-red fluorescence and osmotic potential (Ψ_o) for drought-treated genotypes of winter triticale.

An increase in the emission of blue fluorescence is related to the accumulation of phenolic compounds in the leaf (Schweiger et al., 1996; Bilger et al., 2001). For the sensitive cultivars, the increased emission of blue fluorescence correlated with low $\Psi_{\rm o}$, while the tolerant genotypes showed low emission of blue fluorescence with high Ψ_{0} (Fig. 9). Water deficit can induce protective mechanisms against the destructive action of radiation, which could be absorbed and transformed by phenolic compounds into blue fluorescence that, in turn, could be utilized in the photosynthesis process as well as re-emitted as chlorophyll fluorescence (Caldwell et al., 1983; Schweiger et al., 1996; Bilger et al., 1997, 2001; Buschmann and Lichtenthaler, 1998; Lichtenthaler and Schweiger, 1998). Therefore, plants have the possibility to widen the site of the action of radiation on the photosynthetic apparatus in unfavourable conditions where light may be a stress factor. A positive correlation between the ferulic acid content and Ψ_{o} was noted (Fig. 10A). A higher ferulic acid content was determined for drought-resistant cultivars than for sensitive cultivars, although the highest content of ferulic acid was detected in the cultivar 'Babor', which is sensitive to drought. Ferulic acid, the main source of the blue fluorescence (Lichtenthaler and Schweiger, 1998), can absorb UV and visible radiation (violet and blue light). During the experiments, a statistically significant negative correlation between the intensity of the blue fluorescence and the content of ferulic acid was noted (Fig. 10B), although it was expected that a positive correlation would be found

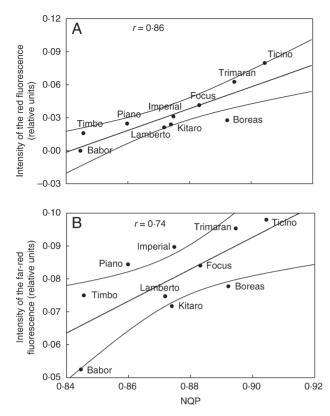


FIG. 8. Correlations between (A) emission of red fluorescence, (B) emission of far-red fluorescence and non-photochemical quenching coefficient (NQP) for drought-treated genotypes of winter triticale.

showing that an increase in the ferulic acid content brings about an increase in the emission of blue fluorescence. The negative correlation could be the result of the re-absorption of the emitted fluorescence by phenolic compounds (Lichtenthaler *et al.*, 1998). Moreover, emission of blue fluorescence can be reduced by partial re-absorption of such fluorescence by chlorophylls and carotenoids possessing a blue absorption band (Lang *et al.*, 1996).

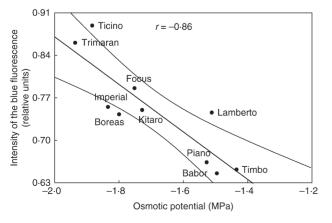


FIG. 9. Correlation between emission of blue fluorescence and osmotic potential of drought-treated genotypes of winter triticale.

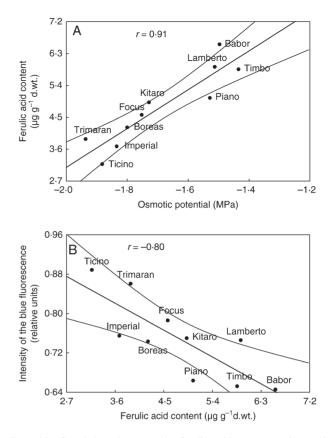


FIG. 10. Correlations between (A) ferulic acid content and osmotic potential and (B) emission of blue fluorescence and ferulic acid content for drought-treated genotypes of winter triticale.

Table 2 shows changes in the total pool of phenolic compounds. An increase in the content of phenolics in drought conditions for the sensitive cultivars 'Trimaran' and 'Ticino' was noticed. The total pool of phenolic compounds did not increase, or increased by only a small amount, in the tolerant cultivars (for 'Kitaro' even a small decrease in the total pool of phenolic compounds was observed) at the expense of the synthesis of ferulic acid as an efficient photoprotector.

TABLE 2. Effect of drought stress on the content of total phenolics (mg g^{-1} d. wt) of winter triticale genotypes

Genotypes	Control	Drought
'Boreas'	37.3 + 1.4	27.9 + 0.5
'Imperial'	45.9 ± 1.2	36.6 ± 1.1
'Kitaro'	41.7 ± 0.6	37.7 ± 1.0
'Trimaran'	22.3 ± 1.0	40.7 ± 0.7
'Babor'	32.7 ± 0.7	41.0 ± 1.2
'Lamberto'	40.7 ± 0.9	41.0 ± 1.1
'Piano'	39.8 ± 1.2	41.9 ± 0.6
'Ticino'	30.0 ± 0.8	45.0 ± 0.5
'Timbo'	39.5 ± 1.0	48.8 ± 1.7
'Focus'	$46 \cdot 1 + 1 \cdot 1$	54.2 + 0.8

Data are means \pm s.e. of five replicates.

TABLE 3. Effect of drought stress on PAL activity (ng cinnamic acid $min^{-1}mg^{-1}$ protein) of winter triticale genotypes

Genotypes	Control	Drought
'Timbo'	2.50 + 0.13	1.69 ± 0.14
'Kitaro'	2.92 ± 0.17	2.60 ± 0.15
'Imperial'	2.31 + 0.13	2.70 + 0.27
'Babor'	1.99 ± 0.19	2.75 ± 0.25
'Trimaran'	1.99 ± 0.17	2.84 ± 0.18
'Focus'	2.58 ± 0.18	2.91 ± 0.18
'Ticino'	2.01 ± 0.11	2.96 ± 0.22
'Lamberto'	2.06 ± 0.06	2.99 ± 0.16
'Boreas'	2.27 ± 0.14	3.17 ± 0.19
'Piano'	2.56 ± 0.15	3.85 ± 0.22

Data are means \pm s.e. of five replicates.

An increase in the activity L-PAL, crucial in the synthesis of phenolic compounds, was observed in all cultivars except for 'Timbo' and 'Kitaro' (Table 3).

CONCLUSIONS

The results of the physiological and biochemical experiments are in good agreement with the classification into drought-tolerant and drought-sensitive genotypes based on field experiments under natural drought stress. The cultivars 'Lamberto', 'Timbo' and 'Piano' belong to the droughttolerant group. For the cultivar 'Babor', recognized as sensitive, the results of the analyses, especially the measurements of the osmotic potential and chlorophyll fluorescence, were close to and in some cases even better than those of the tolerant cultivars. The lack of agreement with the field trials in this case could be due to the effect of stress conditions other than drought working simultaneously on the plant. In field conditions, it is impossible to eliminate the effects of other negative environmental factors, and the indicator of tolerance examined, for example the crop yield, is a resultant indicator of the influence of many stress factors during the growth and development of plants. The cultivars 'Imperial', 'Ticino', 'Trimaran' and 'Boreas' can definitely be classified as drought sensitive on the base of the measurements, while the cultivars 'Focus' and 'Kitaro' belong to the group of moderately drought-tolerant plants. The experiments were carried out during the flowering phase, which is critical for plants as far as water supply is concerned. This phase seems to be suitable for screening of plants tolerant to drought stress. The most important criteria that enable creation of a ranking list of plants, from sensitive to tolerant to drought, are the ability to carry out osmoregulation (measurements of the osmotic potential) and the effective and optimal utilization of excitation energy by the photosynthetic apparatus in drought conditions. Other important factors in drought tolerance are the protective mechanisms involving, among others, phenolic compounds, especially ferulic acid as one of the most effective photoprotectors whose level can be a reliable indicator of drought tolerance. However, determination of ferulic acid levels and PAL enzyme activities should have been monitored at more

than one time point during the stress period to obtain a better understanding of how tolerant and sensitive varieties differ in their response.

Studies carried out in the laboratory based on measurements of the osmotic potential, chlorophyll fluorescence and ferulic acid content enable a greater number of genotypes to be screened in a shorter period of time and may thus become a more economically efficient alternative to long-lasting (several years) and expensive field trials.

LITERATURE CITED

- **Baker NR, Rosenqvist E. 2004.** Applications of chlorophyll fluorescence can improve crop production strategies: an examination of future possibilities. *Journal of Experimental Botany* **55**: 1607–1621.
- Barber J, Andersson B. 1991. Light can be both good and bad for photosynthesis. *Trends in Biochemical Sciences* 17: 61–66.
- Bilger W, Johnsen T, Schreiber U. 2001. UV-excited chlorophyll fluorescence as a tool for the assessment of UV-protection by the epidermis of plants. *Journal of Experimental Botany* 52: 2007–2014.
- Bilger W, Veit M, Schreiber L, Schreiber U. 1997. Measurement of leaf epidermal transmittance of UV radiation by chlorophyll fluorescence. *Physiologia Plantarum* 101: 754–763.
- Bolhàr-Nordenkampf HR, Öquist G. 1993. Chlorophyll fluorescence as a tool in photosynthesis research. In: Hall DO, Scurlock JMO, Bolhàr-Nordenkampf HR, Leegood RC, Long SP, eds. *Photosynthesis and production in a changing environment: a field and laboratory manual*. London: Chapman & Hall, 193–206.
- **Bradford JS. 1976.** A rapid and sensitive method for the quantitation of microgram quantitaties of protein utilizing the principle of protein–dye binding. *Analytical Biochemistry* **72**: 248–254.
- Buschmann C, Lichtenthaler HK. 1998. Principles and characteristics of multi-colour fluorescence imaging of plants. *Journal of Plant Physiology* 152: 297–314.
- Caldwell MM, Robberecht R, Flint SD. 1983. Internal filters: prospects for UV-acclimation in higher plants. *Physiologia Plantarum* 58: 445–450.
- Cerovic ZG, Ounis A, Cartelat A, Latouche G, Goulas Y, Meyer S, Moya I. 2002. The use of chlorophyll fluorescence excitation spectra for the non-destructive *in situ* assessment of UV-absorbing compounds in leaves. *Plant, Cell and Environment* 25: 1663–1676.
- Chaves MM, Oliveira MM. 2004. Mechanisms underlying plant resilience to water deficits: prospects for water-saving agriculture. *Journal of Experimental Botany* 55: 2365–2384.
- **Cornic G, Briantais JM. 1991.** Portioning of photosynthetic electron flow between CO₂ and O₂ reduction in a C₃ leaf (*Phaseolus vulgaris* L.) at different CO₂ concentrations and during drought stress. *Planta* **183**: 178–184.
- Cornic G, Fresneau Ch. 2002. Photosynthetic carbon reduction and carbon oxidation cycles are the main electron sinks for photosystem II activity during a mild drought. *Annals of Botany* 89: 887–894.
- Demmig-Adams B, Adams WW III. 1992. Photoprotection and other responses of plants to high light stress. Annual Review of Plant Physiology and Plant Molecular Biology 43: 599–626.
- Flexas J, Bota J, Galmés J, Medrano H, Ribas-Carbó M. 2006. Keeping a positive carbon balance under adverse conditions: responses of photosynthesis and respiration to water stress. *Physiologia Plantarum* 127: 343–352.
- Garcia-Plazaola JI, Becerril JM. 2000. Effects of drought on photoprotective mechanisms in European beech (*Fagus sylvatica* L.) seedlings from different provenances. *Trees* 14: 485–490.
- Genty B, Briantais JM, Adams WW. 1989. The relationship between the quantum yield of photosynthetic electron transport and quenching of chlorophyll fluorescence. *Biochimica et Biophysica Acta* 990: 87–92.
- Havaux M, Strasser RJ, Greppin H. 1991. A theoretical and experimental analysis of the q_p and q_{np} coefficients of chlorophyll fluorescence quenching and their relation to photochemical and nonphotochemical events. *Photosynthesis Research* 27: 41–55.
- Hura T, Grzesiak S, Hura K, Grzesiak MT, Rzepka A. 2006. Differences in the physiological state between triticale and maize

plants during drought stress and followed rehydration expressed by the leaf gas exchange and spectrofluorimetric methods. *Acta Physiologiae Plantarum* **28**: 433–443.

- Lang M, Lichtenthaler HK, Sowinska M, Heisel F, Miehé JA. 1996. Fluorescence imaging of water and temperature stress in plant leaves. *Journal of Plant Physiology* 148: 613–621.
- Liang J, Zhang J, Wong M. 1997. Can stomatal closure caused by xylem ABA explain the inhibition of leaf photosynthesis under soil drying? *Photosynthesis Research* 51: 149–159.
- Lichtenthaler HK. 1996. Vegetation stress: an introduction to the stress concept in plants. *Journal of Plant Physiology* 148: 4–14.
- Lichtenthaler HK, Schweiger J. 1998. Cell wall bound ferulic acid, the major substance of the blue-green fluorescence emission of plants. *Journal of Plant Physiology* 152: 272–282.
- Lichtenthaler HK, Wenzel O, Buschmann C, Gitelson A. 1998. Plant stress detection by reflectance and fluorescence. Annals of the New York Academy of Sciences 851: 271.
- Llorens L, Peñuelas J, Estiarte M, Bruna P. 2004. Contrasting growth changes in two dominant species of a Mediterranean shrubland submitted to experimental drought and warming. *Annals of Botany* 94: 843–853.
- Loggini B, Scartazza A, Brugnoli E, Navari-Izzo F. 1999. Antioxidative defense system, pigment composition, and photosynthetic efficiency in two wheat cultivars subjected to drought. *Plant Physiology* 119: 1091–1100.
- Ludlow MM. 1987. Contribution of osmotic adjustment to the maintenance of photosynthesis during water stress. *Progress in Photosynthesis Research* 4: 161–168.
- Matthews MA, Boyer JS. 1984. Acclimation of photosynthesis to low water potentials. *Plant Physiology* 74: 161–166.
- Maxwell K, Johnson GN. 2000. Chlorophyll fluorescence a practical guide. *Journal of Experimental Botany* **51**: 659–668.
- Medrano H, Escalona JM, Bota J, Gulías J, Flexas J. 2002. Regulation of photosynthesis of C₃ plants in response to progressive drought: stomatal conductance as a reference parameter. *Annals of Botany* 89: 895–905.
- Meyer S, Cartelat A, Moya I, Cerovic ZG. 2003. UV-induced blue-green and far-red fluorescence along wheat leaves: a potential signature of leaf ageing. *Journal of Experimental Botany* 54: 757–769.
- Morales F, Cartelat A, Álvarez-Fernández A, Moya I, Cerovic ZG. 2005. Time-resolved spectral studies of blue-green fluorescence of

artichoke (*Cynara cardunculus* L. Var. *Scolymus*) leaves: identification of chlorogenic acid as one of the major fluorophores and agemediated changes. *Journal of Agricultural and Food Chemistry* **53**: 9668–9678.

- Nogués S, Baker NR. 2000. Effects of drought on photosynthesis in Mediterranean plants grown under enhanced UV-B radiation. *Journal of Experimental Botany* 51: 1309–1317.
- Peltonen S, Karjalainen R. 1995. Phenylalanine ammonia-lyase activity in barley after infection with *Bipolaris sorokiniana* or treatment with its purified xylanase. *Journal of Phytopathology* 143: 239–245.
- Schweiger J, Lang M, Lichtenthaler HK. 1996. Differences in fluorescence excitation spectra of leaves between stressed and non-stressed plants. *Journal of Plant Physiology* 148: 536–547.
- Sen Gupta A, Berkowitz GA. 1988. Chloroplast osmotic adjustment and water stress effects on photosynthesis. *Plant Physiology* 88: 200–206.
- Šesták Z, Šiffel P. 1997. Leaf-age related differences in chlorophyll fluorescence. *Photosynthetica* 33: 347–369.
- Shangguan Z, Shao M, Dyckmans J. 1999. Interaction of osmotic adjustment and photosynthesis in winter wheat under soil drought. *Journal* of Plant Physiology 154: 753–758.
- Singleton VS, Rossi JA Jr. 1965. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagent. American Journal of Enology and Viticulture 16: 144–157.
- Strack D. 1997. Phenolic metabolism. In: Dey PM, Harborne JB, eds. Plant biochemistry. London: Academic Press, 387–416.
- Verslues PE, Bray EA. 2004. LWR1 and LWR2 are required for osmoregulation and osmotic adjustment in Arabidopsis. Plant Physiology 136: 2831–2842.
- Wardlaw IF. 2002. Interaction between drought and chronic high temperature during kernel filling in wheat in a controlled environment. *Annals* of Botany 90: 469–476.
- Weng JH. 1993. Photosynthesis of different ecotypes of *Miscantus* spp. as affected by water stress. *Photosynthetica* 29: 43–48.
- Yordanov I, Tsonev T, Goltsev V, Kruleva L, Velikova V. 1997. Interactive effect of water deficit and high temperature on photosynthesis of sunflower and maize plants. 1. Changes in parameters of chlorophyll fluorescence induction kinetics and fluorescence quenching. *Photosynthetica* 33: 391–402.
- Yordanov I, Velikova V, Tsonev T. 2000. Plant responses to drought, acclimation, and stress tolerance. *Photosynthetica* 38: 171–186.